

REPORT

of

TEST NO. 862-1 & 3

Toxicity of Freshwater Sediments Using Sediment Bioassays as Part of the Remedial Investigation at the Jacobson Terminals in Seattle, Washington

Submitted to

**Hart Crowser, Inc.
1700 Westlake Ave. North, Suite 200
Seattle, WA 98109**

Submitted by

**Northwestern Aquatic Sciences
3814 Yaquina Bay Road
P.O. Box 1437
Newport, OR 97365**

March 19, 2015

EXECUTIVE SUMMARY OF SEDIMENT BIOASSAYS

Two freshwater sediment bioassays, a 28-day *Hyaella* amphipod survival and growth test and a 10-day *Chironomus dilutus* midge survival and growth test were conducted for Hart Crowser as part of the Remedial Investigation at the Jacobson Terminals property in Seattle, WA. Three test sediments were compared to the control sediment to assess sediment toxicity and to interpret organism response under the Washington State *Sediment Management Standards* (Chapter 173-204 WAC, Last Update: 2/25/13). The test and control sediments tested are listed in Table 1.

TEST AND CONTROL SEDIMENT INFORMATION

The test sediments were provided to Northwestern Aquatic Sciences by Hart Crowser personnel. The negative control sediment was collected by NAS personnel from an area approximately one mile east of the Highway 101 bridge at Beaver Creek, approximately 8 miles south of Newport, Oregon. All sediments were stored at 4°C in the dark until used. Sample identification and collection information is as follows:

Sample description	Hart Crowser Sample Identification	NAS Sample Identification	Collection Date	Receipt Date
Beaver Creek Control*	Control	5195G	1-19-15	1-19-15
Test sediment	JT-SS-06	5192G	1-12-15	1-14-15
Test sediment	JT-SS-08	5193G	1-12-15	1-14-15
Test sediment	JT-SS-10	5194G	1-12-15	1-14-15

*Control used for data interpretation.

BIOASSAY INTERPRETATION CRITERIA

Biological test interpretation for freshwater sediments, as presented in the *Sediment Management Standards*, (SMS) uses biological criteria to identify sediments that have no adverse effects on biological resources, and correspond to no significant health risk to humans. The SMS includes Sediment Cleanup Objectives (SCO) and Cleanup Screening Levels (CSL) biological criteria. The Sediment Cleanup Objectives establish a no adverse effects level, including no acute or chronic adverse effect, to the benthic community. The Cleanup Screening Levels establish a minor adverse effects level, including acute or chronic effects, to the benthic community. The SCO biological criteria are exceeded when one of the biological tests results is above the SCO for that bioassay endpoint. The CSL biological criteria is exceeded when 1) any two bioassay test results are above the SCO criteria; or 2) when one of the bioassay test results is above the CSL criteria.

Sediment Cleanup Objectives Criteria

When any one of the biological test results shows a test sediment response that exceeds the bioassay-specific response guidelines presented below, and that response is statistically different ($p \leq 0.05$) from the control, the test sediment is judged to have exceeded the SCO.

In accordance with the SMS, the bioassay-specific response guidelines for evaluating an exceedance under the SCO criteria are as follows:

Amphipod 28-day Survival/Growth Bioassay. Mean mortality in the test sediment is greater than 10 percent over the mean control and statistically different from the control ($p \leq 0.05$). For the growth endpoint, a mean reduction in the biomass that is greater than 25 percent of the control sediment response and statistically different from the control ($p \leq 0.05$).

Midge 10-Day Survival/Growth Bioassay. Mean mortality in the test sediment is 20 percent over the control mortality and statistically different from control ($p \leq 0.05$). For the growth endpoint, a mean reduction in biomass that is greater than 20 percent compared to the control and statistically different from the control ($p \leq 0.05$).

Cleanup Screening Levels Criteria

When any two biological test results show test sediment responses that are above the SCO listed above, OR when any one of the biological test results is above the Cleanup Screening Level criteria listed below, that sediment is judged to have exceeded the CSL.

In accordance with the SMS, the bioassay-specific response guidelines for evaluating an exceedance under the CSL criteria are as follows:

Amphipod 28-day Survival/Growth Bioassay. Mean mortality in the test sediment is greater than 25 percent over the mean control and statistically different from the control ($p \leq 0.05$). For the growth endpoint, a mean reduction in the biomass that is greater than 40 percent from the control and statistically different from the control ($p \leq 0.05$).

Midge 10-Day Survival/Growth Bioassay. Mean mortality in the test sediment is 30 percent over the control mortality and statistically different from the control ($p \leq 0.05$). For the growth endpoint, a mean reduction in biomass that is greater than 30 percent from the control sediment response and statistically different from the control ($p \leq 0.05$).

RESULTS OF AMPHIPOD, *HYALELLA AZTECA* 28-DAY SURVIVAL TEST (862-1)

All water quality observations were within the protocol specified ranges (Table 1, Section A). The test met applicable test acceptability criteria. Although the reference toxicant (positive control) LC50 result was slightly outside the laboratory's control chart action limits (0.50 g/L; control chart mean \pm 2 S.D. = 0.36 ± 0.12), a review of test conditions and procedures did not detect any unusual circumstances. (Section A).

The control sediment exhibited a mean mortality of 5.0% (Table 2). Mean mortality of test sediments JT-SS-06, JT-SS-08 and JT-SS-10 was 22.5%, 20.0% and 13.8%, respectively. None of these sediments exceeded the criteria for mortality for SCO or CSL. The biomass of test sediments JT-SS-06 and JT-SS-08 were both above that of the control sediment (Table 3). JT-SS-10 did result in an individual biomass of 0.28 mg that was statistically significantly lower than that of the control. However, JT-SS-10 did not exceed either the SCO or CSL criteria for growth.

RESULTS OF MIDGE, *CHIRONOMUS DILUTUS*, 10-DAY SURVIVAL AND GROWTH TEST (862-3)

All water quality observations were within the protocol specified ranges (Table 1, Section B). The test met all other applicable acceptability criteria including positive control performance (Section B).

The mean mortality in the control sediment in the *Chironomid* test was 6.3% (Table 4). Mean mortality of test sediments JT-SS-06, JT-SS-08 and JT-SS-10 was 10.0%, 15.0% and 8.8%, respectively. None of these were more than 20% over the control mortality. Therefore none of the sediments exceeded the SCO or CSL criteria for mortality. The control average individual ash-free dry weight was 1.22 mg (Table 5). All three test sediments resulted in growth that was statistically significantly lower than control growth. Two of the three test sediments, JT-SS-08 and JT-SS-10, resulted in exceedances under the SCO guidelines for growth with ash-free dry weights of 0.93 and 0.91 mg/individual, respectively. No test sediment failed the CSL criteria for growth.

SUMMARY OF SEDIMENT BIOASSAY RESULTS

The Sediment Management Standards includes Sediment Cleanup Objectives (SCO) and Cleanup Screening Levels (CSL) biological criteria to identify sediments that have adverse effects on biological resources. The SCO establishes a no adverse effects level, including no acute or chronic adverse effect, to the benthic community. The CSL establishes a minor adverse effects level, including acute or chronic effects, to the benthic community.

The SCO biological criteria are exceeded when one of the biological tests results is above the SCO for that bioassay endpoint. Two test sediments, JT-SS-08, and JT-SS-10, resulted in an exceedance under the SCO guidelines for growth in the *Chironomus* bioassay.

The CSL biological criteria is exceeded when 1) any two bioassay test results are above the SCO criteria; or 2) when one of the bioassay test results is above the CSL criteria. None of the test sediments exceeded the CSL criteria by resulting in either one test result that was a CSL exceedance or two test results that were SCO exceedances (Table 6).

STUDY APPROVAL


Assistant Laboratory Director Date 3-19-15


 3-19-15
Project Manager Date

Table 2. Mortality results of *Hyalella* 28-day toxicity test and data interpretation using guidelines from the Washington State SMS (2013).

Sample description	Percent mortality (Mean ± SD)	Significantly higher than control sediment at $\alpha=0.05$?	Percent higher (absolute) than control sediment	Exceedance under SCO? ¹	Exceedance under one-test criteria for CSL? ²
Control (NAS# 5195G)	5.0 ± 5.3	---	---	---	---
JT-SS-06 (NAS# 5192G)	22.5 ± 21.9	No	17.5	No	No
JT-SS-08 (NAS# 5193G)	20.0 ± 20.7	No	15.0	No	No
JT-SS-10 (NAS# 5194G)	13.8 ± 16.0	No	8.8	No	No

¹ Sediment Cleanup Objectives (SCO) exceedance if the test sediment mean mortality is significantly higher (1-tailed t-test at $P \leq 0.05$) than the control sediment mean mortality and the absolute difference is >10%.

² Cleanup Screening Levels (CSL) exceedance if the test sediment mean mortality is significantly higher (1-tailed t-test at $P \leq 0.05$) than the control sediment mean mortality and the absolute difference is >25%.

Table 3. Growth results of *Hyalella* 28-day toxicity test and data interpretation using guidelines from the Washington State SMS (2013).

Sample description	Average dry wt/amphipod (Mean ± SD)	Significantly lower than control sediment at $\alpha=0.05$?	Percent lower (absolute) than control sediment	Exceedance under SCO? ¹ ($MIG_c - MIG_T$)/ $MIG_c > 0.25$ and SD	Exceedance under one-test criteria for CSL? ² ($MIG_c - MIG_T$)/ $MIG_c > 0.40$ and SD
Control (NAS# 5195G)	0.32 ± 0.03	---	---	---	---
JT-SS-06 (NAS# 5192G)	0.41 ± 0.08	No	-28.0	No	No
JT-SS-08 (NAS# 5193G)	0.37 ± 0.08	No	-15.6	No	No
JT-SS-10 (NAS# 5194G)	0.28 ± 0.04	Yes	12.5	No	No

¹ Sediment Cleanup Objectives (SCO) exceedance if the test sediment mean growth is significantly lower (1-tailed t-test at $P \leq 0.05$) than the control sediment mean growth and the difference is >25%.

² Cleanup Screening Levels (CSL) exceedance (one-test criteria) if the test sediment mean growth is significantly lower (1-tailed t-test at $P \leq 0.05$) than the control sediment mean growth and the difference is >40%.

Table 4. Mortality results of *Chironomus* toxicity test and data interpretation using guidelines from the Washington State SMS (2013).

Sample description	Percent mortality (Mean ± SD)	Significantly higher than control sediment at $\alpha=0.05$?	Percent higher (absolute) than control sediment	Exceedance under SCO? ¹	Exceedance under one-test criteria for CSL? ²
Control (NAS# 5195G)	6.3 ± 5.2	---	---	---	---
JT-SS-06 (NAS# 5192G)	10.0 ± 14.1	No	3.7	No	No
JT-SS-08 (NAS# 5193G)	15.0 ± 16.9	No	8.7	No	No
JT-SS-10 (NAS# 5194G)	8.8 ± 11.3	No	2.5	No	No

¹ Sediment Cleanup Objectives (SCO) exceedance if the test sediment mean mortality is significantly higher (1-tailed t-test at $P \leq 0.05$) than the control sediment mean mortality and the absolute difference is >20%.

² Cleanup Screening Levels (CSL) exceedance if the test sediment mean mortality is significantly higher (1-tailed t-test at $P \leq 0.05$) than the control sediment mean mortality and the absolute difference is >30%.

Table 5. Growth results of *Chironomus* toxicity test and data interpretation using guidelines from the Washington State SMS (2013).

Sample description	Average ash-free dry w/midge (mg) (Mean ± SD)	Statistically significantly lower than control sediment at $\alpha=0.05$?	Percent lower than control sediment	Exceedance under SCO? ¹ ($MIG_C - MIG_T / MIG_C > 0.20$)	Exceedance under one-test criteria for CSL? ² ($MIG_C - MIG_T / MIG_C > 0.30$)
Control (NAS# 5195G)	1.22 ± 0.09	---	---	---	---
JT-SS-06 (NAS# 5192G)	0.98 ± 0.11	Yes	19.7	No	No
JT-SS-08 (NAS# 5193G)	0.93 ± 0.15	Yes	23.8	Yes	No
JT-SS-10 (NAS# 5194G)	0.91 ± 0.12	Yes	25.4	Yes	No

¹ Sediment Cleanup Objectives (SCO) exceedance if the test sediment mean growth is significantly lower (1-tailed t-test at $P \leq 0.05$) than the control sediment mean growth and the difference is >20%.

² Cleanup Screening Level (CSL) exceedance (one-test criteria) if the test sediment mean growth is significantly lower (1-tailed t-test at $P \leq 0.05$) than the control sediment mean growth and the difference is >30%.

Table 6. Interpretation of bioassay test results for Jacobson Terminals Property based on Washington State Sediment Management Standards - Sediment Cleanup Objectives (SCO) and Cleanup Screening Level (CSL) criteria.

Sample Description	<u>SCO Exceedance / CSL Exceedance¹</u>			<u>CSL Exceedance by two SCO Exceedances²</u>
	Test No. 862-1 <i>Hyalella</i> 28-day Survival	Test No. 862-1 <i>Hyalella</i> 28-day Growth	Test No. 862-3 <i>Chironomus</i> 10-day Survival	
Control (NAS# 5195G)	---	---	---	CSL ---
JT-SS-06 (NAS# 5192G)	No / No	No / No	No / No	No
JT-SS-08 (NAS# 5193G)	No / No	No / No	No / No	No
JT-SS-10 (NAS# 5194G)	No / No	No / No	No / No	No

¹One test result must exceed an SCO or a CSO criterion. Two test sediments exceeded the SCO criterion for *Chironomus* growth. No other single criterion exceedances occurred under either SCO or CSO criteria.

²Two test results must exceed the SCO criteria to exceed the CSL under this interpretation. No test sediments exceeded the CSL by having two SCO exceedances.

SECTION A

Amphipod (*Hyaella azteca*) 28-day sediment bioassay 862-1 data report

TOXICITY TEST REPORT

TEST IDENTIFICATION

Test No.: 862-1

Title: Toxicity of freshwater sediments using a 28-day amphipod, *Hyalella azteca*, sediment bioassay as part of the Remedial Investigation at the Jacobson Terminals Property in Seattle, WA.

Protocol No.: NAS-XXX-HA4c, February 11, 2000. Revision 3 (4-26-05). Based on ASTM 2001 (Standard test methods for measuring the toxicity of sediment-associated contaminants with fresh water invertebrates, E1706-00), Am. Soc. Test. Mat., Phila., PA, and EPA Method 100.1 (Methods for measuring the toxicity and bioaccumulation of sediment-associated contaminants with freshwater invertebrates, EPA/600/R-99/064). Washington State Sediment Management Standards (SMS) (Chapter 173-204 WAC, Last Update: 2/25/13).

STUDY MANAGEMENT

Study Sponsor: Hart Crowser, Inc., 1700 Westlake Ave. North, Suite 200, Seattle, WA 98109.

Sponsor's Study Monitor: Mr. Philip Cordell

Testing Laboratory: Northwestern Aquatic Sciences, P.O. Box 1437, Newport, OR 97365

Test Location: Newport laboratory

Laboratory's Study Personnel: G.J. Irissarri, B.S., Proj. Mngr./ Study Dir.; L.K. Nemeth, B.A., M.B.A., QA Officer; R.S. Caldwell, Ph.D., Sr. Aq. Toxicol.; G.A. Buhler, B.S., Aq. Toxicol.; J.B. Brown, B.S., D.V.M., Assoc. Aq. Toxicol.; Y. Nakahama, Sr. Tech.; L. Brady, Tech.

Study Schedule:

Test Beginning: 1-23-15, 0905 hrs.

Test Ending: 2-20-15, 1000 hrs.

Disposition of Study Records: All raw data, reports, and other study records are stored at Northwestern Aquatic Sciences, 3814 Yaquina Bay Rd., Newport, OR 97365.

Statement of Quality Assurance: The test data were reviewed by the Quality Assurance Unit to assure that the study was performed in accordance with the protocol and standard operating procedures. This report is an accurate reflection of the raw data.

TEST MATERIAL

Test Sediments: Freshwater test sediments collected as part of the Remedial Investigation at the Jacobson Terminals Property in Seattle, WA. Details are as follows:

NAS Sample No.	5192G	5193G	5194G
Description	JT-SS-06	JT-SS-08	JT-SS-10
Collection Date	1/12/15	1/12/15	1/12/15
Receipt Date	1/14/15	1/14/15	1/14/15

Control Sediment: The negative control sediment (NAS#5195G) was collected on 1-19-15 from an area approximately one mile east of the Hwy. 101 bridge at Beaver Creek, approx. 8 miles south of Newport, OR.

Treatments: Homogenized at test set up by mixing using stainless steel implements.

Storage: All test and control sediments were stored at 4°C in the dark in sealed containers until used.

TEST WATER

Source: Dechlorinated municipal tap water.

Dates of Preparation: 1-19-15 and 2-6-15

Water Quality:

pH: 7.7, 7.5

conductivity: 108, 119 µmhos/cm

hardness: 26, 26 mg/L as CaCO₃

alkalinity: 30, 30 mg/L as CaCO₃.

total chlorine: <0.02, <0.02 mg/L

Pretreatment: Dechlorinated and aerated ≥24 hr.

TEST ORGANISMS

Species: *Hyaella azteca*, amphipod.

Age/Size: 7-8 days old

Source: Chesapeake Cultures, Hayes, VA; received 1-21-15

Acclimation: Holding conditions for the three days prior to testing averaged: Temperature, 19.5 ± 1.7 °C; dissolved oxygen, 10.8 ± 3.7 mg/L; pH, 7.2 ± 0.4 ; conductivity, 358 ± 207 μ mhos/cm; hardness, 111 ± 67 mg/L as CaCO₃; and alkalinity, 103 ± 67 mg/L as CaCO₃. Photoperiod, 16:8, L:D. Half of the water was replaced daily with dechlorinated municipal tap water during holding. Animals were fed YTC daily during holding.

TEST PROCEDURES AND CONDITIONS

The following is an abbreviated statement of the test procedures and a statement of the test conditions actually employed. See the test protocol (Appendix I) for a more detailed description of the test procedures used in this study.

Test Chambers: 300 ml high-form glass beakers

Test Volumes: 100 ml sediment layer; 175 ml test water.

Replicates/Treatment: 8

Organisms/Treatment: 80

Water Volume Changes: 2 water volumes per day

Aeration: None.

Feeding: Animals are fed 1.0 ml of YTC suspension per beaker daily.

Acceptance Criteria: Results are valid if mean control mortality does not exceed 20%, and the mean individual dry weight at test termination is ≥ 0.15 mg.

Effects Criteria: 1) survival after 28 days, and 2) average individual dry weight after 28 days. Death is defined as no visible movement or response to tactile stimulation. Missing organisms were considered to be dead.

Water Quality and Other Test Conditions: The temperature, dissolved oxygen, conductivity, pH, hardness, alkalinity, and ammonia-nitrogen were measured in the overlying water of one replicate test container per treatment on days 0 and 28 of the test. Temperature was measured daily, pH and dissolved oxygen three times per week, and conductivity weekly, in the overlying water of one replicate test container per treatment. Hardness and alkalinity were measured with titrimetric methods. Ammonia-N was measured using Hach reagents based on the salicylate (Clin. Chim. Acta 14:403, 1996) colorimetric method; samples were not distilled prior to analysis. The photoperiod was 16:8, L:D.

DATA ANALYSIS METHODS

Percent survival, percent mortality and average individual dry weight were calculated for each replicate as follows:

$$\text{percent survival} = 100 \times (\text{number surviving}/\text{initial number tested})$$

$$\text{percent mortality} = 100 \times (\text{number dead}/\text{initial number tested})$$

$$\text{average individual dry weight} = (\text{final wt.} - \text{tare wt.})/\text{number weighed,}$$

where:

$$\text{final wt.} = \text{tare wt.} + \text{dry weight of organisms recovered on day 28, in mg}$$

Means and standard deviations for the biological endpoints described above, and for water quality data, were computed using Microsoft Excel 2010. Mean individual dry weight and mean percent mortality in each test sediment were statistically compared to the control sediment. Where appropriate, an arcsine square root transformation was performed on percent mortality data before analysis. Following determination of normality and homogeneity of variances, a one-tailed Student T-test, Mann-Whitney or Approximate T test was conducted at the 0.05 level of significance. The statistical software used was BioStat (version Feb 9, 2006 (EXCEL)) bioassay software developed by the U.S. Army Corps of Engineers, Seattle District.

PROTOCOL DEVIATIONS

None

REFERENCE TOXICANT TEST

The reference toxicant test is a multi-concentration toxicity test using potassium chloride, to evaluate the performance of the test organisms used in the sediment toxicity test. The performance is evaluated by comparing the results of this test with historical results obtained at the laboratory. A summary of the reference toxicant test result is given below. The reference toxicant test raw data are found in Appendix III.

Test No.: 999-3381

Reference Toxicant and Source: Potassium Chloride (KCl), Fisher Lot #114689.

Test Date: 1-23-15.

Dilution Water Used: Moderately hard synthetic water prepared from Milli-Q® deionized water.

Result: 96-hr LC50, 0.50 g/L. This result is slightly outside the laboratory's control chart action limits (0.25 – 0.48 g/L). A review of test organisms and test conditions and procedures indicated that there were no unusual circumstances that may have affected the test results.

TEST RESULTS

Observations of water quality in the overlying water throughout the test are summarized in Table 1. A detailed tabulation of the water quality results by sample and test day can be found in Appendix II. The means and standard deviations of percent mortality of *Hyalella* exposed for 28 days to sediments are summarized in Table 2. The means and standard deviations of average individual dry weight of *Hyalella* exposed for 28 days to sediments are summarized in Table 3. Detailed data organized by sample and replicate, and summary statistics for these observations, are given in Appendix II.

All water quality observations of overlying water temperature and dissolved oxygen were within the protocol specified ranges. Ammonia-N in the overlying water ranged between <0.1 and 0.9 mg/L for all day 0 and day 28 measurements.

The test met the acceptability criteria specified in the SMS with 5.0 % mean control mortality ($\leq 20\%$ required) and a final control sediment growth of 0.32 mg/individual (≥ 0.15 mg required). The reference toxicant (positive control) LC50 result was slightly outside the laboratory's control chart action limits (0.50 g/L; control chart mean ± 2 S.D. = 0.36 ± 0.12). A review of test conditions and procedures did not detect any unusual circumstances.

Interpretation was based on guidelines from the Washington State Sediment Management Standards (Chapter 173-204 WAC, Last Update: 2/25/13). The SMS includes Sediment Cleanup Objectives (SCO) and Cleanup Screening Levels (CSL) biological criteria. The Sediment Cleanup Objectives establish a no adverse effects level, including no acute or chronic adverse effect, to the benthic community. The Cleanup Screening Levels establish a minor adverse effects level, including acute or chronic effects, to the benthic community. To qualify as an adverse effect under the SCO for mortality the mean mortality in the test sediment is greater than 10 percent over the mean control and statistically different from the reference ($p = 0.05$). For the growth endpoint, a mean reduction in the biomass that is greater than 25 percent of the control sediment response and statistically different from the control ($p = 0.05$). For the CSL adverse effects criteria mean mortality in the test sediment is greater than 25 percent over the mean control and statistically different from the control ($p = 0.05$). For the growth endpoint, a mean reduction in the biomass that is greater than 40 percent from the control and statistically different from the control ($p = 0.05$).

Although sediment JT-SS-10 resulted in mean dry weight that was statistically significantly lower than that of the control, the mean dry weight did not exceed either the SCO or CSL criteria. None of the test sediments resulted in an exceedance under either the SCO or the CSL of the SMS guidelines.

STUDY APPROVAL

Arnoldo Brissani 3-19-15
Project Manager/Study Director Date

Julie R. Fiore 3-19-15
Quality Assurance Unit Date

Linda Nemeth 3-19-15
Assistant Laboratory Director Date

Table 1. Summary of water quality conditions during tests of the amphipod, *Hyalella azteca*, exposed to freshwater sediments.

Water Quality Parameter	Mean \pm S.D.	Minimum	Maximum	N
Temperature ($^{\circ}$ C)	22.4 \pm 0.4	22.0	23.9	116
Dissolved oxygen (mg/L)	7.0 \pm 0.5	6.1	8.5	52
Conductivity (μ mhos/cm)	123 \pm 22	110	219	24
pH	7.2 \pm 0.2	6.6	8.0	52
Hardness (mg/L as CaCO ₃)	32 \pm 4	26	34	8
Alkalinity (mg/L as CaCO ₃)	35 \pm 5	30	40	8
Total ammonia (mg/L)	---	<0.1	0.9	8

Table 2. Mortality results of *Hyalella* 28-day toxicity test and data interpretation using guidelines from the Washington State SMS (2013).

Sample description	Percent mortality (Mean \pm SD)	Significantly higher than the control sediment at $\alpha=0.05$?	Percent higher (absolute) than control sediment	Exceedance under SCO? ¹	Exceedance under one-test criteria for CSL ²
Control (NAS# 5195G)	5.0 \pm 5.3	---	---	---	---
JT-SS-06 (NAS# 5192G)	22.5 \pm 21.9	No	17.5	No	No
JT-SS-08 (NAS# 5193G)	20.0 \pm 20.7	No	15.0	No	No
JT-SS-10 (NAS# 5194G)	13.8 \pm 16.0	No	8.8	No	No

¹ Sediment Cleanup Objectives (SCO) exceedance if the test sediment mean mortality is significantly higher (1-tailed t-test at $P \leq 0.05$) than the control sediment mean mortality and the absolute difference is $>10\%$.

² Cleanup Screening Levels (CSL) exceedance if the test sediment mean mortality is significantly higher (1-tailed t-test at $P \leq 0.05$) than the control sediment mean mortality and the absolute difference is $>25\%$.

Table 3. Growth results of *Hyalella* 28-day toxicity test and data interpretation using guidelines from the Washington State SMS (2013).

Sample description	Average Individual Dry Weight (mg) (Mean \pm SD) ¹	Statistically significantly lower than control sediment at $\alpha=0.05$?	Percent lower than control sediment	Exceedance under SCO? ¹ (MIG _C - MIG _T /MIG _C >0.25)	Exceedance under one-test criteria for CSL? ² (MIG _C - MIG _T /MIG _C >0.40)
Control (NAS# 5195G)	0.32 \pm 0.03	---	---	---	---
JT-SS-06 (NAS# 5192G)	0.41 \pm 0.08	No	-28.0	No	No
JT-SS-08 (NAS# 5193G)	0.37 \pm 0.08	No	-15.6	No	No
JT-SS-10 (NAS# 5194G)	0.28 \pm 0.04	Yes	12.5	No	No

¹ Sediment Cleanup Objectives (SCO) exceedance if the test sediment mean growth is significantly lower (1-tailed t-test at $P \leq 0.05$) than the control sediment mean growth, and the difference is $>25\%$.

² Cleanup Screening Levels (CSL) exceedance (one-test criteria) if the test sediment mean individual growth is significantly lower (1-tailed t-test at $P \leq 0.05$) than the control sediment mean growth, and the difference is $>40\%$.

APPENDIX I

PROTOCOL

TEST PROTOCOL

FRESHWATER AMPHIPOD, *HYALELLA AZTECA*, 28-DAY SEDIMENT SURVIVAL AND GROWTH TEST

1. INTRODUCTION

1.1 Purpose of Study: The purpose of this study is to characterize the chronic toxicity of freshwater sediments using a 28-day exposure and survival and growth endpoints with the amphipod, *Hyalella azteca*.

1.2 Referenced Method: This protocol is based on ASTM Method E 1706-00 (ASTM 2001) and EPA Method 100.1 (EPA/600/R-99/064)

1.3 Summary of Method: A summary of test conditions for the amphipod 28-day sediment survival and growth test is tabulated below. The test with *Hyalella azteca* is conducted at $23 \pm 1^\circ\text{C}$ with a 16L:8D photoperiod at an illuminance of about 100-1000 lux. Test chambers are 300-mL high-form lipless beakers containing 100 mL of sediment and 175 mL of overlying water. Ten 7-8day old amphipods are used in each replicate. The number of replicates/treatment depends on the objective of the test. Eight replicates are recommended for routine testing. Amphipods in each test chamber are fed 1.0 mL of YCT food daily. Each chamber receives two volume additions per day of overlying water. Test endpoints include survival and growth.

2. STUDY MANAGEMENT

2.1 Sponsor's Name and Address:

2.2 Sponsor's Study Monitor:

2.3 Name of Testing Laboratory:

Northwestern Aquatic Sciences
3814 Yaquina Bay Road, P.O. Box 1437
Newport, OR 97365.

2.4 Test Location:

2.5 Laboratory's Personnel to be Assigned to the Study:

Study Director: _____
Quality Assurance Unit: _____
Aquatic Toxicologist: _____
Aquatic Toxicologist: _____

2.6 Proposed Testing Schedule: Tests are normally begun within 14 days of sample collection. Reference toxicant test to be run concurrently.

2.7 Good Laboratory Practices: The test is conducted following the principles of Good Laboratory Practices (GLP) as defined in the EPA/TSCA Good Laboratory Practice regulations revised August 17, 1989 (40 CFR Part 792).

3. TEST MATERIAL

The test materials are freshwater sediments. The control, reference, and test sediments are placed in solvent cleaned 1 L glass jars fitted with PTFE-lined screw caps. At the laboratory the samples are stored at 4°C in the dark. The original sealed containers may be stored for up to 8 weeks prior to testing, depending on the testing requirements. If jars are not full when received or if sediment is removed for testing, headspaces should be filled with nitrogen to retard deterioration. A negative control sediment is collected from a clean site. In addition, a reference sediment, a clean sediment with physical characteristics similar to the test sediments, may be employed as a comparison station.

4. TEST WATER

Test water (overlying water) at NAS is normally dechlorinated tap water or moderately hard synthetic water. Synthetic dilution water is prepared from Milli-Q reagent grade water and reagent grade chemicals. Test water may also be well water, surface water, site water, or other water depending on the study design. The hardness or other water quality parameters of the dilution water may need to be adjusted to meet the study design.

5. TEST ORGANISMS

5.1 Species: amphipod, *Hyalella azteca*.

5.2 Source: Cultured at NAS. Alternatively, animals may be purchased from a reputable commercial supplier.

5.3 Age: 7-8 days old at start of test

5.4 Acclimation and Pretest Observation: Cultures are maintained at $23 \pm 1^\circ\text{C}$ under a 16:8 L:D photoperiod. Cultured amphipods are fed dried maple leaves with YTC. Rabbit chow, Tetramin® or TetraFin® flakes may also be used. Acclimation of test organisms to the test water may be desirable, depending on culture water, but it is not required. If test organisms are to be acclimated, fifty percent of the holding water is changed daily with the addition of test water.

6. DESCRIPTION OF TEST SYSTEM

6.1 Test Chambers and Environmental Control: Test chambers used in the toxicity test are 300-mL high-form lipless glass beakers. Test chambers are maintained at constant temperature by partial immersion in a temperature-controlled water bath or by placement in a temperature-controlled room. Aeration is not employed unless dissolved oxygen drops below 2.5 mg/L. The test is conducted under an illuminance of 100-1000 lux with a 16L:8D photoperiod.

6.2 Cleaning: All laboratory glassware, including test chambers, is cleaned as described in EPA/600/4-90/027F. New glassware and test systems are soaked 15 minutes in tap water and scrubbed with detergent (or cleaned in automatic dishwasher); rinsed twice with tap water; carefully rinsed once with fresh, dilute (10%, V:V) hydrochloric or nitric acid to remove scale, metals, and bases; rinsed twice with deionized water; rinsed once with acetone to remove organic compounds (using a fume hood or canopy); and rinsed three times with deionized water. Test systems and chambers are rinsed again with dilution water just before use.

7. EXPERIMENTAL DESIGN AND TEST PROCEDURES

7.1 Experimental Design: The test involves exposure of amphipods to test, control, and reference sediments. The sediments are placed on the bottom of the test containers and are overlain with test water. The test exposure is for 28 days. The renewal of overlying water consists of two volume additions per day, either continuous or intermittent. Each treatment consists of eight replicate test containers, each containing 10 organisms. Test chamber positions are completely randomized. Test organisms are randomly distributed to the test chambers. Blind testing is normally used.

7.2 Setup of Test Containers: Sediments are homogenized and placed in test chambers on the day before addition of test organisms. Sediment (100 ml) is placed into each of eight replicate beakers. After addition of the sediment, 175 ml of test water is gently added to each beaker in a manner to prevent resuspension. The overlying water is replaced twice daily. The test begins when amphipods are introduced to the test chambers. Initial water quality measurements are taken prior to the addition of test organisms.

7.3 Effect Criterion: The effect criteria used in the 28-day amphipod bioassay are mortality and growth. Death is defined as the lack of movement of body or appendages on response to tactile stimulation. Growth is measured as change in dry weight.

7.4 Test Conditions: No aeration is employed unless dissolved oxygen falls below 2.5 mg/L. The test temperature employed is $23 \pm 1^\circ\text{C}$. A 16:8, L:D photoperiod is used. Illumination is supplied by daylight fluorescent lamps at 100-1000lux. The overlying water is replaced twice daily.

7.5 Beginning the Test: On the day the test begins, amphipods are impartially counted into small containers of test water (10/container). The test is begun by rinsing test organisms into the equilibrated test containers. For the growth endpoint, time-zero weight data should be collected.

7.6 Feeding: Amphipods are fed 1.0 mL of YCT daily per test chamber. A feeding may be skipped if there is a build up of excess food. However, all beakers must be treated similarly.

7.7 Test Duration, Type and Frequency of Observations, and Methods: The duration of the toxicity test is 28 days. The type and frequency of observations to be made are summarized as follows:

TYPE OF OBSERVATION	TIMES OF OBSERVATION
<i>BIOLOGICAL DATA</i>	
Survival, growth	Day 28
<i>PHYSICAL AND CHEMICAL DATA</i>	
Hardness, alkalinity, conductivity, and ammonia-N	Beginning and end of test in overlying water of one replicate beaker from each treatment.
Temperature	Daily in overlying water of one replicate beaker from each treatment.
Conductivity	Weekly
Dissolved oxygen and pH	3X/week
Optional pore water ammonia and/or sulfide	In test sediments prior to initiating the tests. Optionally in sediments from sacrificial test chambers at test beginning and/or end.

Dissolved oxygen is measured using a polarographic oxygen probe calibrated according to the manufacturer's recommendations. The pH is measured using a pH probe and a properly calibrated meter with scale divisions of 0.1 pH units. Temperature is measured with a calibrated mercury thermometer or telethermometer. Conductivity is measured with a conductivity meter. Hardness and alkalinity are measured using titrimetric methods. Total soluble sulfide and total ammonia-N were

measured using Hach test kits based on the methylene blue (EPA Method 376.2) and salicylate (Clin. Chim. Acta 14:403, 1996) colorimetric methods, respectively; samples were not distilled prior to analysis.

Overlying water should be sampled just before water renewal from about 1 to 2 cm above the sediment surface using a pipet. It may be necessary to pool water samples from individual replicates. The pipet should be checked to make sure no organisms are removed during sampling of overlying water.

7.8 Test Termination: At test termination, the contents of each test container are sieved through a #35 (500 μm mesh) sieve to recover the amphipods. Amphipods from each replicate are put into a 30 mL plastic cup, rinsed with DI water, gently blotted and placed into the appropriate tared aluminum weighing pan. The number of survivors for each container is recorded on the datasheet.

7.9 Growth Measurement: Growth is measured as average dry weight of animals in a test replicate at the end of the test on day 28. Pooled animals from each test replicate are gently blotted and placed into tared aluminum weigh pans. The pans are dried at 60-90°C to constant weight. The dried amphipods are placed into a dessicator and weighed as soon as possible to the nearest 0.01 mg (desirable to use 0.001 mg). The total weight of the dried amphipods in each pan is divided by the number of amphipods weighed to obtain an average dry weight per surviving amphipod per replicate.

8. CRITERIA OF TEST ACCEPTANCE

The test results are acceptable if the minimum survival of organisms in the control treatment at the end of the test is at least 80%.

9. DATA ANALYSIS

The endpoints of the toxicity test are survival and growth. Survival is obtained as a direct count of living organisms in each test container at the end of the test. Average amphipod dry weight, also measured at the end of the test, may be used to compare growth between treatment sediments and the control or reference sediment. Ordinarily the following data analysis is performed. Due to special requirements, alternative methods may be used. The means and standard deviations are calculated for each treatment level. Identification of toxic sediments is established by statistical comparison of test endpoints between test and control or reference sediments. Between treatment comparisons may be made using a Student's t-test or Wilcoxon's Two-Sample test, where each treatment is compared to the control or the reference sediment. An arcsine-square root transformation of proportional data, and tests for normality and heterogeneity of variances, are performed prior to statistical comparisons.

10. REPORTING

The final report of the test results must include all of the following standard information at a minimum: name and identification of the test; the investigator and laboratory; date and time of test beginning and end; information on the test material; information on the source and quality of the overlying/test water; detailed information about the test organisms including acclimation conditions; a description of the experimental design and test chambers and other test conditions including feeding, if any, and water quality; definition of the effect criteria and other observations; responses, if any, in the control treatment; tabulation and statistical analysis of measured responses and a summary table of endpoints; a description of the statistical methods used; any unusual information about the test or deviations from procedures; reference toxicant testing information.

11. STUDY DESIGN ALTERATION

Amendments made to the protocol must be approved by the sponsor and study director and should include a description of the change, the reason for the change, the date the change took effect and the dated signatures of the study director and sponsor. Any deviations in the protocol must be described and recorded in the study raw data.

12. REFERENCE TOXICANT

The reference toxicant test is a standard multi-concentration toxicity test using a specified chemical toxicant to evaluate the performance of test organisms used in the study. Reference toxicant tests are 96-hour, water only exposures, not 28-day sediment exposures. The reference toxicant test is run concurrently. Performance is evaluated by comparing the results of the reference toxicant test with historical results (e.g., control charts) obtained at the laboratory.

13. REFERENCED GUIDELINES

ASTM. 2001. Standard Test Methods for Measuring the Toxicity of Sediment-Associated Contaminants with Fresh Water Invertebrates. ASTM Standard Method No. E 1706-00. Am. Soc. Test. Mat., Philadelphia, PA.

U.S. EPA. 2000. Section 11, Test Method 100.1, *Hyalella azteca* 10-d Survival and Growth Test for Sediments, pp. 47-54 In: Methods for Measuring the Toxicity and Bioaccumulation of Sediment-associated Contaminants with Freshwater Invertebrates (Second Edition). EPA/600/R-99/064.

Weber, C.I. (Ed.) 1993. Methods for Measuring the Acute Toxicity of Effluents and Receiving Waters to Freshwater and Marine Organisms (Fourth Edition). EPA/600/4-90/027F.

14. APPROVALS

_____ for _____
Name Date

_____ for Northwestern Aquatic Sciences
Name Date

Appendix A
Test Conditions Summary

1. Test type	whole sediment toxicity test with renewal of overlying water
2. Test duration	28 days
3. Temperature	23 ± 1°C
4. Light quality	daylight fluorescent light
5. Illuminance	100-1000 lux
6. Photoperiod	16L:8D
7. Test chamber size	300-mL high-form lipless beakers, (Pyrex® 1040 or equivalent)
8. Sediment volume	100 mL
9. Overlying water volume	175 mL
10. Renewal overlying water	2 volume additions/day (continuous or intermittent)
11. Age of test organisms	7-8 days old at test initiation
12. Organisms per test chamber	10
13. Replicates per treatment	8 recommended for routine testing (depends on design)
14. Organisms per treatment	80
15. Feeding regime	YCT food, fed 1.0 mL daily/chamber
16. Cleaning	if screens are used, clean as needed
17. Aeration	None, unless DO falls below 2.5 mg/L
18. Overlying (test) water	Dechlorinated tap water, culture water, well water, surface water, site water or reconstituted water, depending on study design.
19. Water quality	Hardness, alkalinity, conductivity, ammonia-N beginning and end; temperature daily; conductivity weekly; DO & pH 3X/wk
20. Endpoints	Survival & growth (based on weight)
21. Test acceptability criteria	Minimum control survival of 80%
22. Sample holding	14 days at 4°C in the dark (recommended)
23. Sample volume required	1L (800 mL per sediment)
24. Reference toxicant	Concurrent testing required

APPENDIX II

RAW DATA

**TEST DESCRIPTION, MONITORING, AND RESULTS
BENCHSHEETS**

Test No. 862-1 Client Hart Crowser Investigator _____

STUDY MANAGEMENT

Client: Hart Crowser, Inc., 1700 Westlake Ave. North, Suite 200, Seattle, WA 98109

Client's Study Monitor: Mr. Philip Cordell

Testing Laboratory: Northwestern Aquatic Sciences

Test Location: Newport Laboratory

Laboratory's Study Personnel:

Proj. Man./Study Dir. G.J. Irissarri ^{ESL}

QA Officer L.K. Nemeth

1. Yves Kala Namu ^{ES} 2. hA Buhler ^{ES}

3. J. Brown ^{ES} 4. Lauren Brady ^{ES}

5. _____ 6. J.S. Calhoun ^{ES}

7. _____ 8. _____

Study Schedule:

Test Beginning: 1-23-15 0905 Test Ending: 2-20-15 1000

TEST MATERIAL

General description (see sample logbook/chain-of-custody for details):

NAS Sample No.:	5192G	5193G	5194G		
Description:	JT-SS-06	JT-SS-08	JT-SS-10		
Collection Date:	1/12/15	1/12/15	1/12/15		
Receipt Date:	1/14/15	1/14/15	1/14/15		

NAS Sample No.:					
Description:					
Collection Date:					
Receipt Date:					

NAS Sample No.:					
Description:					
Collection Date:					
Receipt Date:					

NAS Sample No.:					
Description:					
Collection Date:					
Receipt Date:					

NAS Sample No.:					
Description:					
Collection Date:					
Receipt Date:					

Error codes: 1) correction of handwriting error
2) written in wrong location; entry deleted
3) wrong date deleted, replaced with correct date
4) error found in measurement; measurement repeated

Test No. 862-1 Client Hart Crowser Investigator _____

TEST WATER

Source: Dechlorinated Newport, OR tap water
 Date of Collection/Preparation: 1-19-15, 2-6-15
 pH 7.8, 7.5
 Cond (umhos/cm²) 108, 119
 Hardness (mg/La0) 26, 26
 Alkalinity (mg/L) 30, 30
 Total Chlorine (mg/l) <0.02, <0.02
 Treatments: Aerated ≥ 24 hrs

TEST ORGANISMS

Species: Hyalella azteca Age: 7-8 DAYS Date received: 1-21-15
 Source: Chesapeake Cultures, Hayes, VA

Acclimation Data:

Date	Temp. (deg.C)	pH	DO (mg/L)	Cond. umhos/cm	Hardness (mg/L)	Alkalinity (mg/L)	Feeding		Water changes
							Amount	description	
1-21-15	17.5	6.8	7.5-0	596	188	180	10mls	YTE	see data - yes 2 20%
1-22-15	20.2	7.3	8.9	255	77	70	"	"	"
1-23-15	20.7	7.5	8.4	223	68	60	"	"	"
Mean	19.5	7.2	10.8	358	111	103			
S.D.	1.7	0.4	3.7	207	67	67			
(N)	3	3	3	3	3	3			

Photoperiod during acclimation: 16:8, L:D

TEST PROCEDURES AND CONDITIONS

Test chambers: 300 ml glass beakers
 Test volumes: 100 ml of test sediment; 275 ml total volume
 Replicates/treatment: (8) 8 Organisms/treatment: (80) 80 (10/REP)
 Test water changes: Twice daily
 Aeration: only if DO falls below 2.5 mg/L
 Feeding: everyday beginning with day zero
 Test temperature (deg.C): 23
 Beaker placement: Total randomization
 Photoperiod: 16:8, L:D

Control Sediment:

Source: From an area approximately one mile east of the Hwy. 101 bridge at Beaver Creek, approx. 8 miles south of Newport, OR.
 Date collected: 1/19/15
 Sieved through 0.5 -mm screen
 Storage: 4°C in the dark in closed containers. NAS# 5195G

MISCELLANEOUS NOTES

Test No. 862-1 Client Hart Crowser Investigator _____

Test conducted in (circle one): room 1 room 2 trailer water bath other: _____

Randomization chart: TOP SHELF

5	10	15	20	25	30				
4	9	14	19	24	29				
3	8	13	18	23	28				
2	7	12	17	22	27	32			
1	6	11	16	21	26	31			

Randomization chart: FRONT

Randomization chart:

Randomization chart:

Test No. 862-1 Client Hart Crowser Investigator _____

DAILY RECORD SHEET

Day 0 (1/23/15) UAS/631

Beaker No.	Temp.* (deg.C)	DO* (ppm)	Cond.* (umhos/cm)	pH*	Hardness* (mg/L)	Alkalinity* (mg/L)	NH3* (ppm)	Comments
3	22.2	7.6	219	6.8	34	30		Each beaker fed 1.0 ml
7	22.2	7.7	113	6.6	34	30		YTC suspension
17	22.3	7.6	133	6.6	34	40		Initials: <u>UAS</u>
18	22.3	7.6	115	6.6	34	40		
								Water changed in all
								beakers.
								Time: <u>0515</u>
								Initials: <u>UAS</u>
								Water changed in all
								beakers.
								Time: <u>1600</u>
								Initials: <u>UAS</u>

*Water quality measurements to be taken.

Day 1 (1/24/15) UAS

Beaker No.	Temp.* (deg.C)	DO (ppm)	Cond. (umhos/cm)	pH	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	22.2							Each beaker fed 1.0 ml
7	22.3							YTC suspension
17	22.2							Initials: <u>UAS</u>
18	22.2							
								Water changed in all
								beakers.
								Time: <u>0515</u>
								Initials: <u>UAS</u>
								Water changed in all
								beakers.
								Time: <u>1630</u>
								Initials: <u>UAS</u>

*Water quality measurements to be taken.

Test No 862-1 Client Hart Crowser Investigator _____

DAILY RECORD SHEET

Day 2 (1/25/15) BSJ

Beaker No.	Temp.* (deg.C)	DO (ppm)	Cond. (umhos/cm)	pH	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	22.2							Each beaker fed 1.0 ml YTC suspension Initials: <u>BSJ</u>
7	22.2							
17	22.1							
18	22.1							
								Water changed in all beakers. Time: <u>0450</u> Initials: <u>BSJ</u>
								Water changed in all beakers. Time: <u>1620</u> Initials: <u>BSJ</u>

*Water quality measurements to be taken.

Day 3 (1/26/15) BSJ

Beaker No.	Temp.* (deg.C)	DO* (ppm)	Cond. (umhos/cm)	pH*	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	22.3	7.0		7.1				Each beaker fed 1.0 ml YTC suspension Initials: <u>BSJ</u>
7	22.2	7.0		7.1				
17	22.1	7.0		7.1				
18	22.1	7.2		7.2				
								Water changed in all beakers. Time: <u>0510</u> Initials: <u>BSJ</u>
								Water changed in all beakers. Time: <u>1625</u> Initials: <u>BSJ</u>

*Water quality measurements to be taken.

Test No 862-1 Client Hart Crowser Investigator _____

DAILY RECORD SHEET

Day 4 (1/27/15) ✓

Beaker No.	Temp.* (deg.C)	DO (ppm)	Cond. (umhos/cm)	pH	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	22.6							Each beaker fed 1.0 ml
7	22.4							YTC suspension
17	22.2							Initials: ✓
18	22.2							
								Water changed in all beakers.
								Time: 0520
								Initials: ✓
								Water changed in all beakers.
								Time: 1645
								Initials: ✓

*Water quality measurements to be taken.

Day 5 (1/28/15) ✓

Beaker No.	Temp.* (deg.C)	DO* (ppm)	Cond.* (umhos/cm)	pH*	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	23.0	7.3	139	7.2				Each beaker fed 1.0 ml
7	22.8	7.1	124	7.2				YTC suspension
17	22.7	7.1	124	7.2				Initials: ✓
18	22.7	7.1	120	7.2				
								Water changed in all beakers.
								Time: 0520
								Initials: ✓
								Water changed in all beakers.
								Time: 1650
								Initials: ✓

*Water quality measurements to be taken.

Test No 862-1 Client Hart Crowser Investigator _____

DAILY RECORD SHEET

Day 6 (1/29/15) YV

Beaker No.	Temp.* (deg.C)	DO (ppm)	Cond. (umhos/cm)	pH	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	23.1							Each beaker fed 1.0 ml
7	23.0							YTC suspension
17	22.8							Initials: <u>YV</u>
18	22.8							
								Water changed in all
								beakers.
								Time: <u>0525</u>
								Initials: <u>Y</u>
								Water changed in all
								beakers.
								Time: <u>1615</u>
								Initials: <u>YS</u>

*Water quality measurements to be taken.

Day 7 (1/30/15) Y

Beaker No.	Temp.* (deg.C)	DO* (ppm)	Cond. (umhos/cm)	pH*	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	22.5	7.3		7.1				Each beaker fed 1.0 ml
7	22.4	6.9		7.0				YTC suspension
17	22.2	6.7		7.0				Initials: <u>YV</u>
18	22.2	6.8		7.1				
								Water changed in all
								beakers.
								Time: <u>0510</u>
								Initials: <u>YV</u>
								Water changed in all
								beakers.
								Time: <u>1645</u>
								Initials: <u>Y</u>

*Water quality measurements to be taken.

Test No 862-1 Client Hart Crowser Investigator _____

DAILY RECORD SHEET

Day 8 (1/31/15) CS

Beaker No.	Temp.* (deg.C)	DO (ppm)	Cond. (umhos/cm)	pH	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	22.5							Each beaker fed 1.0 ml
7	22.2							YTC suspension
17	22.2							Initials: <u>CS</u>
18	22.3							
								Water changed in all beakers.
								Time: <u>0515</u>
								Initials: <u>CS</u>
								Water changed in all beakers.
								Time: <u>1700</u>
								Initials: <u>CS</u>

*Water quality measurements to be taken.

Day 9 (2/1/15) CS

Beaker No.	Temp.* (deg.C)	DO (ppm)	Cond. (umhos/cm)	pH	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	22.3							Each beaker fed 1.0 ml
7	22.1							YTC suspension
17	22.0							Initials: <u>CS</u>
18	22.1							
								Water changed in all beakers.
								Time: <u>0500</u>
								Initials: <u>CS</u>
								Water changed in all beakers.
								Time: <u>1610</u>
								Initials: <u>CS</u>

*Water quality measurements to be taken.

Test No 862-1 Client Hart Crowser Investigator _____

DAILY RECORD SHEET

Day 10 (2/2/15) 631

Beaker No.	Temp.* (deg.C)	DO* (ppm)	Cond. (umhos/cm)	pH*	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	22.4	7.4		7.2				Each beaker fed 1.0 ml
7	22.2	7.4		7.1				YTC suspension
17	22.1	7.0		7.1				Initials: <u>631</u>
18	22.1	7.1		7.1				
								Water changed in all
								beakers.
								Time: <u>0510</u>
								Initials: <u>631</u>
								Water changed in all
								beakers.
								Time: <u>1105</u>
								Initials: <u>631</u>

*Water quality measurements to be taken.

Day 11 (2/3/15) 631

Beaker No.	Temp.* (deg.C)	DO* (ppm)	Cond. (umhos/cm)	pH*	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	22.6							Each beaker fed 1.0 ml
7	22.7							YTC suspension
17	22.5							Initials:
18	22.5							
								Water changed in all
								beakers.
								Time: <u>0505</u>
								Initials: <u>631</u>
								Water changed in all
								beakers.
								Time: <u>1105</u>
								Initials: <u>631</u>

*Water quality measurements to be taken.

Test No. 862-1 Client Hart Crowser Investigator _____

DAILY RECORD SHEET

Day 12 (2/4/15) LB

Beaker No.	Temp.* (deg.C)	DO* (ppm)	Cond.* (umhos/cm)	pH*	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	22.6	6.3	126	7.1				Each beaker fed 1.0 ml
7	22.6	6.3	119	7.0				YTC suspension
17	22.4	6.1	122	7.0				Initials: <u>LB</u>
18	22.4	6.2	119	7.0				
								Water changed in all
								beakers.
								Time: <u>0510</u>
								Initials: <u>LB</u>
								Water changed in all
								beakers.
								Time: <u>1605</u>
								Initials: <u>LB</u>

*Water quality measurements to be taken.

Day 13 (2/5/15) LB

Beaker No.	Temp.* (deg.C)	DO (ppm)	Cond. (umhos/cm)	pH	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	23.0							Each beaker fed 1.0 ml
7	23.1							YTC suspension
17	23.1							Initials: <u>LB</u>
18	23.1							
								Water changed in all
								beakers.
								Time: <u>0500</u>
								Initials: <u>LB</u>
								Water changed in all
								beakers.
								Time: <u>1610</u>
								Initials: <u>LB</u>

*Water quality measurements to be taken.

Test No 862-1 Client Hart Crowser Investigator _____

DAILY RECORD SHEET

Day 14 (2/6/15) YV/bs

Beaker No.	Temp.* (deg.C)	DO* (ppm)	Cond. (umhos/cm)	pH*	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	23.9	7.0		7.6				Each beaker fed 1.0 ml
7	23.6	6.7		7.3				YTC suspension
17	23.4	6.6		7.3				Initials: <u>AS</u>
18	23.4	6.7		7.3				
								Water changed in all
								beakers.
								Time: <u>0505</u>
								Initials: <u>AS</u>
								Water changed in all
								beakers.
								Time: <u>1610</u>
								Initials: <u>Y</u>

*Water quality measurements to be taken.

Day 15 (2/7/15) YV

Beaker No.	Temp.* (deg.C)	DO (ppm)	Cond. (umhos/cm)	pH	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	22.4							Each beaker fed 1.0 ml
7	22.5							YTC suspension
17	22.3							Initials: <u>AS</u>
18	22.3							
								Water changed in all
								beakers.
								Time: <u>0510</u>
								Initials: <u>AS</u>
								Water changed in all
								beakers.
								Time: <u>1615</u>
								Initials: <u>YV</u>

*Water quality measurements to be taken.

Test No 862-1 Client Hart Crowser Investigator _____

DAILY RECORD SHEET

Day 16 (2/8/15) GSJ

Beaker No.	Temp.* (deg.C)	DO (ppm)	Cond. (umhos/cm)	pH	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	22.2							Each beaker fed 1.0 ml
7	22.1							YTC suspension
17	22.1							Initials: <u>GSJ</u>
18	22.0							
								Water changed in all beakers.
								Time: <u>0505</u>
								Initials: <u>GSJ</u>
								Water changed in all beakers.
								Time: <u>1605</u>
								Initials: <u>GSJ</u>

*Water quality measurements to be taken.

Day 17 (2/9/15) GSJ

Beaker No.	Temp.* (deg.C)	DO* (ppm)	Cond. (umhos/cm)	pH*	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	22.6	6.8		7.3				Each beaker fed 1.0 ml
7	22.4	6.6		7.1				YTC suspension
17	22.3	6.6		7.1				Initials: <u>GSJ</u>
18	22.3	6.9		7.1				
								Water changed in all beakers.
								Time: <u>0510</u>
								Initials: <u>GSJ</u>
								Water changed in all beakers.
								Time: <u>1649</u>
								Initials: <u>GSJ</u>

*Water quality measurements to be taken.

HYALELLA AZTECA 28-DAY SOLID PHASE SEDIMENT TEST

Test No 862-1 Client Hart Crowser Investigator _____

DAILY RECORD SHEET

Day 18 (2/10/15) JS

Beaker No.	Temp.* (deg.C)	DO (ppm)	Cond. (umhos/cm)	pH	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	22.2							Each beaker fed 1.0 ml
7	22.0							YTC suspension
17	22.0							Initials: <u>JS</u>
18	22.0							
								Water changed in all
								beakers.
								Time: <u>0505</u>
								Initials: <u>JS</u>
								Water changed in all
								beakers.
								Time: <u>1620</u>
								Initials: <u>JS</u>

*Water quality measurements to be taken.

Day 19 (2/11/15) JS

Beaker No.	Temp.* (deg.C)	DO* (ppm)	Cond.* (umhos/cm)	pH*	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	22.1	6.8	117	7.2				Each beaker fed 1.0 ml
7	22.1	6.6	111	7.3				YTC suspension
17	22.2	6.7	113	7.3				Initials: <u>JS</u>
18	22.2	6.6	110	7.2				
								Water changed in all
								beakers.
								Time: <u>0510</u>
								Initials: <u>JS</u>
								Water changed in all
								beakers.
								Time: <u>1605</u>
								Initials: <u>JS</u>

*Water quality measurements to be taken.

Test No 862-1 Client Hart Crowser Investigator _____

DAILY RECORD SHEET

Day 20 (2/12/15) ✓

Beaker No.	Temp.* (deg.C)	DO (ppm)	Cond. (umhos/cm)	pH	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	22.0							Each beaker fed 1.0 ml
7	22.0							YTC suspension
17	22.0							Initials: ✓
18	22.0							
								Water changed in all beakers.
								Time: 0510
								Initials: ✓
								Water changed in all beakers.
								Time: 1630
								Initials: JS

*Water quality measurements to be taken.

Day 21 (2/13/15) ✓

Beaker No.	Temp.* (deg.C)	DO* (ppm)	Cond. (umhos/cm)	pH*	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	22.7	7.3		7.5				Each beaker fed 1.0 ml
7	22.6	7.1		7.2				YTC suspension
17	22.4	6.7		7.2				Initials: JS
18	22.3	7.1		7.2				
								Water changed in all beakers.
								Time: 0505
								Initials: JS
								Water changed in all beakers.
								Time: 1615
								Initials: ✓

*Water quality measurements to be taken.

Test No 862-1 Client Hart Crowser Investigator _____

DAILY RECORD SHEET

Day 22 (2/14/15) MS

Beaker No.	Temp.* (deg.C)	DO (ppm)	Cond. (umhos/cm)	pH	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	22.1							Each beaker fed 1.0 ml
7	22.1							YTC suspension
17	22.0							Initials: <u>MS</u>
18	22.0							
								Water changed in all beakers.
								Time: <u>0520</u>
								Initials: <u>MS</u>
								Water changed in all beakers.
								Time: <u>1630</u>
								Initials: <u>MS</u>

*Water quality measurements to be taken.

Day 23 (2/15/15) MS

Beaker No.	Temp.* (deg.C)	DO (ppm)	Cond. (umhos/cm)	pH	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	22.3							Each beaker fed 1.0 ml
7	22.3	22.4						YTC suspension
17	22.3							Initials: <u>MS</u>
18	22.2							
								Water changed in all beakers.
								Time: <u>0510</u>
								Initials: <u>MS</u>
								Water changed in all beakers.
								Time: <u>1620</u>
								Initials: <u>MS</u>

*Water quality measurements to be taken.

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2-15-15

Test No. 862-1 Client Hart Crowser Investigator _____

DAILY RECORD SHEET

Day 24 (2/16/15) BSJ

Beaker No.	Temp.* (deg.C)	DO* (ppm)	Cond. (umhos/cm)	pH*	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	22.4	6.9		7.4				Each beaker fed 1.0 ml
7	22.2	6.6		7.2				YTC suspension
17	22.2	6.6		7.2				Initials: <u>BSJ</u>
18	22.1	6.8		7.2				
								Water changed in all beakers.
								Time: <u>0510</u>
								Initials: <u>BSJ</u>
								Water changed in all beakers.
								Time: <u>1600</u>
								Initials: <u>US</u>

*Water quality measurements to be taken.

Day 25 (2/17/15) US/BSJ

Beaker No.	Temp.* (deg.C)	DO (ppm)	Cond. (umhos/cm)	pH	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	22.7							Each beaker fed 1.0 ml
7	22.4							YTC suspension
17	22.4							Initials: <u>US</u>
18	22.2							
								Water changed in all beakers.
								Time: <u>0540</u>
								Initials: <u>US</u>
								Water changed in all beakers.
								Time: <u>1610</u>
								Initials: <u>BSJ</u>

*Water quality measurements to be taken.

Test No 862-1 Client Hart Crowser Investigator _____

DAILY RECORD SHEET

Day 26 (2/18/15) LS

Beaker No.	Temp.* (deg.C)	DO* (ppm)	Cond.* (umhos/cm)	pH*	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	22.9	7.2	118	7.2				Each beaker fed 1.0 ml
7	22.9	6.4	114	7.1				YTC suspension
17	22.8	6.3	115	7.1				Initials: <u>LS</u>
18	22.9	6.9	112	7.2				
								Water changed in all
								beakers.
								Time: <u>0505</u>
								Initials: <u>LS</u>
								Water changed in all
								beakers.
								Time: <u>1610</u>
								Initials: <u>LS</u>

*Water quality measurements to be taken.

Day 27 (2/19/15) LS

Beaker No.	Temp.* (deg.C)	DO (ppm)	Cond. (umhos/cm)	pH	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
3	22.0							Each beaker fed 1.0 ml
7	22.4							YTC suspension
17	22.4							Initials: <u>LS</u>
18	22.3							
								Water changed in all
								beakers.
								Time: <u>0505</u>
								Initials: <u>LS</u>
								Water changed in all
								beakers.
								Time: <u>1610</u>
								Initials: <u>LS</u>

*Water quality measurements to be taken.

Test No. 862-1 Client Hart Crowser Investigator _____

ZERO-TIME WEIGHING DATA SHEET

Tare: Date 1-22-15 Oven temp (C.) 61 Drying time (hr.) 24 Initials JRF
 Standard Weights: 10 mg: 10.007 100mg: 100.014

Final: Date 1-26-15 Oven temp (C.) 62 Drying time (hr.) 24 Initials JRF
 Standard Weights: 10 mg: 10.008 100mg: 100.015

Equip. used: Oven: Blue m #1 Balance: Sartorius M3P

(Dry overnight at 60-90 degrees C)

Pan #	Tare wt. (mg)	Total wt. (mg)	#weighed	Comments
1	32.244	32.891	10	
2	30.550	31.202	10	
3	41.342	42.029	10	
4	35.318	35.956	10	
5	29.978	30.727	10	

Test No. 862-1 Client Hart Crowser Investigator _____

WEIGHING DATA SHEET

Tare: Date 1-22-15 Oven temp (C.) 61 Drying time (hr.) 24 Initials JRF
Standard Weights: 10 mg: 10.007 100mg: 100.014

Final #1: Date 2-15-15 Oven temp (C.) 62 Drying time (hr.) 24 Initials JRF
Standard Weights: 10 mg: 10.008 100mg: 100.019

Final #2: Date 2-24-15 Oven temp (C.) 64 Drying time (hr.) 24 Initials JRF
Standard Weights: 10 mg: 10.007 100mg: 100.018

Equip. used: Oven BLUE M #1 Balance Sartorius M3P
(Dry overnight at 60-90 degrees C)

Bkr. #	Pan #	Tare wt. (mg)	Total wt. (mg)		no. weighed	put into pans-initials	Comments
			1	2			
1	1	34.035	37.127 37.049	37.049	10	LS	
2	2	33.915	36.436 36.436	36.374	9	LS	
3	3	32.674	35.888 35.812	35.812	10	LS	
4	4	33.239	37.397	37.301	10	LS	35.003 JRF
5	5	32.927	36.2141 36.075	36.075	9	LS	
6	6	31.841	34.039	33.992	4	LS	
7	7	32.235	35.172	35.104	10	LS	
8	8	31.388	34.274	34.219	9	LS	
9	9	32.283	34.618	34.580	9	LS	
10	10	31.860	35.142	35.088	10	LS	
11	11	32.240	35.003	34.951	9	LS	
12	12	32.070	34.617	34.577	8	LS	
13	13	33.570	36.182	36.144	10	LS	
14	14	33.263	36.391	36.367	10	LS	
15	15	36.412	40.377	40.319	10	LS	
16	16	31.668	34.408	34.388	8	LS	
17	17	34.924	36.554	36.534	5	LS	
18	18	35.345	37.587	37.558	7	LS	
19	19	28.557	30.975	30.941	8	LS	
20	20	33.640	36.293	36.258	9	LS	
21	21	34.089	38.001	37.952	10	LS	
22	22	32.079	34.675	34.646	8	LS	
23	23	32.435	36.005	35.941	7	LS	
24	24	34.733	37.900	37.854	10	LS	
25	25	36.924	39.485	39.454	9	LS	
26	26	33.049	34.931	34.913	4	LS	
27	27	38.156	41.077	41.043	6	LS	
28	28	34.510	37.985	37.923	10	LS	
29	29	32.536	35.698	35.627	7	LS	
30	30	34.453	37.205	37.139	10	LS	
31	31	30.870	33.051	32.997	9	LS	
32	32	34.577	37.923	37.857	7	LS	

Chesapeake Cultures

P.O. Box 507 Hayes, VA 23072 (804)693-4046 (804)694-4704 fax
www.c-cultures.com
growfish@c-cultures.com

NAS
Shipment Information

RCVD 1-21-15
✓

Species Hyalella azteca Date 1/20/15
Age ~4-5d.; ~1.5mm P.O. No. verbal
Quantity 530+ Invoice No. 8585

Temperature 24°C Salinity - pH 7.88

Notes Thank you!

Biologist [Signature]

* Please inspect shipment and report any problem immediately *

TEST DATA ANALYSIS RECORDS

data entry verified against
laboratory bench sheets 3-9-15 JAF

Endpoints Data Entry and Calculations File

INDEX	BKR	SMPL	NAS	CLIENT	DESCRIP	REPL	INIT	SURV	MORT	PSURV	PMORT	TARE	WT	COUNT	WT	DRY	TWT	WT	INITIAL WEIGHT				Mean
																			tare	wt	final	wt	
1	24	5195G	Contol	1	10	10	0	100.0	0.0	34.733	10	37.854	3.12	0.31									
2	11	5195G	Contol	2	10	9	1	90.0	10.0	32.240	9	34.951	2.71	0.30									
3	2	5195G	Contol	3	10	9	1	90.0	10.0	33.915	9	36.374	2.46	0.27									
4	5	5195G	Contol	4	10	9	1	90.0	10.0	32.927	9	36.075	3.15	0.35									
5	8	5195G	Contol	5	10	9	1	90.0	10.0	31.388	9	34.219	2.83	0.31	Mean	9.5	0.5	95.0	5.0	0.32			
6	21	5195G	Contol	6	10	10	0	100.0	0.0	34.089	10	37.952	3.86	0.39	SD	0.5	0.5	5.3	5.3	0.03			
7	1	5195G	Contol	7	10	10	0	100.0	0.0	34.035	10	37.049	3.01	0.30	n	8	8	8	8	8			
8	3	5195G	Contol	8	10	10	0	100.0	0.0	32.674	10	35.812	3.14	0.31									
9	28	5192G	JT-SS-06	1	10	10	0	100.0	0.0	34.510	10	37.923	3.41	0.34									
10	27	5192G	JT-SS-06	2	10	6	4	60.0	40.0	38.156	6	41.043	2.89	0.48									
11	26	5192G	JT-SS-06	3	10	4	6	40.0	60.0	33.049	4	34.913	1.86	0.47									
12	32	5192G	JT-SS-06	4	10	7	3	70.0	30.0	34.577	7	37.857	3.28	0.47									
13	12	5192G	JT-SS-06	5	10	8	2	80.0	20.0	32.070	8	34.577	2.51	0.31	Mean	7.8	2.3	77.5	22.5	0.41			
14	4	5192G	JT-SS-06	6	10	10	0	100.0	0.0	33.239	10	37.301	4.06	0.41	SD	2.2	2.2	21.9	21.9	0.08			
15	23	5192G	JT-SS-06	7	10	7	3	70.0	30.0	32.435	7	35.941	3.51	0.50	n	8	8	8	8	8			
16	7	5192G	JT-SS-06	8	10	10	0	100.0	0.0	32.235	10	35.104	2.87	0.29									
17	22	5193G	JT-SS-08	1	10	8	2	80.0	20.0	32.079	8	34.646	2.57	0.32									
18	19	5193G	JT-SS-08	2	10	8	2	80.0	20.0	28.557	8	30.941	2.38	0.30									
19	15	5193G	JT-SS-08	3	10	10	0	100.0	0.0	36.412	10	40.319	3.91	0.39									
20	29	5193G	JT-SS-08	4	10	7	3	70.0	30.0	32.536	7	35.627	3.09	0.44									
21	10	5193G	JT-SS-08	5	10	10	0	100.0	0.0	31.860	10	35.088	3.23	0.32	Mean	8.0	2.0	80.0	20.0	0.37			
22	14	5193G	JT-SS-08	6	10	10	0	100.0	0.0	33.263	10	36.367	3.10	0.31	SD	2.1	2.1	20.7	20.7	0.08			
23	6	5193G	JT-SS-08	7	10	4	6	40.0	60.0	31.841	4	33.992	2.15	0.54	n	8	8	8	8	8			
24	18	5193G	JT-SS-08	8	10	7	3	70.0	30.0	35.345	7	37.558	2.21	0.32									
25	20	5194G	JT-SS-10	1	10	9	1	90.0	10.0	33.640	9	36.258	2.62	0.29									
26	30	5194G	JT-SS-10	2	10	10	0	100.0	0.0	34.453	10	37.139	2.69	0.27									
27	25	5194G	JT-SS-10	3	10	9	1	90.0	10.0	36.924	9	39.454	2.53	0.28									
28	13	5194G	JT-SS-10	4	10	10	0	100.0	0.0	33.570	10	36.144	2.57	0.26									
29	9	5194G	JT-SS-10	5	10	9	1	90.0	10.0	32.263	9	34.580	2.30	0.26	Mean	8.6	1.4	86.3	13.8	0.28			
30	16	5194G	JT-SS-10	6	10	8	2	80.0	20.0	31.668	8	34.388	2.72	0.34	SD	1.6	1.6	16.0	16.0	0.04			
31	31	5194G	JT-SS-10	7	10	9	1	90.0	10.0	30.870	9	32.997	2.13	0.24	n	8	8	8	8	8			
32	17	5194G	JT-SS-10	8	10	5	5	50.0	50.0	34.924	5	36.534	1.61	0.32									

Project Name: P862-1 Hyalella % Mortality

Sample: x1
 Samp ID: JT-SS-06
 Alias: NAS# 5192G
 Replicates: 8
 Mean: 22.5
 SD: 21.876
 Tr Mean: 22.873
 Trans SD: 20.152

Ref Samp: x2
 Ref ID: Control
 Alias: NAS# 5195G
 Replicates: 8
 Mean: 5
 SD: 5.345
 Tr Mean: 9.217
 Trans SD: 9.854

Shapiro-Wilk Results:	Levene's Results:	Test Results:
Residual Mean: 0 Residual SD: 13.616 SS: 3522.352 K: 8 b: 56.462 Alpha Level: 0.05 Calculated Value: 0.9051 Critical Value: ≤ 0.887 Normally Distributed: Yes Override Option: N/A	Test Residual Mean: 17.155 Test Residual SD: 8.352 Ref. Residual Mean: 9.217 Ref. Residual SD: 0 Deg. of Freedom: 14 Alpha Level: 0.1 Calculated Value: 2.688 Critical Value: ≥ 1.761 Variances Homogeneous: No	Statistic: Approximate t Balanced Design: Yes Transformation: ArcSin Experimental Hypothesis Null: $x_1 \leq x_2$ Alternate: $x_1 > x_2$ Degrees of Freedom: 10 Experimental Alpha Level: <u>0.05</u> Calculated Value: 1.7219 Critical Value: ≥ 1.812 <u>Accept Null Hypothesis: Yes</u> Power: Min. Difference for Power:

Replicate Number	Test Data	Trans. Test Data	Reference Data	Trans. Reference Data	Levene's Test Residuals	Levene's Reference Residuals	Mann-Whitney Ranks	Rankits	Shapiro-Wilk Residuals
1	0	0	0	0	22.873	9.217			-22.873
2	40	39.232	10	18.435	16.358	9.217			-22.873
3	60	50.768	10	18.435	27.895	9.217			-22.873
4	30	33.211	10	18.435	10.338	9.217			-9.217
5	20	26.565	10	18.435	3.692	9.217			-9.217
6	0	0	0	0	22.873	9.217			-9.217
7	30	33.211	0	0	10.338	9.217			-9.217
8	0	0	0	0	22.873	9.217			3.692
9									9.217
10									9.217
11									9.217
12									9.217
13									10.338
14									10.338
15									16.358
16									27.895

The percent mortality in test sediment JT-SS-06 was not significantly higher than that of the control sediment at $\alpha=0.05$.

Project Name: P862-1 Hyalella % Mortality

Sample: x1
 Samp ID: JT-SS-08
 Alias: NAS# 5193G
 Replicates: 8
 Mean: 20
 SD: 20.702
 Tr Mean: 21.29
 Trans SD: 19.156

Ref Samp: x2
 Ref ID: Control
 Alias: NAS# 5195G
 Replicates: 8
 Mean: 5
 SD: 5.345
 Tr Mean: 9.217
 Trans SD: 9.854

Shapiro-Wilk Results:	Levene's Results:	Test Results:
Residual Mean: 0 Residual SD: 13.075 SS: 3248.336 K: 8 b: 54.155 Alpha Level: 0.05 Calculated Value: 0.9029 Critical Value: <= 0.887 Normally Distributed: Yes Override Option: N/A	Test Residual Mean: 15.968 Test Residual SD: 8.693 Ref. Residual Mean: 9.217 Ref. Residual SD: 0 Deg. of Freedom: 14 Alpha Level: 0.1 Calculated Value: 2.1963 Critical Value: >= 1.761 Variances Homogeneous: No	Statistic: Approximate t Balanced Design: Yes Transformation: ArcSin Experimental Hypothesis Null: $x_1 \leq x_2$ Alternate: $x_1 > x_2$ Degrees of Freedom: 10 <u>Experimental Alpha Level: 0.05</u> Calculated Value: 1.5851 <u>Critical Value: >= 1.812</u> <u>Accept Null Hypothesis: Yes</u> Power: Min. Difference for Power:

Replicate Number	Test Data	Trans. Test Data	Reference Data	Trans. Reference Data	Levene's Test Residuals	Levene's Reference Residuals	Mann-Whitney Ranks	Rankits	Shapiro-Wilk Residuals
1	20	26.565	0	0	5.275	9.217			-21.29
2	20	26.565	10	18.435	5.275	9.217			-21.29
3	0	0	10	18.435	21.29	9.217			-21.29
4	30	33.211	10	18.435	11.921	9.217			-9.217
5	0	0	10	18.435	21.29	9.217			-9.217
6	0	0	0	0	21.29	9.217			-9.217
7	60	50.768	0	0	29.478	9.217			-9.217
8	30	33.211	0	0	11.921	9.217			5.275
9									5.275
10									9.217
11									9.217
12									9.217
13									9.217
14									11.921
15									11.921
16									29.478

The percent mortality in test sediment JT-SS-08 was not significantly higher than that of the control sediment at $\alpha=0.05$. -651

Project Name: P862-1 Hyalella % Mortality

Sample: x1
 Samp ID: JT-SS-10
 Alias: NAS# 5194G
 Replicates: 8
 Mean: 13.75
 SD: 15.98
 Tr Mean: 18.163
 Trans SD: 14.397

Ref Samp: x2
 Ref ID: Control
 Alias: NAS# 5195G
 Replicates: 8
 Mean: 5
 SD: 5.345
 Tr Mean: 9.217
 Trans SD: 9.854

Shapiro-Wilk Results:	Levene's Results:	Test Results:
Residual Mean: 0 Residual SD: 10.589 SS: 2130.599 K: 8 b: 44.42 Alpha Level: 0.05 Calculated Value: 0.9261 Critical Value: <= 0.887 Normally Distributed: Yes Override Option: N/A	Test Residual Mean: 9.082 Test Residual SD: 10.631 Ref. Residual Mean: 9.217 Ref. Residual SD: 0 Deg. of Freedom: 14 Alpha Level: 0.1 Calculated Value: 0.0362 Critical Value: >= 1.761 Variances Homogeneous: Yes	Statistic: Student's t Balanced Design: Yes Transformation: ArcSin Experimental Hypothesis Null: $x_1 \leq x_2$ Alternate: $x_1 > x_2$ Degrees of Freedom: 14 Experimental Alpha Level: 0.05 Calculated Value: 1.4503 Critical Value: >= 1.761 Accept Null Hypothesis: Yes Power: Min. Difference for Power:

Replicate Number	Test Data	Trans. Test Data	Reference Data	Trans. Reference Data	Levene's Test Residuals	Levene's Reference Residuals	Mann-Whitney Ranks	Rankits	Shapiro-Wilk Residuals
1	10	18.435	0	0	0.272	9.217			-18.163
2	0	0	10	18.435	18.163	9.217			-18.163
3	10	18.435	10	18.435	0.272	9.217			-9.217
4	0	0	10	18.435	18.163	9.217			-9.217
5	10	18.435	10	18.435	0.272	9.217			-9.217
6	20	26.565	0	0	8.402	9.217			-9.217
7	10	18.435	0	0	0.272	9.217			0.272
8	50	45	0	0	26.837	9.217			0.272
9									0.272
10									0.272
11									8.402
12									9.217
13									9.217
14									9.217
15									9.217
16									26.837

The percent mortality in test sediment JT-SS-10 was not significantly higher than that of the control sediment at $\alpha=0.05$. -631

Project Name: P862-1 Hyalella Growth (dry wt)

Sample: x1
 Samp ID: JT-SS-06
 Alias: NAS# 5192G
 Replicates: 8
 Mean: 0.409
 SD: 0.084
 Tr Mean: 0.409
 Trans SD: 0.084

Ref Samp: x2
 Ref ID: Control
 Alias: NAS# 5195G
 Replicates: 8
 Mean: 0.318
 SD: 0.037
 Tr Mean: 0.318
 Trans SD: 0.037

Shapiro-Wilk Results:	Levene's Results:	Test Results:
Residual Mean: 0 Residual SD: 0.056 SS: 0.059 K: 8 b: 0.236 Alpha Level: 0.05 Calculated Value: 0.9453 Critical Value: ≤ 0.887 Normally Distributed: Yes Override Option: N/A	Test Residual Mean: 0.072 Test Residual SD: 0.035 Ref. Residual Mean: 0.026 Ref. Residual SD: 0.023 Deg. of Freedom: 14 Alpha Level: 0.1 Calculated Value: 3.0506 Critical Value: ≥ 1.761 Variances Homogeneous: No	Statistic: Approximate t Balanced Design: Yes Transformation: No Transformation Experimental Hypothesis Null: $x1 \geq x2$ Alternate: $x1 < x2$ Degrees of Freedom: 10 Experimental Alpha Level: 0.05 Calculated Value: -2.8151 Critical Value: ≥ 1.812 Accept Null Hypothesis: Yes Power: Min. Difference for Power:

Replicate Number	Test Data	Trans. Test Data	Reference Data	Trans. Reference Data	Levene's Test Residuals	Levene's Reference Residuals	Mann-Whitney Ranks	Rankits	Shapiro-Wilk Residuals
1	0.34	0.34	0.31	0.31	0.069	0.008			-0.119
2	0.48	0.48	0.3	0.3	0.071	0.018			-0.099
3	0.47	0.47	0.27	0.27	0.061	0.048			-0.069
4	0.47	0.47	0.35	0.35	0.061	0.033			-0.048
5	0.31	0.31	0.31	0.31	0.099	0.008			-0.018
6	0.41	0.41	0.39	0.39	0.001	0.073			-0.018
7	0.5	0.5	0.3	0.3	0.091	0.018			-0.008
8	0.29	0.29	0.31	0.31	0.119	0.008			-0.008
9									-0.008
10									0.001
11									0.033
12									0.061
13									0.061
14									0.071
15									0.073
16									0.091

Average individual growth (dry wt) in test sediment JT-SS-06 is not significantly less than that in the control sediment at $\alpha=0.05$. -631

Project Name: P862-1 Hyalella Growth (dry wt)

Sample: x1
 Samp ID: JT-SS-08
 Alias: NAS# 5193G
 Replicates: 8
 Mean: 0.368
 SD: 0.085
 Tr Mean: 0.413
 Trans SD: 0.884

Ref Samp: x2
 Ref ID: Control
 Alias: NAS# 5195G
 Replicates: 8
 Mean: 0.318
 SD: 0.037
 Tr Mean: -0.413
 Trans SD: 0.862

Shapiro-Wilk Results:	Levene's Results:	Test Results:
Residual Mean: Residual SD: SS: K: b: Alpha Level: N/A Calculated Value: N/A Critical Value: N/A Normally Distributed: N/A Override Option: Not Invoked	Test Residual Mean: 0.672 Test Residual SD: 0.515 Ref. Residual Mean: 0.638 Ref. Residual SD: 0.527 Deg. of Freedom: 14 Alpha Level: 0.1 Calculated Value: 0.1307 Critical Value: ≥ 1.761 Variances Homogeneous: Yes	Statistic: Student's t Balanced Design: Yes Transformation: Rankits Experimental Hypothesis Null: $x_1 \geq x_2$ Alternate: $x_1 < x_2$ Degrees of Freedom: 14 Experimental Alpha Level: 0.05 Calculated Value: -1.8911 Critical Value: ≥ 1.761 Accept Null Hypothesis: Yes Power: Min. Difference for Power:

Replicate Number	Test Data	Trans. Test Data	Reference Data	Trans. Reference Data	Levene's Test Residuals	Levene's Reference Residuals	Mann-Whitney Ranks	Rankits	Shapiro-Wilk Residuals
1	0.32	0.236	0.31	-0.319	0.177	0.094		-1.766	
2	0.3	-1.013	0.3	-1.013	1.425	0.6		-1.013	
3	0.39	0.877	0.27	-1.766	0.464	1.353		-1.013	
4	0.44	1.285	0.35	0.57	0.872	0.983		-1.013	
5	0.32	0.236	0.31	-0.319	0.177	0.094		-0.319	
6	0.31	-0.319	0.39	0.877	0.732	1.289		-0.319	
7	0.54	1.766	0.3	-1.013	1.353	0.6		-0.319	
8	0.32	0.236	0.31	-0.319	0.177	0.094		-0.319	
9								0.236	
10								0.236	
11								0.236	
12								0.57	
13								0.877	
14								0.877	
15								1.285	
16								1.766	

Average individual growth (dry wt) in test sediment JT-SS-08 is not significantly less than that in the control sediment at $\alpha=0.05$. -651

Project Name: P862-1 Hyalella Growth (dry wt)

Sample: x1
 Samp ID: JT-SS-10
 Alias: NAS# 5194G
 Replicates: 8
 Mean: 0.283
 SD: 0.033
 Tr Mean: 0.283
 Trans SD: 0.033

Ref Samp: x2
 Ref ID: Control
 Alias: NAS# 5195G
 Replicates: 8
 Mean: 0.318
 SD: 0.037
 Tr Mean: 0.318
 Trans SD: 0.037

Shapiro-Wilk Results:	Levene's Results:	Test Results:
Residual Mean: 0 Residual SD: 0.03 SS: 0.017 K: 8 b: 0.125 Alpha Level: 0.05 Calculated Value: 0.9106 Critical Value: ≤ 0.887 Normally Distributed: Yes Override Option: N/A	Test Residual Mean: 0.026 Test Residual SD: 0.019 Ref. Residual Mean: 0.026 Ref. Residual SD: 0.023 Deg. of Freedom: 14 Alpha Level: 0.1 Calculated Value: 0.0588 Critical Value: ≥ 1.761 Variances Homogeneous: Yes	Statistic: Student's t Balanced Design: Yes Transformation: No Transformation Experimental Hypothesis Null: $x_1 \geq x_2$ Alternate: $x_1 < x_2$ Degrees of Freedom: 14 Experimental Alpha Level: 0.05 Calculated Value: 2.0029 Critical Value: ≥ 1.761 Accept Null Hypothesis: No Power: Min. Difference for Power:

Replicate Number	Test Data	Trans. Test Data	Reference Data	Trans. Reference Data	Levene's Test Residuals	Levene's Reference Residuals	Mann-Whitney Ranks	Ranks	Shapiro-Wilk Residuals
1	0.29	0.29	0.31	0.31	0.007	0.008			-0.048
2	0.27	0.27	0.3	0.3	0.013	0.018			-0.043
3	0.28	0.28	0.27	0.27	0.003	0.048			-0.023
4	0.26	0.26	0.35	0.35	0.023	0.033			-0.023
5	0.26	0.26	0.31	0.31	0.023	0.008			-0.018
6	0.34	0.34	0.39	0.39	0.058	0.073			-0.018
7	0.24	0.24	0.3	0.3	0.043	0.018			-0.013
8	0.32	0.32	0.31	0.31	0.038	0.008			-0.008
9									-0.008
10									-0.008
11									-0.003
12									0.007
13									0.033
14									0.038
15									0.058
16									0.073

Average individual growth (dry wt) in test sediment JT-SS-10 is significantly less than that in the control sediment at $\alpha=0.05$. — 68 J

Water Quality Data												
BKR	NAS SMPL	CLIENT DESCRIP	REPL	DAY	Overlying water							
					TEMP	DO	COND	pH	NH3	HARD	ALK	
3	5195G	Control	8	0	22.2	7.6	219	6.8	0.9	34	30	
7	5192G	JT-SS-06	8	0	22.2	7.7	113	6.6	0.2	34	30	
17	5194G	JT-SS-10	8	0	22.3	7.6	133	6.6	0.3	34	40	
18	5193G	JT-SS-08	8	0	22.3	7.6	115	6.6	0.1	34	40	
3	5195G	Control	8	1	22.2							
7	5192G	JT-SS-06	8	1	22.3							
17	5194G	JT-SS-10	8	1	22.2							
18	5193G	JT-SS-08	8	1	22.2							
3	5195G	Control	8	2	22.2							
7	5192G	JT-SS-06	8	2	22.2							
17	5194G	JT-SS-10	8	2	22.1							
18	5193G	JT-SS-08	8	2	22.1							
3	5195G	Control	8	3	22.3	7.0		7.1				
7	5192G	JT-SS-06	8	3	22.2	7.0		7.1				
17	5194G	JT-SS-10	8	3	22.1	7.0		7.1				
18	5193G	JT-SS-08	8	3	22.1	7.2		7.2				
3	5195G	Control	8	4	22.6							
7	5192G	JT-SS-06	8	4	22.4							
17	5194G	JT-SS-10	8	4	22.2							
18	5193G	JT-SS-08	8	4	22.2							
3	5195G	Control	8	5	23.0	7.3	139	7.2				
7	5192G	JT-SS-06	8	5	22.8	7.1	124	7.2				
17	5194G	JT-SS-10	8	5	22.7	7.1	124	7.2				
18	5193G	JT-SS-08	8	5	22.7	7.1	120	7.2				
3	5195G	Control	8	6	23.1							
7	5192G	JT-SS-06	8	6	23.0							
17	5194G	JT-SS-10	8	6	22.8							
18	5193G	JT-SS-08	8	6	22.8							
3	5195G	Control	8	7	22.5	7.3		7.1				
7	5192G	JT-SS-06	8	7	22.4	6.9		7.0				
17	5194G	JT-SS-10	8	7	22.2	6.7		7.0				
18	5193G	JT-SS-08	8	7	22.2	6.8		7.1				
3	5195G	Control	8	8	22.5							
7	5192G	JT-SS-06	8	8	22.2							
17	5194G	JT-SS-10	8	8	22.2							
18	5193G	JT-SS-08	8	8	22.3							
3	5195G	Control	8	9	22.3							
7	5192G	JT-SS-06	8	9	22.1							
17	5194G	JT-SS-10	8	9	22.0							
18	5193G	JT-SS-08	8	9	22.1							
3	5195G	Control	8	10	22.4	7.4		7.2				
7	5192G	JT-SS-06	8	10	22.2	7.4		7.1				
17	5194G	JT-SS-10	8	10	22.1	7.0		7.1				
18	5193G	JT-SS-08	8	10	22.1	7.1		7.1				
3	5195G	Control	8	11	22.6							
7	5192G	JT-SS-06	8	11	22.7							
17	5194G	JT-SS-10	8	11	22.5							
18	5193G	JT-SS-08	8	11	22.5							

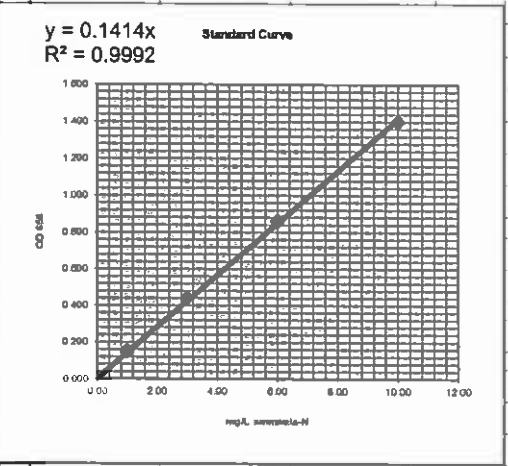
3	5195G	Control	8	12	22.6	6.3	126	7.1
7	5192G	JT-SS-06	8	12	22.6	6.3	119	7.0
17	5194G	JT-SS-10	8	12	22.4	6.1	122	7.0
18	5193G	JT-SS-08	8	12	22.4	6.2	119	7.0
3	5195G	Control	8	13	23.0			
7	5192G	JT-SS-06	8	13	23.1			
17	5194G	JT-SS-10	8	13	23.1			
18	5193G	JT-SS-08	8	13	23.1			
3	5195G	Control	8	14	23.9	7.0		7.6
7	5192G	JT-SS-06	8	14	23.6	6.7		7.3
17	5194G	JT-SS-10	8	14	23.4	6.6		7.3
18	5193G	JT-SS-08	8	14	23.4	6.7		7.3
3	5195G	Control	8	15	22.4			
7	5192G	JT-SS-06	8	15	22.5			
17	5194G	JT-SS-10	8	15	22.3			
18	5193G	JT-SS-08	8	15	22.3			
3	5195G	Control	8	16	22.2			
7	5192G	JT-SS-06	8	16	22.1			
17	5194G	JT-SS-10	8	16	22.1			
18	5193G	JT-SS-08	8	16	22.0			
3	5195G	Control	8	17	22.6	6.8		7.3
7	5192G	JT-SS-06	8	17	22.4	6.6		7.1
17	5194G	JT-SS-10	8	17	22.3	6.6		7.1
18	5193G	JT-SS-08	8	17	22.3	6.9		7.1
3	5195G	Control	8	18	22.2			
7	5192G	JT-SS-06	8	18	22.0			
17	5194G	JT-SS-10	8	18	22.0			
18	5193G	JT-SS-08	8	18	22.0			
3	5195G	Control	8	19	22.1	6.8	117	7.2
7	5192G	JT-SS-06	8	19	22.1	6.6	111	7.3
17	5194G	JT-SS-10	8	19	22.2	6.7	113	7.3
18	5193G	JT-SS-08	8	19	22.2	6.6	110	7.2
3	5195G	Control	8	20	22.0			
7	5192G	JT-SS-06	8	20	22.0			
17	5194G	JT-SS-10	8	20	22.0			
18	5193G	JT-SS-08	8	20	22.0			
3	5195G	Control	8	21	22.7	7.3		7.5
7	5192G	JT-SS-06	8	21	22.6	7.1		7.2
17	5194G	JT-SS-10	8	21	22.4	6.7		7.2
18	5193G	JT-SS-08	8	21	22.3	7.1		7.2
3	5195G	Control	8	22	22.1			
7	5192G	JT-SS-06	8	22	22.1			
17	5194G	JT-SS-10	8	22	22.0			
18	5193G	JT-SS-08	8	22	22.0			
3	5195G	Control	8	23	22.3			
7	5192G	JT-SS-06	8	23	22.4			
17	5194G	JT-SS-10	8	23	22.3			
18	5193G	JT-SS-08	8	23	22.2			
3	5195G	Control	8	24	22.4	6.9		7.4
7	5192G	JT-SS-06	8	24	22.2	6.6		7.2
17	5194G	JT-SS-10	8	24	22.2	6.6		7.2
18	5193G	JT-SS-08	8	24	22.1	6.8		7.2

3	5195G	Control	8	25	22.7							
7	5192G	JT-SS-06	8	25	22.4							
17	5194G	JT-SS-10	8	25	22.4							
18	5193G	JT-SS-08	8	25	22.2							
3	5195G	Control	8	26	22.9	7.2	118	7.2				
7	5192G	JT-SS-06	8	26	22.9	6.4	114	7.1				
17	5194G	JT-SS-10	8	26	22.8	6.3	115	7.1				
18	5193G	JT-SS-08	8	26	22.9	6.9	112	7.2				
3	5195G	Control	8	27	22.0							
7	5192G	JT-SS-06	8	27	22.4							
17	5194G	JT-SS-10	8	27	22.4							
18	5193G	JT-SS-08	8	27	22.3							
3	5195G	Control	8	28	22.7	8.5	122	8.0	<0.1	34		
7	5192G	JT-SS-06	8	28	22.6	7.5	113	7.3	<0.1	26		
17	5194G	JT-SS-10	8	28	22.6	7.5	118	7.3	<0.1	34		
18	5193G	JT-SS-08	8	28	22.5	8.1	113	7.5	<0.1	26		
					Mean	22.4	7.0	123	7.2	---	32	35
					SD	0.4	0.5	22	0.2	---	4	5
					n	116	52	24	52	8	8	8
					Min	22.0	6.1	110	6.6	<0.1	26	30
					Max	23.9	8.5	219	8.0	0.9	34	40

AMMONIA EXPOSURE BENCHSHEETS AND ANALYSIS

Total Ammonia-N in Water: Computation Worksheet Salicylate Method (SOP #5492)

Result						
Sample description	Dilution factor	OD ₆₅₅	NH ₃ -N (mg/L)	pH	Salinity (ppt)	
Blank	----	----	----			
1.0 mg/L NH ₃ -N Std.	----	0.151	1.00			
3.0 mg/L NH ₃ -N Std.	----	0.440	3.00			
6.0 mg/L NH ₃ -N Std.	----	0.862	6.00			
10.0 mg/L NH ₃ -N Std.	----	1.400	10.00			
3.0 mg/L spike	----	0.449	3.17			
3.0 mg/L spike dupl.	----	0.440	3.11			
5.0 mg/L 2nd source		0.710	5.02			
1 Day 0 (1-23-15)						
2 3	1	0.120	0.85			
3 7	1	0.023	0.16			
4 17	1	0.040	0.28			
5 18	1	0.020	0.14			
6						
7 Day 28 (2-20-15)						
8 3	1	0.009	ND			
9 7	1	0.002	ND			
10 17	1	0.000	ND			
11 18	1	0.000	ND			
12						
13						
14						
15						
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34						
35						
36						



Reporting limit (mg/L) = 0.10

Recovery (%) = 104.8

Precision (RPD) = 2.02

2nd source (%) = 100.4

Sample volume (ml): 0.50

Dilution factor 1

Sample Set Description:
 Test No.: 862-1
 Test Day: 0, & 28
 Species: *Hyaella*

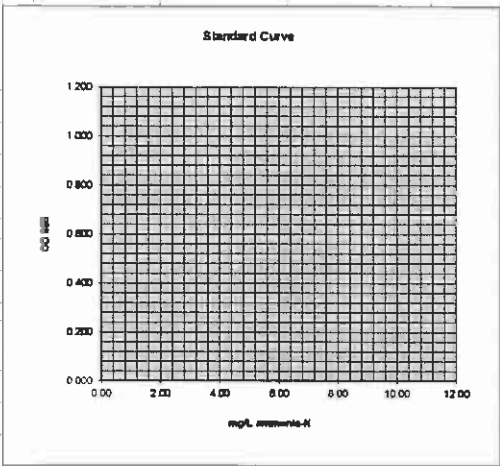
Sample Type (check)
 Bulk Sediment Porewaters
 Test Beaker Porewaters
 Overlying Water

Analyst: JB
 Date analysed: 2/20/2015

Total Ammonia-N in Water: Computation Worksheet Salicylate Method (SOP #5492)

Result

Sample description	Dilution factor	OD655	NH ₃ -N (mg/L)	pH	Salinity (ppt)
Blank	----	----	----		
1.0 mg/L NH ₃ -N Std.	----	.151	1.00		
3.0 mg/L NH ₃ -N Std.	----	.440	3.00		
6.0 mg/L NH ₃ -N Std.	----	.862	6.00		
10.0 mg/L NH ₃ -N Std.	----	1.4	10.00		
3.0 mg/L spike	----	.449			
3.0 mg/L spike dupl.	----	.440			
5.0 mg/L 2nd source		.710			
1 Day 0 (1-23-15)					
2 3	1	.012			
3 7	1	.023			
4 17	1	.040			
5 18	1	.020			
6					
7 Day 28 (2-20-15)					
8 3	1	.009			
9 7	1	.002			
10 17	1	.000			
11 18	1	.000			
12					
13					
14					
15					
16					
17					
18					
19					
20					
21					
22					
23					
24					
25					
26					
27					
28					
29					
30					
31					
32					
33					
34					
35					
36					



Reporting limit (mg/L) = 0.10

Recovery (%) = #VALUE!
 Precision (RPD) = #VALUE!
 2nd source (%) = #VALUE!

Sample volume (ml): 0.50
 Dilution factor: 1

Sample Set Description:
 Test No.: 862-1
 Test Day: 0, & 28
 Species: *Hyaella*

Sample Type (check)
 Bulk Sediment Porewaters
 Test Beaker Porewaters
 Overlying Water

Analyst: JB
 Date analysed: 2/20/2015

CHAIN-OF-CUSTODY RECORDS

Sample Custody Record

Samples Shipped to: Alu Aquatic Services



Hart Crowser, Inc.
1700 Westlake Avenue North, Suite 200
Seattle, Washington 98109-6212
Office: 206.324.9530 • Fax 206.328.5581

JOB 17800-56 LAB NUMBER _____
PROJECT NAME Jacobson Terminals
HART CROWSER CONTACT Phil Cordeak
SAMPLED BY: PRC

REQUESTED ANALYSIS	
10-day <i>Vibrio</i> survival	X
28-day <i>Vibrio</i> survival	X
and growth	X
and growth	X

LAB NO.	SAMPLE ID	DESCRIPTION	DATE	TIME	MATRIX	NO. OF CONTAINERS	OBSERVATIONS/COMMENTS/ COMPOSITING INSTRUCTIONS
JT-95-06	51926	51926	11/2/15	1159	Sediment	4	4 liters sediment
JT-95-08	51936	51936	↓	1058	↓	4	6 "
JT-95-10	51946	51946	↓	1400	↓	4	11 "

RELINQUISHED BY: Phil Cordeak
SIGNATURE
PRINT NAME
COMPANY HIC

RECEIVED BY: Gerald Lissman
SIGNATURE
PRINT NAME
COMPANY NAS

DATE: 11/13/15 TIME: 1000

DATE: 1-14-15 TIME: 1140

RECEIVED BY: _____
SIGNATURE
PRINT NAME
COMPANY _____

DATE: _____ TIME: _____

RECEIVED BY: _____
SIGNATURE
PRINT NAME
COMPANY _____

DATE: _____ TIME: _____

SPECIAL SHIPMENT HANDLING OR STORAGE REQUIREMENTS:
None

TOTAL NUMBER OF CONTAINERS: 12

SAMPLE RECEIPT INFORMATION
CUSTODY SEALS: YES NO N/A
GOOD CONDITION YES NO
TEMPERATURE 2.5 °C
SHIPMENT METHOD: HAND OVERNIGHT
 COURIER

TURNAROUND TIME:
 24 HOURS 1 WEEK
 48 HOURS STANDARD
 72 HOURS OTHER _____

From: (206) 826-4527
Phil Corde!
Hart Crowser, Inc.
1700 Westlake Avenue North
Suite 200
Seattle, WA 98109

Origin ID LKEA



Ship Date: 13JAN15
ActWgt: 75.0 LB
CAD: 4598184/NET3550

Delivery Address Bar Code



SHIP TO: (206) 324-9530
Gerald Irissarri
Northwest Aquatics Scientlsts
3814 Yakima Rd

BILL SENDER

Ref # 17800-56-02
Invoice #
PO #
Dept #

NEWPORT, OR 97365

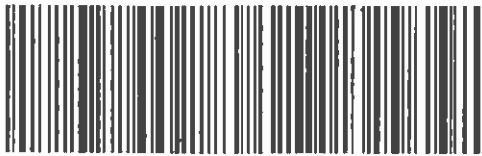
WED - 14 JAN AM
STANDARD OVERNIGHT

TRK# 7725 6121 8674
1 8201



86 ONPA

97365
OR-US
PDX



52216F 15RAC9

Handwritten: 12 CVD 1130 1-14-15

After printing this label

1. Use the 'Print' button on this page to print your label to your laser or inkjet printer
2. Fold the printed page along the horizontal line.
3. Place label in shipping pouch and affix it to your shipment so that the barcode portion of the label can be read and scanned

Warning: Use only the printed original label for shipping. Using a photocopy of this label for shipping purposes is fraudulent and could result in additional billing charges, along with the cancellation of your FedEx account number.

Use of this system constitutes your agreement to the service conditions in the current FedEx Service Guide, available on fedex.com. FedEx will not be responsible for any claim in excess of \$100 per package, whether the result of loss, damage, delay, non-delivery, misdelivery, or misinformation, unless you declare a higher value, pay an additional charge, document your actual loss and file a timely claim. Limitations found in the current FedEx Service Guide apply. Your right to recover from FedEx for any loss, including intrinsic value of the package, loss of sales, income interest, profit, attorney's fees, costs, and other forms of damage whether direct, incidental, consequential, or special is limited to the greater of \$100 or the authorized declared value. Recovery cannot exceed actual documented loss. Maximum for items of extraordinary value is \$1,000, e.g. jewelry, precious metals, negotiable instruments and other items listed in our ServiceGuide. Written claims must be filed within strict time limits, see current FedEx Service Guide.

CUSTODY SEAL

CUSTODY SEAL

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Date 1/13/15

Initials [Signature]

Handwritten: 37 of 37

APPENDIX III

RAW DATA – REFERENCE TOXICANT TEST

REVIEWED
PAGES 1-8
-601

Test No. 999-3381 Client: QC Test Investigator _____
 Test Type (ranging/definitive) definitive Test Length (hr) 96
 Species Hyalella azteca

STUDY MANAGEMENT

Client: QC test
 Client's Study Monitor: QC test
 Testing Laboratory: Northwestern Aquatic Sciences
 Test Location: Newport Laboratory
 Laboratory's Study Personnel:
 Proj. Man./Study Dir. G.J. Irissarri⁶⁰¹
 QA Officer L. K. Nemeth
 1. Yes Nakahama 2. GA Butler
 3. _____ 4. _____
 Test Beginning: 1-23-15 0915 Test Ending: 1-27-15 1030

TEST MATERIAL

Description: Potassium Chloride Crystals - Lot No.: ^{Fisher} 114689
 NAS Sample No. _____
 Date of Collection: _____
 Date of Receipt: _____
 Temperature (deg C): _____
 Dissolved oxygen (mg/L): _____
 pH: _____
 Conductivity (umhos/cm): _____
 Hardness (mg/L): _____
 Alkalinity (mg/L): _____
 Salinity (ppt): _____
 Total chlorine (mg/L): _____
 Total ammonia-N (mg/L): _____

DILUTION WATER

Description: Moderately hard synthetic water
 Date of Preparation/Collection: 1-15-15
 Water Quality: Cond. (umhos/cm): 283 Salinity (ppt) _____ pH 7.5
 Hardness (mg/L as CaCO₃): 77 Alkalinity (mg/L as CaCO₃): 70
 Treatments: Aerated ≥ 24 hrs

TEST LOCATION

Test conducted in (circle one): room 1 room 2 trailer water bath other: _____

Randomization chart:

0.5	0.125	0.063	1.0	0.25	∅				
0.125	0.25	∅	0.063	0.5	1.0				

Error codes: 1) Correction of handwriting error
 2) Written in wrong location; entry deleted
 3) Wrong date deleted; replaced with correct date
 4) Error found in measurement; measurement repeated

Test No. 999-3381 Client _____ QC Test _____ Investigator _____

TEST ORGANISMS

Species: Hyalella azteca Age: 7-8 DAYS Size: _____
Source: Chesapeake Cultures, Hayes, VA Date received: 1-21-15

Acclimation Data:

Date	Temp. (deg.C)	pH	DO (mg/L)	Cond. umhos/cm	Hardness (mg/L)	Alkalinity (mg/L)	Feeding		Water changes
							Amount	description	
1-21-15	17.5	6.8	>15.0	596	188	180	10mls	4 hr	rec'd data - 405 ± 30 °C
1-22-15	20.2	7.3	8.9	255	77	70	"	"	"
1-23-15	20.7	7.5	8.4	223	68	60	"	"	"
Mean	19.5	7.2	10.8	358	111	103			
S.D.	1.7	0.4	3.7	20.7	6.7	6.7			
(N)	3	3	3	3	3	3			

Photoperiod during acclimation: 16:8, L:D

TEST PROCEDURES AND CONDITIONS

Test concentrations (50% series recommended): 1, 0.5, 0.25, 0.125, 0.063 0 g/L

Test chamber: 250 ml glass beakers Test volume: 100 ml
Replicates/treatment: 2 Organisms/treatment: 20 (10/rep)
Test water changes: None Aeration during test: None
Feeding: 0.5 ml YTC suspension per beaker on days 0 and 2

Duration: 24-hr, 48-hr, 96-hr Test temperature (deg.C): 23 ± 1 or 20 ± 1
Beaker placement: Stratified randomization Photoperiod: 16:8, L:D

MISCELLANEOUS NOTES

Test solution preparation:

Working stock: Dissolve 0.5g KCl crystals in dilution water and dilute to 500 mL.
Final conc.: 1.0 g/L.

1-23-15 (99)

Test concentration (g/L)	KCl working stock (ml/200ml)	Dilution water
1	200	Brought up to final volume of 200 ml with dilution water and distributed evenly between two replicates
0.5	100	
0.25	50	
0.125	25	
0.063	12.5	
0	0	

Test No. 999-3381 Client _____ QC Test _____

DAILY RECORD SHEET

Day 0 (1/23/15) AS/CS

Conc. (g/L)	Temp. (deg.C)	pH	Cond. (umhos/cm)	DO (ppm)	Hardness (mg/L)	Alkalinity (mg/L)	Survivors	
							A	B
1. 1	22.7	7.6	1990	8.5	86	70	10	10
2. 0.5	22.7	7.4	1160	8.6			10	10
3. 0.25	22.7	7.3	728	8.7			10	10
4. 0.125	22.8	7.3	510	8.6			10	10
5. 0.063	22.7	7.1	395	8.6			10	10
6. 0	22.4	7.1	282	8.5	86	60	10	10

Each beaker fed 0.5 ml YTC suspension. Initials: AS

Day 1 (1/24/15) AS

Conc. (g/L)	Temp. (deg.C)	pH	Cond. (umhos/cm)	DO (ppm)	Hardness (mg/L)	Alkalinity (mg/L)	Survivors	
							A	B
1. 1	23.2	7.7	1980	8.2			4(10)	5(50)
2. 0.5	23.2	7.6	1208	8.2			10	8(20)
3. 0.25	23.2	7.6	747	8.5			10	10
4. 0.125	23.2	7.6	521	8.3			10	10
5. 0.063	23.1	7.6	412	8.4			10	10
6. 0	23.0	7.5	300	8.2			10	10

Day 2 (1/25/15) CS

Conc. (g/L)	Temp. (deg.C)	pH	Cond. (umhos/cm)	DO (ppm)	Hardness (mg/L)	Alkalinity (mg/L)	Survivors	
							A	B
1. 1	23.3	7.8	2030	8.0			0(40)	0(50)
2. 0.5	23.4	7.8	1264	8.0			7(20)	4(40)
3. 0.25	23.3	7.8	773	8.1			10	10
4. 0.125	23.4	7.7	555	8.1			10	10
5. 0.063	23.4	7.7	442	8.2			10	10
6. 0	23.3	7.5	309	8.2			10	10

Each beaker fed 0.5 ml YTC suspension. Initials: CS

Day 3 (1/26/15) CS

Conc. (g/L)	Temp. (deg.C)	pH	Cond. (umhos/cm)	DO (ppm)	Hardness (mg/L)	Alkalinity (mg/L)	Survivors	
							A	B
1. 1	-	-	-	-				
2. 0.5	23.6	7.7	1282	7.6			INADVERTENTLY	
3. 0.25	23.6	7.7	801	7.6			NOT ASSESSED	
4. 0.125	23.6	7.6	560	7.6			-CS	
5. 0.063	23.5	7.6	454	7.7				
6. 0	23.4	7.5	316	7.8				

Day 4 (1/29/15) CS

Conc. (g/L)	Temp. (deg.C)	pH	Cond. (umhos/cm)	DO (ppm)	Hardness (mg/L)	Alkalinity (mg/L)	Survivors	
							A	B
1. 1	-	-	-	-			0	0
2. 0.5	23.3	7.9	1269	8.1			7	3(10)
3. 0.25	23.2	7.9	785	8.1			10	10
4. 0.125	23.2	7.7	536	8.1			10	10
5. 0.063	23.3	7.8	446	8.1			10	10
6. 0	23.1	7.6	338	8.1	86	70	10	10

Mean
SD
n

(SEE PAGE 5)

Chesapeake Cultures

P.O. Box 507 Hayes, VA 23072 (804)693-4046 (804)694-4704 fax

www.c-cultures.com
growfish@c-cultures.com

NAS
Shipment Information

RCVD 1-21-15
✓

Species Hyalella azteca Date 1/20/15
Age ~4-5 d.; ~1.5 mm P.O. No. verbal
Quantity 530+ Invoice No. 8585

Temperature 24°C Salinity - pH 7.88

Notes Thank you!

Biologist [Signature]

* Please inspect shipment and report any problem immediately *

Hyalella Acute wq 999-3381

Water Quality Data - test #999-338 Hyalella KCl QC test							
Day	Concentration (g/L)	Temperature	pH	Conductivity	DO	Hardness	Alkalinity
0	1	22.7	7.6	1990	8.5	86	70
0	0.5	22.7	7.4	1160	8.6		
0	0.25	22.7	7.3	728	8.7		
0	0.125	22.8	7.3	510	8.6		
0	0.063	22.7	7.1	395	8.6		
0	0	22.4	7.1	282	8.5	86	60
1	1	23.2	7.7	1980	8.2		
1	0.5	23.2	7.6	1208	8.2		
1	0.25	23.2	7.6	747	8.5		
1	0.125	23.2	7.6	521	8.3		
1	0.063	23.1	7.6	412	8.4		
1	0	23.0	7.5	300	8.2		
2	1	23.3	7.8	2080	8.0		
2	0.5	23.4	7.8	1264	8.0		
2	0.25	23.3	7.8	793	8.1		
2	0.125	23.4	7.7	555	8.1		
2	0.063	23.4	7.7	442	8.2		
2	0	23.3	7.5	309	8.2		
3	1						
3	0.5	23.6	7.7	1282	7.6		
3	0.25	23.6	7.7	804	7.6		
3	0.125	23.6	7.6	560	7.6		
3	0.063	23.5	7.6	454	7.7		
3	0	23.4	7.5	316	7.8		
4	1						
4	0.5	23.3	7.9	1269	8.1		
4	0.25	23.2	7.9	785	8.1		
4	0.125	23.2	7.7	536	8.1		
4	0.063	23.3	7.8	446	8.1		
4	0	23.1	7.6	338	8.1	86	70
	MEAN	23.2	7.6		8.2	86	67
	SD	0.3	0.2		0.3	0	6
	N	28	28		28	3	3
	MIN	22.4	7.1		7.6	86	60
	MAX	23.6	7.9		8.7	86	70
		MEAN 1.0 g/L		2017			
		SD					
		N		3			
		MEAN 0 g/L		309			
		SD		21			
		N		5			

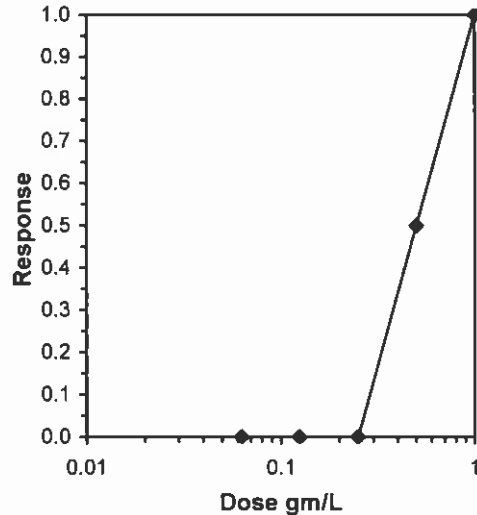
Acute 96-hr Toxicity Test-96 Hr Survival

Start Date: 1/23/2015 09:15 Test ID: 999-3381 Sample ID: REF-Ref Toxicant
 End Date: 1/27/2015 10:30 Lab ID: ORNAS-Northwestern Aquati Sample Type: KCL-Potassium chloride
 Sample Date: Protocol: NASXXXHA1-Hyalaea azteca Test Species: HA-Hyalaea azteca
 Comments:

Conc-gm/L	1	2
D-Control	1.0000	1.0000
0.063	1.0000	1.0000
0.125	1.0000	1.0000
0.25	1.0000	1.0000
0.5	0.7000	0.3000
1	0.0000	0.0000

Conc-gm/L	Transform: Arcsin Square Root							Number Resp	Total Number
	Mean	N-Mean	Mean	Min	Max	CV%	N		
D-Control	1.0000	1.0000	1.4120	1.4120	1.4120	0.000	2	0	20
0.063	1.0000	1.0000	1.4120	1.4120	1.4120	0.000	2	0	20
0.125	1.0000	1.0000	1.4120	1.4120	1.4120	0.000	2	0	20
0.25	1.0000	1.0000	1.4120	1.4120	1.4120	0.000	2	0	20
0.5	0.5000	0.5000	0.7854	0.5796	0.9912	37.050	2	10	20
1	0.0000	0.0000	0.1588	0.1588	0.1588	0.000	2	20	20

Auxiliary Tests				Statistic	Critical	Skew	Kurt
Normality of the data set cannot be confirmed							
Equality of variance cannot be confirmed							
				Trimmed Spearman-Kärber			
Trim Level	EC50	95% CL					
0.0%	0.5000	0.4282	0.5838				
5.0%	0.5000	0.4209	0.5940				
10.0%	0.5000	0.4119	0.6069				
20.0%	0.5000	0.3862	0.6474				
Auto-0.0%	0.5000	0.4282	0.5838				



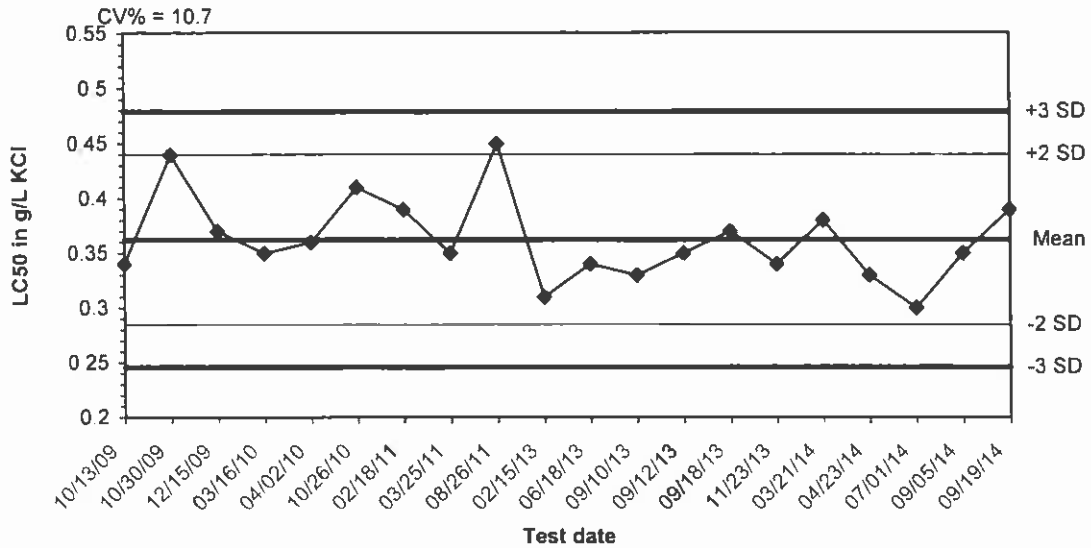
Test: AT-Acute 96-hr Toxicity Test
 Species: HA-Hyaella azteca
 Sample ID: REF-Ref Toxicant
 Start Date: 1/23/2015 09:15

Test ID: 999-3381
 Protocol: NASXXXHA1-Hyaella azteca acute
 Sample Type: KCL-Potassium chloride
 End Date: 1/27/2015 10:3 Lab ID: ORNAS-Northwestern Aquatic Sciences

Pos	ID	Rep	Group	Start	24 Hr	48 Hr	72 Hr	96 Hr	Notes
	1	1	D-Control	10				10	
	2	2	D-Control	10				10	
	3	1	0.063	10				10	
	4	2	0.063	10				10	
	5	1	0.125	10				10	
	6	2	0.125	10				10	
	7	1	0.250	10				10	
	8	2	0.250	10				10	
	9	1	0.500	10				7	
	10	2	0.500	10				3	
	11	1	1.000	10				0	
	12	2	1.000	10				0	

Comments: data entry verified against laboratory bench sheets 3-5-15 JRF

Amphipod, *Hyalella azteca*, acute reference toxicant test



Dates	Values	Mean	-2 SD	-3 SD	+2 SD	+3 SD
10/13/09	0.3400	0.3625	0.2847	0.2458	0.4403	0.4792
10/30/09	0.4400	0.3625	0.2847	0.2458	0.4403	0.4792
12/15/09	0.3700	0.3625	0.2847	0.2458	0.4403	0.4792
03/16/10	0.3500	0.3625	0.2847	0.2458	0.4403	0.4792
04/02/10	0.3600	0.3625	0.2847	0.2458	0.4403	0.4792
10/26/10	0.4100	0.3625	0.2847	0.2458	0.4403	0.4792
02/18/11	0.3900	0.3625	0.2847	0.2458	0.4403	0.4792
03/25/11	0.3500	0.3625	0.2847	0.2458	0.4403	0.4792
08/26/11	0.4500	0.3625	0.2847	0.2458	0.4403	0.4792
02/15/13	0.3100	0.3625	0.2847	0.2458	0.4403	0.4792
06/18/13	0.3400	0.3625	0.2847	0.2458	0.4403	0.4792
09/10/13	0.3300	0.3625	0.2847	0.2458	0.4403	0.4792
09/12/13	0.3500	0.3625	0.2847	0.2458	0.4403	0.4792
09/18/13	0.3700	0.3625	0.2847	0.2458	0.4403	0.4792
11/23/13	0.3400	0.3625	0.2847	0.2458	0.4403	0.4792
03/21/14	0.3800	0.3625	0.2847	0.2458	0.4403	0.4792
04/23/14	0.3300	0.3625	0.2847	0.2458	0.4403	0.4792
07/01/14	0.3000	0.3625	0.2847	0.2458	0.4403	0.4792
09/05/14	0.3500	0.3625	0.2847	0.2458	0.4403	0.4792
09/19/14	0.3900	0.3625	0.2847	0.2458	0.4403	0.4792

SECTION B

Midge (*Chironomus dilutus*) 10-day sediment bioassay 862-3 data report

TOXICITY TEST REPORT

TEST IDENTIFICATION

Test No.: 862-3

Title: Toxicity of freshwater sediments using a 10-day midge, *Chironomus dilutus*, sediment bioassay as part of the Remedial Investigation at the Jacobson Terminals Property in Seattle, WA

Protocol No.: NAS-XXX-CT4b, April 7, 1998. Revision 1 (10-28-03). Based on ASTM 2001 (Standard test methods for measuring the toxicity of sediment-associated contaminants with fresh water invertebrates, E1706-00), Am. Soc. Test. Mat., Phila., PA, and EPA Method 100.2 (Methods for measuring the toxicity and bioaccumulation of sediment-associated contaminants with freshwater invertebrates, EPA/600/R-99/064). Washington State Sediment Management Standards (SMS) (Chapter 173-204 WAC, Last Update: 2/25/13).

STUDY MANAGEMENT

Study Sponsor: Hart Crowser, Inc., 1700 Westlake Ave. North, Suite 200, Seattle, WA 98109.

Sponsor's Study Monitor: Mr. Philip Cordell

Testing Laboratory: Northwestern Aquatic Sciences, P.O. Box 1437, Newport, OR 97365

Test Location: Newport laboratory

Laboratory's Study Personnel: G.J. Irissarri, B.S., Proj. Mngr./ Study Dir.; L.K. Nemeth, B.A., M.B.A., QA Officer; R.S. Caldwell, Ph.D., Sr. Aq. Toxicol.; G.A. Buhler, B.S., Aq. Toxicol.; J.B. Brown, B.S., D.V.M., Assoc. Aq. Toxicol.; Y. Nakahama, Sr. Tech.; L. Brady, Tech.

Study Schedule:

Test Beginning: 2-6-15, 1005 hrs.

Test Ending: 2-16-15, 1045 hrs.

Disposition of Study Records: All raw data, reports, and other study records are stored at Northwestern Aquatic Sciences, 3814 Yaquina Bay Rd., Newport, OR 97365.

Statement of Quality Assurance: The test data were reviewed by the Quality Assurance Unit to assure that the study was performed in accordance with the protocol and standard operating procedures. This report is an accurate reflection of the raw data.

TEST MATERIAL

Test Sediments: Freshwater test sediments collected as part of the Remedial Investigation at the Jacobson Terminals Property in Seattle, WA. Details are as follows:

NAS Sample No.	5192G	5193G	5194G
Description	JT-SS-06	JT-SS-08	JT-SS-10
Collection Date	1/12/15	1/12/15	1/12/15
Receipt Date	1/14/15	1/14/15	1/14/15

Control Sediment: The negative control sediment (NAS#5195G) was collected on 1-19-15 from an area approximately one mile east of the Hwy. 101 bridge at Beaver Creek, approx. 8 miles south of Newport, OR.

Treatments: Homogenized at test set up by mixing using stainless steel implements.

Storage: All test and control sediments were stored at 4°C in the dark in sealed containers until used.

TEST WATER

Source: Dechlorinated municipal tap water.

Dates of Preparation: 1-19-15 and 2-6-15

Water Quality:

pH: 7.7, 7.5

conductivity: 108, 119 µmhos/cm

hardness: 26, 26 mg/L as CaCO₃

alkalinity: 30, 30 mg/L as CaCO₃.

total chlorine: <0.02, <0.02 mg/L

Pretreatment: Dechlorinated and aerated ≥24 hr.

TEST ORGANISMS

Species: *Chironomus dilutus* (formerly *C. tentans*), midge.

Size: 2nd to 3rd instar, mean initial wt: 0.21 ± 0.01 mg

Source: Aquatic BioSystems, Fort Collins, CO

Acclimation: Holding conditions prior to testing averaged: Temperature, 22.7 °C; dissolved oxygen, 6.5 mg/L; pH, 7.3; conductivity, 376 µmhos/cm; hardness, 129 mg/L as CaCO₃; and alkalinity, 90 mg/L as CaCO₃. Photoperiod was 16:8, L:D. Half of the water in culture tanks was replaced with dechlorinated municipal tap water during holding. Animals were fed Tetra Fin suspension and *Selenastrum*.

TEST PROCEDURES AND CONDITIONS

The following is an abbreviated statement of the test procedures and a statement of the test conditions actually employed. See the test protocol (Appendix I) for a more detailed description of the test procedures used in this study.

Test Chambers: 300 ml high-form glass beakers

Test Volumes: 100 ml sediment layer; 175 ml test water.

Replicates/Treatment: 8

Organisms/Treatment: 80

Water Volume Changes: 2 water volumes per day

Aeration: None.

Feeding: Animals were fed 1.5 ml of Tetra Fin suspension (1.5 ml contains 6 mg dry solids) per beaker daily.

Acceptance Criteria: Results are valid if mean control mortality does not exceed 30%, and the mean individual ash-free dry weight at test termination is ≥ 0.48 mg.

Effects Criteria: 1) survival after 10 days, and 2) average individual biomass (based on ash-free dry weight) after 10 days. Death is defined as no visible movement or response to tactile stimulation. Missing organisms were considered to be dead.

Water Quality and Other Test Conditions: The temperature, dissolved oxygen, conductivity, pH, hardness, alkalinity and ammonia-nitrogen were measured in the overlying water of one replicate test container per treatment on days 0 and 10 of the test. Temperature and dissolved oxygen were measured daily in the overlying water of one replicate test container per treatment. Hardness and alkalinity were measured with titrimetric methods. Ammonia-N was measured using Hach reagents based on the salicylate (Clin. Chim. Acta 14:403, 1996) colorimetric method; samples were not distilled prior to analysis. The photoperiod was 16:8, L:D.

DATA ANALYSIS METHODS

Percent survival and average individual ash-free dry weight were calculated for each replicate as follows:

percent survival = 100 x (number surviving/initial number tested)

average individual ash-free dry wt. = (ash-free dry wt.)/number weighed,
where:

ash-free dry wt. = dry weight of organisms recovered on day 10 – ashed dry weight, in mg

Means and standard deviations for the biological endpoints described above, and for water quality data, were computed using Microsoft Excel 2010. The values for mortality and individual ash-free dry wt for the test sediment were statistically compared against the reference sediment. Where appropriate, an arcsine square root transformation was performed on proportional mortality data before analysis. Following determination of normality and homogeneity of variances, a one-tailed Student T-test, Mann-Whitney or Approximate T test was conducted at the 0.05 level of significance. The statistical software used was BioStat (version Feb 9, 2006 (EXCEL)) bioassay software developed by the U.S. Army Corps of Engineers, Seattle District.

PROTOCOL DEVIATIONS

None

REFERENCE TOXICANT TEST

The reference toxicant test is a multi-concentration toxicity test using potassium chloride, to evaluate the performance of the test organisms used in the sediment toxicity test. The performance is evaluated by comparing the results of this test with historical results obtained at the laboratory. A summary of the reference toxicant test result is given below. The reference toxicant test raw data are found in Appendix III.

Test No.: 999-3391

Reference Toxicant and Source: Potassium Chloride (KCl), Fisher Lot #114689.

Test Date: 2-6-15.

Dilution Water Used: Moderately hard synthetic water prepared from Milli-Q® deionized water.

Result: 96-hr LC50, 5.74 g/L. This result is within the laboratory's control chart warning limits (2.71 –7.48 g/L).

TEST RESULTS

Observations of water quality in the overlying water throughout the test are summarized in Table 1. A detailed tabulation of the water quality results by sample and test day can be found in Appendix II. The means and standard deviations of percent mortality and growth (ash-free dry wt.) of midges exposed for 10 days to sediments are summarized in Tables 2 and 3. Detailed data organized by sample and replicate, and summary statistics for these observations, are given in Appendix II.

All water quality observations of overlying water temperature and dissolved oxygen were within the protocol specified ranges. Ammonia-N in the overlying water ranged between <0.1 and 0.3 mg/L for all day 0 and day 10 measurements.

The test met the acceptability criteria specified in the SMS with 6.3% mean control mortality ($\leq 30\%$ required) and a control mean ash-free dry weight of 1.22 mg per larvae (≥ 0.48 mg required). The reference toxicant (positive control) EC50 result was within the laboratory's control chart limits (5.74 g/L; control chart mean ± 2 S.D. = 5.10 ± 2.39).

Interpretation was based on guidelines from the Washington State Sediment Management Standards (SMS) (Chapter 173-204 WAC, Last Update: 2/25/13). The SMS includes Sediment Cleanup Objectives (SCO) and Cleanup Screening Levels (CSL) biological criteria. The Sediment Cleanup Objectives establish a no adverse effects level, including no acute or chronic adverse effect, to the benthic community. The Cleanup Screening Levels establish a minor adverse effects level, including acute or chronic effects, to the benthic community. To qualify as an adverse effect under the SCO for mortality the mean mortality in the test sediment is greater than 20 percent over the mean control and statistically different from the control ($p = 0.05$). For the growth endpoint, a mean reduction in the biomass that is greater than 20 percent of the control sediment response and statistically different from the control ($p = 0.05$). For the CSL adverse effects criteria mean mortality in the test sediment is greater than 30 percent over the mean control and statistically different from the control ($p = 0.05$). For the growth endpoint, a mean reduction in the biomass that is greater than 30 percent and statistically different from the control ($p = 0.05$).

Mean mortality of test sediments JT-SS-06, JT-SS-08 and JT-SS-10 was 10.0%, 15.0% and 8.8%, respectively. None of these were more than 20% above the control mortality; therefore none of the sediments exceeded the SCO or CSL criteria for mortality. For the growth endpoint, all three test sediments were significantly different from the control, but only two test sediments, JT-SS-08, and JT-SS-10, resulted in and exceeded under the SCO guidelines with ash-free dry weights of 0.93 and 0.91 mg/individual, respectively. No sediment failed the CSL criteria for growth since none were >30% different from the control.

STUDY APPROVAL

Maal Lusari 3-19-15
Project Manager/Study Director Date

Julie R. Fane 3-19-15
Quality Assurance Unit Date

for Linda Neeth
Assistant Laboratory Director Date 3-19-15

Table 1. Summary of water quality conditions during tests of the midge, *Chironomus dilutus*, exposed to freshwater sediments.

Water Quality Parameter	Mean \pm S.D.	Minimum	Maximum	N
Temperature ($^{\circ}$ C)	22.7 \pm 0.5	22.0	24.0	44
Dissolved oxygen (mg/L)	6.1 \pm 0.7	4.5	7.4	44
Conductivity (μ hos/cm)	129 \pm 29	108	217	12
pH	7.2 \pm 0.1	7.1	7.4	12
Hardness (mg/L as CaCO ₃)	29 \pm 4	26	34	8
Alkalinity (mg/L as CaCO ₃)	36 \pm 7	30	50	8
Total ammonia (mg/L)	---	<0.1	0.3	8

Table 2. Mortality results of *Chironomus* toxicity test and data interpretation using guidelines from the Washington State SMS.

Sample description	Percent mortality (Mean \pm SD)	Significantly higher than the control sediment at $\alpha=0.05$?	Percent higher (absolute) than control sediment	Exceedance under SCO? ¹	Exceedance under one-test criteria for CSL? ²
Control (NAS# 5195G)	6.3 \pm 5.2	---	---	---	---
JT-SS-06 (NAS# 5192G)	10.0 \pm 14.1	No	3.7	No	No
JT-SS-08 (NAS# 5193G)	15.0 \pm 16.9	No	8.7	No	No
JT-SS-10 (NAS# 5194G)	8.8 \pm 11.3	No	2.5	No	No

¹ **Sediment Cleanup Objectives (SCO) exceedance** if the test sediment mean mortality is significantly higher (1-tailed t-test at $P \leq 0.05$) than the control sediment mean mortality and the absolute difference is $>20\%$.

² **Cleanup Screening Levels (CSL) exceedance** if the test sediment mean mortality is significantly higher (1-tailed t-test at $P \leq 0.05$) than the control sediment mean mortality and the absolute difference is $>30\%$.

Table 3. Growth results of *Chironomus* toxicity test and data interpretation using guidelines from the Washington State SMS.

Sample description	Average ash-free dry wt/midge (mg)* (Mean \pm SD)	Statistically significantly lower than control sediment at $\alpha=0.05$?	Percent lower than control sediment	Exceedance under SCO? ¹ ($MIG_c - MIG_T / MIG_c > 0.20$)	Exceedance under one-test criteria for CSL? ² ($MIG_c - MIG_T / MIG_c > 0.30$)
Control (NAS# 5195G)	1.22 \pm 0.09	---	---	---	---
JT-SS-06 (NAS# 5192G)	0.98 \pm 0.11	Yes	19.7	No	No
JT-SS-08 (NAS# 5193G)	0.93 \pm 0.15	Yes	23.8	Yes	No
JT-SS-10 (NAS# 5194G)	0.91 \pm 0.12	Yes	25.4	Yes	No

* Pupae were not included in the sample to estimate ash-free dry weight (as per EPA/600/R-99/064, p. 59, section 12.3.8.2)

¹ **Sediment Cleanup Objectives (SCO) exceedance** if the test sediment mean growth is significantly lower (1-tailed t-test at $P \leq 0.05$) than the control sediment mean growth, and the difference is $>20\%$.

² **Cleanup Screening Levels (CSL) exceedance** (one-test criteria) if the test sediment mean individual growth is significantly lower (1-tailed t-test at $P \leq 0.05$) than the control sediment mean growth, and the difference is $>30\%$.

APPENDIX I
PROTOCOL

TEST PROTOCOL

**FRESHWATER MIDGE, *CHIRONOMUS TENTANS*,
10-DAY SEDIMENT TOXICITY TEST**

1. **INTRODUCTION**

1.1 **Purpose of Study:** The purpose of this study is to characterize the toxicity of freshwater sediments based on midge survival and growth using the midge, *Chironomus tentans*.

1.2 **Referenced Method:** This protocol is based on EPA Method 100.2 (EPA/600/R-99/064) and ASTM Method E 1706-00 (ASTM 2001).

1.3 **Summary of Method:** A summary of test conditions for the midge 10-day sediment toxicity test is tabulated below. The 10-day sediment toxicity test with *Chironomus tentans* is conducted at 23°C with a 16L:8D photoperiod at an illuminance of about 100-1000 lux. Test chambers are 300-mL high-form lipless beakers containing 100 mL of sediment and 175 mL of overlying water. Ten second to third-instar midges are used in each replicate (all organisms must be third instar or younger and at least 50% of the larvae must be third instar). The number of replicates/treatment depends on the objective of the test. Eight replicates are recommended for routine testing. Midges in each test chamber are fed 1.5 mL of a 4 g/L fish food flakes suspension daily. Each chamber receives two volume additions per day of overlying water. Overlying water can be culture water, well water, surface water, site water, or reconstituted water. Test endpoints include survival and/or growth.

2. **STUDY MANAGEMENT**

2.1 **Sponsor's Name and Address:**

2.2 **Sponsor's Study Monitor:**

2.3 **Name of Testing Laboratory:**

Northwestern Aquatic Sciences
3814 Yaquina Bay Road, P.O. Box 1437
Newport, OR 97365.

2.4 **Test Location:** _____

2.5 **Laboratory's Personnel to be Assigned to the Study:**

Study Director: _____
Quality Assurance Unit: _____
Aquatic Toxicologist: _____
Aquatic Toxicologist: _____

2.6 **Proposed Testing Schedule:** Tests are to begin within 14 days of sample collection. Eight week holding times may apply in some circumstances. Reference toxicant test to be run concurrently.

2.7 Good Laboratory Practices: The test is conducted following the principles of Good Laboratory Practices (GLP) as defined in the EPA/TSCA Good Laboratory Practice regulations revised August 17, 1989 (40 CFR Part 792).

3. TEST MATERIAL

The test materials are freshwater sediments. The control, reference, and test sediments are placed in solvent cleaned 1 L glass jars fitted with PTFE-lined screw caps. At the laboratory the samples are stored at 4°C in the dark. The original sealed containers may be stored for up to 14 days prior to testing. Eight week holding times may apply in some circumstances.- If jars are not full when received or if sediment is removed for testing, headspaces should be filled with nitrogen to retard deterioration. A negative control sediment is collected from a clean site. In addition, a reference sediment, a clean sediment with physical characteristics similar to the test sediments, may be employed as a comparison station.

4. TEST WATER

Test water (overlying water) at NAS is normally *C. tentans* culture water, which is moderately hard synthetic water at a hardness of 80-100 mg/L as CaCO₃ and alkalinity of 60-70 mg/L as CaCO₃. Dilution water is prepared from Milli-Q reagent grade water and reagent grade chemicals. Test water may also be well water, surface water or site water depending on the study design.

5. TEST ORGANISMS

5.1 Species: midge, *Chironomus tentans*.

5.2 Source: Cultured at NAS (Originally obtained from U.S. EPA Environmental Research Lab, Duluth, MN) or purchased from a reputable commercial supplier.

5.3 Age: Third instar or younger larvae (at least 50% of the larvae must be in the third instar at the start of the test). Third instar is normally 9 to 11 days after hatching; head capsule widths range from 0.33 to 0.45 mm; or length ranges from 4-6 mm; or dry weight ranges 0.08 to 0.23 mg/individual.

5.4 Acclimation and Pretest Observation: Cultures are maintained at 23 ± 1°C under a 16:8 L:D photoperiod. The culture water is moderately hard synthetic water. Midge are fed finely ground Tetrafin flakes in suspension (10g Tetrafin in 100 mL Milli-Q water). Mortality during the 48-hr prior to testing should not be excessive.

6. DESCRIPTION OF TEST SYSTEM

6.1 Test Chambers and Environmental Control: Test chambers used in the toxicity test are 300-mL high-form lipless glass beakers (Pyrex® 1040 or equivalent). Test chambers are maintained at constant temperature by partial immersion in a temperature-controlled water bath or by placement in a temperature-controlled room. Aeration is not employed unless dissolved oxygen drops below 2.5 mg/L. The test is conducted under an illuminance of 100 to 1000 lux with a 16L:8D photoperiod.

6.2 Cleaning: All laboratory glassware, including test chambers, is cleaned as described in EPA/600/4-90/027F. New glassware and test systems are soaked 15 minutes in tap water and scrubbed with detergent (or cleaned in automatic dishwasher); rinsed twice with tap water; carefully rinsed once with fresh, dilute (10%, V:V) hydrochloric or nitric acid to remove scale, metals, and bases; rinsed twice with deionized water; rinsed once with acetone to remove organic compounds (using a fume hood or canopy); and rinsed three times with deionized water. Test systems and chambers are rinsed again with dilution water just before use.

7. EXPERIMENTAL DESIGN AND TEST PROCEDURES

7.1 Experimental Design: The test involves exposure of midge larvae to test, control, and reference sediments. The sediments are placed on the bottom of the test containers and are overlain with test water. The test exposure is for 10 days. The renewal of overlying water consists of two volume additions per day, either continuous or intermittent. Each treatment consists of eight replicate test containers, each containing 10 organisms. Test chamber positions are completely randomized. Test organisms are randomly distributed to the test chambers. Blind testing is normally used.

7.2 Setup of Test Containers: Sediments are homogenized and placed in test chambers on the day before addition of test organisms. Sediment (100 ml) is placed into each of eight replicate beakers. After addition of the sediment, 175 ml of test water is gently added to each beaker in a manner to prevent resuspension. The overlying water is replaced twice daily. The test begins when midges are introduced to the test chambers. Initial water quality measurements are taken prior to the addition of test organisms.

7.3 Effect Criterion: The acute effect criterion used in the midge bioassay is mortality, defined as the lack of movement of body or appendages on response to tactile stimulation. The optional chronic effect criterion is growth which is determined by using dry weight measurements.

7.4 Test Conditions: No aeration is employed unless dissolved oxygen falls below 2.5 mg/L. The test temperature employed is 23°C (range of $\pm 1^\circ\text{C}$). A 16:8, L:D photoperiod is used. Illumination is supplied by daylight fluorescent lamps at 100-1000 lux. The overlying water is replaced twice daily.

7.5 Beginning the Test: The test is begun by adding the organisms to the equilibrated test containers as previously described. Three extra replicates of midge larvae should be counted out and randomly selected for drying to determine initial average weight and instar data.

7.6 Feeding: Midge larvae are fed 1.5 mL daily per test chamber (1.5 mL contains 6.0 mg of dry solids). A feeding may be skipped if there is a build up of excess food. However, all beakers must be treated similarly.

7.7 Test Duration, Type and Frequency of Observations, and Methods: The duration of the acute toxicity test is 10 days. The type and frequency of observations to be made are summarized as follows:

Type Of Observation	Times Of Observation
<u>Biological Data</u>	
Survival, growth	Day 10
<u>Physical And Chemical Data</u>	
Hardness, alkalinity, ammonia-N, conductivity, pH, dissolved oxygen, and temperature	Beginning and end of test in overlying water of one replicate beaker from each treatment.
Dissolved oxygen, temperature	Daily in overlying water of one replicate beaker from each treatment.

Dissolved oxygen is measured using a polarographic oxygen probe calibrated according to the manufacturer's recommendations. The pH is measured using a pH probe and a properly calibrated meter with scale divisions of 0.1 pH units. Temperature is measured with a calibrated mercury thermometer or telethermometer. Conductivity is measured with a conductivity meter. Hardness and alkalinity are measured using titrometric methods. Ammonia-nitrogen is measured using the salicylate colorimetric method (Clin. Chim. Acta 14:403, 1996).

7.8 Growth Measurement: Growth is measured as ash-free dry weight (AFDW) of animals in a test replicate at the end of the test on day 10. Pooled animals from each test replicate are rinsed with deionized water, gently blotted and placed into tared aluminum weigh pans. The pans are dried at 60-90°C to constant weight. The dried organisms are placed into a dessicator and weighed as soon as possible to the nearest 0.01 mg (desirable to use

0.001 mg). The total weight of the dried midge in each pan is divided by the number of midge weighed to obtain an average dry weight per midge. The dried larvae in the pan are then ashed at 550°C for two hours. The pan with the ashed larvae is then reweighed and the tissue mass of the larvae is determined as the difference between the weight of the dried larvae plus pan and the weight of the ashed larvae plus pan. Pupae or adult organisms are not included in the sample to estimate AFDW.

8. CRITERIA OF TEST ACCEPTANCE:

The test results are acceptable if the minimum survival of organisms in the control treatment at the end of the test is at least 70% and the average ash-free dry weight of *C. tentans* in the surviving controls is at least 0.48 mg.

9. DATA ANALYSIS

The endpoints of the toxicity test are survival and growth. Survival is obtained as a direct count of living organisms in each test container at the end of the test. Average midge ash-free dry weight, also measured at the end of the test, may be used to compare growth between treatment sediments and the control or reference sediment. Ordinarily the following data analysis is performed. Due to special requirements, alternative methods may be used. The means and standard deviations are calculated for each treatment level. Identification of toxic sediments is established by statistical comparison of test endpoints between test and control or reference sediments. Between treatment comparisons may be made using a Student's t-test or Wilcoxon's Two-Sample test, where each treatment is compared to the control or the reference sediment. An arcsine-square root transformation of proportional data, and tests for normality and heterogeneity of variances, are performed prior to statistical comparisons.

10. REPORTING

The final report of the test results must include all of the following standard information at a minimum: name and identification of the test; the investigator and laboratory; date and time of test beginning and end; information on the test material; information on the source and quality of the overlying/test water; detailed information about the test organisms including acclimation conditions; a description of the experimental design and test chambers and other test conditions including feeding, if any, and water quality; definition of the effect criteria and other observations; responses, if any, in the control treatment; tabulation and statistical analysis of measured responses and a summary table of endpoints; a description of the statistical methods used; any unusual information about the test or deviations from procedures; reference toxicant testing information.

11. STUDY DESIGN ALTERATION

Amendments made to the protocol must be approved by the sponsor and study director and should include a description of the change, the reason for the change, the date the change took effect and the dated signatures of the study director and sponsor. Any deviations in the protocol must be described and recorded in the study raw data.

12. REFERENCE TOXICANT

The reference toxicant test is a standard multi-concentration toxicity test using a specified chemical toxicant to evaluate the performance of test organisms used in the study. Reference toxicant tests are 96-hour, water only exposures, not 10-day sediment exposures. The reference toxicant test is run concurrently. Performance is evaluated by comparing the results of the reference toxicant test with historical results (e.g., control charts) obtained at the laboratory.

13. REFERENCED GUIDELINES

ASTM. 2001. Standard Test Methods for Measuring the Toxicity of Sediment-associated Contaminants with Fresh water Invertebrates. ASTM Standard Method No. E 1706-00. Am. Soc. Test. Mat., Philadelphia, PA.

U.S. EPA. 2000. Section 12, Test Method 100.2, *Chironomus tentans* 10-d Survival and Growth Test for Sediments, pp. 55-62. In: Methods for Measuring the Toxicity and Bioaccumulation of Sediment-associated Contaminants with Freshwater Invertebrates (Second Edition). EPA/600/R-99/064.

Weber, C.I. (Ed.) 1993. Methods for Measuring the Acute Toxicity of Effluents and Receiving Waters to Freshwater and Marine Organisms (Fourth Edition). EPA/600/4-90/027F.

14. APPROVALS

_____ for _____
Name Date

_____ for **Northwestern Aquatic Sciences**
Name Date

Appendix A
Test Conditions Summary

1. Test type	whole sediment toxicity test with renewal of overlying water
2. Test duration	10 days
3. Temperature	23 ± 1°C
4. Light quality	daylight flourescent light
5. Illuminance	100-1000 lux
6. Photoperiod	16L:8D
7. Test chamber size	300-mL high-form lipless beakers (Pyrex® 1040 or equivalent)
8. Sediment volume	100 mL
9. Overlying water volume	175 mL
10. Renewal overlying water	2 volume additions/day (continuous or intermittent)
11. Age of test organisms	2nd to 3rd instar or younger larvae (≥ 50% of organisms must be 3rd instar)
12. Organisms per test chamber	10
13. Replicates per treatment	8 recommended for routine (depends on design)
14. Organisms per treatment	80
15. Feeding regime	Fish food flakes, fed 1.5 mL chamber (1.5 mL contains 6.0 mg of dry solids) daily on days 0 - 9.
16. Aeration	None, unless DO falls below 2.5 mg/L.
17. Overlying (test) water	Culture water, well water, surface water, site water or reconstituted water
18. Water quality	Hardness, alkalinity, conductivity, pH, ammonia-N beginning and end; temperature and DO daily
19. Endpoints	Survival and growth (dry weight)
20. Test acceptability criteria	Minimum control survival of 70%; mean weight of surviving control organisms 0.48 mg AFDW
21. Sample holding	≤14 days at 4°C in the dark Longer under certain conditions
22. Sample volume required	1L (800 mL per sediment)
23. Reference toxicant	Concurrent testing required

APPENDIX II

RAW DATA

**TEST DESCRIPTION, MONITORING, AND RESULTS
BENCHSHEETS**

Test No. 862-3 Client Hart Crowser Investigator _____

STUDY MANAGEMENT

Client: Hart Crowser, Inc., 1700 Westlake Ave. North, Suite 200, Seattle, WA 98109

Client's Study Monitor: Mr. Philip Cordell

Testing Laboratory: Northwestern Aquatic Sciences

Test Location: Newport Laboratory

Laboratory's Study Personnel:

Proj. Man./Study Dir. G.J. Irissarri ⁶³¹

QA Officer L.K. Nemeth

1. NO. Northwestern 2. GAS h by 17

3. J. Brown 4. Lauren Brady LB

5. J.S. Calambokidis PSC 6. _____

7. _____ 8. _____

Study Schedule:

Test Beginning: 2-6-15 1005 Test Ending: 2-16-15 1045

TEST MATERIAL

General description (see sample logbook/chain-of-custody for details):

NAS Sample No.:	5192G	5193G	5194G		
Description:	JT-SS-06	JT-SS-08	JT-SS-10		
Collection Date:	1/12/15	1/12/15	1/12/15		
Receipt Date:	1/14/15	1/14/15	1/14/15		

NAS Sample No.:					
Description:					
Collection Date:					
Receipt Date:					

NAS Sample No.:					
Description:					
Collection Date:					
Receipt Date:					

NAS Sample No.:					
Description:					
Collection Date:					
Receipt Date:					

NAS Sample No.:					
Description:					
Collection Date:					
Receipt Date:					

Error codes: 1) correction of handwriting error
2) written in wrong location; entry deleted
3) wrong date deleted, replaced with correct date
4) error found in measurement; measurement repeated

Test No. 862-3 Client Hart Crowser Investigator _____

TEST WATER

Source: Dechlorinated municipal tap water
 Date of Collection: 1-19-15, 2-6-15
 pH 7.7, 7.5
 Cond (umhos/cm²) 108, 119
 Hardness (mg/L) 26, 26
 Alkalinity (mg/L) 30, 30
 Total Chlorine (mg/L) 20.02, 20.02
 Treatments: Dechlorinated, aerated

TEST ORGANISMS

Species: Chironomus dilutus Age: 2ND TO 3rd instar
 Source: ~~NAS cultures~~ AQUATIC BIOSYSTEMS Date received: 2-5-15

Acclimation Data:

Date	Temp. (deg.C)	pH	DO (mg/L)	Cond. umhos/cm	Hardness (mg/L)	Alkalinity (mg/L)	Feeding	Water changes
2-5-15	20.0	6.9	6.2	436	171	110	Animals fed Tetra Fin	YES
2-6-15	24.8	7.6	6.7	316	96	70	and <i>Selenastrum</i>	
							Details recorded on	
							Chironomid culture	
							data sheets	
Mean	22.7	7.3	6.5	376	129	90		
S.D.	-	-	-	-	-	-		
(N)	2	2	2	2	2	2		

Photoperiod during acclimation: 16:8, L:D

TEST PROCEDURES AND CONDITIONS

Test chambers: 300 ml glass beakers
 Test volumes: 100 ml of test sediment; 275 ml total volume
 Replicates/treatment: (8) 8 Organisms/treatment: (80) 80 (10/REP)
 Test water changes: Twice daily
 Aeration: only if DO falls below 2.5 mg/L
 Feeding: everyday beginning with day zero
 Test temperature (°C): 23 ± 1
 Beaker placement: Total randomization
 Photoperiod: 16:8, L:D

Control Sediment:

Source: From an area approximately one mile east of the Hwy. 101 bridge at Beaver Creek,
approx. 8 miles south of Newport, OR.
 Date collected: 1/19/15
 Sieved through 0.5 -mm screen
 Storage: darkness at 4°C, in sealed containers NAS# 5195G

MISCELLANEOUS NOTES

Test No. 862-3 Client Hart Crowser Investigator _____

Test conducted in (circle one) room 1 room 2 trailer water bath other: _____

Randomization chart:

TOP SHELF

6	12	18	24	30					
5	11	17	23	29					
4	10	16	22	28					
3	9	15	21	27					
2	8	14	20	26	32				
1	7	13	19	25	31				

Randomization chart:

FRONT

Randomization chart:

Test No. 862-3 Client Hart Crowser Investigator _____

DAILY RECORD SHEET

Day 0 (21 615) 1/1/02

Beaker No.	Temp.* (deg.C)	DO* (ppm)	Cond.* (umhos/cm)	pH*	Hardness* (mg/L)	Alkalinity* (mg/L)	NH3* (ppm)	Comments
9	23.6	7.0	118	7.1	26	30		Each beaker fed 1.5 ml
12	24.0	7.4	217	7.4	26	30		Tetra Fin suspension
16	23.8	7.2	120	7.3	34	30		Initials: <u>6SL</u>
28	24.0	6.7	136	7.2	34	30		
								Water changed in all
								beakers.
								Time: <u>0505</u>
								Initials: <u>6SL</u>
								Water changed in all
								beakers.
								Time: <u>1610</u>
								Initials: <u>SL</u>

*Water quality measurements to be taken.

Day 1 (21715) 05/02

Beaker No.	Temp.* (deg.C)	DO* (ppm)	Cond. (umhos/cm)	pH	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
9	22.2	6.6						Each beaker fed 1.5 ml
12	22.6	6.8						Tetra Fin suspension
16	22.3	6.8						Initials: <u>6SL</u>
28	23.0	6.4						
								Water changed in all
								beakers.
								Time: <u>0510</u>
								Initials: <u>6SL</u>
								Water changed in all
								beakers.
								Time: <u>1615</u>
								Initials: <u>SL</u>

*Water quality measurements to be taken.

CHIRONOMUS DILUTUS 10-DAY SOLID PHASE SEDIMENT TEST

Test No. 862-3 Client Hart Crowser Investigator _____

DAILY RECORD SHEET

Day 2 (2/18/15) GSJ

Beaker No.	Temp.* (deg.C)	DO* (ppm)	Cond. (umhos/cm)	pH	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
9	22.2	6.7						Each beaker fed 1.5 ml Tetra Fin suspension Initials: <u>GSJ</u>
12	22.6	6.4						
16	22.4	6.8						
28	22.9	6.8						
								Water changed in all beakers. Time: <u>0505</u> Initials: <u>GSJ</u>
								Water changed in all beakers. Time: <u>1605</u> Initials: <u>GSJ</u>

*Water quality measurements to be taken.

Day 3 (2/19/15) GSJ

Beaker No.	Temp.* (deg.C)	DO* (ppm)	Cond. (umhos/cm)	pH	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
9	22.4	6.5						Each beaker fed 1.5 ml Tetra Fin suspension Initials: <u>GSJ</u>
12	22.9	6.1						
16	22.7	6.2						
28	23.2	6.1						
								Water changed in all beakers. Time: <u>0510</u> Initials: <u>GSJ</u>
								Water changed in all beakers. Time: <u>1606</u> Initials: <u>GSJ</u>

*Water quality measurements to be taken.

Test No. 862-3 Client Hart Crowser Investigator _____

DAILY RECORD SHEET

Day 4 (2/10/15) ✓

Beaker No.	Temp.* (deg.C)	DO* (ppm)	Cond. (umhos/cm)	pH	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
9	22.1	6.5						Each beaker fed 1.5 ml
12	22.4	6.1						Tetra Fin suspension
16	22.2	6.3						Initials: ✓
28	22.7	6.7						
								Water changed in all
								beakers.
								Time: 0505
								Initials: ✓
								Water changed in all
								beakers.
								Time: 1620
								Initials: US

*Water quality measurements to be taken.

Day 5 (2/11/15) ✓

Beaker No.	Temp.* (deg.C)	DO* (ppm)	Cond. (umhos/cm)	pH	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
9	22.0	6.6	108	7.2				Each beaker fed 1.5 ml
12	22.4	6.3	139	7.1				Tetra Fin suspension
16	22.2	6.2	112	7.2				Initials: ✓
28	22.5	6.6	117	7.1				
								Water changed in all
								beakers.
								Time: 0510
								Initials: ✓
								Water changed in all
								beakers.
								Time: 1605
								Initials: ✓

*Water quality measurements to be taken.

Test No. 862-3 Client Hart Crowser Investigator _____

DAILY RECORD SHEET

Day 6 (21 (4/5)) ✓ WS

Beaker No.	Temp.* (deg.C)	DO* (ppm)	Cond. (umhos/cm)	pH	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
9	22.1	4.8						Each beaker fed 1.5 ml
12	22.8	4.6						Tetra Fin suspension
16	22.4	4.0						Initials: ✓
28	23.4	4.5						
								Water changed in all beakers.
								Time: 0510
								Initials: ✓
								Water changed in all beakers.
								Time: 1030
								Initials: WS

*Water quality measurements to be taken.

Day 7 (21 (3/15)) ✓ WS

Beaker No.	Temp.* (deg.C)	DO* (ppm)	Cond. (umhos/cm)	pH	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
9	22.6	5.5						Each beaker fed 1.5 ml
12	23.1	5.3						Tetra Fin suspension
16	22.8	5.3						Initials: WS
28	23.6	5.9						
								Water changed in all beakers.
								Time: 0505
								Initials: WS
								Water changed in all beakers.
								Time: 1615
								Initials: ✓

*Water quality measurements to be taken.

Test No. 862-3 Client Hart Crowser Investigator _____

DAILY RECORD SHEET

Day 8 (2/14/15) US

Beaker No.	Temp.* (deg.C)	DO* (ppm)	Cond. (umhos/cm)	pH	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
9	22.1	5.4						Each beaker fed 1.5 ml
12	22.6	5.6						Tetra Fin suspension
16	22.3	5.7						Initials: <u>OSW</u> <u>GB</u>
28	22.9	5.9						
								Water changed in all
								beakers.
								Time: <u>6:15</u>
								Initials: <u>OSW</u>
								Water changed in all
								beakers.
								Time: <u>16:30</u>
								Initials: <u>JS</u>

*Water quality measurements to be taken.

Day 9 (2/15/15) US

Beaker No.	Temp.* (deg.C)	DO* (ppm)	Cond. (umhos/cm)	pH	Hardness (mg/L)	Alkalinity (mg/L)	NH3 (ppm)	Comments
9	22.4	5.6						Each beaker fed 1.5 ml
12	22.8	5.4						Tetra Fin suspension
16	22.6	5.4						Initials: <u>OSL</u>
28	23.2	5.3						
								Water changed in all
								beakers.
								Time: <u>05:10</u>
								Initials: <u>OSL</u>
								Water changed in all
								beakers.
								Time: <u>16:20</u>
								Initials: <u>OSL</u>

*Water quality measurements to be taken.

Test No. 862-3 Client Hart Crowser Investigator _____

Tare: Date 2-6-15 Oven temp (C.) 550 Drying time (hr.) 2 Initials GSJ
Standard Weights: 10 mg: 10.006 100mg: 100.016

Final: Date 2-9-15 Oven temp (C.) 62 Drying time (hr.) 24 Initials GSJ
#1 Standard Weights: 10 mg: 10.007 100mg: 100.015

Final: Date 2-11-15 Oven temp (C.) 63 Drying time (hr.) 24 Initials GSJ
#2 Standard Weights: 10 mg: 10.006 100mg: 10.016

Ashed Date 2-12-15 Oven temp (C.) 555 Drying time (hr.) 2 Initials GSJ
Standard Weights: 10 mg: 10.007 100mg: 100.015

Equip. used: Oven: BLUE M #1, FISHER Balance: SARTORIUS M3P
ISOTEMP MUFFLE FURNACE

(Dry overnight at 60-90 °C, ash for 2 hrs at 550°C)

Pan #	Tare wt. (mg)	Total wt. (mg)		Ashed total wt. (mg)	#Weighed	Comments
		1	2			
1	50.59	52.65	52.61	50.75	10	
2	45.38	47.68	47.68	45.52	10	
3	47.92	50.08	50.03	48.06	10	
4	48.84	50.81	50.78	49.00	10	
5	41.81	43.80	43.79	41.96	10	

Test No. 862-3 Client Hart Crowser Investigator _____

WEIGHING DATA SHEET

Tare: Date 2-9-15 Oven temp (C.) 550 Drying time (hr.) 2 Initials JRF
Standard Weights: 10 mg: 10.007 100mg: 100.009

Final #1: Date 2-17-15 Oven temp (C.) 62 Drying time (hr.) 24 Initials BSJ
Standard Weights: 10 mg: 10.007 100mg: 100.017

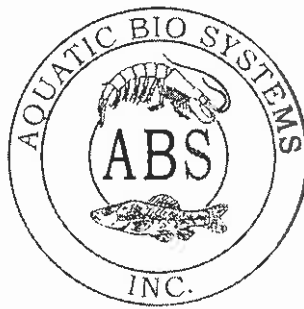
Final #2: Date 2-18-15 Oven temp (C.) 61 Drying time (hr.) 22 Initials BSJ
Standard Weights: 10 mg: 10.005 100mg: 100.015

Final #3: Date 2-19-15 Oven temp (C.) 550 Drying time (hr.) 2 Initials JRF
Standard Weights: 10 mg: 10.006 100mg: 100.008

Equip. used: Oven BLUE M#1, FISHER ISOTEMP MUFFLE FURNACE Balance Sartorius M3P
(Dry overnight at 60-90 degrees C) (Final ashing is at 550 degrees C for 2 hours)

Bkr. #	Pan #	Tare wt. (mg)	Dry total wt. (mg)		no. weighed	put into pans-initials	Ash weight (mg)	Comments
			1	2				
1	1	91.57	99.16	99.12	9	LB	93.18	
2	2	94.72	110.65	110.59	9	LB	99.34	
3	3	91.38	104.78	104.72	10	LB	94.54	
4	4	95.13	110.63	110.57	9	LB	99.60	
5	5	97.48	112.61	112.47	10	LB	101.97	
6	6	86.54	97.96	97.91	10	LB	89.22	
7	7	102.37	119.25	119.18	9	LB	107.17	
8	8	97.82	108.74	108.69	10	LB	100.38	
9	9	95.79	107.71	107.66	10	LB	98.43	
10	10	86.88	96.42	96.39	10	LB	89.04	
11	11	60.69	69.14	69.11	6	LB	62.54	
12	12	91.27	107.62	107.57	10	LB	96.15	
13	13	103.87	111.46	111.44	7	LB	105.52	
14	14	89.36	106.79	106.74	10	LB	94.24	
15	15	92.85	106.20	106.15	9	LB	95.78	
16	16	92.47	101.20	101.14	6	LB	94.32	
17	17	89.93	101.72	101.64	10	LB	92.78	
18	18	93.05	108.27	108.20	9	LB	97.35	
19	19	90.24	102.97	102.91	10	LB	93.19	
20	20	96.56	109.33	109.27	10	LB	99.53	
21	21	95.31	107.66	107.61	9	LB	98.38	
22	22	91.56	104.09	104.05	10	LB	94.44	
23	23	85.64	99.39	99.35	10	LB	88.84	
24	24	95.40	106.30	106.27	8	LB	97.95	
25	25	88.72	105.18	105.13	9	LB	93.38	
26	26	95.60	104.35	104.33	9	LB	97.60	
27	27	94.09	106.45	106.40	10	LB	97.20	
28	28	93.17	102.93	102.89	8	LB	95.36	
29	29	94.74	105.63	105.60	9	LB	97.34	
30	30	101.96	112.46	112.43	8	LB	104.36	
31	31	94.64	102.47	102.45	6	LB	96.44	
32	32	93.81	103.85	103.84	9	LB	96.14	

1300 Blue Spruce Drive, Suite C
Fort Collins, Colorado 80524



Toll Free: 800/331-5916
Tel: 970/484-5091 Fax: 970/484-2514

ORGANISM HISTORY

DATE: 2/4/2015

SPECIES: Chironomus dilutus (formerly C. tentans)

AGE: Deposited 1/24/2015

LIFE STAGE: Second Instar 2/4/2015

HATCH DATE: Emergent date 2/17/2015

BEGAN FEEDING: Immediately

FOOD: Selenastrum sp., Flake slurry

RECEIVED 2-5-15
-WL

Water Chemistry Record:

	Current	Range
TEMPERATURE:	<u>22°C</u>	<u>22-26°C</u>
SALINITY/CONDUCTIVITY:	<u>--</u>	<u>--</u>
TOTAL HARDNESS (as CaCO ₃):	<u>190 mg/l</u>	<u>100-190 mg/l</u>
TOTAL ALKALINITY (as CaCO ₃):	<u>60 mg/l</u>	<u>50-90 mg/l</u>
pH:	<u>7.50</u>	<u>7.50-8.20</u>

Comments:

Facility Supervisor

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TEST DATA ANALYSIS RECORDS

data entry verified against laboratory bench sheets 3-7-15 JRF

Freshwater Sediment Test
10-Day Chironomus dilutus

Test Number: 862-3

Endpoints Data Entry and Calculations File

TARE WT = ashed weight of pan used for that replicate at test termination (mg), or ASHED DRY WT = weight of ashed pan + weight of ashed test organisms recovered
 DRY WT = dry weight of pan if ash-free dry weight is not an endpoint
 WT COUNT = number of test organisms weighed at test end
 DRY WT = TARE WT - dry weight of test organisms recovered at test termination (mg)
 TWT = total biomass = DRY WT + TARE WT
 WT = average individual biomass = TWT / WT COUNT
 PSURV = % survival = 100(SURV/INIT)
 PMORT = % mortality = 100(MORT/INIT)

TAFDW = DRY WT - ASHED DRY WT = total ash-free organism weight for given replicate
 AFDW = average individual ash-free biomass = TAFDW / WT COUNT

INDEX	BKR	SMPL	CLIENT	DESCRIP	REPL	INIT SURV	MORT	PSURV	PMORT	TARE WT (mg)	WT COUNT	DRY WT (mg)	ASHED DRY WT (mg)	TWT (mg)	TAFDW (mg)	AFDW (mg)	SURV	MORT	PSURV	PMORT	WT	AFDW	INITIAL WEIGHT				
																							pan #	wt (mg)	final wt (mg)	count	avg. wt organism (mg)
1	5	5195G	Control	1	10	10	0	100.0	0.0	97.48	10	112.47	101.87	14.99	1.50	10.90	1.05	50.59	52.61	10	0.20	50.75	0.19				
2	4	5195G	Control	2	10	9	1	90.0	10.0	95.13	9	110.57	99.60	15.44	1.72	10.97	1.22	45.38	47.66	10	0.23	45.52	0.22				
3	25	5195G	Control	3	10	9	1	90.0	10.0	88.72	9	105.13	93.38	18.41	1.82	11.75	1.31	47.92	50.03	10	0.21	48.06	0.20				
4	14	5195G	Control	4	10	10	0	100.0	0.0	89.36	10	108.74	94.24	17.38	1.74	12.50	1.25	48.84	50.78	10	0.19	49.00	0.18				
5	2	5195G	Control	5	10	9	1	90.0	10.0	94.72	9	110.59	99.34	15.87	1.76	11.25	1.25	41.81	43.79	10	0.20	41.96	0.18				
6	18	5195G	Control	6	10	9	1	90.0	10.0	93.05	9	108.20	97.35	15.15	1.68	10.85	1.21	Mean	0.6	93.8	6.3	1.72	1.22				
7	7	5195G	Control	7	10	9	1	90.0	10.0	102.37	9	118.18	107.17	16.81	1.87	12.01	1.33	SD	0.5	5.2	5.2	0.12	0.09				
8	12	5195G	Control	8	10	10	0	100.0	0.0	91.27	10	107.57	96.15	16.30	1.63	11.42	1.14	n	8	8	8	8	8				
9	8	5192G	JT-SS-06	1	10	10	0	100.0	0.0	97.82	10	108.69	100.38	10.87	1.09	8.31	0.83	Mean	9.0	1.0	90.0	10.0	1.27	0.98			
10	31	5192G	JT-SS-06	2	10	6	4	60.0	40.0	94.64	6	102.45	96.44	7.81	1.30	6.01	1.00	SD	1.4	1.4	14.1	14.1	0.13	0.11			
11	24	5192G	JT-SS-06	3	10	8	2	80.0	20.0	95.40	8	106.27	97.95	10.87	1.38	8.32	1.04	n	8	8	8	8	8	8			
12	23	5192G	JT-SS-06	4	10	10	0	100.0	0.0	85.64	10	99.35	88.84	13.71	1.37	10.51	1.05	Mean	9.0	1.0	90.0	10.0	1.27	0.98			
13	19	5192G	JT-SS-06	5	10	10	0	100.0	0.0	90.24	10	102.91	93.19	12.67	1.27	9.72	0.97	SD	1.4	1.4	14.1	14.1	0.13	0.11			
14	32	5192G	JT-SS-06	6	10	9	1	90.0	10.0	93.81	9	103.84	96.14	10.03	1.11	7.70	0.86	n	8	8	8	8	8	8			
15	15	5192G	JT-SS-06	7	10	9	1	90.0	10.0	92.85	9	106.15	95.78	13.30	1.48	10.37	1.15	Mean	8.5	1.5	85.0	15.0	1.21	0.93			
16	9	5192G	JT-SS-06	8	10	10	0	100.0	0.0	95.79	10	107.66	98.43	11.87	1.18	9.23	0.92	SD	1.7	1.7	16.9	16.9	0.18	0.15			
17	30	5193G	JT-SS-08	1	10	8	2	80.0	20.0	101.96	8	112.43	104.36	10.47	1.31	8.07	1.01	n	8	8	8	8	8	8			
18	29	5193G	JT-SS-08	2	10	9	1	90.0	10.0	94.74	9	105.60	97.34	10.86	1.21	8.26	0.92	Mean	9.1	0.9	91.3	8.8	1.19	0.91			
19	10	5193G	JT-SS-08	3	10	10	0	100.0	0.0	86.88	10	96.39	89.04	9.51	0.95	7.35	0.73	SD	1.1	1.1	11.3	11.3	0.17	0.12			
20	26	5193G	JT-SS-08	4	10	9	1	90.0	10.0	95.60	9	104.33	97.60	8.73	0.97	6.73	0.75	n	8	8	8	8	8	8			
21	11	5193G	JT-SS-08	5	10	6	4	60.0	40.0	80.69	6	89.11	82.54	8.42	1.40	6.57	1.10	Mean	9.1	0.9	91.3	8.8	1.19	0.91			
22	6	5193G	JT-SS-08	6	10	10	0	100.0	0.0	88.54	10	97.91	89.22	11.37	1.14	8.69	0.87	SD	1.4	1.4	14.1	14.1	0.13	0.11			
23	27	5193G	JT-SS-08	7	10	10	0	100.0	0.0	94.09	10	106.40	97.20	12.31	1.23	9.20	0.92	n	8	8	8	8	8	8			
24	18	5193G	JT-SS-08	8	10	6	4	60.0	40.0	92.47	6	101.14	94.32	8.87	1.45	6.82	1.14	Mean	9.1	0.9	91.3	8.8	1.19	0.91			
25	13	5184G	JT-SS-10	1	10	7	3	70.0	30.0	103.87	7	111.44	105.52	7.57	1.08	5.92	0.85	SD	1.1	1.1	11.3	11.3	0.17	0.12			
26	21	5184G	JT-SS-10	2	10	9	1	90.0	10.0	95.31	9	107.61	98.38	12.30	1.37	8.23	1.03	n	8	8	8	8	8	8			
27	17	5194G	JT-SS-10	3	10	10	0	100.0	0.0	89.93	10	101.64	92.78	11.71	1.17	8.86	0.89	Mean	9.1	0.9	91.3	8.8	1.19	0.91			
28	1	5194G	JT-SS-10	4	10	9	1	90.0	10.0	91.57	9	96.12	83.18	7.55	0.84	5.94	0.66	SD	1.1	1.1	11.3	11.3	0.17	0.12			
29	20	5194G	JT-SS-10	5	10	10	0	100.0	0.0	96.56	10	109.27	98.53	12.71	1.27	9.74	0.97	n	8	8	8	8	8	8			
30	3	5184G	JT-SS-10	6	10	10	0	100.0	0.0	91.38	10	104.72	94.54	13.34	1.33	10.18	1.02	Mean	9.1	0.9	91.3	8.8	1.19	0.91			
31	22	5194G	JT-SS-10	7	10	10	0	100.0	0.0	91.56	10	104.05	94.44	12.49	1.25	9.61	0.96	SD	1.1	1.1	11.3	11.3	0.17	0.12			
32	28	5194G	JT-SS-10	8	10	8	2	80.0	20.0	93.17	8	102.89	95.36	9.72	1.22	7.53	0.94	n	8	8	8	8	8	8			

Project Name: P862-3 Chironomus % Mortality

Sample: x1
 Samp ID: JT-SS-06
 Alias: NAS# 5192G
 Replicates: 8
 Mean: 10
 SD: 14.142
 Tr Mean: N/A
 Trans SD: N/A

Ref Samp: x2
 Ref ID: Control
 Alias: NAS# 5195G
 Replicates: 8
 Mean: 6.25
 SD: 5.175
 Tr Mean: N/A
 Trans SD: N/A

Shapiro-Wilk Results:	Levene's Results:	Test Results:
Residual Mean: 0 Residual SD: 10.868 SS: 2244.172 K: 8 b: 42.996 Alpha Level: 0.05 Calculated Value: 0.8237 Critical Value: ≤ 0.887 Normally Distributed: No Override Option: Not Invoked	Test Residual Mean: 12.833 Test Residual SD: 6.43 Ref. Residual Mean: 8.641 Ref. Residual SD: 2.385 Deg. of Freedom: 14 Alpha Level: 0.1 Calculated Value: 1.7288 Critical Value: ≥ 1.761 Variances Homogeneous: Yes	Statistic: Mann-Whitney Balanced Design: Yes Transformation: rank-order Experimental Hypothesis Null: $x_1 \leq x_2$ Alternate: $x_1 > x_2$ Mann-Whitney N1: 8 Mann-Whitney N2: 8 Degrees of Freedom: Experimental Alpha Level: 0.05 Calculated Value: 33 Critical Value: ≥ 49.000 Accept Null Hypothesis: Yes Power: Min. Difference for Power:

Replicate Number	Test Data	Trans. Test Data	Reference Data	Trans. Reference Data	Levene's Test Residuals	Levene's Reference Residuals	Mann-Whitney Ranks	Rankits	Shapiro-Wilk Residuals
1	0	4	0	4	12.833	11.522	4		-12.833
2	40	16	10	11	26.398	6.913	4		-12.833
3	20	15	10	11	13.732	6.913	4		-12.833
4	0	4	0	4	12.833	11.522	4		-12.833
5	0	4	10	11	12.833	6.913	4		-11.522
6	10	11	10	11	5.602	6.913	4		-11.522
7	10	11	10	11	5.602	6.913	4		-11.522
8	0	4	0	4	12.833	11.522	11		5.602
9							11		5.602
10							11		6.913
11							11		6.913
12							11		6.913
13							11		6.913
14							11		6.913
15							15		13.732
16							16		26.398

The percent mortality in test sediment JT-SS-06 was not significantly higher than that of the control sediment at $\alpha=0.05$. -651

Project Name: P862-3 Chironomus % Mortality

Sample: x1
 Samp ID: JT-SS-08
 Alias: NAS# 5193G
 Replicates: 8
 Mean: 15
 SD: 16.903
 Tr Mean: 17.737
 Trans SD: 16.677

Ref Samp: x2
 Ref ID: Control
 Alias: NAS# 5195G
 Replicates: 8
 Mean: 6.25
 SD: 5.175
 Tr Mean: 11.522
 Trans SD: 9.541

Shapiro-Wilk Results:	Levene's Results:	Test Results:
Residual Mean: 0 Residual SD: 11.662 SS: 2583.955 K: 8 b: 48.025 Alpha Level: 0.05 Calculated Value: 0.8926 Critical Value: <= 0.887 Normally Distributed: Yes Override Option: N/A	Test Residual Mean: 13.303 Test Residual SD: 8.71 Ref. Residual Mean: 8.641 Ref. Residual SD: 2.385 Deg. of Freedom: 14 Alpha Level: 0.1 Calculated Value: 1.4601 Critical Value: >= 1.761 Variances Homogeneous: Yes	Statistic: Student's t Balanced Design: Yes Transformation: ArcSin Experimental Hypothesis Null: $x_1 \leq x_2$ Alternate: $x_1 > x_2$ Degrees of Freedom: 14 Experimental Alpha Level: 0.05 Calculated Value: 0.915 Critical Value: >= 1.761 Accept Null Hypothesis: Yes Power: Min. Difference for Power:

Replicate Number	Test Data	Trans. Test Data	Reference Data	Trans. Reference Data	Levene's Test Residuals	Levene's Reference Residuals	Mann-Whitney Ranks	Rankits	Shapiro-Wilk Residuals
1	20	26.565	0	0	8.828	11.522			-17.737
2	10	18.435	10	18.435	0.698	6.913			-17.737
3	0	0	10	18.435	17.737	6.913			-17.737
4	10	18.435	0	0	0.698	11.522			-11.522
5	40	39.232	10	18.435	21.494	6.913			-11.522
6	0	0	10	18.435	17.737	6.913			-11.522
7	0	0	10	18.435	17.737	6.913			0.698
8	40	39.232	0	0	21.494	11.522			0.698
9									6.913
10									6.913
11									6.913
12									6.913
13									6.913
14									8.828
15									21.494
16									21.494

The percent mortality in test sediment JT-SS-08 was not significantly higher than that of the control sediment at $\alpha=0.05$. -621

Project Name: P862-3 Chironomus % Mortality

Sample: x1
 Samp ID: JT-SS-10
 Alias: NAS# 5194G
 Replicates: 8
 Mean: 8.75
 SD: 11.26
 Tr Mean: 0.058
 Trans SD: 1.072

Ref Samp: x2
 Ref ID: Control
 Alias: NAS# 5195G
 Replicates: 8
 Mean: 6.25
 SD: 5.175
 Tr Mean: -0.058
 Trans SD: 0.662

Shapiro-Wilk Results:	Levene's Results:	Test Results:
Residual Mean: Residual SD: SS: K: b: Alpha Level: N/A Calculated Value: N/A Critical Value: N/A Normally Distributed: N/A Override Option: Not Invoked	Test Residual Mean: 0.916 Test Residual SD: 0.436 Ref. Residual Mean: 0.6 Ref. Residual SD: 0.166 Deg. of Freedom: 14 Alpha Level: 0.1 Calculated Value: 1.9139 Critical Value: >= 1.761 Variances Homogeneous: No	Statistic: Approximate t Balanced Design: Yes Transformation: Ranks Experimental Hypothesis Null: $x_1 \leq x_2$ Alternate: $x_1 > x_2$ Degrees of Freedom: 12 Experimental Alpha Level: 0.05 Calculated Value: 0.2603 Critical Value: >= 1.782 Accept Null Hypothesis: Yes Power: Min. Difference for Power:

Replicate Number	Test Data	Trans. Test Data	Reference Data	Trans. Reference Data	Levene's Test Residuals	Levene's Reference Residuals	Mann-Whitney Ranks	Shapiro-Wilk Residuals
1	30	1.766	0	-0.858	1.708	0.8	-0.858	-0.858
2	10	0.422	10	0.422	0.364	0.48	-0.858	-0.858
3	0	-0.858	10	0.422	0.916	0.48	-0.858	-0.858
4	10	0.422	0	-0.858	0.364	0.8	-0.858	-0.858
5	0	-0.858	10	0.422	0.916	0.48	-0.858	-0.858
6	0	-0.858	10	0.422	0.916	0.48	-0.858	-0.858
7	0	-0.858	10	0.422	0.916	0.48	-0.858	-0.858
8	20	1.285	0	-0.858	1.227	0.8	0.422	0.422
9							0.422	0.422
10							0.422	0.422
11							0.422	0.422
12							0.422	0.422
13							0.422	0.422
14							0.422	0.422
15							1.285	1.285
16							1.766	1.766

The percent mortality in test sediment JT-SS-10 was not significantly higher than that of the control sediment at $\alpha=0.05$. -651

Project Name: P862-3 Chironomus Growth (AFDW)

Sample: x1
 Samp ID: JT-SS-06
 Alias: NAS# 5192G
 Replicates: 8
 Mean: 0.978
 SD: 0.106
 Tr Mean: 0.978
 Trans SD: 0.106

Ref Samp: x2
 Ref ID: Control
 Alias: NAS# 5195G
 Replicates: 8
 Mean: 1.22
 SD: 0.091
 Tr Mean: 1.22
 Trans SD: 0.091

Shapiro-Wilk Results:	Levene's Results:	Test Results:
Residual Mean: 0 Residual SD: 0.085 SS: 0.136 K: 8 b: 0.364 Alpha Level: 0.05 Calculated Value: 0.975 Critical Value: ≤ 0.887 Normally Distributed: Yes Override Option: N/A	Test Residual Mean: 0.083 Test Residual SD: 0.058 Ref. Residual Mean: 0.065 Ref. Residual SD: 0.058 Deg. of Freedom: 14 Alpha Level: 0.1 Calculated Value: 0.6009 Critical Value: ≥ 1.761 Variances Homogeneous: Yes	Statistic: Student's t Balanced Design: Yes Transformation: No Transformation Experimental Hypothesis Null: $x_1 \geq x_2$ Alternate: $x_1 < x_2$ Degrees of Freedom: 14 Experimental Alpha Level: 0.05 Calculated Value: 4.9253 Critical Value: ≥ 1.761 Accept Null Hypothesis: No Power: Min. Difference for Power:

Replicate Number	Test Data	Trans. Test Data	Reference Data	Trans. Reference Data	Levene's Test Residuals	Levene's Reference Residuals	Mann-Whitney Ranks	Rankits	Shapiro-Wilk Residuals
1	0.83	0.83	1.05	1.05	0.148	0.17			-0.17
2	1	1	1.22	1.22	0.023	0			-0.148
3	1.04	1.04	1.31	1.31	0.063	0.09			-0.118
4	1.05	1.05	1.25	1.25	0.073	0.03			-0.08
5	0.97	0.97	1.25	1.25	0.008	0.03			-0.058
6	0.86	0.86	1.21	1.21	0.118	0.01			-0.01
7	1.15	1.15	1.33	1.33	0.173	0.11			-0.008
8	0.92	0.92	1.14	1.14	0.058	0.08			0
9									0.023
10									0.03
11									0.03
12									0.063
13									0.073
14									0.09
15									0.11
16									0.173

Average individual growth (AFDW) in test sediment JT-SS-06 is significantly less than that in the control sediment at $\alpha=0.05$. --631

Project Name: P862-3 Chironomus Growth (AFDW)

Sample: x1
 Samp ID: JT-SS-08
 Alias: NAS# 5193G
 Replicates: 8
 Mean: 0.93
 SD: 0.149
 Tr Mean: 0.93
 Trans SD: 0.149

Ref Samp: x2
 Ref ID: Control
 Alias: NAS# 5195G
 Replicates: 8
 Mean: 1.22
 SD: 0.091
 Tr Mean: 1.22
 Trans SD: 0.091

Shapiro-Wilk Results:	Levene's Results:	Test Results:
Residual Mean: 0 Residual SD: 0.106 SS: 0.213 K: 8 b: 0.453 Alpha Level: 0.05 Calculated Value: 0.9628 Critical Value: <= 0.887 Normally Distributed: Yes Override Option: N/A	Test Residual Mean: 0.115 Test Residual SD: 0.084 Ref. Residual Mean: 0.065 Ref. Residual SD: 0.058 Deg. of Freedom: 14 Alpha Level: 0.1 Calculated Value: 1.3811 Critical Value: >= 1.761 Variances Homogeneous: Yes	Statistic: Student's t Balanced Design: Yes Transformation: No Transformation Experimental Hypothesis Null: $x_1 \geq x_2$ Alternate: $x_1 < x_2$ Degrees of Freedom: 14 Experimental Alpha Level: 0.05 Calculated Value: 4.7022 Critical Value: >= 1.761 Accept Null Hypothesis: No Power: Min. Difference for Power:

Replicate Number	Test Data	Trans. Test Data	Reference Data	Trans. Reference Data	Levene's Test Residuals	Levene's Reference Residuals	Mann-Whitney Ranks	Rankits	Shapiro-Wilk Residuals
1	1.01	1.01	1.05	1.05	0.08	0.17			-0.2
2	0.92	0.92	1.22	1.22	0.01	0			-0.18
3	0.73	0.73	1.31	1.31	0.2	0.09			-0.17
4	0.75	0.75	1.25	1.25	0.18	0.03			-0.08
5	1.1	1.1	1.25	1.25	0.17	0.03			-0.06
6	0.87	0.87	1.21	1.21	0.06	0.01			-0.01
7	0.92	0.92	1.33	1.33	0.01	0.11			-0.01
8	1.14	1.14	1.14	1.14	0.21	0.08			-0.01
9									0
10									0.03
11									0.03
12									0.08
13									0.09
14									0.11
15									0.17
16									0.21

Average individual growth (AFDW) in test sediment JT-SS-08 is significantly less than that in the control sediment at $\alpha=0.05$.

-631

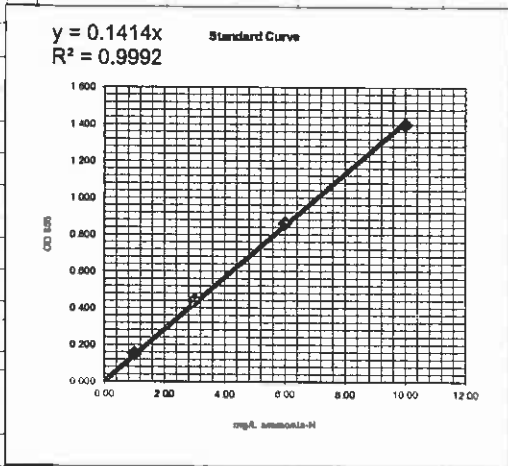
Water Quality Data											
NAS		CLIENT	Overlying water								
BKR	SMPL	DESCRIP	REPL	DAY	TEMP	DO	COND	pH	NH3	HARD	ALK
9	5192G	JT-SS-06	8	0	23.6	7.0	118	7.1	<0.1	26	30
12	5195G	Control	8	0	24.0	7.4	217	7.4	0.3	26	30
16	5193G	JT-SS-08	8	0	23.8	7.2	120	7.3	0.1	34	30
28	5194G	JT-SS-10	8	0	24.0	6.7	136	7.2	0.2	34	30
9	5192G	JT-SS-06	8	1	22.2	6.6					
12	5195G	Control	8	1	22.6	6.8					
16	5193G	JT-SS-08	8	1	22.3	6.8					
28	5194G	JT-SS-10	8	1	23.0	6.4					
9	5192G	JT-SS-06	8	2	22.2	6.7					
12	5195G	Control	8	2	22.6	6.4					
16	5193G	JT-SS-08	8	2	22.4	6.8					
28	5194G	JT-SS-10	8	2	22.9	6.8					
9	5192G	JT-SS-06	8	3	22.4	6.5					
12	5195G	Control	8	3	22.9	6.1					
16	5193G	JT-SS-08	8	3	22.7	6.2					
28	5194G	JT-SS-10	8	3	23.2	6.1					
9	5192G	JT-SS-06	8	4	22.1	6.5					
12	5195G	Control	8	4	22.4	6.1					
16	5193G	JT-SS-08	8	4	22.2	6.3					
28	5194G	JT-SS-10	8	4	22.7	6.7					
9	5192G	JT-SS-06	8	5	22.0	6.6	108	7.2			
12	5195G	Control	8	5	22.4	6.3	139	7.1			
16	5193G	JT-SS-08	8	5	22.2	6.2	112	7.2			
28	5194G	JT-SS-10	8	5	22.5	6.6	117	7.1			
9	5192G	JT-SS-06	8	6	22.1	4.8					
12	5195G	Control	8	6	22.8	4.6					
16	5193G	JT-SS-08	8	6	22.4	4.6					
28	5194G	JT-SS-10	8	6	23.4	4.5					
9	5192G	JT-SS-06	8	7	22.6	5.5					
12	5195G	Control	8	7	23.1	5.3					
16	5193G	JT-SS-08	8	7	22.8	5.3					
28	5194G	JT-SS-10	8	7	23.6	5.9					
9	5192G	JT-SS-06	8	8	22.1	5.4					
12	5195G	Control	8	8	22.6	5.6					
16	5193G	JT-SS-08	8	8	22.3	5.7					
28	5194G	JT-SS-10	8	8	22.9	5.9					
9	5192G	JT-SS-06	8	9	22.4	5.6					
12	5195G	Control	8	9	22.8	5.4					
16	5193G	JT-SS-08	8	9	22.6	5.4					
28	5194G	JT-SS-10	8	9	23.2	5.3					
9	5192G	JT-SS-06	8	10	22.3	6.5	116	7.1	0.2	26	40
12	5195G	Control	8	10	22.9	6.3	129	7.1	0.1	26	40
16	5193G	JT-SS-08	8	10	22.6	6.4	117	7.1	0.1	26	50
28	5194G	JT-SS-10	8	10	23.4	6.2	123	7.1	0.2	34	40
				Mean	22.7	6.1	129	7.2	—	29	36
				SD	0.5	0.7	29	0.1	—	4	7
				n	44	44	12	12	8	8	8
				Min	22.0	4.5	108	7.1	<0.1	26	30
				Max	24.0	7.4	217	7.4	0.3	34	50

AMMONIA EXPOSURE BENCHSHEETS AND ANALYSIS

Total Ammonia-N in Water: Computation Worksheet Salicylate Method (SOP #5492)

Result

Sample description	Dilution factor	OD655	NH ₃ -N (mg/L)	pH	Salinity (ppt)
Blank	----	----	----		
1.0 mg/L NH ₃ -N Std.	----	0.151	1.00		
3.0 mg/L NH ₃ -N Std.	----	0.440	3.00		
6.0 mg/L NH ₃ -N Std.	----	0.862	6.00		
10.0 mg/L NH ₃ -N Std.	----	1.400	10.00		
3.0 mg/L spike	----	0.449	3.17		
3.0 mg/L spike dupl.	----	0.440	3.11		
5.0 mg/L 2nd source		0.710	5.02		



1	Day 0 (2-6-15)				
2	9	1	0.014	ND	
3	12	1	0.044	0.31	
4	16	1	0.020	0.14	
5	28	1	0.029	0.21	
6					
7	Day 10 (2-16-15)				
8	9	1	0.022	0.16	
9	12	1	0.019	0.13	
10	16	1	0.019	0.13	
11	28	1	0.021	0.15	
12					
13					
14					
15					
16					
17					
18					
19					
20					
21					
22					
23					
24					
25					
26					
27					
28					
29					
30					
31					
32					
33					
34					
35					
36					

Reporting limit (mg/L) = 0.10
 Recovery (%) = 104.8
 Precision (RPD) = 2.02
 2nd source (%) = 100.4
 Sample volume (ml): 0.50
 Dilution factor 1

Sample Set Description:

Test No.: 862-3
 Test Day: 0 & 10
 Species: *Chironomus*

Sample Type (check)

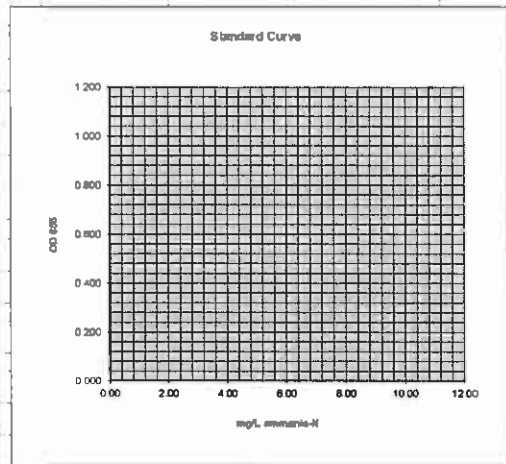
- Bulk Sediment Porewaters
- Test Beaker Porewaters
- Overlying Water

Analyst: JB
 Date analysed: 2/20/2015

Total Ammonia-N in Water: Computation Worksheet Salicylate Method (SOP #5492)

Result

Sample description	Dilution factor	OD655	NH ₃ -N (mg/L)	pH	Salinity (ppt)
Blank	---	---	---		
1.0 mg/L NH ₃ -N Std.	---	.151	1.00		
3.0 mg/L NH ₃ -N Std.	---	.440	3.00		
6.0 mg/L NH ₃ -N Std.	---	.862	6.00		
10.0 mg/L NH ₃ -N Std.	---	1.4	10.00		
3.0 mg/L spike	---	.449			
3.0 mg/L spike dupl.	---	.440			
5.0 mg/L 2nd source		.710			
1 Day 0 (2-6-15)					
2 9	1	.014			
3 12	1	.044			
4 16	1	.020			
5 28	1	.029			
6					
7 Day 10 (2-16-15)					
8 9	1	.022			
9 12	1	.019			
10 16	1	.019			
11 28		.021			
12					
13					
14					
15					
16					
17					
18					
19					
20					
21					
22					
23					
24					
25					
26					
27					
28					
29					
30					
31					
32					
33					
34					
35					
36					



Reporting limit (mg/L) = 0.10

Recovery (%) = #VALUE!

Precision (RPD) = #VALUE!

2nd source (%) = #VALUE!

Sample volume (ml): 0.50

Dilution factor 1

Sample Set Description:

Test No.: 862-3

Test Day: 0 & 10

Species: *Chironomus*

Sample Type (check)

Bulk Sediment Porewaters

Test Beaker Porewaters

Overlying Water

Analyst: JB

Date analysed: 2/20/2015

CHAIN-OF-CUSTODY RECORDS



Sample Custody Record

Samples Shipped to: Alu Aquatic Services

JOB 17800-56 LAB NUMBER _____

PROJECT NAME Jacobson Terminals

HART CROWSER CONTACT Phil Cordeiro

SAMPLED BY: PRC

OBSERVATIONS/COMMENTS/
COMPOSING INSTRUCTIONS

LAB NO.	SAMPLE ID	DESCRIPTION	DATE	TIME	MATRIX	NO. OF CONTAINERS
JT-95-06	51926	11/2/15	1159	Sediment	4	
JT-95-08	51936	↓	1058		4	
JT-95-10	51946	↓	1400		4	

10-day *Chironomus* and growth
 28-day *Hypobrya* and growth
 42-day *Sarothra* and growth

SPECIAL SHIPMENT HANDLING OR STORAGE REQUIREMENTS:
~~None~~

COOLER NO.: _____ STORAGE LOCATION: _____

TURNAROUND TIME:
 24 HOURS 1 WEEK
 48 HOURS STANDARD
 72 HOURS OTHER _____

RELINQUISHED BY: Phil Cordeiro
 SIGNATURE: _____
 PRINT NAME: Phil Cordeiro
 COMPANY: HIC

RECEIVED BY: Gerald Lassner
 SIGNATURE: _____
 PRINT NAME: GERALD LASSNER
 COMPANY: NAS

DATE: 11/13/15 TIME: 1000

DATE: 1-14-15 TIME: 1140

TOTAL NUMBER OF CONTAINERS: 12

SAMPLE RECEIPT INFORMATION
 CUSTODY SEALS: YES NO N/A
 GOOD CONDITION YES NO
 TEMPERATURE: 2.5°C
 SHIPMENT METHOD: HAND OVERNIGHT
 COURIER

From (206) 828-4527
Phil Cordell
Hart Crowser, Inc
1700 Westlake Avenue North
Suite 200
Seattle WA 98109

Origin ID LKEA



Ship Date 13JAN15
ActWgt: 75.0 LB
CAD 4598184/NET3550

Delivery Address Bar Code



SHIP TO: (206) 324-9530
Gerald Irissarri
Northwest Aquatics Scientists
3814 Yakima Rd

BILL SENDER

Ref # 17800-56-02
Invoice #
PO #
Dept #

NEWPORT, OR 97365

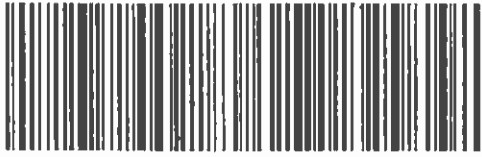
WED - 14 JAN AM
STANDARD OVERNIGHT

TRK# 7725 6121 8674
0201



86 ONPA

97365
OR-US
PDX



522G1RF158A0G

REC'D 11:30 AM 1-14-15

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2. Fold the printed page along the horizontal line.
3. Place label in shipping pouch and affix it to your shipment so that the barcode portion of the label can be read and scanned.

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CUSTODY SEAL

CUSTODY SEAL

CUSTODY SEAL

CUSTODY SEAL

Date 1/13/15

Initials [Signature]

26 of 26

APPENDIX III

RAW DATA – REFERENCE TOXICANT TEST

NORTHWESTERN AQUATIC SCIENCES
ACUTE TOXICITY TEST (ALL SPECIES)

PROTOCOL NO. NAS-

REVIEWED
PAGES 1-5
-6-11

Test No. 999-3391 Client: QC Test Investigator _____
 Test Type (ranging/definitive) _____ Test Length (hr) 96
 Species Chironomus dilutus

STUDY MANAGEMENT

Client: QC test
 Client's Study Monitor: QC test
 Testing Laboratory: Northwestern Aquatic Sciences
 Test Location: Newport Laboratory
 Laboratory's Study Personnel:
 Proj. Man./Study Dir. G.J. Irissarri
 QA Officer L. K. Nemeth
 1. YVES NALCAHAMA 2. URSULA BAZ
 3. _____ 4. _____
 Study Schedule:
 Test Beginning: 2-6-15 0945 Test Ending: 2-10-15 1045

TEST MATERIAL

Description: Potassium Chloride Crystals - ^{Fisher} Lot No.: 114689
 NAS Sample No. _____
 Date of Collection: _____
 Date of Receipt: _____
 Temperature (deg C): _____
 Dissolved oxygen (mg/L): _____
 pH: _____
 Conductivity (umhos/cm): _____
 Hardness (mg/L): _____
 Alkalinity (mg/L): _____
 Salinity (ppt): _____
 Total chlorine (mg/L): _____
 Total ammonia-N (mg/L): _____

DILUTION WATER

Description: Moderately hard synthetic water
 Date of Preparation/Collection: 1-30-15
 Water Quality: Cond. (umhos/cm): 248 Salinity (ppt) _____ pH 8.1
 Hardness (mg/L as CaCO₃): 86 Alkalinity (mg/L as CaCO₃): 70
 Treatments: Aerated ≥ 24 hrs

TEST LOCATION

Test conducted in (circle one): room 1 room 2 trailer water bath other: _____

Randomization chart:

5	20	2.5	1.25	∅	5	5	10	0	0
∅	5	5	5	20	10	2.5	1.25	20	10
2.5	10	10	∅	10	20	∅	∅	1.25	5
1.25	∅	1.25	20	1.25	1.25	1.25	5	10	2.5
10	2.5	20	10	5	∅	10	2.5	5	1.25
20	1.25	∅	2.5	2.5	2.5	20	20	2.5	20

Error codes: 1) Correction of handwriting error
 2) Written in wrong location; entry deleted
 3) Wrong date deleted; replaced with correct date
 4) Error found in measurement; measurement repeated

Test No. 999-3391 Client _____ QC Test _____ Investigator _____

TEST ORGANISMS

Species: Chironomus dilutus Age: ⁶² ~~2ND~~ ^{3rd} instar
 Source: ~~NAS cultures~~ AQUATIC BIOSYSTEMS Date received: 2-5-15
FORT COLLINS, CO

Acclimation Data:

Date	Temp. (deg.C)	pH	DO (mg/L)	Cond. umhos/cm	Hardness (mg/L)	Alkalinity (mg/L)	Feeding	Water changes
2-5-15	20.6	6.9	6.2	436	171	110	Animals fed Tetra Fin	YES
2-6-15	24.8	7.6	6.7	316	86	70	and <i>Selenastrum</i>	-
							Details recorded on	
							Chironomid culture	
							data sheets	
Mean	22.7	7.3	6.5	376	129	90		
S.D.	-	-	-	-	-	-		
(N)	2	2	2	2	2	2		

Photoperiod during acclimation: 16:8, L:D

TEST PROCEDURES AND CONDITIONS

Test concentrations (50% series recommended): 20, 10, 5, 2.5, 1.25, 0 g/L

Test chamber: 30 ml plastic cups Test volume: 20 ml
 Replicates/treatment: 10 Organisms/treatment: 10 (1/rep)
 Test water changes: None Aeration during test: None
 Feeding: 0.25 ml Prime Tropical Flakes (4g/L) suspension per cup on days 0 and 2

Duration: 24-hr, 48-hr, 96-hr Test temperature (deg.C): 23 ± 1
 Beaker placement: Stratified randomization Photoperiod: 16:8, L:D

MISCELLANEOUS NOTES

Test solution preparation:

Working stock: Dissolve 10g KCl crystals in dilution water and dilute to 500 mL.
 Final conc.: 20 g/L.

Test concentration (g/L)	KCl working stock (ml/200ml)	ml of dilution water per 200 ml
20	200	0
10	100	100
5	50	150
2.5	25	175
1.25	12.5	187.5
0	0	0

20-15
0.97

Test No. 999-3391 Client _____ QC Test _____

DAILY RECORD SHEET

Day 0 (2/6/15) 651

Temp Beaker (°C): 23.3

Conc. (g/L)	Temp. (deg.C)	pH	Cond. (umhos/cm)	DO (ppm)	Hardness (mg/L)	Alkalinity (mg/L)	Comments
1. 20	23.4	7.9	220000	8.4	86	60	
2. 10	23.4	8.0	14380	8.3			
3. 5	23.4	8.0	7830	8.3			
4. 2.5	23.4	8.0	4220	8.3			
5. 1.25	23.5	8.0	2320	8.4			
6. 0	23.5	7.9	273	8.4	86	70	

Each replicate fed 0.25 ml Tetra Fin suspension. Initials: 651

Day 1 (2/7/15) 651

Temp Beaker (°C): 22.9

Conc. (g/L)	Temp. (deg.C)	pH	Cond. (umhos/cm)	DO (ppm)	Hardness (mg/L)	Alkalinity (mg/L)	Comments
1. 20							
2. 10							
3. 5							
4. 2.5							
5. 1.25							
6. 0							

Day 2 (2/8/15) 651

Temp Beaker (°C): 22.6

Conc. (g/L)	Temp. (deg.C)	pH	Cond. (umhos/cm)	DO (ppm)	Hardness (mg/L)	Alkalinity (mg/L)	Comments
1. 20							
2. 10							
3. 5							
4. 2.5							
5. 1.25							
6. 0							

Each replicate fed 0.25 ml Tetra Fin suspension. Initials: 651

Day 3 (2/9/15) 651

Temp Beaker (°C): 23.0

Conc. (g/L)	Temp. (deg.C)	pH	Cond. (umhos/cm)	DO (ppm)	Hardness (mg/L)	Alkalinity (mg/L)	Comments
1. 20							
2. 10							
3. 5							
4. 2.5							
5. 1.25							
6. 0							

Day 4 (2/10/15) 651

Temp Beaker (°C): 22.6

Conc. (g/L)	Temp. (deg.C)	pH	Cond. (umhos/cm)	DO (ppm)	Hardness (mg/L)	Alkalinity (mg/L)	Comments
1. 20	-	-	-	-	-	-	
2. 10	-	-	-	-	-	-	
3. 5	23.1	7.8	8960	8.0			
4. 2.5	23.2	7.8	5120	8.1			
5. 1.25	23.5	7.9	2800	8.2			
6. 0	23.3	7.9	364	8.1	94	80	

Mean 23.4 7.9 CONTROL 319 8.3 89 70
SD 0.1 0.1 - 0.1 5 10
n 10 10 2 10 3 3

Test No. 999-3391 Client _____ QC Test _____ Investigator _____

DAILY RECORD SHEET - Survivors

Day 0 (2/6/15) 632

Conc. (g/L)	Survivors in Replicate:										Total
	1	2	3	4	5	6	7	8	9	10	
1. 20											10
2. 10											10
3. 5											10
4. 2.5											10
5. 1.25											10
6. 0											10

Day 1 (2/7/15) 632

Conc. (g/L)	Survivors in Replicate:										Total
	1	2	3	4	5	6	7	8	9	10	
1. 20											10
2. 10											10
3. 5											10
4. 2.5											10
5. 1.25											10
6. 0											10

Day 2 (2/8/15) 631

Conc. (g/L)	Survivors in Replicate:										Total
	1	2	3	4	5	6	7	8	9	10	
1. 20	0	0	0	0	0	0	0	0	0	0	0 (10)
2. 10	0	0	0	0	0	0	0	0	0	0	0 (10)
3. 5											10
4. 2.5											10
5. 1.25											10
6. 0											10

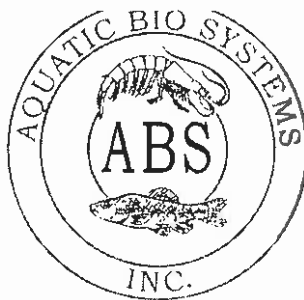
Day 3 (2/9/15) 631

Conc. (g/L)	Survivors in Replicate:										Total
	1	2	3	4	5	6	7	8	9	10	
1. 20	0	0	0	0	0	0	0	0	0	0	0
2. 10	0	0	0	0	0	0	0	0	0	0	0
3. 5			0								9 (10)
4. 2.5											10
5. 1.25											10
6. 0											10

Day 4 (2/10/15) 631

Conc. (g/L)	Survivors in Replicate:										Total
	1	2	3	4	5	6	7	8	9	10	
1. 20	0	0	0	0	0	0	0	0	0	0	0
2. 10	0	0	0	0	0	0	0	0	0	0	0
3. 5			0		0	0					7 (10)
4. 2.5											10
5. 1.25											10
6. 0											10

1300 Blue Spruce Drive, Suite C
Fort Collins, Colorado 80524



Toll Free: 800/331-5916
Tel: 970/484-5091 Fax: 970/484-2514

ORGANISM HISTORY

DATE: 2/4/2015

SPECIES: Chironomus dilutus (formerly C. tentans)

AGE: Deposited 1/24/2015

LIFE STAGE: Second Instar 2/4/2015

HATCH DATE: Emergent date 2/17/2015

BEGAN FEEDING: Immediately

FOOD: Selenastrum sp., Flake slurry

RECEIVED 2-5-15
-GJL

Water Chemistry Record:

	Current	Range
TEMPERATURE:	<u>22°C</u>	<u>22-26°C</u>
SALINITY/CONDUCTIVITY:	<u>--</u>	<u>--</u>
TOTAL HARDNESS (as CaCO ₃):	<u>190 mg/l</u>	<u>100-190 mg/l</u>
TOTAL ALKALINITY (as CaCO ₃):	<u>60 mg/l</u>	<u>50-90 mg/l</u>
pH:	<u>7.50</u>	<u>7.50-8.20</u>

Comments:



Facility Supervisor

Acute 96-hr Toxicity Test-96 Hr Survival

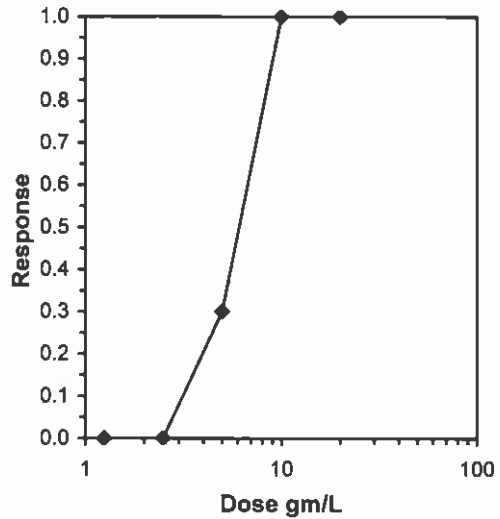
Start Date: 2/6/2015 09:45 Test ID: 999-3391 Sample ID: REF-Ref Toxicant
 End Date: 2/10/2015 10:45 Lab ID: ORNAS-Northwestern Aquati Sample Type: KCL-Potassium chloride
 Sample Date: Protocol: EPAF 91-EPA Freshwater Test Species: CT-Chironomus tentans
 Comments:

Conc-gm/L	1
D-Control	1.0000
1.25	1.0000
2.5	1.0000
5	0.7000
10	0.0000
20	0.0000

Conc-gm/L	Mean	N-Mean	Resp	Not Resp	Total	N	Fisher's Exact P	1-Tailed Critical	Number Resp	Total Number
D-Control	1.0000	1.0000	0	10	10	1			0	10
1.25	1.0000	1.0000	0	10	10	1	1.0000	0.0500	0	10
2.5	1.0000	1.0000	0	10	10	1	1.0000	0.0500	0	10
5	0.7000	0.7000	3	7	10	1	0.1053	0.0500	3	10
10	0.0000	0.0000	10	0	10	1			10	10
20	0.0000	0.0000	10	0	10	1			10	10

Hypothesis Test (1-tail, 0.05)	NOEC	LOEC	ChV	TU
Fisher's Exact Test	5	10	7.07107	

Trim Level	EC50	95% CL	
0.0%	5.7435	4.6982	7.0214
5.0%	5.8220	4.6455	7.2964
10.0%	5.8972	4.5356	7.6674
20.0%	6.0284	4.0589	8.9534
Auto-0.0%	5.7435	4.6982	7.0214

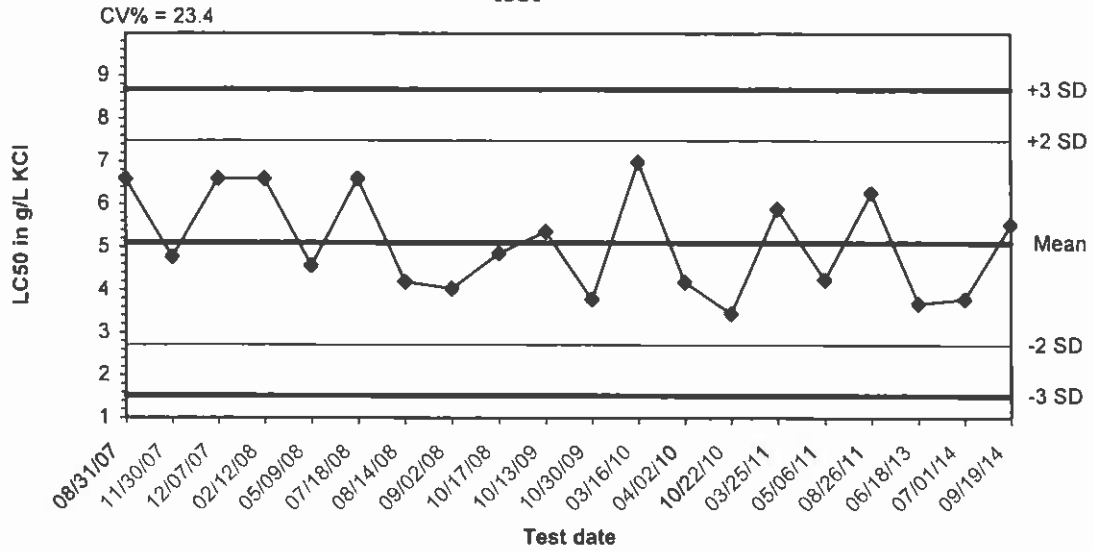


Test: AT-Acute 96-hr Toxicity Test	Test ID: 999-3391
Species: CT-Chironomus tentans	Protocol: EPAF 91-EPA Freshwater
Sample ID: REF-Ref Toxicant	Sample Type: KCL-Potassium chloride
Start Date: 2/6/2015 09:45	End Date: 2/10/2015 10:45 Lab ID: ORNAS-Northwestern Aquatic Sciences

Pos	ID	Rep	Group	Start	24 Hr	48 Hr	72 Hr	96 Hr	Notes
	1	1	D-Control	10				10	
	2	1	1.250	10				10	
	3	1	2.500	10				10	
	4	1	5.000	10				7	
	5	1	10.000	10				0	
	6	1	20.000	10				0	

Comments: *data entry verified against laboratory bench sheets 3-5-15 JDF*

Third instar midge larvae, *Chironomus dilutus*, acute reference toxicant test



Dates	Values	Mean	-2 SD	-3 SD	+2 SD	+3 SD
08/31/07	6.6000	5.0980	2.7119	1.5188	7.4841	8.6772
11/30/07	4.7700	5.0980	2.7119	1.5188	7.4841	8.6772
12/07/07	6.6000	5.0980	2.7119	1.5188	7.4841	8.6772
02/12/08	6.6000	5.0980	2.7119	1.5188	7.4841	8.6772
05/09/08	4.5600	5.0980	2.7119	1.5188	7.4841	8.6772
07/18/08	6.6000	5.0980	2.7119	1.5188	7.4841	8.6772
08/14/08	4.1900	5.0980	2.7119	1.5188	7.4841	8.6772
09/02/08	4.0300	5.0980	2.7119	1.5188	7.4841	8.6772
10/17/08	4.8500	5.0980	2.7119	1.5188	7.4841	8.6772
10/13/09	5.3600	5.0980	2.7119	1.5188	7.4841	8.6772
10/30/09	3.7700	5.0980	2.7119	1.5188	7.4841	8.6772
03/16/10	6.9900	5.0980	2.7119	1.5188	7.4841	8.6772
04/02/10	4.1900	5.0980	2.7119	1.5188	7.4841	8.6772
10/22/10	3.4500	5.0980	2.7119	1.5188	7.4841	8.6772
03/25/11	5.8900	5.0980	2.7119	1.5188	7.4841	8.6772
05/06/11	4.2400	5.0980	2.7119	1.5188	7.4841	8.6772
08/26/11	6.2700	5.0980	2.7119	1.5188	7.4841	8.6772
06/18/13	3.6800	5.0980	2.7119	1.5188	7.4841	8.6772
07/01/14	3.7900	5.0980	2.7119	1.5188	7.4841	8.6772
09/19/14	5.5300	5.0980	2.7119	1.5188	7.4841	8.6772