

# TOXICOLOGY TESTING RESULTS

## PORT GAMBLE BAY RESTORATION MONITORING 2018

### PORT GAMBLE, WASHINGTON

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All testing reported herein was performed consistent with our laboratory's quality assurance program. All results are intended to be considered in their entirety, and EcoAnalysts is not responsible for use of less than the complete report. The test results summarized in this report apply only to the sample(s) evaluated.

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## CONTENTS

<b>1.</b>	<b>INTRODUCTION .....</b>	<b>1</b>
<b>2.</b>	<b>METHODS.....</b>	<b>1</b>
2.1	Sample Collection and Organism Receipt .....	1
2.2	Sample Testing Plan .....	1
2.3	Sample Grain Size and Reference Comparison .....	2
2.4	10-day Amphipod Bioassay .....	4
2.5	20-day Juvenile Polychaete Bioassay .....	4
2.6	Larval Developmental Bioassay .....	5
2.7	Data Analysis and QA/QC .....	6
<b>3.</b>	<b>RESULTS .....</b>	<b>8</b>
3.1	10-day Amphipod Bioassay .....	8
3.2	20-day Juvenile Polychaete Bioassay .....	12
3.3	Larval Development Bioassay .....	17
<b>4.</b>	<b>DISCUSSION.....</b>	<b>23</b>
4.1	Amphipod Test Suitability Determination.....	23
4.2	Juvenile Polychaete Test Suitability Determination .....	23
4.3	Larval Test Suitability Determination.....	25
<b>5.</b>	<b>SUMMARY.....</b>	<b>26</b>
<b>6.</b>	<b>REFERENCES .....</b>	<b>27</b>

## TABLES

Table 2-1. Bioassay Sample Assignments .....	2
Table 2-2. Sample and Reference Grain Size Comparison .....	3
Table 2-3. Reference Station Coordinates .....	3
Table 3-1. Test Results for <i>Eohaustorius estuarius</i> .....	9
Table 3-2. Water Quality Summary for <i>Eohaustorius estuarius</i> .....	10
Table 3-3. Ammonia and Sulfide Summary for <i>Eohaustorius estuarius</i> .....	10
Table 3-4. Test Condition Summary for <i>Eohaustorius estuarius</i> .....	11
Table 3-5. Test Results for <i>Neanthes arenaceodentata</i> .....	13
Table 3-6. Water Quality Summary for <i>Neanthes arenaceodentata</i> .....	14
Table 3-7. Ammonia Summary for <i>Neanthes arenaceodentata</i> .....	14
Table 3-8. Sulfide Summary for <i>Neanthes arenaceodentata</i> .....	15
Table 3-9. Test Condition Summary for <i>Neanthes arenaceodentata</i> .....	16
Table 3-10. Test Results for <i>Mytilus galloprovincialis</i> .....	18
Table 3-11. Water Quality Summary for <i>Mytilus galloprovincialis</i> .....	20
Table 3-12. Ammonia and Sulfide Summary for <i>Mytilus galloprovincialis</i> .....	21
Table 3-13. Test Condition Summary for <i>Mytilus galloprovincialis</i> .....	22
Table 4-1. SMS Comparison for <i>Eohaustorius estuarius</i> .....	23
Table 4-2. SMS Comparison for <i>Neanthes arenaceodentata</i> .....	24
Table 4-3. SMS Comparison for <i>Mytilus galloprovincialis</i> .....	25
Table 5-1. Summary of SMS Evaluation .....	26

## APPENDICES

Appendix A:	Test and Reference Toxicant Test Results
Appendix B:	Statistical Comparisons
Appendix C:	Supporting Documents

## ACRONYMS AND ABBREVIATIONS

AFDW:	Ash-free dry weight
cm:	Centimeter
CSL:	Cleanup Screening Level
°C:	Degrees Celsius
EC <sub>50</sub> :	Effective Concentration that results in a 50% reduction in a sub-lethal endpoint
g:	Grams
LC <sub>50</sub> :	Lethal Concentration that results in a 50% reduction in survival
L:	Liter
µm:	Micrometer
mg:	Milligram
mg/L:	Milligrams per liter
mL:	Milliliter
mm:	Millimeter
NELAP:	National Environmental Laboratory Accreditation Program
NOEC:	No Observed Effect Concentration
OR:	Oregon
ppt:	parts per thousand
PSEP:	Puget Sound Estuary Protocols (PSEP 1995)
SCO:	Sediment Cleanup Objective
SCUM II:	Sediment Cleanup User's Manual II
SMS:	Sediment Management Standards
SOP:	Standard operation procedure
SMARM:	Sediment Management Annual Review Meeting
UIA:	Un-ionized ammonia
USACE:	United States Army Corps of Engineers
USEPA:	United States Environmental Protection Agency
WA:	Washington State
WAC:	Washington Administrative Code
WDOE:	Washington (State) Department of Ecology

## 1. INTRODUCTION

EcoAnalysts conducted biological toxicity testing with sediment samples collected by Anchor QEA, LLC. (Anchor) as part of the restoration monitoring activities being performed at the Port Gamble Bay Cleanup Site in Port Gamble, Washington. Sediments were evaluated for biological effects following guidance provided by the Washington State Department of Ecology (WDOE) Sediment Management Standards (SMS) under the Washington Administrative Code (WAC) 173-204-315. This report presents the results of the toxicity testing portion of the Port Gamble Bay sediment investigation.

## 2. METHODS

This section summarizes the test methods followed for this biological characterization. Test methods followed guidance provided by the Puget Sound Estuary Program (PSEP 1995), the Sediment Cleanup User's Manual II (SCUM II; WDOE 2017), and the various updates presented during the Sediment Management Annual Review Meeting (SMARM). Sediment toxicity was evaluated using three standard PSEP bioassays; the 10-day amphipod test, the 20-day juvenile polychaete survival and growth test, and the 48-hour benthic larval development test.

### 2.1 Sample Collection and Organism Receipt

Nineteen test sediments were collected between September 6 and 19, 2018 and were received at EcoAnalysts on same day as collection. Reference sediments from Carr Inlet, WA were collected by EcoAnalysts on September 28, 2018 and received on the same day. Sediment samples were stored in a walk-in cold room at  $4 \pm 2^{\circ}\text{C}$  in the dark. The test sediment was not sieved prior to testing. All tests were conducted within the eight-week holding time.

Amphipods (*Eohaustorius estuarius*) were supplied by Northwest Amphipod in Newport, Oregon. Animals were held in native sediment at  $15^{\circ}\text{C}$  prior to test initiation. Juvenile polychaete worms (*Neanthes arenaceodentata*) were obtained from Aquatic Toxicology Support in Bremerton, Washington. Juvenile polychaetes were held in seawater at  $20^{\circ}\text{C}$  (*Neanthes* were cultured in water-only and were not held in sediment prior to testing). *Mytilus galloprovincialis* (mussel) broodstock were provided by Taylor Shellfish in Shelton, WA. Broodstock were held in unfiltered seawater at  $14 - 16^{\circ}\text{C}$  prior to spawning.

Native *Eohaustorius* sediment from Yaquina Bay, Oregon was also provided by Northwest Amphipod for use as control sediment treatments for the amphipod and juvenile polychaete tests.

### 2.2 Sample Testing Plan

Based on instruction from Anchor representatives, 12 of the 19 samples collected from the site were selected for bioassay testing. Five samples were identified to be evaluated with the full bioassay suite (three species testing) while seven samples were to be evaluated with the larval benthic test only. Sample locations and their corresponding bioassays are outlined in Table 2-1.

**Table 2-1. Bioassay Sample Assignments**

Sample ID	Benthic Larval Development Test	Juvenile Polychaete Survival and Growth Test	Amphipod Survival Test
SMA1B-IT2-0-10-180907	X	X	X
SMA1B-IT3-0-10-180919	X	X	X
SMA2C-IT3-0-10-180907	X	X	X
SMA2C-IT6-0-10-180907	X	X	X
SMA2C-IT9-0-10-180906	X	X	X
SMA1-ST-0-10-COMP-180917	X		
SMA1A-IT-0-10-COMP-180917	X		
SMA2A-IT-0-10-COMP-180919	X		
SMA2A-ST-0-10-COMP-180918	X		
SMA2B-IT-0-10-COMP-180918	X		
SMA2B-ST-0-10-COMP-180918	X		
BW-15-0-10-180917	X		
SMA2C-IT2-0-10-180906	Sample not analyzed for sediment toxicity		
SMA2C-IT5-0-10-180906			
SMA2C-IT8-0-10-180906			
SMA2C-IT4-0-10-180906			
SMA2C-IT1-0-10-180906			
SMA2C-IT7-0-10-180906			
SMA1B-IT1-0-10-180907			

X = Indicates test to be conducted with associated samples

### 2.3 Sample Grain Size and Reference Comparison

Sediment grain size is one of the characteristics used in selecting the appropriate reference sediment(s) to compare the chemical and biological responses of project sediments. The percent fines value is defined as the amount of sediment that passes through a 62.5- $\mu\text{m}$  sieve, expressed as a percentage of the total sample analyzed. This is also the sum of the silt and clay fraction of sediment. Wet-sieve grain size results for the reference samples was conducted in the field (at the time of collection) and after receipt at the EcoAnalysts laboratory. The percent-fines determination of the project sediments is summarized in Table 2-2.

**Table 2-2. Sample and Reference Grain Size Comparison**

Treatment	Sample Abbreviation	Percent Fines <sup>1</sup>	Treatment Compared To:
CARR-20 (Reference)		40%	
CR-21 (Reference)		14%	
SMA1B-IT2-0-10-180907	SMA-1B-IT2	2%	CR-21
SMA1B-IT3-0-10-180919	SMA-1B-IT3	4%	CR-21
SMA2C-IT3-0-10-180907	SMA-2C-IT3	10%	CR-21
SMA2C-IT6-0-10-180907	SMA-2C-IT6	8%	CR-21
SMA2C-IT9-0-10-180906	SMA-2C-IT9	4%	CR-21
SMA1-ST-0-10-COMP-180917	SMA-1-ST	14%	CR-21
SMA1A-IT-0-10-COMP-180917	SMA-1A-IT	6%	CR-21
SMA2A-IT-0-10-COMP-180919	SMA-2A-IT	0%	CR-21
SMA2A-ST-0-10-COMP-180918	SMA-2A-ST	6%	CR-21
SMA2B-IT-0-10-COMP-180918	SMA-2B-IT	0%	CR-21
SMA2B-ST-0-10-COMP-180918	SMA-2B-ST	0%	CR-21
BW-15-0-10-180917	BW-15	36%	CARR-20

<sup>1</sup> Wet sieve results

Project sample BW-15-0-10-180917 was compared to the Carr Inlet reference CARR-20 for the purposes of evaluating the sediment under the sediment management standards. All other project samples were compared to reference station CR-21.

Station coordinates for the reference samples are summarize in Table 2-3.

**Table 2-3. Reference Station Coordinates**

Station	Latitude	Longitude
CARR-20	47.33298	-122.66981
CR-21	47.33111	-122.67889



## 2.4 10-day Amphipod Bioassay

The 10-day amphipod acute toxicity test was conducted with *Eohaustorius estuarius*. Test organisms were supplied by Northwestern Aquatic Sciences in Newport, OR and held in native sediment at 15°C prior to test initiation. Organisms were fed during the holding period a diet of ground TetraMarin® ad libitum. Native sediment from Yaquina Bay, OR was provided by the organism supplier for use as the control treatment sediment in the test. This matrix has been used successfully in prior tests with this species and is known to support positive organism health and survival.

The amphipod bioassay was conducted as 10-day static exposures with five replicates for each test treatment, reference treatment, and control. Two centimeters of sediment (approximately 175 mL) were placed into each 1-L glass chamber with 775 mL of overlying water. Trickle-flow aeration was provided through glass pipettes, and care was taken to avoid disturbing the sediment surface. Test chambers were placed into randomly assigned positions and allowed to equilibrate to test conditions overnight.

Prior to the test initiation, water quality measurements were taken in all replicates for each test treatment and included dissolved oxygen, temperature, salinity, and pH. Ammonia and sulfide concentrations were measured in both interstitial (pore water) and overlying water at initiation and termination. These measurements were made from a sacrificial surrogate chamber for each test treatment. Sediment pore water was extracted via centrifugation. During the test, water quality was monitored daily in one surrogate replicate per treatment. All water quality instruments were calibrated daily or on their recommended schedule. Records of instrument calibration were retained in the laboratory logs.

To initiate the test, organisms were randomly allocated to each of the test chambers. Initial stocking densities were 20 organisms per test chamber. Amphipods that did not bury within approximately one hour were replaced with healthy amphipods. No food was provided during the 10-day exposure for the amphipod test.

At test termination, sediment from each test chamber was sieved through a 0.5-mm screen to recover all organisms. The number of surviving and dead amphipods was then enumerated.

## 2.5 20-day Juvenile Polychaete Bioassay

The 20-day polychaete survival and growth test was conducted with juvenile polychaete worms (*Neanthes arenaceodentata*). Test organisms were obtained from Aquatic Toxicology Support in Bremerton, WA and held in seawater at 20°C (Neanthes were cultured in water-only and were not held in sediment prior to testing). Organisms were fed during the holding period a diet of ground TetraMarin® ad libitum. Native sediment of the amphipod *Eohaustorius estuarius* from Yaquina Bay, OR was provided by Northwestern Aquatic Sciences for use as the control treatment sediment for the test. This coarse sand control sediment has been routinely tested in conjunction with this species at this laboratory and results from historical testing have demonstrated acceptable organism health and sediment quality.

The polychaete bioassay was conducted as a 20-day static-renewal test, with overlying exchanges of 300 mL of water occurring every third day. Each test treatment, reference treatment, and control consisted of five replicates of 1-L glass chambers, which were filled with two centimeters of sediment (approximately 175 mL) and 775 mL of overlying water. Trickle-flow aeration was provided through glass pipettes, and care was taken to avoid disturbing the sediment surface. Test chambers were then randomly assigned positions and allowed to equilibrate to test conditions overnight.

Prior to the test initiation, water quality measurements were taken in a surrogate chamber for each test treatment and included dissolved oxygen, temperature, salinity, and pH. Ammonia and sulfide concentrations were measured in both interstitial (pore water) and overlying water at initiation and termination. These measurements were made from a sacrificial surrogate chamber for each test treatment. Sediment pore water was extracted via centrifugation. During the test, water quality was monitored daily in one surrogate replicate per treatment. All water quality instruments were calibrated daily or on their recommended schedule. Records of instrument calibration were retained in the laboratory logs.

To initiate the test, organisms were randomly allocated to each of the test chambers. Initial stocking densities were five worms per test chamber. During the test, organisms were fed a diet of 40-mg of TetraMarin® slurry every other day (approximately 8-mg dry weight per worm). Pre-test initial biomass was determined by taking dry weight and ash-free dry weight (AFDW) measurements of three replicates of five worms each on Day 0.

At test termination, sediment from each test chamber was sieved through a 0.5-mm screen. All worms were recovered, enumerated, rinsed in deionized water (to remove salt), and transferred to pre-weighed aluminum foil weigh boats. After drying in an oven at 60°C for approximately 24 hours, each weigh-boat was removed, cooled in a desiccator and weighed to obtain dry weight measurements. They were then heated to 550°C for 2 hours to determine the ashed weight. Ash-free dry weights (AFDW) were calculated to correct for the influence of sediment grain size differences between treatments:

$$\text{AFDW} = \text{Dry weight} - \text{Ashed weight}$$

Both dry weight and AFDW were used to determine individual worm weight and growth rates.

## 2.6 Larval Developmental Bioassay

The bivalve larval development test was conducted with the mussel, *Mytilus galloprovincialis*. Adult organisms were obtained from Taylor Shellfish in Shelton, WA and were held under flowing natural seawater at 12°C prior to spawning induction. Adult mussels were fed during the holding period a marine algal suspension ad libitum. The control treatment for this assay employed a clean seawater treatment (no sediment).

The larval development bioassay was conducted as a static exposure with five replicates for each test treatment, reference treatment, and control. Approximately 18 g ( $\pm 1$  g) of sediment was placed into each 1-L glass chamber with 900 mL of overlying water. Trickle-flow aeration was provided through glass pipettes, and care was taken to avoid disturbing the sediment surface. Test chambers were then shaken for 10 seconds and placed into randomly assigned positions. The larval test was performed without aeration unless dissolved oxygen levels fell below threshold levels for action.

Prior to the test initiation, water quality measurements were taken in the surrogate chamber for each test treatment and included dissolved oxygen, temperature, salinity, and pH. Ammonia and sulfide concentrations were measured in the overlying water at initiation and termination. These measurements were made from a sacrificial surrogate chamber for each test treatment. During the test, water quality was monitored daily in one surrogate replicate per treatment. All water quality instruments were calibrated daily or on their recommended schedule. Records of instrument calibration were retained in the laboratory logs.

To initiate the test, mussels were placed in clean seawater and acclimated at 16°C for approximately 20 minutes. The water bath temperature was then increased over a period of 15 minutes to 20°C. Mussels were held at 20°C and monitored for spawning individuals. Spawning females and males were removed

from the water bath and placed in individual containers with seawater. These individuals were allowed to spawn until sufficient gametes were available to initiate the test. After the spawning period, eggs were transferred to fresh seawater and filtered through a 0.5 mm Nitex® mesh screen to remove large debris, feces, and excess gonadal matter. A composite was made of the sperm and diluted with fresh seawater. The fertilization process was initiated by adding sperm to the isolated egg containers. Egg-sperm solutions were periodically homogenized with a perforated plunger during the fertilization process and sub-samples observed under the microscope for egg and sperm viability. Approximately one to one and a half hours after fertilization, embryo solutions were checked for fertilization rate. Only those embryo stocks with >90% fertilization were used to initiate the tests. Embryo solutions were rinsed free of excess sperm and then combined to create one embryo stock solution. Density of the embryo stock solution was determined by counting the number of embryos in a subsample of homogenized stock solution. This was used to determine the volume of embryo stock solution to deliver approximately 27,000 embryos to each test chamber.

The protocol calls for test termination when 95% of the embryos in the control have reached the prodissoconch I stage (approximately 48-60 hours). At termination, the overlying seawater was decanted into a clean 1-L jar and mixed with a perforated plunger. From this container, a 10 mL subsample was transferred to a scintillation vial and preserved in 5% buffered formalin. Larvae were subsequently stained with a dilute solution of Rose Bengal in 70% alcohol to help visualization of larvae. The number of normal and abnormal larvae was enumerated on an inverted microscope. Normal larvae included all D-shaped prodissoconch I stage larvae. Abnormal larvae included abnormally shaped prodissoconch I larvae and all early stage larvae.

## **2.7 Data Analysis and QA/QC**

All water quality and endpoint data were entered into Excel spreadsheets. Water quality parameters were summarized by calculating the mean, minimum, and maximum values for each test treatment. Endpoint data were calculated for each replicate and the mean values and standard deviations were determined for each test treatment.

All hand-entered data was reviewed for data entry errors, which were corrected prior to summary calculations. A minimum of 10% of all calculations and data sorting were reviewed for errors. Review counts were conducted on any apparent outliers.

For the larval test, the normalized combined mortality and abnormality endpoint was used to evaluate the test sediment. This was based on the number of normal larvae in each treatment and reference sample divided by the mean number of normal larvae in the control replicates, as defined in the SCUM II guidance document (Ecology 2015).

For SMS and CSL suitability determinations, comparisons were made according to DMMP User Manual (DMMP, 2016) and Fox et al. (1998), using BioStat software. Data reported as percent mortality or survival were transformed using an arcsine square root transformation prior to statistical analysis. All data were tested for normality using the Wilk-Shapiro test and equality of variance using Levene's test. Determinations of statistical significance were based on one-tailed Student's t-tests with an alpha of 0.05. A comparison of the larval endpoint relative to the reference was made using an alpha level of 0.10. For samples failing to meet assumptions of normality, a Mann-Whitney test was conducted to determine significance. For those samples failing to meet the assumptions of normality and equality of variance, a t-test on rankits was used.

To evaluate the relative sensitivity of the organisms, reference toxicity tests were performed using standard reference toxicants (Lee 1980). A water-only reference-toxicant test was conducted concurrently with the sediment tests using ammonium chloride. The ammonium chloride reference-toxicant test was used to ensure animals used in the test were healthy and of similar sensitivity to prior tests. This test also provided information on the sensitivity to ammonia concentrations that would possibly be present in the sediments.

Statistical analyses of all dose-response tests were performed using CETIS Comprehensive Toxicity Data Analysis and Database Software version 1.9.2.6. Comparisons between the lab control and each test concentration were performed following recommended USEPA decision matrices (USEPA 2002).

### 3. RESULTS

The results of the sediment testing, including a summary of test results and water quality observations are presented in this section. Data for each of the replicates, as well as laboratory bench sheets are provided Appendix A and statistical analyses are provided in Appendix B.

#### 3.1 10-day Amphipod Bioassay

The bioassay test with *E. estuarius* was validated with 4% mortality in the native sediment control, which met the performance criterion of  $\leq 10\%$  mortality for SMS evaluations. This result indicates that the test conditions were suitable for adequate amphipod survival. Mean mortality in the reference treatments CARR-20 and CR-21 was 4% and 2%, respectively, which met the performance criteria ( $\leq 25\%$  mortality) and indicated that the reference sediments were acceptable for suitability determination. Mean mortality in the five project samples ranged from 1 – 15%. All endpoint results are summarized in Table 3-1. Summaries of water quality measurements, ammonia and sulfide concentrations, and test conditions are presented in Table 3-2, Table 3-3, and Table 3-4.

Water quality parameters were within the acceptable limits throughout the duration of the test.

A reference-toxicant test (positive control) was performed on the batch of test organisms utilized for this study. The  $LC_{50}$  value was within  $\pm 2$  standard deviations from the laboratory historical mean. This result indicates that the test organisms used in this study were of similar sensitivity to those previously tested at EcoAnalysts.

Ammonia concentrations observed in the *E. estuarius* test were below the No Observed Effect Concentration (NOEC) value derived from the concurrent ammonia reference-toxicant test (Table 3-3; compare to NOEC of 134 mg/L). Values were also below the published threshold concentration of 15 mg/L total ammonia (Barton 2002). Therefore, ammonia concentrations within the sediment samples should not have been a contributor to any adverse biological effects observed in the test treatments. Initial sulfide concentrations in interstitial water were unable to be measured due to an insufficient volume of porewater collected from the coarse-grained sediments.

**Table 3-1. Test Results for *Eohaustorius estuarius***

Treatment	Replicate	Number Initiated	Number Surviving	Number Missing or Dead	Percentage Mortality	Mean Percentage Mortality	SD
Control	1	20	19	1	5	4	4.2
	2	20	19	1	0		
	3	20	20	0	5		
	4	20	18	2	0		
	5	20	20	0	0		
CARR-20	1	20	18	2	5	4	4.2
	2	20	20	0	25		
	3	20	19	1	5		
	4	20	20	0	5		
	5	20	19	1	0		
CR-21	1	20	20	0	0	2	2.7
	2	20	19	1	20		
	3	20	20	0	20		
	4	20	20	0	20		
	5	20	19	1	5		
SMA-1B-IT2	1	20	15	5	10	7	10.4
	2	20	19	1	10		
	3	20	19	1	15		
	4	20	20	0	15		
	5	20	20	0	0		
SMA-1B-IT3	1	20	16	4	10	15	7.1
	2	20	16	4	0		
	3	20	16	4	0		
	4	20	19	1	0		
	5	20	18	2	0		
SMA-2C-IT3	1	20	18	2	5	10	6.1
	2	20	17	3	15		
	3	20	17	3	0		
	4	20	20	0	5		
	5	20	18	2	0		
SMA-2C-IT6	1	20	20	0	0	1	2.2
	2	20	20	0	5		
	3	20	20	0	0		
	4	20	20	0	5		
	5	20	19	1	0		
SMA-2C-IT9	1	20	17	3	0	4	6.5
	2	20	20	0	5		
	3	20	19	1	25		
	4	20	20	0	5		
	5	20	20	0	5		

**Table 3-2. Water Quality Summary for *Eohaustorius estuarius***

Treatment	Dissolved Oxygen (mg/L) ≥5.1 mg/L			Temperature (°C) 15 ± 1°C			Salinity (ppt) 28 ± 2 ppt			pH 7 - 9 units		
	Mean	Min	Max	Mean	Min	Max	Mean	Min	Max	Mean	Min	Max
Control	8.3	8.1	8.5	15.3	15.1	15.7	27	27	28	8.0	7.8	8.0
CARR-20	8.3	8.0	8.6	15.4	15.1	15.8	27	27	28	8.0	7.8	8.2
CR-21	8.3	8.1	8.4	15.3	15.1	15.5	28	27	28	8.1	7.6	8.4
SMA-1B-IT2	8.3	7.2	8.5	15.4	15.2	15.7	27	27	28	8.0	7.8	8.1
SMA-1B-IT3	8.3	8.1	8.5	15.3	15.1	15.6	28	27	28	8.1	7.9	8.3
SMA-2C-IT3	8.3	8.2	8.5	15.3	15.1	15.5	27	27	28	8.0	7.9	8.2
SMA-2C-IT6	8.4	8.2	8.6	15.3	15.1	15.5	27	27	28	8.0	7.8	8.1
SMA-2C-IT9	8.2	8.0	8.4	15.3	15.1	15.6	28	28	29	8.0	7.8	8.1

**Table 3-3. Ammonia and Sulfide Summary for *Eohaustorius estuarius***

Treatment	Overlying Ammonia (mg/L Total) <sup>1</sup> NOEC = 134 mg/L		Interstitial Ammonia (mg/L Total) <sup>1</sup> NOEC = 134 mg/L		Overlying Sulfides (mg/L Total) <sup>2</sup> Trigger Value = 1.9 mg/L		Interstitial Sulfides (mg/L Total) <sup>2</sup> Trigger Value = 1.9 mg/L	
	Day 0	Day 10	Day 0	Day 10	Day 0	Day 10	Day 0	Day 10
Control	0.00	0.00	0.00	NM	ND	0.000	NM	NM
CARR-20	0.969	1.31	8.42	3.94	0.001	0.004	NM	0.140
CR-21	0.695	4.01	NM	6.60	ND	ND	NM	NM
SMA-1B-IT2	0.152	0.00	NM	NM	ND	ND	NM	NM
SMA-1B-IT3	0.279	0.00	NM	NM	ND	0.000	NM	NM
SMA-2C-IT3	0.299	0.00	NM	NM	0.012	ND	NM	NM
SMA-2C-IT6	0.00	0.00	NM	NM	ND	ND	NM	NM
SMA-2C-IT9	0.00	8.89	1.53	NM	ND	ND	NM	NM

<sup>1</sup>NOEC (concurrent reference-toxicant test derived) = 134 mg/L total ammonia

<sup>2</sup>Inouye 2015: Total sulfide value 1.9 mg/L derived from hydrogen sulfide dissociation (0.122 mg/L H<sub>2</sub>S @ 15°C, 28 ppt, and 8.1 pH)

ND = not detected; measurement below detection limit

NM = not measured; insufficient porewater recovered for analysis

**Table 3-4. Test Condition Summary for *Eohaustorius estuarius***

Test Conditions: PSEP <i>E. estuarius</i>		
Date sampled	September 6 – 19, 2018	
Date received	September 6 – 19, 2018	
Test dates	October 9 – 19, 2018	
Sample storage conditions	4°C, dark	
Days of holding Recommended: ≤8 weeks (56 days)	20 – 33 Days	
Source of control sediment	Yaquina Bay, OR	
Test Species	<i>Eohaustorius estuarius</i>	
Supplier	Northwest Amphipod, Newport, OR	
Date acquired	October 4, 2018	
Age class	Mature adult, 3-5 mm	
Test Procedures	PSEP 1995 with SMARM revisions, SCUM II (2017) SOP No. SED002.09	
Test location	EcoAnalysts Port Gamble Laboratory	
Test type/duration	10-Day static	
Control water	North Hood Canal seawater, 0.45µm filtered	
Test dissolved oxygen	Recommended: > 5.1 mg/L	Observed: 7.2 – 8.6 mg/L
Test temperature	Recommended: 15 ± 1 °C	Observed: 15.1 – 15.8°C
Test Salinity	Recommended: 28 ± 2 ppt	Observed: 27 – 29 ppt
Test pH	Recommended: 7 - 9	Observed: 7.6 – 8.4
Control Performance Standard SMS	Recommended: Control ≤ 10% mortality	Observed: 4% mortality; Pass
Reference Performance Standard SMS	Recommended: Reference ≤ 25% mortality	Observed mortality: 2%, 4%; Pass
Reference Toxicant LC50 (total ammonia)	LC <sub>50</sub> = 189.1 mg/L	
Mean; Acceptable Range (total ammonia)	148.0; 77.4 – 282.7 mg/L	
NOEC (total ammonia)	134 mg/L	
NOEC (unionized ammonia)	0.74 mg /L	
Test Lighting	50 – 100 foot candles (ambient and constant)	
Test chamber	1-Liter Glass Chamber	
Replicates/treatment	5 + 2 surrogates (one used for WQ measurements throughout the test)	
Organisms/replicate	20	
Exposure volume	175 mL sediment/ 775 mL water	
Feeding	None	
Water renewal	None	
Deviations from Test Protocol	None	



### 3.2 20-day Juvenile Polychaete Bioassay

No mortality was observed in the *N. arenaceodentata* control sediment and mean individual growth (MIG) in the control was 0.808 mg/ind/day (dry weight) and 0.455 mg/ind/day (AFDW). These values fall within the test acceptability criteria of <10% mean mortality and  $\geq 0.38$  mg/ind/day dry weight (Kendall 1996), indicating that the test conditions were suitable for adequate polychaete survival and growth. A summary of the test results for all samples is shown in Table 3-5. Summaries of water quality measurements, ammonia and sulfide concentrations, and test conditions are presented in Table 3-6, Table 3-7, Table 3-8, and Table 3-9.

Mean mortality in the reference treatments ranged from 0 – 8%, meeting the reference performance standard of  $\leq 10\%$  (WDOE 2017; USACE 2015). Mean individual growth for the reference treatments ranged from 0.735 to 0.840 mg/ind/day (dry weight) and 0.473 to 0.475 mg/ind/day (AFDW). When compared to the control, MIG expressed as AFDW ranged from 1.039 to 1.044, which met the reference performance standard of  $\geq 0.80$  (WDOE 2017).

Mortality in the project sediments ranged from 0% to 4%. Mean individual growth (as dry weight) in the test treatments ranged from 0.695 to 0.860 mg/ind/day. Mean individual growth in the AFDW assessment, which removes variability caused by gut contents, ranged from 0.429 to 0.529 mg/ind/day as AFDW.

All water quality parameters were within the acceptable limits throughout the duration of the test. Initial mean individual biomass (pretest) of the test organisms met the recommended criterion of 0.25 – 1.0 mg/individual at 0.859 mg/ind dry weight and 0.760 mg/ind AFDW.

A reference-toxicant test (positive control) was performed on the batch of test organisms utilized for this study. The  $LC_{50}$  value was within control chart limits ( $\pm 2$  standard deviations from the laboratory historical mean). This result indicates that the test organisms used in this study were of similar sensitivity to those previously tested at EcoAnalysts.

Ammonia concentrations observed in the *N. arenaceodentata* test were below the No Observed Effect Concentration (NOEC) value derived from the concurrent ammonia reference-toxicant test (Table 3-7; compare to NOEC of 173 mg/L). Initial sulfide concentrations in the interstitial water were below the NOEC of 3.4 mg/L total sulfides (Kendall and Barton 2004) for all samples.

Table 3-5. Test Results for *Neanthes arenaceodentata*

Treatment	Rep	Number Initiated	Survivors	Mean Mortality (%)	Individual Growth (mg/ind/day)					
					Dry Weight	Mean	Std Dev	AFDW	Mean	Std Dev
Control	1	5	5	0	0.773	0.808	0.142	0.456	0.455	0.060
	2	5	5		0.909			0.487		
	3	5	5		0.736			0.423		
	4	5	5		0.633			0.375		
	5	5	5		0.990			0.534		
CARR-20	1	5	5	0	1.092	0.840	0.264	0.611	0.475	0.114
	2	5	5		0.478			0.317		
	3	5	5		1.094			0.552		
	4	5	5		0.703			0.423		
	5	5	5		0.833			0.473		
CR-21	1	5	5	8	0.775	0.735	0.147	0.461	0.473	0.099
	2	5	5		0.496			0.335		
	3	5	5		0.802			0.530		
	4	5	3		0.886			0.599		
	5	5	5		0.717			0.440		
SMA-1B-IT2	1	5	5	4	0.888	0.860	0.150	0.535	0.527	0.059
	2	5	5		0.802			0.535		
	3	5	5		1.066			0.590		
	4	5	5		0.654			0.429		
	5	5	4		0.890			0.546		
SMA-1B-IT3	1	5	5	0	0.926	0.748	0.140	0.569	0.499	0.066
	2	5	5		0.695			0.480		
	3	5	5		0.741			0.496		
	4	5	5		0.826			0.551		
	5	5	5		0.554			0.401		
SMA-2C-IT3	1	5	5	0	0.788	0.746	0.032	0.499	0.514	0.033
	2	5	5		0.704			0.477		
	3	5	5		0.729			0.565		
	4	5	5		0.761			0.513		
	5	5	5		0.746			0.515		
SMA-2C-IT6	1	5	5	4	0.608	0.782	0.099	0.362	0.466	0.067
	2	5	5		0.801			0.506		
	3	5	5		0.839			0.438		
	4	5	5		0.831			0.530		
	5	5	4		0.832			0.493		
SMA-2C-IT9	1	5	5	0	0.574	0.695	0.098	0.375	0.429	0.040
	2	5	5		0.844			0.482		
	3	5	5		0.678			0.429		
	4	5	5		0.708			0.449		
	5	5	5		0.670			0.412		

**Table 3-6. Water Quality Summary for *Neanthes arenaceodentata***

Treatment	Dissolved Oxygen (mg/L) ≥4.6 mg/L			Temperature (°C) 20 ± 1°C			Salinity (ppt) 28 ± 2 ppt			pH 7 - 9 units		
	Mean	Min	Max	Mean	Min	Max	Mean	Min	Max	Mean	Min	Max
Control	7.4	6.6	7.8	19.7	19.3	20.4	28	27	29	7.8	7.7	8.0
CARR-20	7.2	6.6	7.7	19.9	19.6	20.6	28	28	29	7.8	7.6	8.0
CR-21	6.6	5.5	7.4	19.8	19.4	20.5	28	27	29	7.7	7.4	7.9
SMA-1B-IT2	7.5	7.1	7.7	20.0	19.7	20.6	28	28	29	8.0	7.8	8.1
SMA-1B-IT3	7.4	7.0	7.7	19.9	19.5	20.4	28	27	29	8.0	7.8	8.1
SMA-2C-IT3	7.5	7.2	7.7	19.6	19.3	20.1	28	27	28	7.9	7.8	8.0
SMA-2C-IT6	7.3	6.6	7.7	19.9	19.5	20.5	28	27	28	7.8	7.6	7.9
SMA-2C-IT9	7.4	7.1	7.7	20.0	19.7	20.6	28	28	29	7.8	7.7	8.0

**Table 3-7. Ammonia Summary for *Neanthes arenaceodentata***

Treatment	Overlying Ammonia (mg/L Total) <sup>1</sup> NOEC = 173 mg/L		Interstitial Ammonia (mg/L Total) <sup>1</sup> NOEC = 173 mg/L	
	Day 0	Day 20	Day 0	Day 20
	Control	0.00	1.17	0.00
CARR-20	0.410	1.36	3.21	NM
CR-21	0.304	7.50	NM	NM
SMA-1B-IT2	0.585	0.420	8.75	NM
SMA-1B-IT3	0.622	0.326	NM	NM
SMA-2C-IT3	0.282	1.84	NM	NM
SMA-2C-IT6	0.00	2.77	0.096	NM
SMA-2C-IT9	0.00	1.44	0.296	1.81

<sup>1</sup>NOEC (concurrent reference-toxicant test derived) = 173 mg/L total ammonia

NM = not measured; insufficient porewater recovered for analysis

**Table 3-8. Sulfide Summary for *Neanthes arenaceodentata***

Treatment	Overlying Sulfides (mg/L Total) <sup>1</sup> Trigger Value = 3.4 mg/L		Interstitial Sulfides (mg/L Total) <sup>1</sup> Trigger Value = 3.4 mg/L	
	Day 0	Day 20	Day 0	Day 20
	Control	0.008	0.001	ND
CARR-20	0.014	0.000	0.012	ND
CR-21	0.017	0.003	ND	0.070
SMA-1B-IT2	0.007	0.000	0.240	NM
SMA-1B-IT3	0.011	0.000	NM	NM
SMA-2C-IT3	0.009	0.001	0.110	NM
SMA-2C-IT6	0.008	0.003	0.130	ND
SMA-2C-IT9	0.004	0.001	0.310	0.000

<sup>1</sup>Kendall and Barton 2004

ND = not detected; measurement below detection limit

NM = not measured; insufficient porewater recovered for analysis

**Table 3-9. Test Condition Summary for *Neanthes arenaceodentata***

Test Conditions: PSEP <i>N. arenaceodentata</i>		
Date sampled	September 6 – 19, 2018	
Date received	September 6 – 19, 2018	
Test dates	October 3 – 23, 2018	
Sample storage conditions	4°C, dark	
Days of holding Recommended: ≤8 weeks (56 days)	14 – 27 days	
Source of control sediment	Yaquina Bay, OR	
Test Species	<i>Neanthes arenaceodentata</i>	
Supplier	Aquatic Toxicology Support	
Date acquired	September 28, 2018	
Age class	Juvenile; 23 - 28 Days post emergence	
Test Procedures	PSEP 1995 with SMARM revisions, SCUM II (2017) SOP No. SED009.08	
Test location	EcoAnalysts Port Gamble Laboratory	
Test type/duration	20-Day static renewal	
Control water	North Hood Canal seawater, 0.45µm filtered	
Test dissolved oxygen	Recommended: > 4.6 mg/L	Observed: 5.5 – 7.8 mg/L
Test temperature	Recommended: 20 ± 1 °C	Observed: 19.3 – 20.6 °C
Test Salinity	Recommended: 28 ± 2 ppt	Observed: 27 – 29 ppt
Test pH	Recommended: 7 - 9	Observed: 7.4 – 8.1
Initial biomass	Recommended: 0.5 - 1.0 mg Minimum: 0.25 mg	0.859 mg (dry weight) 0.760 mg (AFDW)
Control Performance Standard	Recommended: Control < 10% mortality	Observed: 0% Pass
	Recommended: ≥ 0.72 mg/ind/day Minimum: ≥ 0.38 mg/ind/day (as Dry Weight)	Observed: 0.808 mg/ind/day; Pass
Reference performance standard (SMS)	Recommended: Mortality ≤20% MIG <sub>Reference</sub> /MIG <sub>Control</sub> ≥ 0.80	0 – 8%; Pass 1.039 – 1.044; Pass
Reference Toxicant LC <sub>50</sub> (total ammonia)	LC <sub>50</sub> = 197.3 mg/L	
Mean; Acceptable Range (total ammonia)	174.9; 122.5 – 249.6 mg/L	
NOEC (total ammonia)	173 mg/L	
NOEC (unionized ammonia)	2.059 mg /L	
Test Lighting	50 – 100 foot candles	
Test chamber	1-Liter Glass Chamber	
Replicates/treatment	5 + 2 surrogates (one used for WQ measurements throughout the test)	
Organisms/replicate	5	
Exposure volume	175 mL sediment/ 775 mL water	
Feeding	40 mg/jar every other day (8 mg/ind every other day)	
Water renewal	Water renewed every third day (1/3 volume of exposure chamber)	
Deviations from Test Protocol	None	

### 3.3 Larval Development Bioassay

The larval development test with *M. galloprovincialis* was validated by 0.977 proportion normal survivorship, defined as the mean number of normal larvae within the control divided by the stocking density. This value was within both the SMS acceptability criteria of  $\geq 0.70$ . A summary of the test results for all samples is shown in Table 3-10. Summaries of water quality measurements, ammonia and sulfide concentrations, and test conditions are presented in, Table 3-11, Table 3-12, and Table 3-13.

Mean normal survival of the reference sediments were between 0.894 and 0.920 of the control response, which met the SMS reference acceptability criteria ( $N_R/N_C$ ) of  $\geq 0.65$ . This is defined as the number of normal larvae in the reference sample(s) divided by the number of normal larvae in the control. The test mean chamber stocking density (measured at test initiation) was 33.6 embryos/mL and was within the test objective of 20 – 40 embryos/mL.

Water quality parameters were within acceptable limits throughout the duration of the test.

A reference-toxicant test (positive control) was performed on the batch of test organisms utilized for this study. The  $LC_{50}$  value was within control chart limits ( $\pm 2$  standard deviations from the laboratory historical mean). This result indicates that the test organisms used in this study were of similar sensitivity to those previously tested at EcoAnalysts.

Ammonia concentrations observed in the *M. galloprovincialis* test were below the No Observed Effect Concentration (NOEC) value derived from the concurrent ammonia reference-toxicant test (Table 3-12; compare to NOEC 6.0 mg/L total ammonia). Initial sulfide concentrations were below the trigger value of 0.009 mg/L total sulfides, derived from hydrogen sulfide dissociation (0.0025 mg/L  $H_2S$ ) and mean water quality data (Inouye 2015). This indicates that ammonia and sulfide concentrations within the sediment samples should not have contributed to any adverse biological effects observed in the test treatments.

Table 3-10. Test Results for *Mytilus galloprovincialis*

Treatment	Rep	Number Normal	Number Abnormal	Mean # Normal (N)	Std. Dev.	Control Normal Survival N <sub>c</sub> /I	Reference Normal Survival Relative to Control N <sub>R</sub> /N <sub>C</sub>	Performance Standard
Control	1	345	14	333.2	15.6	0.977		≥0.70; Meets Criterion
	2	343	11					
	3	344	11					
	4	310	7					
	5	324	12					
CARR-20	1	250	8	297.8	33.7		0.894	≥0.65; Meets Criterion
	2	332	5					
	3	294	6					
	4	285	14					
	5	328	9					
CR-21	1	264	11	310.4	32.6		0.920	≥0.65; Meets Criterion
	2	352	7					
	3	298	7					
	4	325	6					
	5	313	12					
SMA-1B-IT2	1	337	6	302.2	25.2			
	2	274	5					
	3	316	6					
	4	300	3					
	5	284	5					
SMA-1B-IT3	1	318	6	320.4	13.6			
	2	299	8					
	3	336	10					
	4	324	11					
	5	325	4					
SMA-2C-IT3	1	305	10	313.8	21.0			
	2	342	13					
	3	305	8					
	4	289	4					
	5	328	10					
SMA-2C-IT6	1	286	8	303.6	21.0			
	2	318	10					
	3	286	5					
	4	333	5					
	5	295	11					
SMA-2C-IT9	1	287	11	293.2	9.0			
	2	291	14					
	3	283	7					
	4	304	9					
	5	301	9					
SMA-1-ST	1	266	9	277.8	18.1			
	2	289	11					
	3	304	2					
	4	269	13					
	5	261	7					

See Section 4.3 for Larval Test Suitability Determination

Treatment	Rep	Number Normal	Number Abnormal	Mean # Normal (N)	Std. Dev.	Control Normal Survival N <sub>c</sub> /I	Reference Normal Survival Relative to Control N <sub>R</sub> /N <sub>c</sub>	Performance Standard
SMA-1A-IT	1	328	4	307.6	18.7			
	2	292	13					
	3	328	10					
	4	297	10					
	5	293	11					
SMA-2A-IT	1	323	6	317.6	18.9			
	2	287	5					
	3	313	7					
	4	333	4					
	5	332	10					
SMA-2A-ST	1	275	14	304.2	25.3			
	2	336	11					
	3	285	6					
	4	303	8					
	5	322	6					
SMA-2B-IT	1	280	7	298.0	20.3			
	2	297	10					
	3	318	4					
	4	276	10					
	5	319	10					
SMA-2B-ST	1	320	8	301.2	16.1			
	2	302	12					
	3	277	7					
	4	310	5					
	5	297	2					
BW-15	1	268	10	294.4	29.3			
	2	287	30					
	3	340	21					
	4	305	16					
	5	272	19					

I = Mean Initial count (Stocking density); 336

N<sub>c</sub> = Mean Control Normal

N<sub>R</sub> = Mean Reference Normal



**Table 3-11. Water Quality Summary for *Mytilus galloprovincialis***

Treatment	Dissolved Oxygen (mg/L) ≥5.0 mg/L			Temperature (°C) 16± 1°C			Salinity (ppt) 28 ± 1 ppt			pH 7 - 9 units		
	Mean	Min	Max	Mean	Min	Max	Mean	Min	Max	Mean	Min	Max
Control	7.6	7.5	7.7	16.7	16.5	16.9	28	28	28	7.8	7.7	7.8
CARR-20	6.8	6.2	7.5	16.6	16.5	16.8	28	28	28	7.7	7.6	7.7
CR-21	6.7	6.5	7.0	16.6	16.4	16.8	28	28	28	7.7	7.6	7.7
SMA-1B-IT2	7.1	6.9	7.3	16.7	16.5	16.8	28	28	28	7.7	7.7	7.7
SMA-1B-IT3	7.2	7.0	7.7	16.8	16.5	17.0	28	28	28	7.7	7.7	7.7
SMA-2C-IT3	7.2	7.1	7.5	16.6	16.4	16.9	28	28	28	7.7	7.7	7.7
SMA-2C-IT6	7.4	7.2	7.6	16.8	16.4	17.0	28	27	28	7.7	7.7	7.8
SMA-2C-IT9	7.6	7.4	7.7	16.6	16.5	16.7	28	28	28	7.8	7.7	7.8
SMA-1-ST	6.3	5.7	6.9	16.8	16.6	16.9	28	28	28	7.7	7.6	7.7
SMA-1A-IT	6.1	5.7	6.7	16.8	16.7	17.0	28	28	28	7.7	7.7	7.7
SMA-2A-IT	6.8	6.4	7.4	16.9	16.7	17.0	28	28	28	7.7	7.7	7.7
SMA-2A-ST	6.7	6.4	7.4	16.7	16.3	17.0	28	28	28	7.7	7.6	7.7
SMA-2B-IT	6.0	5.7	6.5	16.7	16.4	16.8	28	28	28	7.7	7.6	7.7
SMA-2B-ST	7.1	6.9	7.4	16.7	16.5	16.9	28	28	28	7.7	7.7	7.7
BW-15	6.6	6.1	7.4	16.6	16.3	16.8	28	28	28	7.7	7.6	7.7

**Table 3-12. Ammonia and Sulfide Summary for *Mytilus galloprovincialis***

Treatment	Overlying Ammonia (mg/L Total) <sup>1</sup> NOEC = 6.0 mg/L		Overlying Sulfides (mg/L Total) <sup>2</sup> Trigger Value = 0.009 mg/L	
	Day 0	Final (Day 2)	Day 0	Final (Day 2)
Control	0.00	0.00	ND	0.002
CARR-20	0.00	0.00	0.008	0.005
CR-21	0.00	0.00	0.004	0.002
SMA-1B-IT2	0.00	0.00	ND	0.007
SMA-1B-IT3	0.00	0.00	0.002	0.003
SMA-2C-IT3	0.00	0.00	0.001	ND
SMA-2C-IT6	0.00	0.00	ND	0.003
SMA-2C-IT9	0.00	0.00	0.000	ND
SMA-1-ST	0.00	0.00	0.005	0.003
SMA-1A-IT	0.00	0.00	ND	0.001
SMA-2A-IT	0.00	0.00	ND	0.002
SMA-2A-ST	0.00	0.00	ND	0.001
SMA-2B-IT	0.00	0.00	0.004	0.006
SMA-2B-ST	0.00	0.00	0.007	ND
BW-15	0.00	0.00	0.005	0.004

<sup>1</sup>NOEC (concurrent reference-toxicant test derived) = 6.0 mg/L total ammonia

<sup>2</sup>Inouye 2015: Total sulfide value 0.009 mg/L derived from hydrogen sulfide dissociation (0.0025 mg/L H<sub>2</sub>S @ 16°C, 28 ppt, and 7.7 pH)

ND = Non-detect

**Table 3-13. Test Condition Summary for *Mytilus galloprovincialis***

Test Conditions: PSEP <i>M. galloprovincialis</i>		
Date sampled	September 6 – 19, 2018	
Date received	September 6 – 19, 2018	
Test dates	October 3 – 5, 2018	
Sample storage conditions	4°C, dark	
Holding time Recommended: < 8 weeks (56 days)	14 – 27 Days	
Test Species	<i>Mytilus galloprovincialis</i>	
Supplier	Taylor Shellfish, Shelton, Wa	
Date acquired	October 2, 2018	
Age class	<4-h old embryos	
Test Procedures	PSEP 1995 with SMARM revisions, SCUM II (2017) SOP No. SED005.06	
Test location	EcoAnalysts Port Gamble Laboratory	
Test type/duration	48-60 Hour static test (Actual: 48.5 hours)	
Control water	North Hood Canal sea water, 0.45µm filtered	
Test dissolved oxygen	Recommended: >5.0 mg/L	Observed: 5.7 – 7.7 mg/L
Test temperature	Recommended: 16 ± 1 °C	Observed: 16.3 – 17.0 °C
Test Salinity	Recommended: 28 ± 1 ppt	Observed: 27 – 28 ppt
Test pH	Recommended: 7 - 9	Observed: 7.6 – 7.8
Stocking Density	Recommended: 20 – 40 embryos/mL	Observed: 33.6 embryos/mL
Control performance standard (SMS)	Recommended: Control normal survival ≥ 0.70	Observed: 0.977 Pass
Reference performance standard (SMS)	Recommended: Reference normal survival relative to control ≥ 0.65	Observed: 0.894 – 0.920; Pass
Reference Toxicant	Total Ammonia	Unionized Ammonia
Reference Toxicant EC <sub>50</sub> (total ammonia)	EC <sub>50</sub> = 8.20 mg/L	EC <sub>50</sub> = 0.106 mg/L
Mean; Acceptable Range (total ammonia)	6.81; 3.38 – 13.71 mg/L	0.111; 0.049 – 0.252 mg/L
NOEC Combined proportion normal (total ammonia)	6.0 mg/L	0.086 mg/L
Test Lighting	50 – 100 foot candles	
Test chamber	1-Liter Glass Chamber	
Replicates/treatment	5 + 1 surrogate (used for WQ measurements throughout the test)	
Exposure volume	18 g sediment/ 900 mL water	
Feeding	None	
Water renewal	None	
Deviations from Test Protocol	None	

## 4. DISCUSSION

Sediments were evaluated based on Sediment Management Standards (SMS) criteria. The biological criteria are based on both statistical significance (a statistical comparison) and the degree of biological response (a numerical comparison). The SMS criteria are derived from the Washington Department of Ecology’s Sediment Cleanup User’s Manual II (SCUM II; WDOE 2017). Comparisons were made for each treatment against the reference sample. Two numerical comparisons were made under SMS, the Sediment Cleanup Objective (SCO) and the Cleanup Screening Level (CSL).

### 4.1 Amphipod Test Suitability Determination

Under the SMS program, a treatment will fail SCO if mean mortality in the test sediment relative to the reference sediment is >25% and the difference between mean mortality in the treatment compared to mean mortality in the reference is statistically significant ( $p < 0.05$ ). Treatments fail the CSL if mean mortality in the test treatment >30% relative to the reference sediment and the difference is statistically significant.

Project sediments from the Port Gamble Bay restoration site do not fail the SCO and CSL criteria for the amphipod test as shown in Table 4-1.

**Table 4-1. SMS Comparison for *Eohaustorius estuarius***

Treatment	Mean Mortality (%)	Compared To:	Statistically Different than Reference? (P=0.05)	Mortality Comparison to Reference $M_T - M_R$ (%)	Fails SCO? <sup>1</sup> > 25 %	Fails CSL? <sup>2</sup> > 30 %
Control	4					
CARR-20	4					
CR-21	2					
SMA-1B-IT2	7					
SMA-1B-IT3	15	CR-21	No	13	No	No
SMA-2C-IT3	10	CR-21	No	8	No	No
SMA-2C-IT6	1	CR-21	No	1	No	No
SMA-2C-IT9	4	CR-21	No	2	No	No

<sup>1</sup>SCO: Statistical Significance and  $M_T > 25\%$

<sup>2</sup>CSL: Statistical Significance and  $M_T - M_R > 30\%$

$M_T$  = Treatment Mortality

$M_R$  = Reference Mortality

### 4.2 Juvenile Polychaete Test Suitability Determination

Suitability determinations for the juvenile polychaete test were based on mean individual growth (MIG). A test treatment fails SCO criteria if MIG is statistically lower in the test treatment, relative to the reference, and the ratio of the MIG in the test treatment is <0.70 that of the reference. The treatments will fail CSL criteria if the MIG is significantly lower than the reference treatment and the ratio between the MIG of the treatment and the MIG of the reference is <0.50.

All Port Gamble Bay project sediments pass the SCO and CSL criteria when evaluated on a dry weight and AFDW basis (Table 4-2).

Table 4-2. SMS Comparison for *Neanthes arenaceodentata*

Treatment	MIG (mg/ind/day)	Comparison To:	Statistically Less than Reference? (p=0.05)	MIG Relative to Reference MIG <sub>T</sub> /MIG <sub>R</sub>	Fails SCO? <sup>1</sup> < 0.70	Fails CSL? <sup>2</sup> < 0.50
<b>Dry Weight</b>						
Control	0.808					
CARR-20	0.840					
CR-21	0.735					
SMA-1B-IT2	0.860	CR-21	No	1.17	No	No
SMA-1B-IT3	0.748	CR-21	No	1.02	No	No
SMA-2C-IT3	0.746	CR-21	No	1.01	No	No
SMA-2C-IT6	0.782	CR-21	No	1.06	No	No
SMA-2C-IT9	0.695	CR-21	No	0.95	No	No
<b>Ash-Free Dry Weight</b>						
Control	0.455					
CARR-20	0.475					
CR-21	0.473					
SMA-1B-IT2	0.527	CR-21	No	1.11	No	No
SMA-1B-IT3	0.499	CR-21	No	1.06	No	No
SMA-2C-IT3	0.514	CR-21	No	1.09	No	No
SMA-2C-IT6	0.466	CR-21	No	0.98	No	No
SMA-2C-IT9	0.429	CR-21	No	0.91	No	No

<sup>1</sup>SCO: Statistical Significance and MIG<sub>T</sub>/MIG<sub>R</sub> < 0.70

<sup>2</sup>CSL: Statistical Significance and MIG<sub>T</sub>/MIG<sub>R</sub> < 0.50

MIG<sub>T</sub> = Treatment Mean Individual Growth

MIG<sub>R</sub> = Reference Mean Individual Growth

### 4.3 Larval Test Suitability Determination

Larval test treatments fail SCO criteria if the number of normal larvae in the test treatment is significantly lower ( $p < 0.10$ ) than that of the reference and if the ratio between the normal larval development in the test treatment is less than 0.85 of the normal development in the reference. Treatments fail CSL criteria if the number of normal larvae in the test treatment is significantly lower ( $p < 0.10$ ) than that of the reference and if the ratio between the normal larval development in the test treatment is less than 0.70 of the normal development in the reference.

All Port Gamble Bay project sediments pass both the SCO and CSL criteria for larval bivalve development evaluation (Table 4-3).

**Table 4-3. SMS Comparison for *Mytilus galloprovincialis***

Treatment	Mean Normal Survival (%) <sup>1</sup>	Mean Number Normal	Compared To:	Statistically Less than Reference? ( $p=0.10$ )	Normal Survival to Reference $N_T/N_R$	Fails SCO? <sup>2</sup> <0.85	Fails CSL? <sup>3</sup> <0.70
Control	97.7	333.2					
CARR-20	89.4	297.8					
CR-21	92.0	310.4					
SMA-1B-IT2	90.5	302.2	CR-21	No	0.97	No	No
SMA-1B-IT3	96.0	320.4	CR-21	No	1.03	No	No
SMA-2C-IT3	93.6	313.8	CR-21	No	1.01	No	No
SMA-2C-IT6	91.1	303.6	CR-21	No	0.98	No	No
SMA-2C-IT9	88.0	293.2	CR-21	No	0.94	No	No
SMA-1-ST	83.4	277.8	CR-21	Yes	0.89	No	No
SMA-1A-IT	92.3	307.6	CR-21	No	0.99	No	No
SMA-2A-IT	95.3	317.6	CR-21	No	1.02	No	No
SMA-2A-ST	91.1	304.2	CR-21	No	0.98	No	No
SMA-2B-IT	89.4	298.0	CR-21	No	0.96	No	No
SMA-2B-ST	90.4	301.2	CR-21	No	0.97	No	No
BW-15	87.9	294.4	CARR-20	No	0.99	No	No

<sup>1</sup> Control data is normalized to the stocking density; reference and project treatments are normalized to the control

<sup>2</sup> SCO: Statistical Significance and  $(N_T/N_R) < 0.85$

<sup>3</sup> CSL: Statistical Significance and  $(N_T/N_R) < 0.70$

$N_T$  = Treatment Mean Number Normal

$N_R$  = Reference Mean Number Normal

$N_C$  = Control Mean Number Normal

## 5. SUMMARY

A summary of the biological tests conducted on the Port Gamble Bay restoration site sediments evaluated under the SMS sediment quality criteria (Table 5-1) are provided below.

All project sediments pass the SCO and CSL performance criteria for all tests performed on the Port Gamble Bay restoration site sediments.

**Table 5-1. Summary of SMS Evaluation**

Treatment	Sediment Cleanup Objectives			Cleanup Screening Levels		
	Amphipod	Polychaete	Larval	Amphipod	Polychaete	Larval
SMA-1B-IT2	Pass	Pass	Pass	Pass	Pass	Pass
SMA-1B-IT3	Pass	Pass	Pass	Pass	Pass	Pass
SMA-2C-IT3	Pass	Pass	Pass	Pass	Pass	Pass
SMA-2C-IT6	Pass	Pass	Pass	Pass	Pass	Pass
SMA-2C-IT9	Pass	Pass	Pass	Pass	Pass	Pass
SMA-1-ST			Pass			Pass
SMA-1A-IT			Pass			Pass
SMA-2A-IT			Pass			Pass
SMA-2A-ST			Pass			Pass
SMA-2B-IT			Pass			Pass
SMA-2B-ST			Pass			Pass
BW-15			Pass			Pass

\*Shaded cells represent tests not conducted as part of the test design

## 6. REFERENCES

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**APPENDIX A. .... TEST AND REFERENCE TOXICANT TEST RESULTS**

1. *Eohaustorius estuarius* 10-Day Test
2. *Neanthes arenaceodentata* 20-Day Test
3. *Mytilus galloprovincialis* Bivalve Larval Test

**APPENDIX B. .... STATISTICAL COMPARISONS**

STATISTICAL RESULTS: *EOHAUSTORIUS ESTUARIUS* TEST  
STATISTICAL RESULTS: *NEANTHES ARENACEODENTATA* TEST  
STATISTICAL RESULTS: *MYTILUS GALLOPROVNCIALIS* LARVAL TEST

**APPENDIX C. .... CHAIN-OF-CUSTODY LOGS**

## **APPENDIX A. TEST AND REFERENCE TOXICANT TEST RESULTS**

# **1. *Eohaustorius estuarius* 10-Day Test**

### 10 DAY SOLID PHASE TEST DATA

CLIENT Anchor QEA	PROJECT Port Gamble Monitoring	JOB NO. PG1123	PROJECT MAN. Brian Hester	LABORATORY Port Gamble /	PROTOCOL PSEP 1995	SPECIES <i>Eohaustorius estuarius</i>
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ENDPOINT DATA & OBSERVATIONS

Sample ID	REP	JAR #	INITIAL #	DAY 1	DAY 2	DAY 3	DAY 4	DAY 5	DAY 6	DAY 7	DAY 8	DAY 9	DAY 10	NUMBER REMAINING
				DATE	DATE	DATE	DATE	DATE	DATE	DATE	DATE	DATE	DATE	
				TECHNICIAN	TECHNICIAN	TECHNICIAN	TECHNICIAN	TECHNICIAN	TECHNICIAN	TECHNICIAN	TECHNICIAN	TECHNICIAN	TECHNICIAN	
#S= Number on the Surface #M= Number of Mortality L=Anoxic Surface F=Fungal Patches D=No Air Flow (DO?) U=Excess food N=Normal B=No Burrows <div style="border: 1px solid black; padding: 5px; display: inline-block; margin-top: 10px;">INITIAL # OF ORGANISMS 20</div>				10/10/18	10/11	10/12	10/13	10/14	10/15	10/16	10/17	10/18	10/19	
				WD	SW	RG	MK	MK	SW	SW	SW	MK	SW	
				OBSRVNS.	OBSRVNS.	OBSRVNS.	OBSRVNS.	OBSRVNS.	OBSRVNS.	OBSRVNS.	OBSRVNS.	OBSRVNS.	OBSRVNS.	
Control /	1			N	N	N	N	N	N	N	N	N	N	19
	2			↓	↓	↓	↓	↓	↓	↓	↓	↓	↓	19
	3			↓	↓	↓	↓	↓	↓	↓	↓	↓	↓	20
	4			↓	↓	↓	↓	↓	↓	↓	↓	↓	↓	18
	5			↓	↓	↓	↓	↓	↓	↓	↓	↓	↓	20
CARR-20 /	1			N	N	N	N	N	N	N	N	N	N	18
	2			↓	↓	↓	↓	↓	↓	↓	↓	↓	↓	20
	3			↓	↓	↓	↓	↓	↓	↓	↓	↓	↓	19
	4			↓	↓	↓	↓	↓	↓	↓	↓	↓	↓	20
	5			↓	↓	↓	↓	↓	↓	↓	↓	↓	↓	19
CR-21 /	1			5S	4S	N	N	N	N	N	N	N	N	20
	2			N	N	↓	↓	↓	↓	↓	↓	↓	↓	19
	3			↓	N	↓	↓	↓	↓	↓	↓	↓	↓	20
	4			↓	2S	↓	↓	↓	↓	↓	↓	↓	↓	20
	5			↓	N	↓	↓	↓	↓	↓	↓	↓	↓	19
SMA-1B-IT2 /	1			N	N	N	N	N	N	N	N	N	N	15
	2			↓	↓	↓	↓	↓	↓	↓	↓	↓	↓	19
	3			↓	↓	1S	↓	↓	↓	↓	↓	↓	↓	19
	4			↓	↓	↓	↓	↓	↓	↓	↓	↓	↓	20
	5			↓	↓	↓	↓	↓	↓	↓	↓	↓	↓	20

### 10 DAY SOLID PHASE TEST DATA

CLIENT Anchor QEA	PROJECT Port Gamble Monitoring	JOB NO. PG1123	PROJECT MAN. Brian Hester	LABORATORY Port Gamble /	PROTOCOL PSEP 1995	SPECIES <i>Eohaustorius estuarius</i>
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ENDPOINT DATA & OBSERVATIONS

Sample ID	REP	JAR #	INITIAL #	ENDPOINT DATA & OBSERVATIONS										NUMBER REMAINING
				DAY 1	DAY 2	DAY 3	DAY 4	DAY 5	DAY 6	DAY 7	DAY 8	DAY 9	DAY 10	
				DATE	DATE	DATE	DATE	DATE	DATE	DATE	DATE	DATE	DATE	
#S= Number on the Surface #M= Number of Mortality L=Anoxic Surface F=Fungal Patches D=No Air Flow (DO?) U=Excess food N=Normal B=No Burrows INITIAL # OF ORGANISMS 20				10/10/18	10/11	10/12	10/13	10/14	10/15	10/16	10/17	10/18	10/19	
				VB	SW	RE	ME	ME	SW	SW	SW	ME	SW	
				OBSERVNS.	OBSERVNS.	OBSERVNS.	OBSERVNS.	OBSERVNS.	OBSERVNS.	OBSERVNS.	OBSERVNS.	OBSERVNS.	OBSERVNS.	
SMA-1B-IT3 /	1			N	2S	N	N	N	N	N	N	N	N	16, 1M
	2			↓	N	↓	1S	↓	1S	↓	↓	↓	↓	16
	3			↓	↓	↓	N	↓	2	↓	1S	↓	↓	16
	4			↓	↓	↓	↓	↓	↓	N	↓	↓	↓	19
	5			↓	↓	↓	↓	↓	↓	N	1S	↓	↓	18
SMA-2C-IT3 /	1			1S	2N	N	↓	↓	2	N	N	N	N	18
	2			N	2N	↓	↓	1S	↓	↓	N	↓	↓	17
	3			↓	1S	↓	↓	N	↓	↓	1S	↓	↓	17
	4			↓	2N	↓	↓	↓	↓	↓	N	↓	↓	20
	5			↓	2N	↓	↓	↓	↓	↓	N	↓	↓	18, 1M
SMA-2C-IT6 /	1			N	2N	2N	↓	↓	2	N	N	N	N	20
	2			↓	↓	↓	↓	↓	↓	↓	↓	↓	↓	20
	3			↓	↓	↓	↓	↓	↓	↓	↓	↓	↓	20
	4			↓	↓	↓	↓	↓	↓	↓	↓	↓	↓	20
	5			↓	↓	↓	↓	↓	↓	↓	↓	↓	↓	19
SMA-2C-IT9 /	1			N	2N	2N	↓	↓	2	N	N	N	N	17
	2			↓	↓	↓	↓	↓	↓	↓	↓	↓	↓	20
	3			↓	↓	↓	↓	↓	↓	↓	↓	↓	↓	19
	4			↓	↓	↓	↓	↓	↓	↓	↓	↓	↓	20
	5			↓	↓	↓	↓	↓	↓	↓	↓	↓	↓	20

### 10 DAY SOLID PHASE TEST DATA

CLIENT Anchor QEA	PROJECT Port Gamble Monitoring	SPECIES <i>Eohaustorius estuarius</i>	TEST START DATE Port Gamble	TEST START DATE 09Oct18	TEST END DATE 19Oct18
JOB NUMBER PG1123	PROJECT MANAGER Brian Hester	LABORATORY Port Gamble /	DILUTION WATER BATCH FSW100518.01	PROTOCOL PSEP 1995	

#### WATER QUALITY DATA

TEST CONDITIONS			D.O. (mg/L) >5.1		TEMP (°C) 15 ± 1		SALINITY (ppt) 28 ± 2		pH (pH units) 7 - 9		TECH.	Date
SAMPLE ID	DAY	REP	D.O.		TEMP		SALINITY		pH			
			meter	mg/L	meter	°C	meter	ppt	meter	unit		
Control /	0	1	8	8.3	8	15.3	8	27	8	8.0	MK	10/9/18
Control /	0	2		8.4		15.2		27		8.0		
Control /	0	3		8.3		15.7		27		8.0		
Control /	0	4		8.4		15.2		27		8.0		
Control /	0	5	↓	8.3	↓	15.3	↓	27	↓	8.0	↓	↓
Control /	1	Surr	8	8.3	8	15.3	8	28	8	7.8	US	10/10
Control /	2	Surr	8	8.2	8	15.4	8	28	8	7.9	MARU	10/11
Control /	3	Surr	8	8.1	8	15.5	8	28	8	7.9	RE	10/12
Control /	4	Surr	8	<del>8.0</del> 8.5	8	15.1	8	28	8	8.0	MK	10/13
Control /	5	Surr	8	8.4	8	15.3	8	28	8	8.0	MK	10/14
Control /	6	Surr	8	8.4	8	15.2	8	27	8	7.9	SW	10/15
Control /	7	Surr	8	8.4	8	15.2	8	27	8	8.0	SW	10/16
Control /	8	Surr	8	8.4	8	15.2	8	27	8	7.9	SW	10/17
Control /	9	Surr	8	8.5	8	15.1	8	28	8	7.9	JL	10/18
Control /	10	1	8	8.3	8	15.4	8	27	8	8.0	JK	10/18
Control /	10	2		8.3		15.4		28		8.0		
Control /	10	3		8.3		15.5		28		8.0		
Control /	10	4		8.3		15.4		28		8.0		
Control /	10	5	↓	8.3	↓	15.3	↓	27	↓	8.0	↓	↓

① WC. MK 10/13.

**10 DAY SOLID PHASE TEST DATA**

<b>CLIENT</b> Anchor QEA	<b>PROJECT</b> Port Gamble Monitoring	<b>SPECIES</b> <i>Eohaustorius estuarius</i>	<b>TEST START DATE</b> Port Gamble	<b>TEST START DATE</b> 09Oct18	<b>TEST END DATE</b> 19Oct18
<b>JOB NUMBER</b> PG1123	<b>PROJECT MANAGER</b> Brian Hester	<b>LABORATORY</b> Port Gamble /	<b>DILUTION WATER BATCH</b> FSW100518.01	<b>PROTOCOL</b> PSEP 1995	

**WATER QUALITY DATA**

TEST CONDITIONS	D.O. (mg/L)		TEMP (°C)		SALINITY (ppt)		pH (pH units)		TECH.	Date		
	>5.1		15 ± 1		28 ± 2		7 - 9					
	meter	mg/L	meter	°C	meter	ppt	meter	unit				
CARR-20 /	0	1	8	8.4	8	15.1	8	27	8	8.0	MK	<del>08/10</del> 10/9/18
CARR-20 /	0	2	↓	8.4	↓	15.2	↓	27	↓	8.0	↓	↓
CARR-20 /	0	3	↓	8.2	↓	15.4	↓	27	↓	8.0	↓	↓
CARR-20 /	0	4	↓	8.3	↓	15.4	↓	27	↓	8.0	↓	↓
CARR-20 /	0	5	↓	8.3	↓	15.4	↓	27	↓	8.0	↓	↓
CARR-20 /	1	Surr	8	8.3	8	15.5	8	28	8	7.8	WS	10/10
CARR-20 /	2	Surr	8	8.6	8	15.8	8	28	8	7.9	MAR24	10/11
CARR-20 /	3	Surr	8	8.2	8	15.6	8	28	8	7.9	RE	10/12
CARR-20 /	4	Surr	8	8.5	8	15.2	8	28	8	8.0	MK	10/13
CARR-20 /	5	Surr	8	8.3	8	15.4	8	28	8	8.0	MK	10/14
CARR-20 /	6	Surr	8	8.3	8	15.4	8	27	8	8.0	SW	10/15
CARR-20 /	7	Surr	8	8.2	8	15.4	8	27	8	8.0	SW	10/16
CARR-20 /	8	Surr	8	8.3	8	15.3	8	27	8	8.0	SW	10/17
CARR-20 /	9	Surr	8	8.3	8	15.2	8	27	8	8.0	JL	10/18
CARR-20 /	10	1	8	8.0	8	15.6	8	27	8	8.2	JL	10/19
CARR-20 /	10	2	↓	8.3	↓	15.4	↓	28	↓	8.2	↓	↓
CARR-20 /	10	3	↓	8.2	↓	15.5	↓	27	↓	8.2	↓	↓
CARR-20 /	10	4	↓	8.3	↓	15.5	↓	27	↓	8.1	↓	↓
CARR-20 /	10	5	↓	8.3	↓	15.4	↓	28	↓	8.2	↓	↓

① W.C. MK 10/19.

### 10 DAY SOLID PHASE TEST DATA

CLIENT Anchor QEA	PROJECT Port Gamble Monitoring	SPECIES <i>Eohaustorius estuarius</i>	TEST START DATE Port Gamble	TEST START DATE 09Oct18	TEST END DATE 19Oct18
JOB NUMBER PG1123	PROJECT MANAGER Brian Hester	LABORATORY Port Gamble /	DILUTION WATER BATCH FSW100518.01	PROTOCOL PSEP 1995	

#### WATER QUALITY DATA

TEST CONDITIONS			D.O. (mg/L) >5.1		TEMP (°C) 15 ± 1		SALINITY (ppt) 28 ± 2		pH (pH units) 7 - 9		TECH.	Date
SAMPLE ID	DAY	REP	D.O.		TEMP		SALINITY		pH			
			meter	mg/L	meter	°C	meter	ppt	meter	unit		
CR-21 /	0	1	8	8.3	8	15.3	8	27	8	8.0	MK	10/9/18
CR-21 /	0	2	↓	8.1	↓	15.2	↓	27	↓	7.6	↓	↓
CR-21 /	0	3	↓	8.3	↓	15.5	↓	27	↓	7.9	↓	↓
CR-21 /	0	4	↓	8.2	↓	15.2	↓	27	↓	7.8	↓	↓
CR-21 /	0	5	↓	8.3	↓	15.3	↓	27	↓	8.0	↓	↓
CR-21 /	1	Surr	8	8.4	8	15.1	8	28	8	7.9	WS	10/10
CR-21 /	2	Surr	8	8.3	8	15.4	8	28	8	8.0	MARLH	10/11
CR-21 /	3	Surr	8	8.2	8	15.4	8	28	8	7.9	RE	10/12
CR-21 /	4	Surr	8	8.4	8	15.1	8	28	8	8.1	MK	10/13
CR-21 /	5	Surr	8	8.1	8	15.3	8	28	8	8.2	MK	10/14
CR-21 /	6	Surr	8	8.3	8	15.3	8	28	8	8.2	SW	10/15
CR-21 /	7	Surr	8	8.3	8	15.3	8	28	8	8.3	SW	10/16
CR-21 /	8	Surr	8	8.2	8	15.2	8	28	8	8.3	SW	10/17
CR-21 /	9	Surr	8	8.3	8	15.1	8	28	8	8.3	JL	10/18
CR-21 /	10	1	8	8.3	8	15.5	8	27	8	8.3	↓	10/19
CR-21 /	10	2	↓	8.3	↓	15.4	↓	28	↓	8.4	↓	↓
CR-21 /	10	3	↓	8.3	↓	15.2	↓	28	↓	8.4	↓	↓
CR-21 /	10	4	↓	8.3	↓	15.3	↓	28	↓	8.3	↓	↓
CR-21 /	10	5	↓	8.3	↓	15.4	↓	28	↓	8.3	↓	↓



### 10 DAY SOLID PHASE TEST DATA

CLIENT Anchor QEA	PROJECT Port Gamble Monitoring	SPECIES <i>Eohaustorius estuarius</i>	TEST START DATE Port Gamble	TEST START DATE 09Oct18	TEST END DATE 19Oct18
JOB NUMBER PG1123	PROJECT MANAGER Brian Hester	LABORATORY Port Gamble /	DILUTION WATER BATCH FSW100518.01	PROTOCOL PSEP 1995	

#### WATER QUALITY DATA

TEST CONDITIONS			D.O. (mg/L) >5.1		TEMP (°C) 15 ± 1		SALINITY (ppt) 28 ± 2		pH (pH units) 7 - 9		TECH.	Date
SAMPLE ID	DAY	REP	D.O.		TEMP		SALINITY		pH			
			meter	mg/L	meter	°C	meter	ppt	meter	unit		
SMA-1B-IT2 /	0	1	8	7.8	8	15.4	8	27	8	7.9	MK	10/9/18
SMA-1B-IT2 /	0	2	↓	8.3	↓	15.4	↓	27	↓	8.0	↓	↓
SMA-1B-IT2 /	0	3	↓	8.4	↓	15.3	↓	27	↓	8.0	↓	↓
SMA-1B-IT2 /	0	4	↓	8.4	↓	15.2	↓	27	↓	8.0	↓	↓
SMA-1B-IT2 /	0	5	↓	8.1	↓	15.4	↓	27	↓	8.0	↓	↓
SMA-1B-IT2 /	1	Surr	8	8.4	8	15.4	8	27	8	7.8	WB	10/10
SMA-1B-IT2 /	2	Surr	8	8.3	8	15.6	8	28	8	8.0	MARH	10/11
SMA-1B-IT2 /	3	Surr	8	8.4	8	15.4	8	28	8	7.9	RE	10/12
SMA-1B-IT2 /	4	Surr	8	8.5	8	15.2	8	28	8	8.0	MK	10/13
SMA-1B-IT2 /	5	Surr	8	8.5	8	15.3	8	28	8	8.0	MK	10/14
SMA-1B-IT2 /	6	Surr	8	8.4	8	15.3	8	27	8	8.0	SW	10/15
SMA-1B-IT2 /	7	Surr	8	8.3	8	15.3	8	27	8	8.0	SW	10/16
SMA-1B-IT2 /	8	Surr	8	8.5	8	15.3	8	27	8	8.0	SW	10/17
SMA-1B-IT2 /	9	Surr	8	8.5	8	15.3	8	27	8	8.0	~	10/18
SMA-1B-IT2 /	10	1	8	7.2	8	15.7	8	27	8	8.1	~	10/19
SMA-1B-IT2 /	10	2	↓	8.3	↓	15.6	↓	27	↓	8.0	↓	↓
SMA-1B-IT2 /	10	3	↓	8.3	↓	15.3	↓	28	↓	8.1	↓	↓
SMA-1B-IT2 /	10	4	↓	8.3	↓	15.4	↓	27	↓	8.0	↓	↓
SMA-1B-IT2 /	10	5	↓	8.2	↓	15.3	↓	28	↓	8.1	↓	↓

### 10 DAY SOLID PHASE TEST DATA

CLIENT Anchor QEA	PROJECT Port Gamble Monitoring	SPECIES <i>Eohaustorius estuarius</i>	TEST START DATE Port Gamble	TEST START DATE 09Oct18	TEST END DATE 19Oct18
JOB NUMBER PG1123	PROJECT MANAGER Brian Hester	LABORATORY Port Gamble /	DILUTION WATER BATCH FSW100518.01	PROTOCOL PSEP 1995	

#### WATER QUALITY DATA

TEST CONDITIONS			D.O. (mg/L) >5.1		TEMP (°C) 15 ± 1		SALINITY (ppt) 28 ± 2		pH (pH units) 7 - 9		TECH.	Date
SAMPLE ID	DAY	REP	D.O.		TEMP		SALINITY		pH			
			meter	mg/L	meter	°C	meter	ppt	meter	unit		
SMA-1B-IT3 /	0	1	8	8.3	8	15.2	8	27	8	8.0	MK	10/9/18
SMA-1B-IT3 /	0	2	↓	8.4	↓	15.2	↓	27	↓	8.0	↓	↓
SMA-1B-IT3 /	0	3	↓	8.4	↓	15.3	↓	27	↓	8.0	↓	↓
SMA-1B-IT3 /	0	4	↓	8.3	↓	15.2	↓	27	↓	8.0	↓	↓
SMA-1B-IT3 /	0	5	↓	8.2	↓	15.4	↓	27	↓	8.0	↓	↓
SMA-1B-IT3 /	1	Surr	8	8.4	8	15.2	8	28	8	7.9	UB	10/10
SMA-1B-IT3 /	2	Surr	8	8.4	8	15.4	8	28	8	8.0	MARU	10/11
SMA-1B-IT3 /	3	Surr	8	8.2	8	15.5	8	28	8	7.9	RE	10/12
SMA-1B-IT3 /	4	Surr	8	8.5	8	15.2	8	28	8	8.1	MK	10/13
SMA-1B-IT3 /	5	Surr	8	8.5	8	15.2	8	28	8	8.1	MK	10/14
SMA-1B-IT3 /	6	Surr	8	8.4	8	15.2	8	27	8	8.1	SW	10/15
SMA-1B-IT3 /	7	Surr	8	8.2	8	15.3	8	27	8	8.1	RJ	10/16
SMA-1B-IT3 /	8	Surr	8	8.4	8	15.2	8	28	8	8.1	SW	10/17
SMA-1B-IT3 /	9	Surr	8	8.5	8	15.1	8	28	8	8.1	UL	10/18
SMA-1B-IT3 /	10	1	8	8.2	8	15.4	8	28	8	8.2	UL	10/19
SMA-1B-IT3 /	10	2	↓	8.3	↓	15.2	↓	27	↓	8.2	↓	↓
SMA-1B-IT3 /	10	3	↓	8.4	↓	15.2	↓	28	↓	8.3	↓	↓
SMA-1B-IT3 /	10	4	↓	8.1	↓	15.2	↓	28	↓	8.2	↓	↓
SMA-1B-IT3 /	10	5	↓	8.3	↓	15.6	↓	27	↓	8.1	↓	↓

### 10 DAY SOLID PHASE TEST DATA

CLIENT Anchor QEA	PROJECT Port Gamble Monitoring	SPECIES <i>Eohaustorius estuarius</i>	TEST START DATE Port Gamble	TEST START DATE 09Oct18	TEST END DATE 19Oct18
JOB NUMBER PG1123	PROJECT MANAGER Brian Hester	LABORATORY Port Gamble /	DILUTION WATER BATCH FSW100518.01	PROTOCOL PSEP 1995	

#### WATER QUALITY DATA

TEST CONDITIONS			D.O. (mg/L) >5.1		TEMP (°C) 15 ± 1		SALINITY (ppt) 28 ± 2		pH (pH units) 7 - 9		TECH.	Date
SAMPLE ID	DAY	REP	D.O.		TEMP		SALINITY		pH			
			meter	mg/L	meter	°C	meter	ppt	meter	unit		
SMA-2C-IT3 /	0	1	8	8.3	8	15.3	8	27	8	8.0	MK	10/9/18
SMA-2C-IT3 /	0	2	↓	8.3	↓	15.3	↓	27	↓	8.0	↓	↓
SMA-2C-IT3 /	0	3	↓	8.2	↓	15.4	↓	27	↓	8.0	↓	↓
SMA-2C-IT3 /	0	4	↓	8.3	↓	15.3	↓	27	↓	8.0	↓	↓
SMA-2C-IT3 /	0	5	↓	8.3	↓	15.4	↓	27	↓	8.0	↓	↓
SMA-2C-IT3 /	1	Surr	8	8.3	8	15.3	8	27	8	7.9	WB	10/10
SMA-2C-IT3 /	2	Surr	8	8.4	8	15.4	8	28	8	8.0	MARU	10/11
SMA-2C-IT3 /	3	Surr	8	8.2	8	15.5	8	28	8	7.9	RE	10/12
SMA-2C-IT3 /	4	Surr	8	8.5	8	15.1	8	28	8	8.1	MK	10/13
SMA-2C-IT3 /	5	Surr	8	8.3	8	15.3	8	28	8	8.1	MK	10/14
SMA-2C-IT3 /	6	Surr	8	8.4	8	15.3	8	27	8	8.0	SW	10/15
SMA-2C-IT3 /	7	Surr	8	8.3	8	15.3	8	27	8	8.1	SW	10/16
SMA-2C-IT3 /	8	Surr	8	8.4	8	15.2	8	27	8	8.0	SW	10/17
SMA-2C-IT3 /	9	Surr	8	8.4	8	15.3	8	27	8	7.9	J	10/18
SMA-2C-IT3 /	10	1	8	8.3	8	15.3	8	27	8	8.2	↓	10/19
SMA-2C-IT3 /	10	2	↓	8.3	↓	15.3	↓	28	↓	8.1	↓	↓
SMA-2C-IT3 /	10	3	↓	8.4	↓	15.3	↓	27	↓	8.2	↓	↓
SMA-2C-IT3 /	10	4	↓	8.3	↓	15.3	↓	27	↓	8.1	↓	↓
SMA-2C-IT3 /	10	5	↓	8.2	↓	15.4	↓	27	↓	8.1	↓	↓

### 10 DAY SOLID PHASE TEST DATA

CLIENT Anchor QEA	PROJECT Port Gamble Monitoring	SPECIES <i>Eohaustorius estuarius</i>	TEST START DATE Port Gamble	TEST START DATE 09Oct18	TEST END DATE 19Oct18
JOB NUMBER PG1123	PROJECT MANAGER Brian Hester	LABORATORY Port Gamble /	DILUTION WATER BATCH FSW100518.01	PROTOCOL PSEP 1995	

#### WATER QUALITY DATA

TEST CONDITIONS			D.O. (mg/L) >5.1		TEMP (°C) 15 ± 1		SALINITY (ppt) 28 ± 2		pH (pH units) 7 - 9		TECH.	Date
SAMPLE ID	DAY	REP	D.O.		TEMP		SALINITY		pH			
			meter	mg/L	meter	°C	meter	ppt	meter	unit		
SMA-2C-IT6 /	0	1	8	8.3	8	15.3	8	27	8	8.0	MK	10/9/18
SMA-2C-IT6 /	0	2	↓	8.3	↓	15.4	↓	27	↓	8.0	↓	↓
SMA-2C-IT6 /	0	3	↓	8.3	↓	15.2	↓	27	↓	8.0	↓	↓
SMA-2C-IT6 /	0	4	↓	8.3	↓	15.4	↓	27	↓	8.0	↓	↓
SMA-2C-IT6 /	0	5	↓	8.3	↓	15.3	↓	27	↓	8.0	↓	↓
SMA-2C-IT6 /	1	Surr	8	8.4	8	15.3	8	27	8	7.8	UB	10/10
SMA-2C-IT6 /	2	Surr	8	8.4	8	15.4	8	28	8	7.9	MARU	10/11
SMA-2C-IT6 /	3	Surr	8	8.4	8	15.3	8	28	8	7.9	RE	10/12
SMA-2C-IT6 /	4	Surr	8	8.5	8	15.3	8	28	8	8.0	MK	10/13
SMA-2C-IT6 /	5	Surr	8	8.5	8	15.2	8	28	8	8.0	MK	10/14
SMA-2C-IT6 /	6	Surr	8	8.5	8	15.2	8	27	8	8.0	SW	10/15
SMA-2C-IT6 /	7	Surr	8	8.4	8	15.2	8	27	8	8.0	SW	10/14
SMA-2C-IT6 /	8	Surr	8	8.6	8	15.2	8	27	8	8.0	SW	10/17
SMA-2C-IT6 /	9	Surr	8	8.6	8	15.1	8	27	8	8.0	↓	10/18
SMA-2C-IT6 /	10	1	8	8.3	8	15.3	8	27	8	8.1	✓	10/19
SMA-2C-IT6 /	10	2	↓	8.3	↓	15.3	↓	27	↓	8.1	↓	↓
SMA-2C-IT6 /	10	3	↓	8.3	↓	15.4	↓	28	↓	8.1	↓	↓
SMA-2C-IT6 /	10	4	↓	8.2	↓	15.5	↓	27	↓	8.1	↓	↓
SMA-2C-IT6 /	10	5	↓	8.3	↓	15.2	↓	28	↓	8.1	↓	↓

**10 DAY SOLID PHASE TEST DATA**

CLIENT Anchor QEA	PROJECT Port Gamble Monitoring	SPECIES <i>Eohaustorius estuarius</i>	TEST START DATE Port Gamble	TEST START DATE 09Oct18	TEST END DATE 19Oct18
JOB NUMBER PG1123	PROJECT MANAGER Brian Hester	LABORATORY Port Gamble /	DILUTION WATER BATCH FSW100518.01	PROTOCOL PSEP 1995	

**WATER QUALITY DATA**

TEST CONDITIONS			D.O. (mg/L) >5.1		TEMP (°C) 15 ± 1		SALINITY (ppt) 28 ± 2		pH (pH units) 7 - 9		TECH.	Date
SAMPLE ID	DAY	REP	D.O.		TEMP		SALINITY		pH			
			meter	mg/L	meter	°C	meter	ppt	meter	unit		
SMA-2C-IT9 /	0	1	8	8.4	8	15.2	8	28	8	8.0	MK	10/9/18
SMA-2C-IT9 /	0	2	↓	8.4	↓	15.1	↓	28	↓	8.0	↓	↓
SMA-2C-IT9 /	0	3	↓	8.2	↓	15.3	↓	28	↓	8.0	↓	↓
SMA-2C-IT9 /	0	4	↓	8.3	↓	15.3	↓	28	↓	8.0	↓	↓
SMA-2C-IT9 /	0	5	↓	8.4	↓	15.3	↓	28	↓	8.0	↓	↓
SMA-2C-IT9 /	1	Surr	8	8.3	8	15.2	8	28	8	7.8	WS	10/10
SMA-2C-IT9 /	2	Surr	8	8.2	8	15.4	8	29	8	8.0	MARU	10/11
SMA-2C-IT9 /	3	Surr	8	8.0	8	15.5	8	29	8	7.8	RC	10/12
SMA-2C-IT9 /	4	Surr	8	8.1	8	15.1	8	29	8	7.9	MK	10/13
SMA-2C-IT9 /	5	Surr	8	8.4	8	15.2	8	29	8	8.0	MK	10/14
SMA-2C-IT9 /	6	Surr	8	8.0	8	15.3	8	28	8	8.7.9	SW	10/15
SMA-2C-IT9 /	7	Surr	8	8.1	8	15.3	8	28	8	8.0	SW	10/16
SMA-2C-IT9 /	8	Surr	8	8.1	8	15.2	8	28	8	7.9	SW	10/17
SMA-2C-IT9 /	9	Surr	8	8.1	8	15.1	8	28	8	8.0	JL	10/18
SMA-2C-IT9 /	10	1	8	8.3	8	15.4	8	28	8	8.0	✓	10/19
SMA-2C-IT9 /	10	2	↓	8.3	↓	15.4	↓	28	↓	8.1	↓	↓
SMA-2C-IT9 /	10	3	↓	8.3	↓	15.2	↓	28	↓	8.1	↓	↓
SMA-2C-IT9 /	10	4	↓	8.4	↓	15.6	↓	28	↓	8.1	↓	↓
SMA-2C-IT9 /	10	5	↓	8.3	↓	15.3	↓	28	↓	8.0	↓	↓

① IE 10-15-18 SW

# Ammonia and Sulfide Analysis Record

<b>Client/Project:</b> Anchor / Port Gamble	<b>Organism:</b> Eoh	<b>Test Duration (days):</b> 10
------------------------------------------------	-------------------------	------------------------------------

PRETEST / INITIAL / FINAL / OTHER (circle one)  
 OVERLYING (OV) / POREWATER (PW) (circle one) / Comments: \_\_\_\_\_

Calibration Standards Temperature		Sample temperature should be within $\pm 1^\circ\text{C}$ of standards temperature at time and date of analysis.
<b>Date:</b> 10/9/18	<b>Temperature:</b> 21.6°C	

	Sample ID or Description	Conc. or Rep	Date of Sampling and Initials	Ammonia Value (mg/L)		Meter #/ Temp (°C)		Date of Reading and Initials	Sample Preserved (Y/N)	pH	Sal (ppt)	Sample Volume (mL)	Measured Sulf. (mg/L)	Multiplier	Calculated Sulf. (mg/L)
OV	Ø	SWR	10/9/18 CP	0.00	10	22.2	10/9/18 MK	N				10	ND	1	ND
	CHRP-20			0.969									0.001		0.001
	CP-21			0.695									ND		ND
	SMA IB-1T2			0.152											
	IB-1T3			0.279											
	2C-1T3			0.299											0.012
	2C-1T6			0.00											ND
2C-1T9			0.00											ND	
PW	Ø	SWR	10/9/18 CP	0.00						7.9	28	①			
	CHRP-20			8.42						7.8	29	①			
	CP-21			①											
	SMA IB-1T2			①											
	IB-1T3			①											
	2C-1T3			①											
	2C-1T6			①											
2C-1T9			1.53						7.9	28	①				

① Insufficient pw for analysis. MK 10/9.      ② MK JK 10/09.

# Ammonia and Sulfide Analysis Record

Client/Project: <u>Anchor / Port Gamble</u>	Organism: <u>Eoh</u>	Test Duration (days): <u>10</u>
PRETEST / INITIAL / <b>FINAL</b> / OTHER (circle one)		DAY of TEST: <u>10</u>
OVERLYING (OV) / <b>POREWATER (PW)</b> (circle one) / Comments: _____		

Calibration Standards Temperature		Sample temperature should be within $\pm 1^\circ\text{C}$ of standards temperature at time and date of analysis.
Date:	Temperature:	
<u>10/19/18</u>	<u>21.3 °C</u>	

Sample ID or Description	Conc. or Rep	Date of Sampling and Initials	Ammonia Value (mg/L)	Meter #/ Temp (°C)		Date of Reading and Initials	Sample Preserved (Y/N)	pH	Sal (ppt)	Sample Volume (mL)	Measured Sulf. (mg/L)	Multiplier	Calculated Sulf. (mg/L)
∅	Surr	10/19 RE	0.0	T1	20.3	10/19 RE	N			10	0	1	0
CARR-20	↓	↓	1.31	↓	20.3	↓	↓	↘		↓	0.004	↓	0.004
CR-21	↓	↓	4.01	↓	20.4	↓	↓			↓	ND	↓	ND
SMA 1B-IT2	↓	↓	0.0	↓	20.3	↓	↓			↓	ND	↓	ND
↓ 1B-IT3	↓	↓	0.0	↓	20.3	↓	↓			↓	0	↓	0
↓ 2C-IT3	↓	↓	0.0	↓	20.4	↓	↓			↓	ND	↓	ND
↓ 2C-IT6	↓	↓	0.0	↓	20.4	↓	↓			↓	ND	↓	ND
↓ 2C-IT9	↓	↓	8.89	↓	20.4	↓	↓			↓	ND	↓	ND
∅	②												
CARR-20	Surr	10/19 RE	3.94	T1	21.4	10/19 RE	N	7.07	28	2	0.028	5	0.14
CR-21	↓	↓	6.60	↓	<del>20.3</del>	↓	↓	7.11	29	②		↓	
SMA 1B-IT2	↓	↓		↓		↓	↓					↓	
↓ 1B-IT3	↓	↓		↓		↓	↓					↓	
↓ 2C-IT3	↓	↓		↓		↓	↓					↓	
↓ 2C-IT6	↓	↓		↓		↓	↓					↓	
↓ 2C-IT9	↓	↓		↓		↓	↓					↓	

OV

PW

① IE 10/19 RE 20.3

② Insufficient Volume

## ORGANISM RECEIPT LOG

<b>Date:</b> 10/04/18		<b>Time:</b> 1230		<b>Batch No.</b> NWA100418			
<b>Organism:</b> Eohs							
<b>Source / Supplier:</b> Northwest Amphipod							
<b>No. Ordered:</b> 1910		<b>No. Received:</b> 1910 + 10.1		<b>Source Batch:</b> Collection date, hatch date, etc.): Collected 10/02/18			
<b>Condition of Organisms:</b> Good				<b>Approximate Size or Age:</b> (Days from hatch, life stage, size class, etc.): 3-5mm			
<b>Shipper:</b> Fed Ex				<b>B of L (Tracking No.)</b> 0215 8112 <sup>7869</sup> 9210			
<b>Condition of Container:</b> Good				<b>Received By:</b> JL			
Container	D.O. (mg/L)	Temp. (°C)	Cond. or Sal. (Include Units)	pH (Units)	# Dead	% Dead*	Tech. (Initials)
1	*	7.8	*	—————→			JL
<small>*if &gt;10% contact lab manager</small>							
<b>Notes:</b> * Recvd Dry.							



**Northwest Amphipod**

3814 Yaquina Bay Rd., P.O. Box 542, Newport, OR 97365  
 Tel: 541-867-7225, nwamphipod@gmail.com

<b>SUBJECT: Animal Collection Data Sheet (shipping)</b>			
<b>SOLD TO: EcoAnalysts</b> 4770 NE View Dr. P.O. Box 216 Port Gamble WA 98364		<b>Brian Hester/Collin Ray/Hillary Eicholer</b> 360.297.6040 <b>Julia Baum</b> 360.509.4141	
FedEx# 1817-5747-7		P.O. # PGL	
<b>DATE OF SHIPMENT: 10-3-18</b>			
<b>ANIMAL HISTORY</b>			
<b>Species</b>	<b>Age/Size</b>	<b>Number Shipped</b>	
<i>Eohaustorius estuarius</i>	3-5mm	1910 + 10%	
<b>WATER QUALITY AT TIME OF SHIPMENT</b>			
Temperature (°C): 14.7	pH: 8.0	Salinity (ppt): 35.0	D.O. (mg/L): 8.0
Other:			
<b>PACKAGED BY:</b> GB		<b>DATE:</b> 10-3-18	
<b>FIELD COLLECTION/CULTURE NOTES</b>			
Collected 10-2-18 from Yaquina Bay, OR. Interstitial WQ: Temp: 11.0°C, Salinity 34.5 ppt; salinity adjusted ~5 ppt up or down as needed. Held at 15°C in aerated water.			
<b>ADDITIONAL COMMENTS</b>			
4-liters of 0.5 mm sieved home sediment included.			

**PLEASE RETURN ALL SHIPPING MATERIALS**

If you have any questions, Please call Gary Buhler or Gerald Irissarri at (541) 265-7225. Thank You.





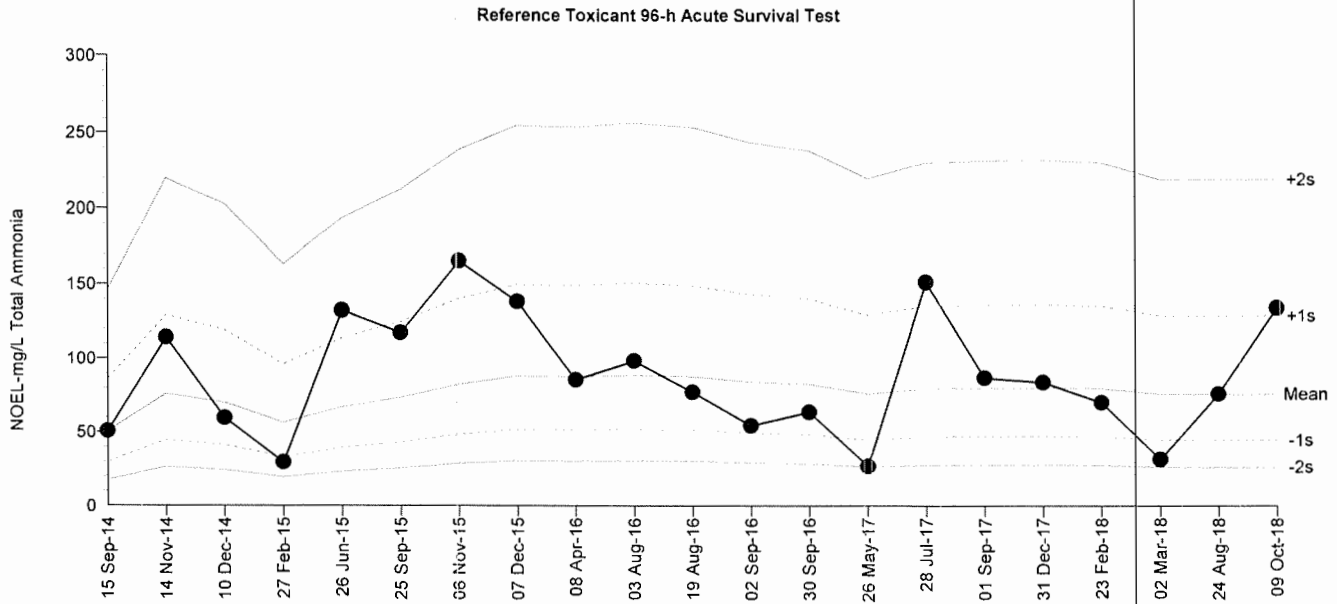
CETIS QC Plot

Report Date: 22 Oct-18 16:15 (1 of 1)

Reference Toxicant 96-h Acute Survival Test

All Matching Labs

Test Type: Survival Organism: Eohaustorius estuarius (Amphipod) Material: Total Ammonia  
 Protocol: EPA/600/R-94/025 (1994) Endpoint: Proportion Survived Source: Reference Toxicant-REF

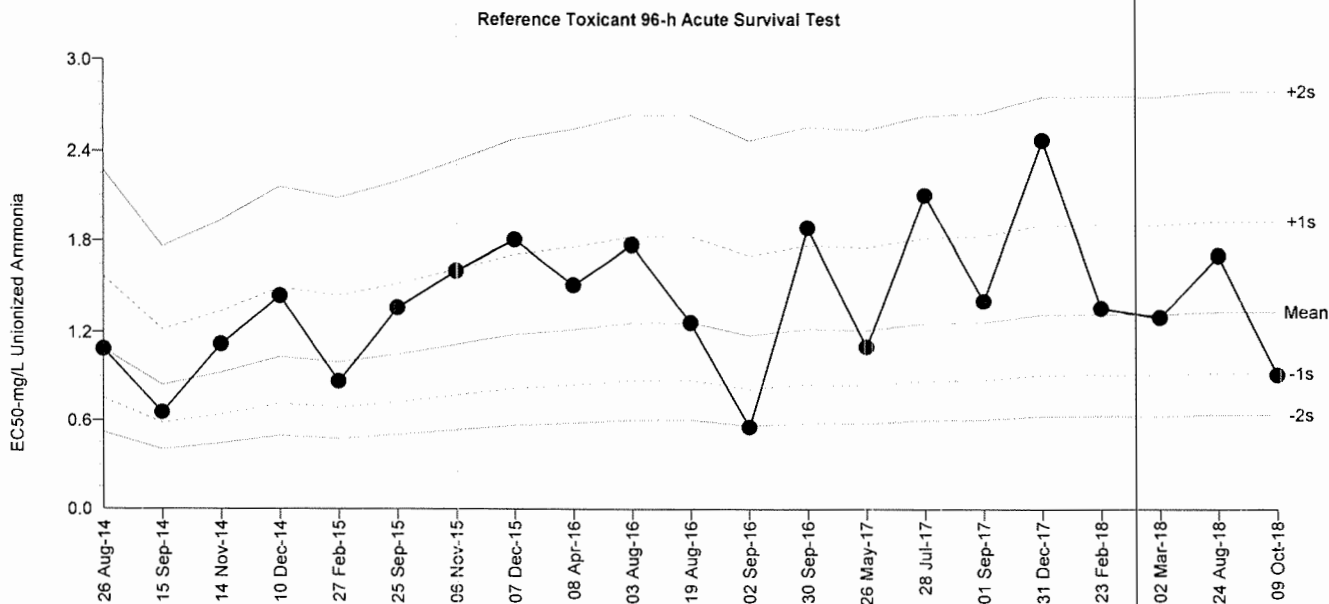


Mean: 75.57 Count: 20 -1s Warning Limit: 44.4 -2s Action Limit: 26.08  
 Sigma: n/a CV: 57.20% +1s Warning Limit: 128.6 +2s Action Limit: 219

Quality Control Data

Point	Year	Month	Day	Time	QC Data	Delta	Sigma	Warning	Action	Test ID	Analysis ID	Laboratory
1	2014	Sep	15	15:10	50.5	-25.07	-0.7577			07-1282-2061	16-3885-0935	ENVIRON
2		Nov	14	14:25	114	38.43	0.7731			09-0717-5355	07-0500-8008	ENVIRON
3		Dec	10	15:50	59.4	-16.17	-0.4525			19-3485-9112	07-0579-1018	ENVIRON
4	2015	Feb	27	12:35	29.3	-46.27	-1.781	(-)		19-3876-5860	19-7961-3594	ENVIRON
5		Jun	26	13:20	132	56.43	1.049	(+)		00-5720-1886	15-3704-4199	ENVIRON
6		Sep	25	17:30	117	41.43	0.8219			05-7835-3625	21-0939-3919	ENVIRON
7		Nov	6	15:30	165	89.43	1.468	(+)		07-0462-4762	19-7906-3673	ENVIRON
8		Dec	7	15:58	138	62.43	1.132	(+)		18-5380-2632	00-7335-5231	ENVIRON
9	2016	Apr	8	14:40	85.2	9.634	0.2256			20-3339-4511	16-7438-0764	ENVIRON
10		Aug	3	16:55	98	22.43	0.4888			15-5854-7986	05-8855-9934	ENVIRON
11			19	14:25	76.9	1.334	0.03291			10-0746-9736	12-8850-4495	ENVIRON
12		Sep	2	16:25	54.1	-21.47	-0.6282			06-2389-4542	18-8647-7799	ENVIRON
13			30	15:00	63.2	-12.37	-0.336			16-2341-4864	17-9345-6065	ENVIRON
14	2017	May	26	13:00	26.6	-48.97	-1.963	(-)		06-2743-8362	12-3565-7845	EcoAnalysts
15		Jul	28	14:20	151	75.43	1.302	(+)		14-8451-4586	09-8418-8824	EcoAnalysts
16		Sep	1	15:45	86.5	10.93	0.2541			02-8963-0820	17-0422-4621	EcoAnalysts
17		Dec	31	15:47	83.6	8.034	0.19			09-7306-1854	06-1883-5465	EcoAnalysts
18	2018	Feb	23	13:35	69.7	-5.866	-0.1519			21-0530-3984	16-8514-1800	EcoAnalysts
19		Mar	2	11:45	31.2	-44.37	-1.663	(-)		11-4485-4691	15-7110-1336	EcoAnalysts
20		Aug	24	15:55	75.6	0.03449	0.0008578			16-5739-3244	15-9917-9528	EcoAnalysts
21		Oct	9	15:00	134	58.43	1.077	(+)		20-4720-6933	14-7074-1873	EcoAnalysts

Reference Toxicant 96-h Acute Survival Test			All Matching Labs
Test Type: Survival	Organism: Eohaustorius estuarius (Amphipod)	Material: Unionized Ammonia	
Protocol: EPA/600/R-94/025 (1994)	Endpoint: Proportion Survived	Source: Reference Toxicant-REF	



Mean: 1.338      Count: 20      -1s Warning Limit: 0.9263      -2s Action Limit: 0.6412  
 Sigma: n/a      CV: 38.10%      +1s Warning Limit: 1.933      +2s Action Limit: 2.792

Quality Control Data

Point	Year	Month	Day	Time	QC Data	Delta	Sigma	Warning	Action	Test ID	Analysis ID	Laboratory
1	2014	Aug	26	15:45	1.087	-0.251	-0.5648			16-9917-4183	13-7453-5343	ENVIRON
2		Sep	15	15:10	0.6543	-0.6837	-1.945	(-)		04-2286-3837	03-1229-8693	ENVIRON
3		Nov	14	14:25	1.119	-0.2189	-0.4857			07-5753-6828	00-1415-6148	ENVIRON
4		Dec	10	15:50	1.441	0.1025	0.2008			04-0714-3304	08-0742-5225	ENVIRON
5	2015	Feb	27	12:35	0.8668	-0.4712	-1.18	(-)		10-1977-7129	06-3048-0232	ENVIRON
6		Sep	25	17:30	1.361	0.02276	0.04586			00-7510-8480	16-9779-9851	ENVIRON
7		Nov	6	15:30	1.605	0.2673	0.4952			14-1974-2437	14-7486-0204	ENVIRON
8		Dec	7	15:58	1.807	0.4686	0.8163			12-1918-7694	00-1085-2209	ENVIRON
9	2016	Apr	8	14:40	1.512	0.1736	0.3318			17-7738-6530	02-5159-2977	ENVIRON
10		Aug	3	16:55	1.775	0.4365	0.7677			15-5470-2613	20-0153-1348	ENVIRON
11			19	14:25	1.264	-0.07434	-0.1554			11-7594-3529	18-2266-1841	ENVIRON
12		Sep	2	16:25	0.5558	-0.7822	-2.389	(-)	(-)	20-2236-1025	01-7459-0032	ENVIRON
13			30	15:00	1.885	0.5468	0.9316			12-0597-8760	12-1436-9613	ENVIRON
14	2017	May	26	13:00	1.101	-0.2374	-0.5312			15-8049-8093	00-1911-6893	EcoAnalysts
15		Jul	28	14:20	2.103	0.7648	1.229	(+)		11-4327-6237	03-7130-7368	EcoAnalysts
16		Sep	1	15:45	1.407	0.06921	0.1371			18-6405-8290	07-0280-8863	EcoAnalysts
17		Dec	31	15:47	2.466	1.128	1.662	(+)		16-6781-4604	13-3844-2137	EcoAnalysts
18	2018	Feb	23	13:35	1.361	0.0228	0.04594			00-2027-3508	17-9124-7622	EcoAnalysts
19		Mar	2	11:45	1.301	-0.03713	-0.07651			02-5827-5523	13-4470-6509	EcoAnalysts
20		Aug	24	15:55	1.708	0.3702	0.664			17-5739-4878	03-7944-2577	EcoAnalysts
21		Oct	9	15:00	0.9139	-0.4241	-1.036	(-)		17-3069-9320	16-8151-1630	EcoAnalysts



# CETIS Summary Report

Report Date: 22 Oct-18 16:14 (p 1 of 1)  
 Test Code/ID: 7A05E615 / 20-4720-6933

Reference Toxicant 96-h Acute Survival Test				EcoAnalysts
Batch ID: 16-5970-2885	Test Type: Survival	Analyst:		
Start Date: 09 Oct-18 15:00	Protocol: EPA/600/R-94/025 (1994)	Diluent: Laboratory Seawater		
Ending Date: 13 Oct-18 13:50	Species: Eohaustorius estuarius	Brine: Not Applicable		
Test Length: 95h	Taxon: Malacostraca	Source: Northwestern Aquatic Scienc/Ages		
Sample ID: 11-5195-8884	Code: 44A97F64	Project: Reference Toxicant		
Sample Date: 15 May-17	Material: Total Ammonia	Source: Reference Toxicant		
Receipt Date: 15 May-17	CAS (PC):	Station: p170515.110		
Sample Age: 512d 15h	Client: Internal Lab			

Multiple Comparison Summary						
Analysis ID	Endpoint	Comparison Method	✓ NOEL	LOEL	TOEL	TU PMSD S
14-7074-1873	Proportion Survived	Steel Many-One Rank Sum Test	✓ 134	✓ 276	192.3	24.6% 1

Point Estimate Summary						
Analysis ID	Endpoint	Point Estimate Method	✓ Level	mg/L	95% LCL	95% UCL TU S
11-3401-1593	Proportion Survived	Spearman-Kärber	EC50	✓ 189.1	166.3	215.1 1

Proportion Survived Summary											
Conc-mg/L	Code	Count	Mean	95% LCL	95% UCL	Min	Max	Std Err	Std Dev	CV%	%Effect
0	D	4	0.9000	0.5818	1.0000	0.6000	1.0000	0.1000	0.2000	22.22%	0.00%
17.1		4	1.0000	1.0000	1.0000	1.0000	1.0000	0.0000	0.0000	0.00%	-11.11%
34.8		4	0.8500	0.4712	1.0000	0.5000	1.0000	0.1190	0.2380	28.01%	5.56%
69		4	0.9500	0.7909	1.0000	0.8000	1.0000	0.0500	0.1000	10.53%	-5.56%
134		4	0.8500	0.7581	0.9419	0.8000	0.9000	0.0289	0.0577	6.79%	5.56%
276		4	0.1750	0.0227	0.3273	0.1000	0.3000	0.0479	0.0957	54.71%	80.56%
541		4	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000		100.00%

Proportion Survived Detail					
Conc-mg/L	Code	Rep 1	Rep 2	Rep 3	Rep 4
0	D	1.0000	1.0000	0.6000	1.0000
17.1		1.0000	1.0000	1.0000	1.0000
34.8		1.0000	0.5000	1.0000	0.9000
69		0.8000	1.0000	1.0000	1.0000
134		0.8000	0.8000	0.9000	0.9000
276		0.3000	0.1000	0.1000	0.2000
541		0.0000	0.0000	0.0000	0.0000

Proportion Survived Binomials					
Conc-mg/L	Code	Rep 1	Rep 2	Rep 3	Rep 4
0	D	10/10	10/10	6/10	10/10
17.1		10/10	10/10	10/10	10/10
34.8		10/10	5/10	10/10	9/10
69		8/10	10/10	10/10	10/10
134		8/10	8/10	9/10	9/10
276		3/10	1/10	1/10	2/10
541		0/10	0/10	0/10	0/10

**CETIS Test Data Worksheet**

Report Date: 22 Oct-18 16:14 (p 1 of 2)  
 Test Code/ID: 7A05E615 / 20-4720-6933

Reference Toxicant 96-h Acute Survival Test							EcoAnalysts
Start Date:	09 Oct-18 15:00	Species:	Eohaustorius estuarius	Sample Code:	44A97F64		
End Date:	13 Oct-18 13:50	Protocol:	EPA/600/R-94/025 (1994)	Sample Source:	Reference Toxicant		
Sample Date:	15 May-17	Material:	Total Ammonia	Sample Station:	p170515.110		
Conc-mg/L	Code	Rep	Pos	# Exposed	# Survived	Notes	
0	D	1	1	10	10		
0	D	2	12	10	10		
0	D	3	21	10	6		
0	D	4	27	10	10		
17.1		1	14	10	10		
17.1		2	20	10	10		
17.1		3	11	10	10		
17.1		4	25	10	10		
34.8		1	15	10	10		
34.8		2	3	10	5		
34.8		3	17	10	10		
34.8		4	10	10	9		
69		1	4	10	8		
69		2	18	10	10		
69		3	8	10	10		
69		4	26	10	10		
134		1	7	10	8		
134		2	9	10	8		
134		3	24	10	9		
134		4	28	10	9		
276		1	19	10	3		
276		2	22	10	1		
276		3	2	10	1		
276		4	23	10	2		



# CETIS Test Data Worksheet

Report Date: 22 Oct-18 16:14 (p 2 of 2)  
Test Code/ID: 7A05E615 / 20-4720-6933

Conc-mg/L	Code	Rep	Pos	# Exposed	# Survived	Notes
✓ 541		1	6	10	0 ✓	
541		2	13	10	0 ✓	
541		3	16	10	0 ✓	
541		4	5	10	0 ✓	

# CETIS Summary Report

Report Date: 22 Oct-18 16:20 (p 1 of 1)  
 Test Code/ID: 67286038 / 17-3069-9320

Reference Toxicant 96-h Acute Survival Test				EcoAnalysts
Batch ID: 02-7500-5202	Test Type: Survival	Analyst:		
Start Date: 09 Oct-18 15:00	Protocol: EPA/600/R-94/025 (1994)	Diluent: Laboratory Seawater		
Ending Date: 13 Oct-18 13:50	Species: Eohaustorius estuarius	Brine: Not Applicable		
Test Length: 95h	Taxon: Malacostraca	Source: Northwestern Aquatic Sci	Age: <i>stat</i>	
Sample ID: 10-5025-6844	Code: 3E99A5CC	Project: Reference Toxicant		
Sample Date: 15 May-17	Material: Unionized Ammonia	Source: Reference Toxicant		
Receipt Date: 15 May-17	CAS (PC):	Station: p170515.110		
Sample Age: 512d 15h	Client: Internal Lab			

Multiple Comparison Summary									
Analysis ID	Endpoint	Comparison Method	✓ NOEL	LOEL	TOEL	TU	PMSD	S	
05-9114-6332	Proportion Survived	Steel Many-One Rank Sum Test	0.74	1.221	0.9505		24.6%	1	

Point Estimate Summary									
Analysis ID	Endpoint	Point Estimate Method	✓ Level	mg/L	95% LCL	95% UCL	TU	S	
16-8151-1630	Proportion Survived	Spearman-Kärber	EC50	0.9139	0.8403	0.994		1	

Proportion Survived Summary											
Conc-mg/L	Code	Count	Mean	95% LCL	95% UCL	Min	Max	Std Err	Std Dev	CV%	%Effect
0	D	4	0.9000	0.5818	1.0000	0.6000	1.0000	0.1000	0.2000	22.22%	0.00%
0.141		4	1.0000	1.0000	1.0000	1.0000	1.0000	0.0000	0.0000	0.00%	-11.11%
0.288		4	0.8500	0.4712	1.0000	0.5000	1.0000	0.1190	0.2380	28.01%	5.56%
0.475		4	0.9500	0.7909	1.0000	0.8000	1.0000	0.0500	0.1000	10.53%	-5.56%
0.74		4	0.8500	0.7581	0.9419	0.8000	0.9000	0.0289	0.0577	6.79%	5.56%
1.221		4	0.1750	0.0227	0.3273	0.1000	0.3000	0.0479	0.0957	54.71%	80.56%
1.535		4	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000		100.00%

Proportion Survived Detail						
Conc-mg/L	Code	Rep 1	Rep 2	Rep 3	Rep 4	
0	D	1.0000	1.0000	0.6000	1.0000	
0.141		1.0000	1.0000	1.0000	1.0000	
0.288		1.0000	0.5000	1.0000	0.9000	
0.475		0.8000	1.0000	1.0000	1.0000	
0.74		0.8000	0.8000	0.9000	0.9000	
1.221		0.3000	0.1000	0.1000	0.2000	
1.535		0.0000	0.0000	0.0000	0.0000	

Proportion Survived Binomials						
Conc-mg/L	Code	Rep 1	Rep 2	Rep 3	Rep 4	
0	D	10/10	10/10	6/10	10/10	
0.141		10/10	10/10	10/10	10/10	
0.288		10/10	5/10	10/10	9/10	
0.475		8/10	10/10	10/10	10/10	
0.74		8/10	8/10	9/10	9/10	
1.221		3/10	1/10	1/10	2/10	
1.535		0/10	0/10	0/10	0/10	



**CETIS Test Data Worksheet**

Report Date: 22 Oct-18 16:20 (p 1 of 2)  
 Test Code/ID: 67286038 / 17-3069-9320

Reference Toxicant 96-h Acute Survival Test							EcoAnalysts
Start Date:	09 Oct-18 15:00	Species:	Eohaustorius estuarius	Sample Code:	3E99A5CC		
End Date:	13 Oct-18 13:50	Protocol:	EPA/600/R-94/025 (1994)	Sample Source:	Reference Toxicant		
Sample Date:	15 May-17	Material:	Unionized Ammonia	Sample Station:	p170515.110		
Conc-mg/L	Code	Rep	Pos	# Exposed	# Survived		Notes
0	D	1	7	10	10	/	
0	D	2	4	10	10	/	
0	D	3	25	10	6	/	
0	D	4	20	10	10	/	
0.141		1	2	10	10	/	
0.141		2	23	10	10	/	
0.141		3	14	10	10	/	
0.141		4	19	10	10	/	
0.288		1	16	10	10	/	
0.288		2	28	10	5	/	
0.288		3	17	10	10	/	
0.288		4	9	10	9	/	
0.475		1	22	10	8	/	
0.475		2	10	10	10	/	
0.475		3	5	10	10	/	
0.475		4	18	10	10	/	
0.74		1	24	10	8	/	
0.74		2	6	10	8	/	
0.74		3	8	10	9	/	
0.74		4	26	10	9	/	
1.221		1	1	10	3	/	
1.221		2	27	10	1	/	
1.221		3	13	10	1	/	
1.221		4	11	10	2	/	

# CETIS Test Data Worksheet

Report Date: 22 Oct-18 16:20 (p 2 of 2)  
Test Code/ID: 67286038 / 17-3069-9320

Conc-mg/L	Code	Rep	Pos	# Exposed	# Survived	Notes
1.535		1	21	10	0 ✓	
1.535		2	3	10	0 ✓	
1.535		3	12	10	0 ✓	
1.535		4	15	10	0 ✓	

**Ammonia Reference Toxicant  
Spiking Worksheet**

Reference Toxicant ID: P170515.110  
 Date Prepared: 10/9/18  
 Technician Initials: CR

# Amp/Eoh NH<sub>3</sub> RT

Assumptions in Model  
 Stock ammonia concentration is 10,000 mg/L = 10 mg/mL

Date: 10/2/2018  
 Measurement: 9,833

Test Solutions			Volume of stock to reach desired concentration	
Measured Concentration	Desired Concentration	Volume		
mg/L	mg/L	mL	mL stock to increase	
				SALT WATER
541	500	3200		244.076
276	250	3200		122.038
134	125	3200		61.019
69.0	62.5	3200		30.510
34.8	31.2	3200		15.230
17.1	15.6	3200		7.615
0.00	0			

# Ammonia Reference Toxicant Test Water Quality Data Sheet

CLIENT [REDACTED]	PROJECT [REDACTED]	<i>Eohaustorius estuarius</i>			LABORATORY Port Gamble	PROTOCOL USEPA 1994 / ASTM 2006
TEST ID <b>P170515.110</b>	LOT #: <b>2986C510</b>	DILUTION PREP INITIALS: <b>CR</b>				
CHAMBER SIZE/TYPE 1L Plastic	EXPOSURE VOLUME 800mL	TEST START DATE 09Oct18	INITIALS MK, CR	TIME 1500	TEST END DATE 13Oct18	TIME 1350

## WATER QUALITY DATA

TEST CONDITIONS				DO (mg/L)		TEMP(C)		SAL (ppt)		pH		TECHNICIAN	AMMONIA			
SAMPLE ID	CONCENTRATION		DAY	REP	D.O.		TEMP.		SALINITY		pH		AMMONIA		Tech	
	value	units			meter	mg/L	meter	°C	meter	ppt	meter	unit	WQ TECH/ DATE	METER		mg/L
	Ref.Tox.-ammonia	0			mg/L	0	Stock	8	7.0	8	13.7	8	31	8		7.6
			2	1	8	6.9	8	16.6	8	31	8	7.8	MAR 10/11			
			4	2	8	7.5	8	15.1	8	31	8	7.8	MK 10/13	10	0.414	JL
Ref.Tox.-ammonia	15.6	mg/L	0	Stock	8	6.9	8	13.9	8	31	8	7.6	JL 10/09	10	17.1	SW
			2	1	8	7.2	8	16.6	8	31	8	7.7	MAR 10/11			
			4	2	8	7.4	8	15.0	8	31	8	7.8	MK 10/13	10	17.8	JL
Ref.Tox.-ammonia	31.2	mg/L	0	Stock	8	7.0	8	14.0	8	31	8	7.6	JL 10/09	10	34.8	SW
			2	1	8	7.2	8	16.4	8	31	8	7.7	MAR 10/11			
			4	2	8	7.2	8	15.3	8	31	8	7.8	MK 10/13	10	36.5	JL
Ref.Tox.-ammonia	62.5	mg/L	0	Stock	8	7.3	8	14.6	8	31	8	7.5	JL 10/09	10	69.0	SW
			2	1	8	7.1	8	16.7	8	31	8	7.7	MAR 10/11			
			4	2	8	7.3	8	15.1	8	31	8	7.7	MK 10/13	10	69.7	JL
Ref.Tox.-ammonia	125	mg/L	0	Stock	8	7.3	8	14.7	8	31	8	7.4	JL 10/09	10	124.134	SW
			2	1	8	7.5	8	16.8	8	31	8	7.7	MAR 10/11			
			4	2	8	7.6	8	15.3	8	31	8	7.7	MK 10/13	10	130	JL
Ref.Tox.-ammonia	250	mg/L	0	Stock	8	7.3	8	14.8	8	31	8	7.3	JL 10/09	10	235.276	SW
			2	1	8	7.1	8	16.6	8	31	8	7.5	MAR 10/11			
			4	2	8	7.4	8	15.1	8	31	8	7.6	MK 10/13	10	269	JL

## Ammonia Reference Toxicant Test Water Quality Data Sheet

CLIENT [REDACTED]	PROJECT [REDACTED]	<i>Eohaustorius estuarius</i>			LABORATORY Port Gamble	PROTOCOL USEPA 1994 / ASTM 2006
TEST ID P170515.110	LOT #: 29866510	DILUTION PREP INITIALS:				
CHAMBER SIZE/TYPE 1L Plastic	EXPOSURE VOLUME 800mL	TEST START DATE 09Oct18	INITIALS mk, CR	TIME 1500	TEST END DATE 13Oct18	TIME 1350

### WATER QUALITY DATA

TEST CONDITIONS				DO (mg/L)	TEMP(C)	SAL (ppt)	pH	TECHNICIAN	AMMONIA						
				> 8.0	15 ± 2	32 ± 2	7.7 - 8.3								
Ref.Tox.-ammonia	500 mg/L	0	Stock	8	7.2	8	15.0	8	31	8	7.1	10/09 JL	10	541	JW
		2	1	①	<del>_____</del>										
		4	2	②	<del>_____</del>										

① NO surviving animals. JL 10/22/18.



SPECIES	<i>Eohaustorius estuarius</i>
CLIENT	[REDACTED]
PROJECT	[REDACTED]
PROJECT MANAGER	B. Hester
LABORATORY	Port Gamble
PROTOCOL	USEPA 1994 / ASTM 2006

**SURVIVAL & BEHAVIOR DATA**

OBSERVATION KEY N = Normal LOE = Loss of equilibrium Q = Quiescent DC = Discoloration NB = No body F = Floating on surface				DAY 1			DAY 2			DAY 3			DAY 4		
				DATE	TECHNICIAN	INITIAL # OF ORGANISMS	DATE	TECHNICIAN	INITIAL # OF ORGANISMS	DATE	TECHNICIAN	INITIAL # OF ORGANISMS	DATE	TECHNICIAN	INITIAL # OF ORGANISMS
Ref.Tox. - Ammonia	0 mg/L	10	1	10	0	N	10	0	N	10	0	1F	10	0	1F
			2	10	0	N	10	0	N	10	0	3F	10	0	N
			3	10	0	N	10	0	N	10	0	1F	6	2	2NB
			4	10	0	N	10	0	N	10	0	3F	10	0	N
Ref.Tox. - Ammonia	15.6 mg/L	10	1	10	0	N	10	0	N	10	0	2F	10	0	3F
			2	10	0	2F	10	0	N	10	0	N	10	0	N
			3	10	0	1F	10	0	N	10	0	N	10	0	N
			4	10	0	N	10	0	N	10	0	N	10	0	N
Ref.Tox. - Ammonia	31.2 mg/L	10	1	10	0	1F	10	0	N	10	0	1F	10	0	N
			2	10	0	2F	10	0	2F	10	0	3F	5	1	4F, 4NB
			3	10	0	2F	10	0	2F	10	0	N	10	0	N
			4	10	0	3F	10	0	N	10	0	1F	9	0	1F, 1NB
Ref.Tox. - Ammonia	62.5 mg/L	10	1	10	0	N	10	0	1Q	10	0	1Q	8	1	1F, 1NB
			2	10	0	N	10	0	N	10	0	1Q	10	0	N
			3	10	0	2F	10	0	N	10	0	N	10	0	N
			4	10	0	N	10	0	1F	10	0	3F	10	0	1F
Ref.Tox. - Ammonia	125 mg/L	10	1	10	0	1F	10	0	1Q	10	0	2F	8	2	1F
			2	10	0	2F	9	1	2F	9	0	2Q	8	1	1F
			3	10	0	2F	10	0	N	10	0	1F	9	1	2F
			4	10	0	2F	9	1	N	9	0	1Q	9	0	1F
Ref.Tox. - Ammonia	250 mg/L	10	1	9	1	1F	9	0	Q	5	4	Q	3	2	Q
			2	7	3	Q	5	2	Q	4	1	Q	1	3	N
			3	9	1	Q	7	2	Q	4	3	Q	1	3	N
			4	8	2	Q	7	1	Q	3	4	Q	2	1	N

① WC. MB 10/13

① IE - RE 10/12

SPECIES

*Eohaustorius estuarius*

CLIENT [REDACTED]	PROJECT [REDACTED]	PROJECT MANAGER B. Hester	LABORATORY Port Gamble	PROTOCOL USEPA 1994 / ASTM 2006
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**SURVIVAL & BEHAVIOR DATA**

OBSERVATION KEY N = Normal LOE = Loss of equilibrium Q = Quiescent DC = Discoloration NB = No body F = Floating on surface  INITIAL # OF ORGANISMS 10				DAY 1			DAY 2			DAY 3			DAY 4				
				DATE	TECHNICIAN		DATE	TECHNICIAN		DATE	TECHNICIAN		DATE	TECHNICIAN			
SAMPLE ID	CONC.		REP	INITIAL NUMBER	#ALIVE	#DEAD	OBS	#ALIVE	#DEAD	OBS	#ALIVE	#DEAD	OBS	#ALIVE	#DEAD	OBS	
	value	units			#ALIVE	#DEAD	OBS	#ALIVE	#DEAD	OBS	#ALIVE	#DEAD	OBS	#ALIVE	#DEAD	OBS	
Ref.Tox. - Ammonia	500 mg/L		1	10	0	10	-										
			2	↓	1	9	Q	0	1	-							
			3	↓	3	17	Q	0	3	-							
			4	↓	1	9	Q	0	1	-							

## **2. *Neanthes arenaceodentata* 20-Day Test**

CLIENT		PROJECT		JOB NO.		PROJECT MANAGER		LABORATORY		PROTOCOL		SPECIES																		
Anchor QEA		Port Gamble Monitoring		PG1123		B. Hester		Port Gamble		PSEP 1995		Neanthes arenaceodentata																		
ENDPOINT DATA & OBSERVATIONS														NUMBER REMAINING	TARE WEIGHT (mg)	TOTAL WEIGHT (mg)	ASHED WEIGHT (mg)													
CLIENT/ENVIRON	REP	JAR	INITIAL # (if diff)	Date and Initials	1	2	3	4	5	6	7	8	9					10	11	12	13	14	15	16	17	18	19	20		
Control /	1				N	N	N	N	N	N	N	G	G	G	U	G	G	G	IE	G	G	G	N	N	5	1	209.99	291.59	242.15	
	2				N	N	N	N	N	N	N	G	G	G	U	G	IE	G	G	U	N	N	N	N	5	2	200.35	295.52	243.07	
	3				N	N	N	N	N	N	N	U	N	IE	U	N	N	N	IE	U	IE	N	N	N	5	3	181.29	259.22	213.13	
	4				N	N	N	N	N	N	N	N	N	N	N	N	G	G	G	G	U	N	N	IE	U	5	4	214.16	281.74	240.40
	5				N	N	N	N	N	N	N	N	G	IE	U	U	IE	G	G	G	U	G	G	N	5	5	208.23	311.51	254.32	
CARR-20 /	1				N	N	N	G	G	G	G	G	G	G	G	G	G	G	IE	G	G	G	G	G	5	6	195.34	308.85	243.96	
	2				N	N	N	G	G	U	U	G	U	U	U	U	U	U	U	U	U	U	U	U	5	7	224.77	276.84	241.38	
	3		13		10W	N	N	N	N	U	N	G	U	U	U	U	U	U	G	G	U	U	U	U	5	8	231.35	345.08	286.08	
	4				N	N	N	N	N	N	N	G	U	U	U	U	U	U	G	U	U	U	U	U	5	9	216.30	290.85	244.80	
	5				N	N	N	N	N	N	N	G	U	U	U	U	U	U	G	U	U	U	U	U	5	10	233.01	320.63	269.58	
Initial Biomass	Rep	Number	Tare Weight (mg)	Dry Weight (mg)	Ashed Weight (mg)	Comments:																								
	1	5	46.72	50.94	47.12	Boat #																58	222.47							
	2	5	46.73	50.76	47.20	101																60	220.28							
3	5	47.38	52.02	48.00	102																65	312.02								
						103																								

0 WC 10/24/18 SW

CLIENT		PROJECT		JOB NO.		PROJECT MANAGER		LABORATORY		PROTOCOL		SPECIES													
Anchor QEA		Port Gamble Monitoring		PG1123		B. Hester		Port Gamble		PSEP 1995		Neanthes arenaceodentata													
ENDPOINT DATA & OBSERVATIONS																									
CLIENT EXPOSURE ID	REP	JAR	INITIAL # (if differs)	Date and Initials																	NUMBER REMAINING	TARE WEIGHT (mg)	TOTAL WEIGHT (mg)	ASHED WEIGHT (mg)	
				10/4 SW	10/5 SW	10/6 CR	10/7 CR	10/8 US	10/9 MK	10/10 US	10/11 SW	10/12 RE	10/13 MK	10/14 MK	10/15 SW	10/16 SW	10/17 SW	10/18 SW	10/19 MK	10/20 SW					10/21 SW
CR-21 /	1			N	N	N	N	N	N	G	G	G	G	U	G	G	G	G	G	G	5	11	195.51	277.26	227.33
	2			N	N	N	N	N	N	G	G	G	G	U	G	G	G	G	G	G	5	12	193.88	247.77	210.52
	3			N	N	N	N	N	N	G	G	G	G	U	G	G	G	G	G	G	5	13	212.04	296.53	239.77
	4			N	N	N	N	N	N	G	G	G	G	U	G	G	G	G	G	G	3	14	212.26	268.01	229.77
	5	7		10W	N	N	N	N	N	N	G	G	G	G	U	G	G	G	G	G	G	5	15	239.45	315.40
SMA-1B-IT2 /	1			N	N	N	N	N	N	U	G	G	G	G	G	G	G	G	G	G	5	16	195.97	289.07	231.81
	2			N	N	N	N	N	N	U	G	G	G	G	G	G	G	G	G	G	5	17	204.38	288.89	231.59
	3			N	N	N	N	N	N	U	G	G	G	G	G	G	G	G	G	G	5	18	197.34	308.25	245.46
	4			N	N	N	N	N	N	U	G	G	G	G	G	G	G	G	G	G	5	19	198.68	268.33	221.63
	5			N	N	N	N	N	N	U	G	G	G	G	G	G	G	G	G	G	4	20	199.45	274.12	227.39
SMA-1B-IT3 /	1			N	N	N	N	N	N	N	N	N	N	G	G	G	G	G	G	G	5	21	197.86	294.76	234.07
	2			N	N	N	N	N	N	N	N	N	N	G	G	G	G	G	G	G	5	22	186.58	260.36	208.57
	3			N	N	N	N	N	N	N	N	N	N	G	G	G	G	G	G	G	5	23	185.54	263.98	210.57
	4			N	N	N	N	N	N	N	N	N	N	G	G	G	G	G	G	G	5	24	181.27	268.21	209.34
	5			N	N	N	N	N	N	N	N	N	N	G	G	G	G	G	G	G	5	25	176.05	235.71	191.84

CLIENT		PROJECT		JOB NO.		PROJECT MANAGER		LABORATORY		PROTOCOL		SPECIES															
Anchor QEA		Port Gamble Monitoring		PG1123		B. Hester		Port Gamble		PSEP 1995		Neanthes arenaceodentata															
ENDPOINT DATA & OBSERVATIONS																											
CLIENT-ENVIRON ID	REP	JAR	INITIAL # (if differs)	Date and Initials																				NUMBER REMAINING	TARE WEIGHT (mg)	TOTAL WEIGHT (mg)	ASHED WEIGHT (mg)
				10/4 SW	10/5 SW	10/6 CR	10/7 CR	10/8 US	10/9 MK	10/10 US	10/11 SW	10/12 RE	10/13 MK	10/14 MK	10/15 SW	10/16 SW	10/17 SW	10/18 SW	10/19 MK	10/20 SW	10/21 SW	10/22 SW	10/23 CR				
SMA-2C-IT3 /	1			N	N	N	N	N	N	N	G	G	G	G	G	G	G	G	G	G	G	5	<sup>26</sup> 197.13	280.26	226.55		
	2			N	N	N	N	N	N	N	G	G	G	G	G	G	G	G	G	G	G	5	<sup>27</sup> 189.34	264.02	212.53		
	3			N	N	N	N	N	N	N	G	G	G	G	G	G	G	G	G	G	G	5	<sup>28</sup> 203.36	280.55	220.22		
	4			N	N	N	N	N	N	N	G	G	G	G	G	G	G	G	G	G	G	5	<sup>29</sup> 209.06	289.45	234.33		
	5			N	N	N	N	N	N	N	G	G	G	G	G	G	G	G	G	G	G	5	<sup>30</sup> 193.34	272.20	216.92		
SMA-2C-IT6 /	1			N	N	N	N	N	N	N	G	G	G	G	G	G	G	G	G	G	G	5	<sup>31</sup> 196.24	261.29	221.32		
	2			N	N	N	N	N	N	N	G	G	G	G	G	G	G	G	G	G	G	5	<sup>32</sup> 174.21	258.62	204.26		
	3			N	N	N	N	N	N	N	G	G	G	G	G	G	G	G	G	G	G	5	<sup>33</sup> 196.85	285.03	237.48		
	4			N	N	N	N	N	N	N	G	G	G	G	G	G	G	G	G	G	G	5	<sup>34</sup> 225.30	312.66	255.91		
	5			N	N	N	N	N	N	N	G	G	G	G	G	G	G	G	G	G	G	4	<sup>35</sup> 218.91	288.89	246.38		
SMA-2C-IT9 /	1			N	N	N	N	N	N	N	G	G	G	G	G	G	G	G	G	G	G	5	<sup>36</sup> 194.27	255.92	214.58		
	2			N	N	N	N	N	N	N	G	G	G	G	G	G	G	G	G	G	G	5	<sup>37</sup> 255.88	344.62	292.65		
	3			N	N	N	N	N	N	N	G	G	G	G	G	G	G	G	G	G	G	5	<sup>38</sup> 236.02	308.13	261.47		
	4			N	N	N	N	N	N	N	G	G	G	G	G	G	G	G	G	G	G	5	<sup>39</sup> 294.25	369.39	320.73		
	5			N	N	N	N	N	N	N	G	G	G	G	G	G	G	G	G	G	G	5	<sup>40</sup> 182.64	253.91	208.96		

CLIENT Anchor QEA	PROJECT Port Gamble Monitoring	START TIME/ END TIME 1100 / 0960	DILUTION WATER BATCH FSW100218.01	PROTOCOL PSEP 1995	TEST START DATE 3-Oct-2018
JOB NUMBER PG1123	PROJECT MANAGER B. Hester	LABORATORY Port Gamble	TEMP. RECDR./HOBO#	TEST SPECIES <i>Neanthes arenaceodentata</i>	TEST END DATE 23-Oct-2018

### WATER QUALITY DATA

Test Location:
Test Acceptability:
> 80% Survival in Control

Dilution Water Batch:
Supplier:
Organism Batch: Age:
Test Chamber Size:

TEST CONDITIONS				DO (mg/L) > 4.6 D.O.		TEMP (C) 20 ± 1 TEMP		SALINITY (ppt) 28 ± 2 SALINITY		pH 8.0 ± 1.0 pH		WATER RENEWAL	Feeding	TECH/DATE
CLIENT/ENVIRON ID	DAY	REP	JAR	meter	mg/L	meter	°C	meter	ppt	meter	unit			
Control /	0	Surr	8	8	7.5	8	20.1	8	29	8	8.0		SW	MK 10/3
Control /	1	Surr	8	8	7.4	8	20.2	8	29	8	7.7			SW 10/4
Control /	2	Surr	8	8	7.3	8	20.2	8	29	8	7.8		JL	SW 10/5
Control /	3	Surr	8	8	7.4	8	20.4	8	29	8	7.8	CR		CR 10/6
Control /	4	Surr	8	8	6.6	8	19.9	8	28	8	7.8		CR	CR 10/7
Control /	5	Surr	8	8	7.0	8	19.9	8	27	8	7.8			WS 10/8
Control /	6	Surr	8	8	7.4	8	19.9	8	28	8	7.9	JL	JL	MK 10/9
Control /	7	Surr	8	8	7.1	8	19.6	8	28	8	7.7			WS 10/10
Control /	8	Surr	8	8	7.6	8	19.6	8	28	8	7.9		JL	SW 10/11
Control /	9	Surr	8	8	7.3	8	19.6	8	28	8	7.7	SW		PE 10/12
Control /	10	Surr	8	8	7.7	8	19.5	8	28	8	7.9		MK	MK 10/13
Control /	11	Surr	8	8	7.5	8	19.6	8	28	8	7.9			MK 10/14
Control /	12	Surr	8	8	7.4	8	19.6	8	28	8	7.9	SW	MK	SW 10/15
Control /	13	Surr	8	8	7.7	8	19.4	8	28	8	7.9			SW 10/16
Control /	14	Surr	8	8	7.8	8	19.4	8	28	8	7.8		SW	SW 10/17
Control /	15	Surr	8	8	7.3	8	19.4	8	28	8	7.8	SW		SW 10/18
Control /	16	Surr	8	8	7.4	8	19.5	8	28	8	7.9		SW	MK 10/19
Control /	17	Surr	8	8	7.2	8	19.5	8	28	8	7.8			SW 10/20
Control /	18	Surr	8	8	7.2	8	19.5	8	28	8	7.8	SW	SW	SW 10/21
Control /	19	Surr	8	8	7.3	8	19.4	8	28	8	7.8			SW 10/22
Control /	20	Surr	8	8	7.3	8	19.3	8	28	8	7.9			SW 10/23

CLIENT Anchor QEA	PROJECT Port Gamble Monitoring	START TIME/ END TIME —————	DILUTION WATER BATCH FSW100218.01	PROTOCOL PSEP 1995	TEST START DATE 3-Oct-2018
JOB NUMBER PG1123	PROJECT MANAGER B. Hester	LABORATORY Port Gamble	TEMP. RECDR./HOBO#	TEST SPECIES <i>Neanthes arenaceodentata</i>	TEST END DATE 23-Oct-2018

### WATER QUALITY DATA

Test Location: —————	Dilution Water Batch: —————
Test Acceptability: —————	Supplier: —————
90% Survival in Control	Organism Batch: Age —————
	Test Chamber Size: —————

CLIENT/ENVIRON ID	DAY	REP	JAR	DO (mg/L)		TEMP (C)		SALINITY (ppt)		pH		WATER RENEWAL	Feeding	TECH/DATE
				meter	mg/L	meter	TEMP °C	meter	ppt	meter	unit			
CARR-20 /	0	Surr	16	8	7.5	8	20.2	8	29	8	7.9		SW	MK 10/3
CARR-20 /	1	Surr	1	8	7.0	8	20.4	8	29	8	7.6			SW 10/4
CARR-20 /	2	Surr	1	8	7.2	8	20.3	8	29	8	7.7		JL	SW 10/5
CARR-20 /	3	Surr	1	8	6.6	8	20.6	8	29	8	7.7	CR		CR 10/6
CARR-20 /	4	Surr	1	8	7.4	8	19.8	8	28	8	7.8		CR	CR 10/7
CARR-20 /	5	Surr	1	8	6.9	8	20.1	8	28	8	7.7			UB 10/8
CARR-20 /	6	Surr	1	8	7.4	8	20.1	8	28	8	7.8	JL	JL	MK 10/9
CARR-20 /	7	Surr	1	8	7.2	8	19.7	8	28	8	7.6			UB 10/10
CARR-20 /	8	Surr	1	8	7.4	8	19.8	8	28	8	7.9		JL	SW 10/11
CARR-20 /	9	Surr	1	8	7.0	8	19.9	8	28	8	7.7	SW		RE 10/12
CARR-20 /	10	Surr	1	8	7.7	8	19.6	8	28	8	8.0			MK MK 10/13
CARR-20 /	11	Surr	16	8	7.5	8	19.9	8	28	8	7.9			MK 10/14
CARR-20 /	12	Surr	1	8	7.5	8	19.8	8	28	8	8.0	SW	MK	SW 10/15
CARR-20 /	13	Surr	1	8	7.3	8	19.6	8	28	8	7.9			SW 10/16
CARR-20 /	14	Surr	1	8	7.7	8	19.6	8	28	8	7.9		SW	SW 10/17
CARR-20 /	15	Surr	1	8	7.1	8	19.6	8	28	8	7.8	SW		SW 10/18
CARR-20 /	16	Surr	1	8	7.4	8	19.7	8	28	8	7.9		SW	MK 10/19
CARR-20 /	17	Surr	1	8	7.1	8	19.7	8	28	8	7.9			SW 10/20
CARR-20 /	18	Surr	1	8	6.9	8	19.9	8	28	8	7.8	SW	SW	SW 10/21
CARR-20 /	19	Surr	1	8	7.1	8	19.6	8	28	8	7.8			SW 10/22
CARR-20 /	20	Surr	1	8	7.0	8	19.6	8	28	8	7.9			SW 10/23



CLIENT Anchor QEA	PROJECT Port Gamble Monitoring	START TIME/ END TIME 	DILUTION WATER BATCH FSW100218.01	PROTOCOL PSEP 1995	TEST START DATE 3-Oct-2018
JOB NUMBER PG1123	PROJECT MANAGER B. Hester	LABORATORY Port Gamble	TEMP. RECDR./HOBO#	TEST SPECIES <i>Neanthes arenaceodentata</i>	TEST END DATE 23-Oct-2018

**WATER QUALITY DATA**

Test Location: 	Dilution Water Batch: 
Test Acceptability: ____ > 90% Survival in Control	Supplier: 
	Organism Batch: Age: 
	Test Chamber Size: 

TEST CONDITIONS				DO (mg/L) > 4.6	TEMP (C) 20 ± 1	SALINITY (ppt) 28 ± 2	pH 8.0 ± 1.0							
CLIENT/ENVIRON ID	DAY	REP	JAR	D.O.	TEMP	SALINITY	pH	WATER RENEWAL	Feeding	TECH/DATE				
				meter mg/L	meter °C	meter ppt	meter unit							
CR-21 /	0	Surr	4	8	7.3	8	20.1	8	29	8	7.9		SW	MK 10/3
CR-21 /	1	Surr	1	8	6.7	8	20.3	8	29	8	7.5			SW 10/4
CR-21 /	2	Surr	1	8	6.3	8	20.2	8	29	8	7.7		JL	SW 10/5
CR-21 /	3	Surr	1	8	6.3	8	20.5	8	29	8	7.7	CR		CR 10/6
CR-21 /	4	Surr	1	8	6.6	8	19.8	8	28	8	7.6		CR	CR 10/7
CR-21 /	5	Surr	1	8	5.5	8	20.1	8	28	8	7.5			UB 10/8
CR-21 /	6	Surr	1	8	6.1	8	19.9	8	28	8	7.7	JL	JL	MK 10/9
CR-21 /	7	Surr	1	8	5.6	8	19.8	8	28	8	7.4			UB 10/10
CR-21 /	8	Surr	1	8	6.9	8	19.8	8	28	8	7.8		JL	SW 10/11
CR-21 /	9	Surr	1	8	6.1	8	19.8	8	28	8	7.5	SW		RE 10/12
CR-21 /	10	Surr	1	8	7.0	8	19.6	8	28	8	7.8		MK	MK 10/13
CR-21 /	11	Surr	4	8	6.7	8	19.8	8	28	8	7.8			MK 10/14
CR-21 /	12	Surr	1	8	7.3	8	19.6	8	28	8	7.8	SW	MK	SW 10/15
CR-21 /	13	Surr	1	8	6.8	8	19.6	8	27	8	7.7			SW 10/16
CR-21 /	14	Surr	1	8	7.4	8	19.5	8	27	8	7.8		SW	SW 10/17
CR-21 /	15	Surr	1	8	6.1	8	19.5	8	27	8	7.7	SW		SW 10/18
CR-21 /	16	Surr	1	8	6.7	8	19.7	8	27	8	7.8		SW	MK 10/19
CR-21 /	17	Surr	1	8	6.8	8	19.5	8	28	8	7.7			SW 10/20
CR-21 /	18	Surr	1	8	6.6	8	19.6	8	27	8	7.8	SW	SW	SW 10/21
CR-21 /	19	Surr	1	8	6.6	8	19.5	8	28	8	7.7			SW 10/22
CR-21 /	20	Surr	1	8	6.5	8	19.4	8	28	8	7.9			SW 10/23





CLIENT Anchor QEA	PROJECT Port Gamble Monitoring	START TIME/ END TIME <del>                    </del>	DILUTION WATER BATCH FSW100218.01	PROTOCOL PSEP 1995	TEST START DATE 3-Oct-2018
JOB NUMBER PG1123	PROJECT MANAGER B. Hester	LABORATORY Port Gamble	TEMP. RECDR./HOBO#	TEST SPECIES <i>Neanthes arenaceodentata</i>	TEST END DATE 23-Oct-2018

### WATER QUALITY DATA

Test Location:	Dilution Water Batch:
Test Acceptability: - > 99% Survival in Control	Supplier:
	Organism Batch: Age:
	Test Chamber Size:

TEST CONDITIONS				DO (mg/L) > 4.6 D.O.		TEMP (C) 20 ± 1 TEMP		SALINITY (ppt) 28 ± 2 SALINITY		pH 8.0 ± 1.0 pH		WATER RENEWAL	Feeding	TECH/DATE
CLIENT/ENVIRON ID	DAY	REP	JAR	meter	meter	meter	meter	meter	meter	unit				
SMA-2C-IT3 /	0	Surr	44	8	7.6	8	19.5	8	28	8	8.0		SW	MK 10/3
SMA-2C-IT3 /	1	Surr	1	8	7.4	8	19.6	8	28	8	7.9			SW 10/4
SMA-2C-IT3 /	2	Surr		8	7.4	8	19.7	8	28	8	8.0		JL	SW 10/5
SMA-2C-IT3 /	3	Surr		8	7.4	8	20.1	8	28	8	8.0	CR		CR 10/6
SMA-2C-IT3 /	4	Surr		8	7.5	8	19.5	8	28	8	8.0		CR	CR 10/7
SMA-2C-IT3 /	5	Surr		8	7.2	8	19.8	8	27	8	7.9			UB 10/8
SMA-2C-IT3 /	6	Surr		8	7.6	8	19.6	8	27	8	8.0	JL	JL	MK 10/9
SMA-2C-IT3 /	7	Surr		8	7.4	8	19.5	8	28	8	7.8			UB 10/10
SMA-2C-IT3 /	8	Surr		8	7.4	8	19.6	8	28	8	8.0		JL	SW 10/11
SMA-2C-IT3 /	9	Surr		8	7.5	8	19.7	8	28	8	7.8	SW		RE 10/12
SMA-2C-IT3 /	10	Surr	2	8	7.7	8	19.5	8	28	8	8.0		MK	MK 10/13
SMA-2C-IT3 /	11	Surr	44	8	7.6	8	19.6	8	28	8	7.9			MK 10/14
SMA-2C-IT3 /	12	Surr	1	8	7.7	8	19.5	8	28	8	8.0	SW	MK	SW 10/15
SMA-2C-IT3 /	13	Surr		8	7.5	8	19.4	8	27	8	7.9			SW 10/16
SMA-2C-IT3 /	14	Surr		8	7.6	8	19.4	8	28	8	7.9		SW	SW 10/17
SMA-2C-IT3 /	15	Surr		8	7.5	8	19.3	8	28	8	7.9	SW		SW 10/18
SMA-2C-IT3 /	16	Surr		8	7.5	8	19.5	8	28	8	8.0		SW	MK 10/19
SMA-2C-IT3 /	17	Surr		8	7.4	8	19.5	8	28	8	7.9			SW 10/20
SMA-2C-IT3 /	18	Surr		8	7.4	8	19.6	8	28	8	7.8	SW	SW	SW 10/21
SMA-2C-IT3 /	19	Surr		8	7.4	8	19.5	8	28	8	7.8			SW 10/22
SMA-2C-IT3 /	20	Surr	1	8	7.3	8	19.4	8	28	8	7.9			SW 10/23

CLIENT Anchor QEA	PROJECT Port Gamble Monitoring	START TIME/ END TIME <del>                    </del>	DILUTION WATER BATCH FSW100218.01	PROTOCOL PSEP 1995	TEST START DATE 3-Oct-2018
JOB NUMBER PG1123	PROJECT MANAGER B. Hester	LABORATORY Port Gamble	TEMP. RECDR./HOB0#	TEST SPECIES <i>Neanthes arenaceodentata</i>	TEST END DATE 23-Oct-2018

### WATER QUALITY DATA

Test Location: <del>                    </del>	Dilution Water Batch: <del>                    </del>
Test Acceptability: 90% Survival in Control	Supplier: <del>                    </del>
	Organism Batch: Age: <del>                    </del>
	Test Chamber Size: <del>                    </del>

CLIENT/ENVIRON ID	DAY	REP	JAR	DO (mg/L)		TEMP (C)		SALINITY (ppt)		pH		WATER RENEWAL	Feeding	TECH/DATE
				> 4.6		20 ± 1		28 ± 2		8.0 ± 1.0				
				meter	mg/L	meter	TEMP °C	meter	SALINITY ppt	meter	unit			
SMA-2C-IT6 /	0	Surr	37	8	7.6	8	20.0	8	28	8	7.9		SW	MK 10/3
SMA-2C-IT6 /	1	Surr		8	6.6	8	20.3	8	28	8	7.6			SW 10/4
SMA-2C-IT6 /	2	Surr		8	7.0	8	20.2	8	28	8	7.7		JL	SW 10/5
SMA-2C-IT6 /	3	Surr		8	7.0	8	20.5	8	28	8	7.8	CR		CR 10/6
SMA-2C-IT6 /	4	Surr		8	7.4	8	19.8	8	28	8	7.8		CR	CR 10/7
SMA-2C-IT6 /	5	Surr		8	7.0	8	20.1	8	27	8	7.7			UB 10/8
SMA-2C-IT6 /	6	Surr		8	7.4	8	20.1	8	28	8	7.9	JL	JL	MK 10/9
SMA-2C-IT6 /	7	Surr		8	7.2	8	19.7	8	28	8	7.7			UB 10/10
SMA-2C-IT6 /	8	Surr		8	7.4	8	19.9	8	28	8	7.9		JL	SW 10/11
SMA-2C-IT6 /	9	Surr		8	7.2	8	20.0	8	28	8	7.7	SW		Re 10/12
SMA-2C-IT6 /	10	Surr		8	7.7	8	19.7	8	28	8	7.9			MK 10/13
SMA-2C-IT6 /	11	Surr	37	8	7.5	8	19.9	8	28	8	7.8			MK 10/14
SMA-2C-IT6 /	12	Surr		8	7.5	8	19.8	8	28	8	7.8	SW	MK	SW 10/15
SMA-2C-IT6 /	13	Surr		8	7.4	8	19.6	8	27	8	7.8			SW 10/16
SMA-2C-IT6 /	14	Surr		8	7.5	8	19.6	8	28	8	7.8		SW	SW 10/17
SMA-2C-IT6 /	15	Surr		8	7.2	8	19.5	8	28	8	7.7	SW		SW 10/18
SMA-2C-IT6 /	16	Surr		8	7.0	8	19.6	8	28	8	7.8		SW	MK 10/19
SMA-2C-IT6 /	17	Surr		8	7.1	8	19.7	8	27	8	7.8			SW 10/20
SMA-2C-IT6 /	18	Surr		8	7.2	8	19.7	8	28	8	7.7	SW	SW	SW 10/21
SMA-2C-IT6 /	19	Surr		8	7.2	8	19.7	8	28	8	7.7			SW 10/22
SMA-2C-IT6 /	20	Surr		8	7.3	8	19.6	8	28	8	7.8			SW 10/23



CLIENT Anchor QEA	PROJECT Port Gamble Monitoring	START TIME/ END TIME _____	DILUTION WATER BATCH FSW100218.01	PROTOCOL PSEP 1995	TEST START DATE 3-Oct-2018
JOB NUMBER PG1123	PROJECT MANAGER B. Hester	LABORATORY Port Gamble	TEMP. RECDR./HOBO#	TEST SPECIES <i>Neanthes arenaceodentata</i>	TEST END DATE 23-Oct-2018

**WATER QUALITY DATA**

Test Location:
Test Acceptability:
90% Survival in Control

Dilution Water Batch:
Supplier:
Organism Batch: Age:
Test Chamber Size:

TEST CONDITIONS				DO (mg/L)		TEMP (C)		SALINITY (ppt)		pH		WATER RENEWAL	Feeding	TECH/DATE
CLIENT/ENVIRON ID	DAY	REP	JAR	> 4.6		20 ± 1		28 ± 2		8.0 ± 1.0				
				meter	mg/L	meter	TEMP °C	meter	ppt	meter	unit			
SMA-2C-IT9 /	0	Surr	9	8	7.5	8	20.3	8	28	8	8.0		SW	MK 10/3
SMA-2C-IT9 /	1	Surr	1	8	7.1	8	20.5	8	28	8	7.7			SW 10/4
SMA-2C-IT9 /	2	Surr	1	8	7.4	8	20.3	8	28	8	7.8		JV	SW 10/5
SMA-2C-IT9 /	3	Surr	1	8	7.1	8	20.6	8	29	8	7.8	CR		CR 10/6
SMA-2C-IT9 /	4	Surr	1	8	7.1	8	19.9	8	28	8	7.8		CR	CR 10/7
SMA-2C-IT9 /	5	Surr	1	8	7.2	8	20.2	8	28	8	7.8			UB 10/8
SMA-2C-IT9 /	6	Surr	1	8	7.5	8	20.2	8	28	8	7.9	JV	JV	MK 10/9
SMA-2C-IT9 /	7	Surr	1	8	7.3	8	19.9	8	28	8	7.7			UB 10/10
SMA-2C-IT9 /	8	Surr	1	8	7.5	8	20.1	8	28	8	7.9		JV	SW 10/11
SMA-2C-IT9 /	9	Surr	1	8	7.5	8	20.0	8	28	8	7.7	SW		PC 10/12
SMA-2C-IT9 /	10	Surr	2	8	7.7	8	19.8	8	28	8	7.9			MK MK 10/13
SMA-2C-IT9 /	11	Surr	9	8	7.6	8	20.1	8	28	8	7.9			MK 10/14
SMA-2C-IT9 /	12	Surr	1	8	7.5	8	19.9	8	28	8	7.8	SW	MK	SW 10/15
SMA-2C-IT9 /	13	Surr	1	8	7.5	8	19.8	8	28	8	7.8			SW 10/16
SMA-2C-IT9 /	14	Surr	1	8	7.6	8	19.8	8	28	8	7.8		SW	SW 10/17
SMA-2C-IT9 /	15	Surr	1	8	7.3	8	19.7	8	28	8	7.8	SW		SW 10/18
SMA-2C-IT9 /	16	Surr	1	8	7.4	8	19.9	8	28	8	7.8		SW	MK 10/19
SMA-2C-IT9 /	17	Surr	1	8	7.2	8	19.9	8	28	8	7.8			SW 10/20
SMA-2C-IT9 /	18	Surr	1	8	7.2	8	19.9	8	28	8	7.7	SW	SW	SW 10/21
SMA-2C-IT9 /	19	Surr	1	8	7.2	8	19.8	8	28	8	7.7			SW 10/22
SMA-2C-IT9 /	20	Surr	2	8	7.4	8	19.7	8	28	8	7.8			SW 10/23

# Ammonia and Sulfide Analysis Record

<b>Client/Project:</b> ANCHOR / Port Embankment	<b>Organism:</b> Neanthea	<b>Test Duration (days):</b> 20
<b>PRETEST</b> / <b>INITIAL</b> / <b>FINAL</b> / <b>OTHER</b> (circle one)		<b>DAY of TEST:</b> <u>0</u>
<b>OVERLYING (OV)</b> / <b>POREWATER (PW)</b> (circle one) / <b>Comments:</b> _____		

Calibration Standards Temperature		Sample temperature should be within $\pm 1^\circ\text{C}$ of standards temperature at time and date of analysis.
<b>Date:</b> 10/03/18	<b>Temperature:</b> 22.0°C	

Sample ID or Description	Conc. or Rep	Date of Sampling and Initials	Ammonia Value (mg/L)		Meter #/ Temp (°C)		Date of Reading and Initials	Sample Preserved (Y/N)	pH	Sal (ppt)	Sample Volume (mL)	Measured Sulf. (mg/L)	Multiplier	Calculated Sulf. (mg/L)
OV $\otimes$	Surr.	10/03/18 JL	0.00		T1	21.0	10/03/18 JL	N			10	0.008		
CAP 20			0.410									0.014		
CR 21			0.304									0.017		
1B-IT2			0.585									0.007		
IT3			0.622									0.011		
2C IT3			0.282									0.009		
IT6			0.00									0.008		
IT9			0.00									0.004		
PW $\otimes$	Surr		0.00		T1	21.0	10/03 RE		7.2	29	5	ND	2	ND
CAP 20			3.21		T1	21.0			7.3	29	5	0.006	2	0.012
CR 21			①								2	ND	5	ND
1B-IT2			8.75		T1	21.0			7.4	26	2	0.048	5	② 0.090.240
IT3			①											
2C IT3			①		T1				7.5	24	1	0.011	10	0.110
IT6			0.096			21.0			7.2	26	1	0.013	10	0.130
IT9			0.296			21.0			7.3	27	1	0.031	10	0.310

① Insufficient volume RE 10/03. ② RE JL 10/03.

# Ammonia and Sulfide Analysis Record

<b>Client/Project:</b> Anchor / Port Embankment	<b>Organism:</b> Neantnes	<b>Test Duration (days):</b> 20
<b>PRETEST / INITIAL / FINAL / OTHER (circle one)</b>		<b>DAY of TEST:</b> 20
<b>OVERLYING (OV) / POREWATER (PW) (circle one) / Comments:</b> _____		

Calibration Standards Temperature	
<b>Date:</b> 10/23/18	<b>Temperature:</b> 22.2°C
Sample temperature should be within ±1°C of standards temperature at time and date of analysis.	

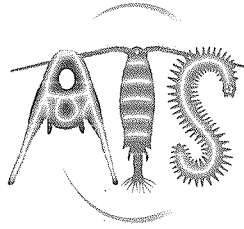
Sample ID or Description	Conc. or Rep	Date of Sampling and Initials	Ammonia Value (mg/L)	Meter #/ Temp (°C)		Date of Reading and Initials	Sample Preserved (Y/N)	pH	Sal (ppt)	Sample Volume (mL)	Measured Sulf. (mg/L)	Multiplier	Calculated Sulf. (mg/L)
OV ⊕	Surv.	10/23/18 JL	1.17	T1	21.4	10/23 JL	N			10			0.001
CRP 20			1.36										0.000
CR 21			7.50										0.003
IB 1T2			0.420										0.000
IB 1T3			0.326										0.000
2C 1T3			1.84										0.001
2C 1T6			2.77										0.003
2C 1T9			1.44										0.001
PW ⊕			1.79	T1	21.2			7.5	27	2	0.004	5	0.020
CRP 20			⓪					7.0	27	5	ND	2	ND
CR 21			⓪					7.9	27	1	0.007	10	0.070
IB 1T2			⓪										
IB 1T3			⓪										
2C 1T3			⓪										
2C 1T6			⓪					7.6	27	2	⓪ ND	5	ND
2C 1T9			1.81					7.5	27	2	0.00	5	0.00

⓪ insufficient volume for analysis. JL 10/23      ⓪ 1E JL 10/23 JL



## ORGANISM RECEIPT LOG

<b>Date:</b> 9-28-18	<b>Time:</b> 1020	<b>Batch No.</b> ATSO92818					
<b>Organism:</b> Neanthes arenaceodentata							
<b>Source / Supplier:</b> Aquatic Toxicology Support							
<b>No. Ordered:</b> 840	<b>No. Received:</b> 924	<b>Source Batch:</b> Collection date, hatch date, etc.): Emergence Date 09-5-10-2018					
<b>Condition of Organisms:</b> Good		<b>Approximate Size or Age:</b> (Days from hatch, life stage, size class, etc.): 23-28 days old					
<b>Shipper:</b> Courier		<b>B of L (Tracking No.):</b> NA					
<b>Condition of Container:</b> Good		<b>Received By:</b> BH, SW					
Container	D.O. (mg/L)	Temp. (°C)	Cond. or Sal. (Include Units) <sup>ph</sup>	pH (Units)	# Dead	% Dead*	Tech. (Initials)
Comp	14.0	19.1	31	7.3	—	—	SW, BH
*if >10% contact lab manager							
<b>Notes:</b>							



Aquatic Toxicology Support  
1849 Charleston Beach Road West  
Bremerton, Washington 98312  
(360) 813-1202

Order Summary

Species: <i>Neanthes arenaceodentata</i> *	Emerge Date: Sept 5-10 '18
Number Ordered: 840	Number Shipped: 840 + 104.
Date Shipped: Sept 28 '18	Salinity (ppt): 31

\*Smith 1964. CSU Long Beach strain. Feed upon arrival.

## MAINTENANCE LOG FOR CULTURES

ORGANISM: Nearnes

LOCATION: Bath 2

Batch Number: <u>ATS092018</u>	Date Received: <u>9/28/18</u>	Initial # of Organisms:
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Date	Feed AM/PM	Tub No.	D.O.	Temp (°C)	Cond/ (Sal) ppt	pH	H <sub>2</sub> O Change	Organisms appear healthy (Y/N)	# Mort	% Mort*	Init.	Comments
2019 9/29	—	10-d 1	6.9	19.7	32	7.8	N	Y	—	—	UL	
↓	—	2	6.9	19.9	32	7.8	↓	↓	—	—	↓	
↓	—	20-d	6.2	19.9	32	7.5	↓	↓	—	—	↓	
9/30	✓	10-d 1	6.9	19.9	32	7.8	Y	Y	—	—	u	
↓	✓	2	6.9	20.0	32	7.8	↓	↓	—	—	↓	
↓	✓	20-d	6.5	19.8	31	7.6	↓	↓	—	—	↓	
10/01	—	10-d 1	7.0	20.0	32	7.9	N	Y	—	—	UL	
↓	—	2	6.8	20.1	32	7.8	↓	↓	—	—	↓	
↓	—	20-d	6.3	19.7	32	7.7	↓	↓	—	—	↓	
10/2	—	10-d 1	7.0	19.5	32	7.9	N	Y	—	—	MK	
↓	—	2	7.1	19.5	32	7.8	↓	↓	—	—	↓	
↓	—	20-d 1	6.7	19.4	32	7.7	↓	↓	—	—	↓	
10.3	—	20-d 1	7.1	19.1	32	7.8	N	Y	—	—	BH	

FT = Flow-through  
 \*if >10% notify lab manager



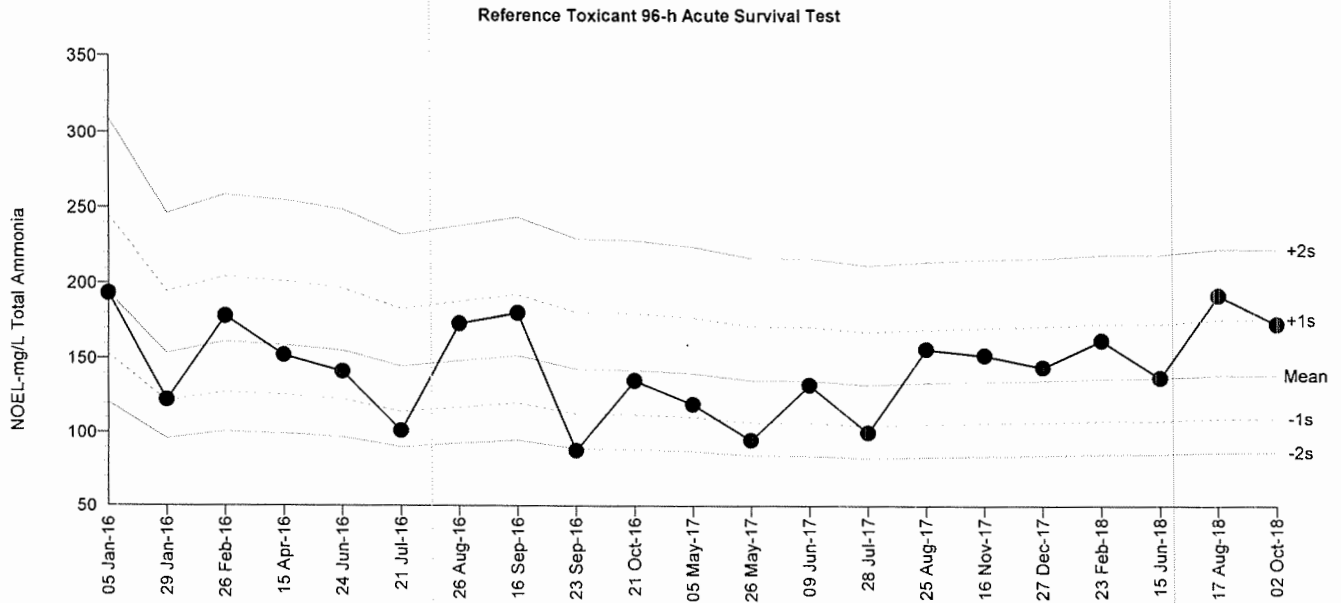
Reference Toxicant 96-h Acute Survival Test

All Matching Labs

Test Type: Survival  
Protocol: PSEP (1995)

Organism: Neanthes arenaceodentata (Polycha  
Endpoint: Proportion Survived

Material: Total Ammonia  
Source: Reference Toxicant-REF



Mean: 138.9      Count: 20      -1s Warning Limit: 109.8      -2s Action Limit: 86.69  
Sigma: n/a      CV: 23.90%      +1s Warning Limit: 176      +2s Action Limit: 222.8

Quality Control Data

Point	Year	Month	Day	Time	QC Data	Delta	Sigma	Warning	Action	Test ID	Analysis ID	Laboratory
1	2016	Jan	5	15:40	193	54.07	1.393	(+)		08-2089-5605	14-5672-8302	ENVIRON
2			29	10:55	122	-16.93	-0.5508			17-5198-4435	20-2746-8183	ENVIRON
3		Feb	26	13:05	178	39.07	1.05	(+)		12-4659-9912	04-9112-4627	ENVIRON
4		Apr	15	11:20	152	13.07	0.3808			14-5662-2397	08-8399-3634	ENVIRON
5		Jun	24	14:10	141	2.066	0.06254			18-4503-3329	05-9137-4303	ENVIRON
6		Jul	21	14:00	101	-37.93	-1.351	(-)		03-2252-3368	05-6001-6512	ENVIRON
7		Aug	26	17:00	173	34.07	0.9292			03-0001-3671	13-0809-8319	ENVIRON
8		Sep	16	13:45	180	41.07	1.097	(+)		15-1361-3636	20-6898-0370	ENVIRON
9			23	14:00	87.6	-51.33	-1.954	(-)		11-8849-2684	14-0354-7159	ENVIRON
10		Oct	21	12:20	135	-3.934	-0.1217			07-3517-7142	04-1673-2094	ENVIRON
11	2017	May	5	11:10	119	-19.93	-0.6563			19-9695-8635	08-7557-3106	EcoAnalysts
12			26	11:20	94.7	-44.23	-1.624	(-)		02-4398-8901	20-6641-5688	EcoAnalysts
13		Jun	9	13:12	132	-6.934	-0.2169			06-5936-3810	02-9842-7509	EcoAnalysts
14		Jul	28	10:45	99.9	-39.03	-1.398	(-)		04-6413-3650	18-1225-7941	EcoAnalysts
15		Aug	25	10:40	156	17.07	0.4909			03-6651-4426	09-2206-9228	EcoAnalysts
16		Nov	16	13:40	152	13.07	0.3808			03-0415-6136	18-4343-4696	EcoAnalysts
17		Dec	27	15:35	144	5.066	0.1518			01-8821-8905	01-3003-6293	EcoAnalysts
18	2018	Feb	23	16:20	162	23.07	0.6508			13-7905-8989	16-4206-5191	EcoAnalysts
19		Jun	15	14:35	137	-1.934	-0.0594			04-1274-1602	00-8204-0817	EcoAnalysts
20		Aug	17	14:15	192	53.07	1.371	(+)		07-8700-6256	00-3442-0298	EcoAnalysts
21		Oct	2	14:10	173	34.07	0.9292			21-2226-7904	05-0269-1791	EcoAnalysts



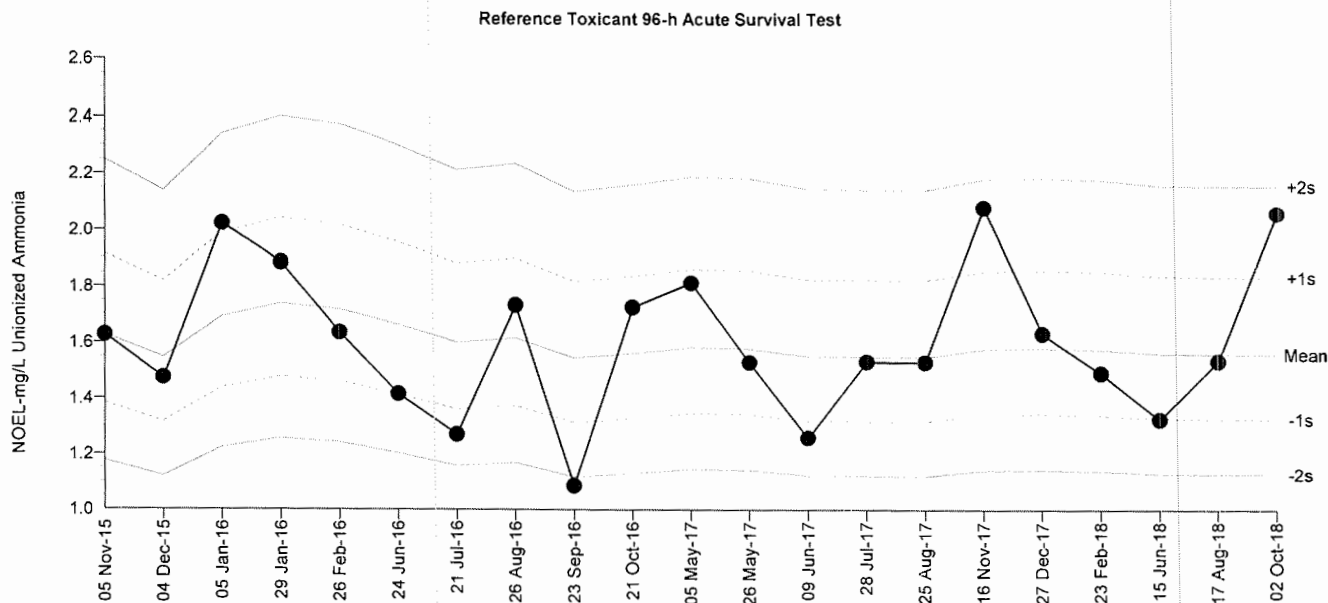
Reference Toxicant 96-h Acute Survival Test

All Matching Labs

Test Type: Survival  
Protocol: PSEP (1995)

Organism: Neanthes arenaceodentata (Polycha  
Endpoint: Proportion Survived

Material: Unionized Ammonia  
Source: Reference Toxicant-REF



Mean: 1.56      Count: 20      -1s Warning Limit: 1.327      -2s Action Limit: 1.129  
 Sigma: n/a      CV: 16.30%      +1s Warning Limit: 1.835      +2s Action Limit: 2.157

Quality Control Data

Point	Year	Month	Day	Time	QC Data	Delta	Sigma	Warning	Action	Test ID	Analysis ID	Laboratory
1	2015	Nov	5	16:00	1.627	0.06667	0.2584			13-9158-6969	18-5085-3785	ENVIRON
2		Dec	4	15:55	1.473	-0.08733	-0.3558			05-0232-3049	09-1115-6716	ENVIRON
3	2016	Jan	5	15:40	2.023	0.4627	1.604	(+)		16-5879-5239	13-0355-9173	ENVIRON
4			29	10:55	1.883	0.3227	1.161	(+)		02-3774-6836	16-2829-1192	ENVIRON
5		Feb	26	13:05	1.635	0.07467	0.2887			18-2733-1978	17-6331-1700	ENVIRON
6		Jun	24	14:10	1.415	-0.1453	-0.6039			14-5937-9292	10-8537-0051	ENVIRON
7		Jul	21	14:00	1.27	-0.2903	-1.272	(-)		13-0851-4355	06-2505-9350	ENVIRON
8		Aug	26	17:00	1.732	0.1717	0.6447			18-0730-6378	04-2606-0638	ENVIRON
9		Sep	23	14:00	1.085	-0.4753	-2.244	(-)	(-)	16-0277-5330	10-1484-1501	ENVIRON
10		Oct	21	12:20	1.725	0.1647	0.6197			15-4953-5653	10-3980-3312	ENVIRON
11	2017	May	5	11:10	1.812	0.2517	0.9236			06-1983-2716	14-3198-2813	EcoAnalysts
12			26	11:20	1.529	-0.03133	-0.1253			11-9977-1019	08-2373-9544	EcoAnalysts
13		Jun	9	13:12	1.258	-0.3023	-1.33	(-)		20-5746-1828	13-3286-2330	EcoAnalysts
14		Jul	28	10:45	1.532	-0.02833	-0.1132			11-9488-2902	00-8692-4177	EcoAnalysts
15		Aug	25	10:40	1.53	-0.03033	-0.1213			04-3451-1040	07-6966-4955	EcoAnalysts
16		Nov	16	13:40	2.079	0.5187	1.773	(+)		21-2485-6236	00-9775-4668	EcoAnalysts
17		Dec	27	15:35	1.633	0.07267	0.2812			14-3251-3795	04-4998-7500	EcoAnalysts
18	2018	Feb	23	16:20	1.491	-0.06933	-0.2807			08-4313-9079	12-5767-7210	EcoAnalysts
19		Jun	15	14:35	1.325	-0.2353	-1.01	(-)		13-9420-2469	12-4705-5925	EcoAnalysts
20		Aug	17	14:15	1.535	-0.02533	-0.1011			16-0114-0602	03-4717-2264	EcoAnalysts
21		Oct	2	14:10	2.059	0.4987	1.713	(+)		02-7017-5318	12-2051-3096	EcoAnalysts

**CETIS Summary Report**

Report Date: 22 Oct-18 16:37 (p 1 of 1)  
 Test Code/ID: 7E7F3D00 / 21-2226-7904

**Reference Toxicant 96-h Acute Survival Test**

EcoAnalysts

<b>Batch ID:</b> 20-2661-2066	<b>Test Type:</b> Survival	<b>Analyst:</b>
<b>Start Date:</b> 02 Oct-18 14:10 ✓	<b>Protocol:</b> PSEP (1995)	<b>Diluent:</b> Laboratory Seawater
<b>Ending Date:</b> 06 Oct-18 14:50 ✓	<b>Species:</b> Neanthes arenaceodentata ✓	<b>Brine:</b> Not Applicable
<b>Test Length:</b> 4d 1h ✓	<b>Taxon:</b> Polychaeta ✓	<b>Source:</b> Aquatic Toxicology Support Age:
<b>Sample ID:</b> 17-6336-1456	<b>Code:</b> 691AC2B0	<b>Project:</b> Reference Toxicant ✓
<b>Sample Date:</b> 15 May-17 ✓	<b>Material:</b> Total Ammonia	<b>Source:</b> Reference Toxicant ✓
<b>Receipt Date:</b> 15 May-17 ✓	<b>CAS (PC):</b>	<b>Station:</b> P170515.103 ✓
<b>Sample Age:</b> 505d 14h	<b>Client:</b> Internal Lab	

**Multiple Comparison Summary**

Analysis ID	Endpoint	Comparison Method	✓ NOEL	LOEL	TOEL	TU	PMSD	S
05-0269-1791	Proportion Survived	Fisher Exact Test	173	223	196.4		n/a	1

**Point Estimate Summary**

Analysis ID	Endpoint	Point Estimate Method	✓ Level	mg/L	95% LCL	95% UCL	TU	S
00-1709-8680	Proportion Survived	Trimmed Spearman-Kärber	EC50	197.3	195.5	199		1

**Proportion Survived Summary**

Conc-mg/L	Code	Count	Mean	95% LCL	95% UCL	Min	Max	Std Err	Std Dev	CV%	%Effect
0	D	3	1.0000	1.0000	1.0000	1.0000	1.0000	0.0000	0.0000	0.00%	0.00%
63.6		3	1.0000	1.0000	1.0000	1.0000	1.0000	0.0000	0.0000	0.00%	0.00%
116		3	1.0000	1.0000	1.0000	1.0000	1.0000	0.0000	0.0000	0.00%	0.00%
148		3	1.0000	1.0000	1.0000	1.0000	1.0000	0.0000	0.0000	0.00%	0.00%
173		3	1.0000	1.0000	1.0000	1.0000	1.0000	0.0000	0.0000	0.00%	0.00%
223		3	0.0333	0.0000	0.1768	0.0000	0.1000	0.0333	0.0577	173.21%	96.67%

**Proportion Survived Detail**

Conc-mg/L	Code	Rep 1	Rep 2	Rep 3
0	D	1.0000	1.0000	1.0000
63.6		1.0000	1.0000	1.0000
116		1.0000	1.0000	1.0000
148		1.0000	1.0000	1.0000
173		1.0000	1.0000	1.0000
223		0.0000	0.1000	0.0000

**Proportion Survived Binomials**

Conc-mg/L	Code	Rep 1	Rep 2	Rep 3
0	D	10/10	10/10	10/10
63.6		10/10	10/10	10/10
116		10/10	10/10	10/10
148		10/10	10/10	10/10
173		10/10	10/10	10/10
223		0/10	1/10	0/10



**CETIS Test Data Worksheet**

Report Date: 22 Oct-18 16:36 (p 1 of 1)  
 Test Code/ID: 7E7F3D00 / 21-2226-7904

<b>Reference Toxicant 96-h Acute Survival Test</b>				<b>EcoAnalysts</b>	
<b>Start Date:</b> 02 Oct-18 14:10	<b>Species:</b> Neanthes arenaceodentata	<b>Sample Code:</b> 691AC2B0			
<b>End Date:</b> 06 Oct-18 14:50	<b>Protocol:</b> PSEP (1995)	<b>Sample Source:</b> Reference Toxicant			
<b>Sample Date:</b> 15 May-17	<b>Material:</b> Total Ammonia	<b>Sample Station:</b> P170515.103			

Conc-mg/L	Code	Rep	Pos	# Exposed	# Survived	Notes
0	D	1	13	10	10	
0	D	2	10	10	10	
0	D	3	3	10	10	
63.6		1	8	10	10	
63.6		2	6	10	10	
63.6		3	7	10	10	
116		1	16	10	10	
116		2	15	10	10	
116		3	2	10	10	
148		1	14	10	10	
148		2	17	10	10	
148		3	9	10	10	
173		1	12	10	10	
173		2	11	10	10	
173		3	1	10	10	
223		1	18	10	0	
223		2	4	10	1	
223		3	5	10	0	

# CETIS Summary Report

Report Date: 26 Oct-18 09:27 (p 1 of 1)  
 Test Code/ID: 101A8C56 / 02-7017-5318

## Reference Toxicant 96-h Acute Survival Test

EcoAnalysts

Batch ID: 16-8756-8795	Test Type: Survival	Analyst:
Start Date: 02 Oct-18 14:10	Protocol: PSEP (1995)	Diluent: Laboratory Seawater
Ending Date: 06 Oct-18 14:50	Species: Neanthes arenaceodentata	Brine: Not Applicable
Test Length: 4d 1h	Taxon: Polychaeta	Source: Aquatic Toxicology Support Age:
Sample ID: 07-0788-5901	Code: 2A317B4D	Project: Reference Toxicant
Sample Date: 15 May-17	Material: Unionized Ammonia	Source: Reference Toxicant
Receipt Date: 15 May-17	CAS (PC):	Station: P170515.103
Sample Age: 505d 14h	Client: Internal Lab	

## Multiple Comparison Summary

Analysis ID	Endpoint	Comparison Method	✓ NOEL	LOEL	TOEL	TU	PMSD	S
12-2051-3096	Proportion Survived	Fisher Exact Test	2.059	2.114	2.086		n/a	1

## Point Estimate Summary

Analysis ID	Endpoint	Point Estimate Method	✓ Level	mg/L	95% LCL	95% UCL	TU	S
01-2654-4798	Proportion Survived	Trimmed Spearman-Kärber	EC50	2.087	2.085	2.089		1

## Proportion Survived Summary

Conc-mg/L	Code	Count	Mean	95% LCL	95% UCL	Min	Max	Std Err	Std Dev	CV%	%Effect
0	D	3	1.0000	1.0000	1.0000	1.0000	1.0000	0.0000	0.0000	0.00%	0.00%
1.192		3	1.0000	1.0000	1.0000	1.0000	1.0000	0.0000	0.0000	0.00%	0.00%
1.733		3	1.0000	1.0000	1.0000	1.0000	1.0000	0.0000	0.0000	0.00%	0.00%
1.974		3	1.0000	1.0000	1.0000	1.0000	1.0000	0.0000	0.0000	0.00%	0.00%
2.059		3	1.0000	1.0000	1.0000	1.0000	1.0000	0.0000	0.0000	0.00%	0.00%
2.114		3	0.0333	0.0000	0.1768	0.0000	0.1000	0.0333	0.0577	173.21%	96.67%

## Proportion Survived Detail

Conc-mg/L	Code	Rep 1	Rep 2	Rep 3
0	D	1.0000	1.0000	1.0000
1.192		1.0000	1.0000	1.0000
1.733		1.0000	1.0000	1.0000
1.974		1.0000	1.0000	1.0000
2.059		1.0000	1.0000	1.0000
2.114		0.0000	0.1000	0.0000

## Proportion Survived Binomials

Conc-mg/L	Code	Rep 1	Rep 2	Rep 3
0	D	10/10	10/10	10/10
1.192		10/10	10/10	10/10
1.733		10/10	10/10	10/10
1.974		10/10	10/10	10/10
2.059		10/10	10/10	10/10
2.114		0/10	1/10	0/10

**CETIS Test Data Worksheet**

Report Date: 26 Oct-18 09:26 (p 1 of 1)  
 Test Code/ID: 101A8C56 / 02-7017-5318

Reference Toxicant 96-h Acute Survival Test							EcoAnalysts
Start Date: 02 Oct-18 14:10		Species: Neanthes arenaceodentata		Sample Code: 2A317B4D			
End Date: 06 Oct-18 14:50		Protocol: PSEP (1995)		Sample Source: Reference Toxicant			
Sample Date: 15 May-17		Material: Unionized Ammonia		Sample Station: P170515.103			
Conc-mg/L	Code	Rep	Pos	# Exposed	# Survived	Notes	
0	D	1	8	10	10		
0	D	2	5	10	10		
0	D	3	1	10	10		
1.192		1	4	10	10		
1.192		2	2	10	10		
1.192		3	3	10	10		
1.733		1	11	10	10		
1.733		2	14	10	10		
1.733		3	17	10	10		
1.974		1	15	10	10		
1.974		2	12	10	10		
1.974		3	18	10	10		
2.059		1	10	10	10		
2.059		2	13	10	10		
2.059		3	16	10	10		
2.114		1	9	10	0		
2.114		2	7	10	1		
2.114		3	6	10	0		



# Ammonia Reference Toxicant Test Survival Data Sheet

CLIENT ADH			PROJECT Crescent City		JOB NO. PG1160		SPECIES <i>Neanthes arenaceodentata</i>		PROJECT MANAGER B. Hester		LABORATORY Port Gamble		PROTOCOL USACE 1991	
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## SURVIVAL & BEHAVIOR DATA

OBSERVATION KEY N = Normal LOE = Loss of equilibrium Q = Quinscent DC = Discoloration NB = No body F = Floating on surface				DAY 1			DAY 2			DAY 3			DAY 4						
				DATE 10/3/18			DATE 10/4/18			DATE 10/5/18			DATE 10/6/18						
				TECHNICIAN RE			TECHNICIAN MK			TECHNICIAN MK			TECHNICIAN CR						
CLIENT/ENVIRON ID	CONC.		REP	INITIAL NUMBER	INITIAL # OF ORGANISMS 10			#ALIVE: #DEAD: OBS			#ALIVE: #DEAD: OBS			#ALIVE: #DEAD: OBS			#ALIVE: #DEAD: OBS		
	value	units			#ALIVE	#DEAD	OBS	#ALIVE	#DEAD	OBS	#ALIVE	#DEAD	OBS	#ALIVE	#DEAD	OBS			
Ref.Tox.- Ammonia	0 mg/L		1	10	0	N	10	0	N	10	0	N	10	0	N				
			2	10	0	N	10	0	N	10	0	N	10	0	N				
			3	10	0	N	10	0	N	10	0	N	10	0	N				
Ref.Tox.- Ammonia	60 mg/L		1	10	0	N	10	0	N	10	0	N	10	0	N				
			2	10	0	N	10	0	N	10	0	N	10	0	N				
			3	10	0	N	10	0	N	10	0	N	10	0	N				
Ref.Tox.- Ammonia	100 mg/L		1	10	0	N	10	0	N	10	0	N	10	0	N				
			2	10	0	N	10	0	N	10	0	N	10	0	N				
			3	10	0	N	10	0	N	10	0	N	10	0	N				
Ref.Tox.- Ammonia	140 mg/L		1	10	0	N	10	0	N	10	0	N	10	0	N				
			2	10	0	N	10	0	N	10	0	N	10	0	N				
			3	10	0	N	10	0	N	10	0	N	10	0	N				
Ref.Tox.- Ammonia	180 mg/L		1	10	0	N	10	0	Q	10	0	Q	10	0	N				
			2	10	0	N	10	0	N	10	0	N	10	0	N				
			3	10	0	N	10	0	N	10	0	N	10	0	N				
Ref.Tox.- Ammonia	220 mg/L		1	10	0	N	10	0	Q	10	0	Q	0	10	N				
			2	10	0	N	10	0	N	10	0	N	1	9	N				
			3	10	0	N	10	0	N	10	0	N	0	10	N				

## Ammonia Reference Toxicant Test Survival Data Sheet

CLIENT ADH	PROJECT Crescent City	SPECIES <i>Neanthes arenaceodentata</i>	LABORATORY Port Gamble	PROTOCOL USACE 1991
TEST ID P170515.103	LOT #: 2986C510	TEST START DATE 02Oct18	TIME 1410 MK	4-DAY END DATE 06Oct18
CHAMBER SIZE/TYPE Glass pint jar	EXPOSURE VOLUME 250 mL			TIME 1450

### WATER QUALITY DATA

TEST CONDITIONS				DO (mg/L)		TEMP(C)		SAL (ppt)		pH		TECHNICIAN	AMMONIA		SULFIDES		
				> 4.6		20 + 1		30 + 2		7.8 + 0.5							
CLIENT/ENVIRON ID	CONCENTRATION		DAY	REP	D.O.		TEMP.		SALINITY		pH		WQ TECH/ DATE	AMMONIA		SULFIDES	
	value	units			meter	mg/L	meter	°C	meter	ppt	meter	unit		METER	mg/L	Tech	meter
Ref.Tox.-ammonia	0	mg/L	0	Stock	8	7.4	8	19.0	8	32	8	8.0	MK 10/2	10	0.00		
			4	1	8	7.3	8	20.0	8	32	8	8.0	CR 10/6				
Ref.Tox.-ammonia	60	mg/L	0	Stock	8	7.3	8	19.0	8	32	8	7.8	MK 10/2	10	63.6		
			4	1	8	7.1	8	20.1	8	33	8	7.9	CR 10/6				
Ref.Tox.-ammonia	100	mg/L	0	Stock	8	7.4	8	19.0	8	32	8	7.7	MK 10/2	10	116		
			4	1	8	7.0	8	20.1	8	33	8	7.8	CR 10/6				
Ref.Tox.-ammonia	140	mg/L	0	Stock	8	7.4	8	19.0	8	32	8	7.7	MK 10/2	10	148		
			4	1	8	7.0	8	20.1	8	33	8	7.8	CR 10/6				
Ref.Tox.-ammonia	180	mg/L	0	Stock	8	7.4	8	19.0	8	32	8	7.6	MK 10/2	10	173		
			4	1	8	7.1	8	20.1	8	33	8	7.8	CR 10/6				
Ref.Tox.-ammonia	220	mg/L	0	Stock	8	7.4	8	19.0	8	32	8	7.5	MK 10/2	10	223		
			4	1	8	6.7	8	20.1	8	33	8	7.8	CR 10/6				

**Ammonia Reference Toxicant  
Spiking Worksheet**

Reference Toxicant ID: 9170515.103  
 Date Prepared: 10/2/18  
 Technician Initials: mk

# Neanthes NH<sub>3</sub> RT

Assumptions in Model  
 Stock ammonia concentration is 10,000 mg/L = 10 mg/mL

Date: 10/2/2018  
 Measurement: 8/30/2018  
 9833

Test Solutions			Volume of stock to reach desired concentration	
Measured Concentration	Desired Concentration	Volume	mL stock to increase	
mg/L	mg/L	mL	FRESH WATER (mL)	SALT WATER (mL)
0				
63.6	60	750		6.86
116	100	750		11.44
148	140	750		16.02
173	180	750		20.59
223	220	750		25.17

### **3. *Mytilus galloprovincialis* Bivalve Larval Test**



CLIENT Anchor QEA	PROJECT Port Gamble Monitoring	JOB NUMBER 0	PROJECT MANAGER Brian Hester	LABORATORY Port Gamble	PROTOCOL PSEP (1995)
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**TEST ORGANISM SPAWNING DATA**

SPECIES <i>Mytilus spp</i>		TEST START DATE 3-Oct-18	
SUPPLIER Taylor shellfish		ORGANISM BATCH TS100218	
SPAWNING METHOD heatshock	INITIAL SPAWNING TIME 1225	FINAL SPAWNING TIME 1321	
MALES 6	FEMALES 4	SPERM VIABILITY ✓	EGG CONDITION Good
BEGIN FERTILIZATION 1321	END FERTILIZATION 1600	CONDITION OF EMBRYOS 790% div	

SAMPLE STORAGE 4 Degrees Celsius - dark
SEDIMENT TREATMENT none
TEST CHAMBERS 1 L Mason Jars
EXPOSURE VOLUME 900mL seawater / 18g Sediment
TIME OF SHAKE 1000-1040
TIME OF INITIATION 1620

**SPECIAL CONDITIONS**

UV LIGHT EXPOSURE (YES/NO) N	AERATION FROM TEST INITIATION (YES/NO) N
SCREEN TUBE TEST (YES/NO) N	OTHER (EXPLAIN) Resuspension

<p><b>EMBRYO DENSITY CALCULATIONS</b></p> <p>85x100 = 8500 eggs/mL</p> <p>Target <math>\frac{27,000 \text{ eggs/mL} / \text{jar}}{8500}</math></p> <p>= 3.17</p> <p>Deliver 3.2 mL/jar</p>	<p>RT <math>\frac{2700}{8500} = 0.317</math></p> <p>x 70 mL =</p> <p>22 mL egg stock</p> <p>48 mL seawater</p> <p>Deliver 0.100 mL/vial</p>
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SPECIES  
Mytilus spp

CLIENT Anchor QEA	PROJECT Port Gamble Monitoring	JOB NUMBER 0	PROJECT MANAGER Brian Hester	LAB / LOCATION Port Gamble /	PROTOCOL PSEP (1995)
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LARVAL OBSERVATION DATA

CLIENT / ID	REP	NUMBER NORMAL	NUMBER	DATE	TECHNICIAN	COMMENTS
STOCKING DENSITY	1		0245371	10/25/18	UB	
	2		338			
	3		326			
	4		322			
	5		323			
Control /	1	345	14	10/16/18	JW	
	2	343	11			
	3	344	11			
	4	310	7			312/4 OA
	5	324	12			
CARR-20 /	1	250	8			
	2	332	5			
	3	294	6			
	4	285	14			
	5	328	9			
CR-21 /	1	264	11			
	2	352	7			
	3	298	7			
	4	325	6			325/6 OA
	5	313	12			

① WC, UB 10/25

SPECIES <i>Mytilus spp</i>	
CLIENT Anchor QEA	PROJECT Port Gamble Monitoring
JOB NUMBER 0	PROJECT MANAGER Brian Hester
LAB / LOCATION Port Gamble /	PROTOCOL PSEP (1995)

**LARVAL OBSERVATION DATA**

CLIENT/ ID	REP	NUMBER NORMAL	NUMBER	DATE	TECHNICIAN	COMMENTS
SMA-1B-IT2 /	1	337	6	10/16/18	JW	
	2	274	5	↓	↓	
	3	316	6			
	4	300	3			293/4 OA
	5	284	5			
SMA-1B-IT3 /	1	318	6			
	2	299	8			
	3	336	10			
	4	324	11			
	5	325	4	↓		
SMA-2C-IT3 /	1	305	10	10/17/18		
	2	342	13	↓	↓	341/12 OA
	3	305	8			
	4	289	4			
	5	328	10			
SMA-2C-IT6 /	1	286	8			
	2	318	10			
	3	286	5			
	4	333	5			
	5	295	11	↓	↓	

## SPECIES

*Mytilus spp*

CLIENT Anchor QEA	PROJECT Port Gamble Monitoring	JOB NUMBER 0	PROJECT MANAGER Brian Hester	LAB / LOCATION Port Gamble /	PROTOCOL PSEP (1995)
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## LARVAL OBSERVATION DATA

CLIENT/ ID	REP	NUMBER NORMAL	NUMBER	DATE	TECHNICIAN	COMMENTS
SMA-2C-IT9 /	1	287	11	10/17/18	JW	
	2	291	14			
	3	283	7			
	4	304	9			
	5	301	9			286/12 OA
SMA-1-ST /	1	266	9			
	2	289	11			
	3	304	2			
	4	269	13			
	5	261	7			
SMA-1A-IT /	1	328	4			
	2	292	13			
	3	328	10			
	4	297	10			
	5	293	11			
SMA-2A-IT /	1	323	6			
	2	287	5			
	3	313	7			
	4	333	4			
	5	332	10			

## SPECIES

*Mytilus spp*

CLIENT Anchor QEA	PROJECT Port Gamble Monitoring	JOB NUMBER 0	PROJECT MANAGER Brian Hester	LAB / LOCATION Port Gamble /	PROTOCOL PSEP (1995)
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## LARVAL OBSERVATION DATA

CLIENT/ ID	REP	NUMBER NORMAL	NUMBER	DATE	TECHNICIAN	COMMENTS
SMA-2A-ST /	1	275	14	10/22/10	JW	
	2	336	11			
	3	285	6			
	4	303	8			
	5	322	6			
SMA-2B-IT /	1	280	7			
	2	297	10			
	3	318	4			
	4	276	10			
	5	319	10			
SMA-2B-ST /	1	320	8			
	2	302	12			
	3	277	7			
	4	310	5			
	5	297	2			281 / 2 OA
BW-15 /	1	268	10			
	2	287	30			
	3	340	21			
	4	305	16			
	5	272	19			

CLIENT Anchor QEA	PROJECT Port Gamble Monitoring	SPECIES Mytilus spp	LAB / LOCATION Port Gamble /	PROTOCOL PSEP (1995)
JOB NUMBER 0	PROJECT MANAGER Brian Hester	TEST START DATE 03Oct18	TIME 1620	TEST END DATE 10.5.18
TIME 1650				

\* Day 3&4 observations needed only if development endpoint not met by day 2

**WATER QUALITY DATA**

TEST CONDITIONS				DO (mg/L)		Temp (°C)		Sal (ppt)		pH		Ammonia		Sulfide		TECH	DATE
SAMPLE ID	DAY	Random #	REP	>5.0		16 ± 1		28 ± 1		7 - 9		NA		NA			
				D.O.		TEMP.		SALINITY		pH		AMMONIA		SULFIDE			
				meter	mg/L	meter	°C	meter	ppt	meter	unit	Techn. mg/L (total)	Techn. mg/L (Total)				
Control /	0	37	WQ Surr	8	7.7	8	16.5	8	28	8	7.7	RE	0.00	MK	ND	RE	10/3/18
Control /	1	↓	WQ Surr	8	7.5	8	16.7	8	28	8	7.8					MK	10/4
Control /	2	↓	WQ Surr	8	7.5	8	16.9	8	28	8	7.8	SW	0.00	JL	0.002	SW	10/5
Control /	3		WQ Surr														
Control /	4		WQ Surr														
CARR-20 /	0	45	WQ Surr	8	7.5	8	16.5	8	28	8	7.7	RE	0.00	MK	0.008	RE	10/3/18
CARR-20 /	1	↓	WQ Surr	8	6.6	8	16.6	8	28	8	7.7					MK	10/4
CARR-20 /	2	↓	WQ Surr	8	6.2	8	16.8	8	28	8	7.6	SW	0.00	JL	0.005	SW	10/5
CARR-20 /	3		WQ Surr														
CARR-20 /	4		WQ Surr														
CR-21 /	0	65	WQ Surr	8	7.0	8	16.4	8	28	8	7.7	RE	0.00	MK	0.004	RE	10/3/18
CR-21 /	1	↓	WQ Surr	8	6.5	8	16.5	8	28	8	7.7					MK	10/4
CR-21 /	2	↓	WQ Surr	8	6.5	8	16.8	8	28	8	7.6	SW	0.00	JL	0.002	SW	10/5
CR-21 /	3		WQ Surr														
CR-21 /	4		WQ Surr														

CLIENT Anchor QEA	PROJECT Port Gamble Monitoring	SPECIES Mytilus spp	LAB / LOCATION Port Gamble /	PROTOCOL PSEP (1995)
JOB NUMBER 0	PROJECT MANAGER Brian Hester	TEST START DATE 03Oct18	TIME 1620	TEST END DATE 10.5.18
TIME 1650				

\* Day 3&4 observations needed only if development endpoint not met by day 2

**WATER QUALITY DATA**

TEST CONDITIONS				DO (mg/L)		Temp (°C)		Sal (ppt)		pH		Ammonia		Sulfide		TECH	DATE
SAMPLE ID	DAY	Random #	REP	>5.0		16 ± 1		28 ± 1		7-9		NA		NA			
				D.O.	mg/L	TEMP.	°C	SALINITY	ppt	pH	unit	AMMONIA	SULFIDE				
				meter	mg/L	meter	°C	meter	ppt	meter	unit	Techn. mg/L (total)	Techn. mg/L (Total)				
SMA-1B-IT2 /	0	78	WQ Surr	8	7.3	8	16.5	8	28	8	7.7	RE	0.00	MK	ND	RE	10/3/18
SMA-1B-IT2 /	1	↓	WQ Surr	8	7.1	8	16.8	8	28	8	7.7					MK	10/4
SMA-1B-IT2 /	2	↓	WQ Surr	8	6.9	8	16.7	8	28	8	7.7	SW	0.00	JL	0.007	SW	10/5
SMA-1B-IT2 /	3		WQ Surr														
SMA-1B-IT2 /	4		WQ Surr														
SMA-1B-IT3 /	0	32	WQ Surr	8	7.7	8	16.5	8	28	8	7.7	RE	0.00	MK	0.002 ND	RE	10/3/18
SMA-1B-IT3 /	1	↓	WQ Surr	8	7.0	8	17.0	8	28	8	7.7					MK	10/4
SMA-1B-IT3 /	2	↓	WQ Surr	8	7.0	8	16.8	8	28	8	7.7	SW	0.00	JL	0.003	SW	10/5
SMA-1B-IT3 /	3		WQ Surr														
SMA-1B-IT3 /	4		WQ Surr														
SMA-2C-IT3 /	0	76	WQ Surr	8	7.5	8	16.4	8	28	8	7.7	RE	0.00	MK	0.001	RE	10/3/18
SMA-2C-IT3 /	1	↓	WQ Surr	8	7.1	8	16.6	8	28	8	7.7					MK	10/4
SMA-2C-IT3 /	2	↓	WQ Surr	8	7.1	8	16.9	8	28	8	7.7	SW	0.00	JL	ND	SW	10/5
SMA-2C-IT3 /	3		WQ Surr														
SMA-2C-IT3 /	4		WQ Surr														
SMA-2C-IT6 /	0	6	WQ Surr	8	7.4	8	16.4	8	27	8	7.7	RE	0.00	MK	ND	RE	10/3/18
SMA-2C-IT6 /	1	↓	WQ Surr	8	7.2	8	17.0	8	28	8	7.7					MK	10/4
SMA-2C-IT6 /	2	↓	WQ Surr	8	7.5	8	17.0	8	28	8	7.8	SW	0.00	JL	0.003	SW	10/5
SMA-2C-IT6 /	3		WQ Surr														
SMA-2C-IT6 /	4		WQ Surr														

① WC - 10/3/18 RE

② WC MK 10/3.

③ MK SW 10-5-18 DO = 7.6

CLIENT Anchor QEA	PROJECT Port Gamble Monitoring	SPECIES Mytilus spp	LAB / LOCATION Port Gamble /	PROTOCOL PSEP (1995)
JOB NUMBER 0	PROJECT MANAGER Brian Hester	TEST START DATE 03Oct18	TIME 1620	TEST END DATE 10.5.18
TIME 1650				

\* Day 3&4 observations needed only if development endpoint not met by day 2

**WATER QUALITY DATA**

TEST CONDITIONS				DO (mg/L)		Temp (°C)		Sal (ppt)		pH		Ammonia		Sulfide		TECH	DATE
SAMPLE ID	DAY	Random #	REP	>5.0		16 ± 1		28 ± 1		7 - 9		NA		NA			
				D.O.		TEMP.		SALINITY		pH		AMMONIA		SULFIDE			
				meter	mg/L	meter	°C	meter	ppt	meter	unit	Techn.	mg/L (total)	Techn.	mg/L (Total)		
SMA-2C-IT9 /	0	30	WQ Surr	8	7.7	8	16.5	8	28	8	7.7	Re	0.00	MK	0.00	Re	10/3/18
SMA-2C-IT9 /	1	↓	WQ Surr	8	7.4	8	16.7	8	28	8	7.8					MK	10/4
SMA-2C-IT9 /	2	↓	WQ Surr	8	7.6	8	16.6	8	28	8	7.8	SW	0.00	JL	ND	SW	10/5
SMA-2C-IT9 /	3		WQ Surr														
SMA-2C-IT9 /	4		WQ Surr														
SMA-1-ST /	0	82	WQ Surr	8	6.2	8	16.6	8	28	8	7.7	Re	0.00	MK	0.005	Re	10/3/18
SMA-1-ST /	1	↓	WQ Surr	8	6.9	8	16.8	8	28	8	7.7					MK	10/4
SMA-1-ST /	2	↓	WQ Surr	8	5.7	8	16.9	8	28	8	7.6	SW	0.00	JL	0.003	SW	10/5
SMA-1-ST /	3		WQ Surr														
SMA-1-ST /	4		WQ Surr														
SMA-1A-IT /	0	88	WQ Surr	8	6.7	8	16.7	8	28	8	7.7	Re	0.00	MK	ND	Re	10/3/18
SMA-1A-IT /	1	↓	WQ Surr	8	6.8	8	16.7	8	28	8	7.7					MK	10/4
SMA-1A-IT /	2	↓	WQ Surr	8	5.9	8	17.0	8	28	8	7.7	SW	0.00	JL	0.00	SW	10/5
SMA-1A-IT /	3		WQ Surr														
SMA-1A-IT /	4		WQ Surr														
SMA-2A-IT /	0	12	WQ Surr	8	7.4	8	16.7	8	28	8	7.7	Re	0.00	MK	ND	Re	10/3/18
SMA-2A-IT /	1	↓	WQ Surr	8	6.4	8	17.0	8	28	8	7.7					MK	10/4
SMA-2A-IT /	2	↓	WQ Surr	8	6.6	8	16.9	8	28	8	7.7	SW	0.00	JL	0.002	SW	10/5
SMA-2A-IT /	3		WQ Surr														
SMA-2A-IT /	4		WQ Surr														

① MP. Actual = 5.7 MK 10/4.  
 ② 1E. MK 10/4.



CLIENT Anchor QEA	PROJECT Port Gamble Monitoring	SPECIES Mytilus spp	LAB / LOCATION Port Gamble /	PROTOCOL PSEP (1995)
JOB NUMBER 0	PROJECT MANAGER Brian Hester	TEST START DATE 03Oct18	TIME 1620	TEST END DATE 10.5.18
TIME 1650				

\* Day 3&4 observations needed only if development endpoint not met by day 2

**WATER QUALITY DATA**

TEST CONDITIONS				DO (mg/L)		Temp (°C)		Sal (ppt)		pH		Ammonia		Sulfide		TECH	DATE
SAMPLE ID	DAY	Random #	REP	>5.0		16 ± 1		28 ± 1		7 - 9		NA		NA			
				meter	mg/L	meter	°C	meter	ppt	meter	unit	AMMONIA		SULFIDE			
				D.O.		TEMP.		SALINITY		pH		Techn. mg/L (total)		Techn. mg/L (Total)			
SMA-2A-ST /	0	18	WQ Surr	8	7.4	8	16.3	8	28	8	7.7	RE	0.00	MK	ND	RE	10/3/18
SMA-2A-ST /	1	↓	WQ Surr	8	6.4	8	17.0	8	28	8	7.7					MK	10/4
SMA-2A-ST /	2	↓	WQ Surr	8	6.4	8	16.8	8	28	8	7.6	SW	0.00	JL	0.001	SW	10/5
SMA-2A-ST /	3		WQ Surr														
SMA-2A-ST /	4		WQ Surr														
SMA-2B-IT /	0	59	WQ Surr	8	6.5	8	16.4	8	28	8	7.7	RE	0.00	MK	0.004	RE	10/3/18
SMA-2B-IT /	1	↓	WQ Surr	8	5.7	8	16.8	8	28	8	7.7					MK	10/4
SMA-2B-IT /	2	↓	WQ Surr	8	5.9	8	16.8	8	28	8	7.6	SW	0.00	JL	0.006	SW	10/5
SMA-2B-IT /	3		WQ Surr														
SMA-2B-IT /	4		WQ Surr														
SMA-2B-ST /	0	23	WQ Surr	8	7.4	8	16.5	8	28	8	7.7	RE	0.00	MK	0.007	RE	10/3/18
SMA-2B-ST /	1	↓	WQ Surr	8	6.9	8	16.9	8	28	8	7.7					MK	10/4
SMA-2B-ST /	2	↓	WQ Surr	8	6.9	8	16.8	8	28	8	7.7	SW	0.00	JL	ND	SW	10/5
SMA-2B-ST /	3		WQ Surr														
SMA-2B-ST /	4		WQ Surr														
BW-15 /	0	46	WQ Surr	8	7.4	8	16.3	8	28	8	7.7	RE	0.00	MK	0.005	RE	10/3/18
BW-15 /	1	↓	WQ Surr	8	6.3	8	16.8	8	28	8	7.7					MK	10/4
BW-15 /	2	↓	WQ Surr	8	6.1	8	16.7	8	28	8	7.6	SW	0.00	JL	0.004	SW	10/5
BW-15 /	3		WQ Surr														
BW-15 /	4		WQ Surr														

## ORGANISM RECEIPT LOG

<b>Date:</b> 10.2.18		<b>Time:</b> 1015		<b>Batch No.</b> TS100218			
<b>Organism:</b> Mytilus gallo							
<b>Source / Supplier:</b> Taylor Shellfish							
<b>No. Ordered:</b> 8 lbs		<b>No. Received:</b> 8 lbs		<b>Source Batch:</b> Collection date, hatch date, etc.): Collected 10.2.18 Totten Inlet			
<b>Condition of Organisms:</b> Good				<b>Approximate Size or Age:</b> (Days from hatch, life stage, size class, etc.): Adult			
<b>Shipper:</b> Courier				<b>B of L (Tracking No.)</b> N/A			
<b>Condition of Container:</b> Good				<b>Received By:</b> MARLH			
Container	D.O. (mg/L)	Temp. (°C)	Cond. or Sal. (Include Units)	pH (Units)	# Dead	% Dead*	Tech. (Initials)
1	—	①	—————	—————	—————		MARLH
*if >10% contact lab manager							
<b>Notes:</b> ① Shipped dry with ice packs MARLH 10.2.18 Temp not recorded							

### MAINTENANCE LOG FOR CULTURES

ORGANISM: Mytilus galloprovincialis  
 LOCATION: Bath 1

Batch Number: <sup>①</sup> <del>TS100318</del> TS100218	Date Received: 10.2.18	Initial # of Organisms: 8lbs
---------------------------------------------------------	------------------------	------------------------------

Date	Feed AM/PM	Tub No.	D.O.	Temp (°C)	Cond/Sal	pH	H <sub>2</sub> O Change	Organisms appear healthy (Y/N)	# Mort	% Mort*	Init.	Comments
10.3	X	1	5.6	13.7	31	7.4	FT	Y	0	—	BH	
↓	X	2	6.5	13.5	31	7.6	FT	Y	0	—	↓	
10/5	✓ <del>OX</del>	1	5.4	13.5	31	7.4	FT	Y	0	—	SW	
↓	✓ <del>OX</del>	2	7.4	13.1	32	7.7	FT	Y	0	—	↓	
10/8	✓	1	6.2	13.0	31	7.6	FT	Y	1	—	SW	
↓	✓	2	7.4	13.3	31	7.7	FT	Y	0	—	SW	
10/10	✓	1	6.5	12.8	31	7.4	FT	Y	0	—	SW	
↓	✓	2	7.5	12.8	31	7.6	↓	↓	0	—	SW	
10/12		1	6.4	13.0	31	7.4	FT	Y	0	—	RE	
↓		2	7.3	12.7	31	7.6	FT	Y	0	—	RE	
10/15	✓	1	6.6	13.3	30	7.5	FT	Y	0	—	US	
↓	✓	2	6.8	12.9	31	7.7	↓	Y	0	—	↓	
10/17	✓	1	6.8	13.0	31	7.5	FT	Y	0	—	SW	
↓	✓	2	7.6	12.7	31	7.6	↓	↓	↓	—	↓	
10/19	✓	1	6.8	13.0	30	7.6	FT	Y	0	—	SW	
↓	✓	2	7.7	12.7	31	7.8	↓	Y	0	—	SW	
10/22	✓	1	7.0	12.7	31	7.6	FT	Y	0	—	SW	
↓	✓	2	7.8	12.6	31	7.8	FT	Y	0	—	SW	

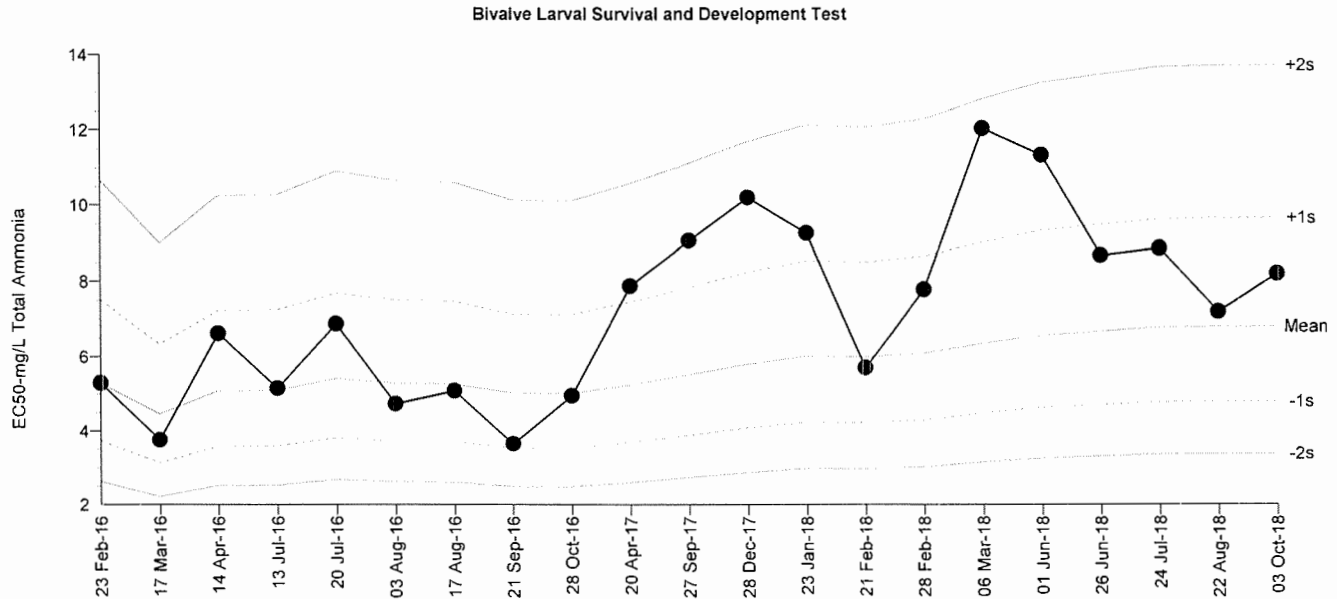
FT = Flow-through  
 \*if >10% notify lab manager

① IE 10.3.18 BH @ WC 10.5.18 AM feed SW

Bivalve Larval Survival and Development Test

All Matching Labs

Test Type: Development-Survival      Organism: Mytilus galloprovincialis (Bay Mussel)      Material: Total Ammonia  
 Protocol: EPA/600/R-95/136 (1995)      Endpoint: Combined Proportion Normal      Source: Reference Toxicant-REF



Mean: 6.807      Count: 20      -1s Warning Limit: 4.794      -2s Action Limit: 3.377  
 Sigma: n/a      CV: 36.10%      +1s Warning Limit: 9.66      +2s Action Limit: 13.71

Quality Control Data

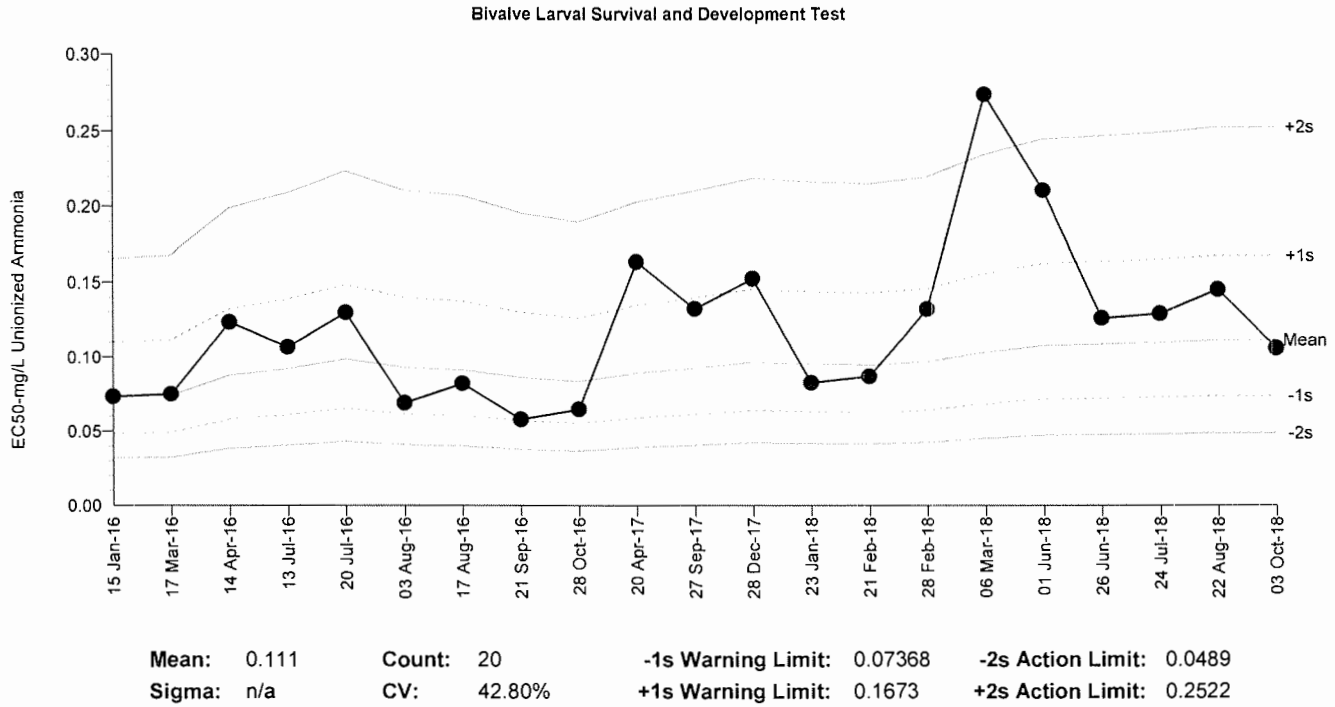
Point	Year	Month	Day	Time	QC Data	Delta	Sigma	Warning	Action	Test ID	Analysis ID	Laboratory
1	2016	Feb	23	17:50	5.292	-1.516	-0.7189			18-1470-2153	19-1029-0373	ENVIRON
2		Mar	17	19:30	3.757	-3.05	-1.697	(-)		15-5000-9198	13-5264-4225	ENVIRON
3		Apr	14	17:15	6.621	-0.186	-0.07907			20-6935-4588	11-5576-6536	ENVIRON
4		Jul	13	19:55	5.147	-1.66	-0.7981			21-3594-7965	13-9208-2204	ENVIRON
5			20	19:37	6.871	0.0637	0.02659			15-8198-2198	11-4931-7833	ENVIRON
6		Aug	3	18:20	4.732	-2.075	-1.038	(-)		01-0657-3943	18-0523-9298	ENVIRON
7			17	17:05	5.081	-1.726	-0.8348			12-6418-6345	06-5970-9287	ENVIRON
8		Sep	21	17:05	3.644	-3.163	-1.784	(-)		12-2755-6335	12-7771-4113	ENVIRON
9		Oct	28	16:55	4.947	-1.86	-0.9112			11-5556-2644	13-8974-7601	ENVIRON
10	2017	Apr	20	17:20	7.867	1.06	0.4132			10-4553-7194	08-5173-1627	EcoAnalysts
11		Sep	27	17:50	9.055	2.248	0.8145			17-5033-2538	21-1690-3737	EcoAnalysts
12		Dec	28	15:06	10.19	3.381	1.151	(+)		14-5351-9068	16-2506-4728	EcoAnalysts
13	2018	Jan	23	15:10	9.255	2.448	0.8769			09-7170-7085	18-2220-1310	EcoAnalysts
14		Feb	21	16:50	5.699	-1.109	-0.5074			11-9789-0593	04-0007-2351	EcoAnalysts
15			28	15:50	7.78	0.9724	0.3812			07-6747-7949	18-9659-8443	EcoAnalysts
16		Mar	6	17:25	12.04	5.23	1.627	(+)		07-4370-2173	07-8492-0076	EcoAnalysts
17		Jun	1	18:20	11.33	4.525	1.455	(+)		12-7842-7476	20-1810-5029	EcoAnalysts
18			26	19:10	8.678	1.871	0.6931			17-9894-2551	06-4820-9232	EcoAnalysts
19		Jul	24	14:29	8.87	2.062	0.7554			03-7590-2196	04-9004-0822	EcoAnalysts
20		Aug	22	17:20	7.194	0.387	0.1579			16-3013-6338	06-6775-4461	EcoAnalysts
21		Oct	3	16:45	8.195	1.388	0.5296			12-1625-9203	05-0256-3584	EcoAnalysts



Bivalve Larval Survival and Development Test

All Matching Labs

Test Type: Development-Survival Organism: Mytilus galloprovincialis (Bay Mussel) Material: Unionized Ammonia  
 Protocol: EPA/600/R-95/136 (1995) Endpoint: Combined Proportion Normal Source: Reference Toxicant-REF



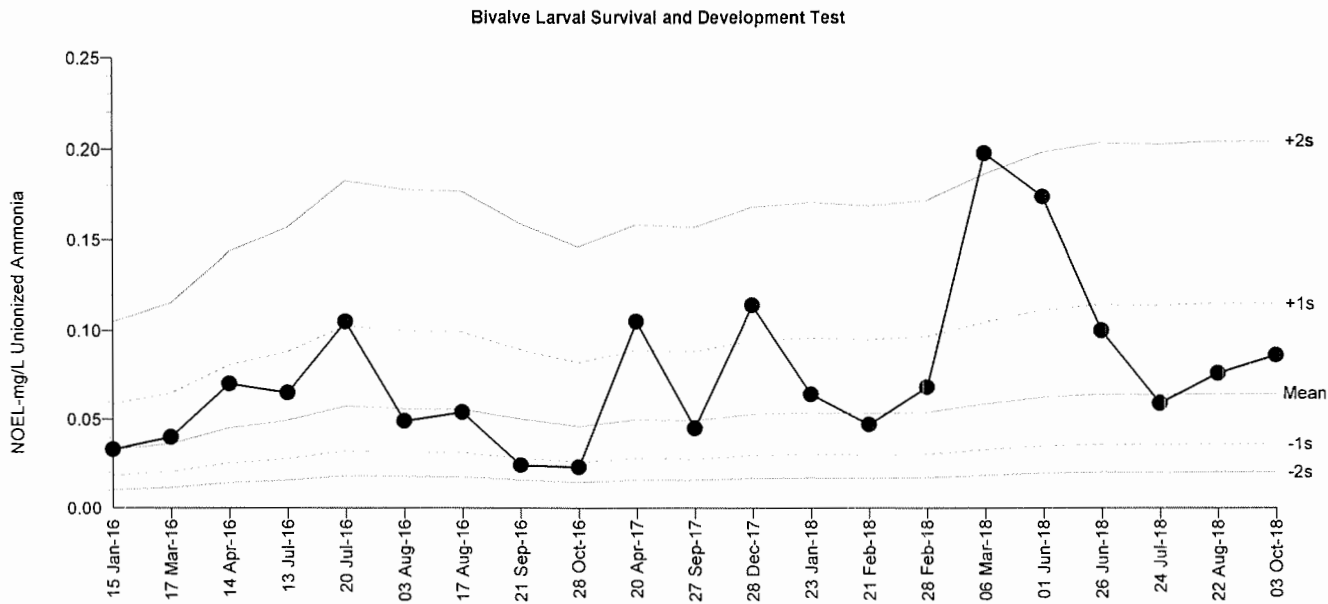
Quality Control Data

Point	Year	Month	Day	Time	QC Data	Delta	Sigma	Warning	Action	Test ID	Analysis ID	Laboratory
1	2016	Jan	15	18:45	0.07307	-0.03795	-1.02	(-)		14-3705-6085	19-2478-0688	ENVIRON
2		Mar	17	19:30	0.0747	-0.03632	-0.9661			04-9304-7933	11-8311-5806	ENVIRON
3		Apr	14	17:15	0.1231	0.01209	0.252			21-0694-5716	16-1625-7449	ENVIRON
4		Jul	13	19:55	0.1066	-0.004396	-0.09852			17-3624-8523	09-8766-4801	ENVIRON
5			20	19:37	0.1295	0.01852	0.3761			16-4492-6137	18-6316-6428	ENVIRON
6		Aug	3	18:20	0.0688	-0.04222	-1.167	(-)		09-2291-5121	03-3596-0456	ENVIRON
7			17	17:05	0.08185	-0.02917	-0.7433			07-3188-3147	05-9364-3209	ENVIRON
8		Sep	21	17:05	0.05781	-0.05321	-1.591	(-)		12-6262-3928	08-6003-5732	ENVIRON
9		Oct	28	16:55	0.06459	-0.04644	-1.321	(-)		05-5636-6924	16-1083-2604	ENVIRON
10	2017	Apr	20	17:20	0.1634	0.05235	0.9419			05-0815-7437	21-2956-9302	EcoAnalysts
11		Sep	27	17:50	0.1319	0.02091	0.4208			20-9743-5500	21-3701-9130	EcoAnalysts
12		Dec	28	15:06	0.1522	0.04115	0.7688			11-9837-6366	14-7670-9371	EcoAnalysts
13	2018	Jan	23	15:10	0.08248	-0.02854	-0.7247			14-9653-2794	09-5264-1795	EcoAnalysts
14		Feb	21	16:50	0.08676	-0.02426	-0.6012			15-2546-6673	13-8547-7245	EcoAnalysts
15			28	15:50	0.1317	0.02067	0.4162			15-8114-9389	20-6854-2996	EcoAnalysts
16		Mar	6	17:25	0.2738	0.1628	2.201	(+)	(+)	00-7847-6930	00-7321-8707	EcoAnalysts
17		Jun	1	18:20	0.2103	0.09926	1.557	(+)		05-1801-7066	04-7994-1641	EcoAnalysts
18			26	19:10	0.1255	0.01446	0.2985			06-7139-6937	06-3245-7683	EcoAnalysts
19		Jul	24	14:29	0.1286	0.01757	0.3582			16-0134-7952	19-7798-4523	EcoAnalysts
20		Aug	22	17:20	0.1449	0.03384	0.6487			07-1087-5324	12-5339-9193	EcoAnalysts
21		Oct	3	16:45	0.1056	-0.005424	-0.1221			01-0403-3220	16-2351-0172	EcoAnalysts

Bivalve Larval Survival and Development Test

All Matching Labs

Test Type: Development-Survival      Organism: Mytilus galloprovincialis (Bay Mussel)      Material: Unionized Ammonia  
 Protocol: EPA/600/R-95/136 (1995)      Endpoint: Combined Proportion Normal      Source: Reference Toxicant-REF



Mean: 0.06451      Count: 20      -1s Warning Limit: 0.03622      -2s Action Limit: 0.02033  
 Sigma: n/a      CV: 62.90%      +1s Warning Limit: 0.115      +2s Action Limit: 0.2048

Quality Control Data

Point	Year	Month	Day	Time	QC Data	Delta	Sigma	Warning	Action	Test ID	Analysis ID	Laboratory
1	2016	Jan	15	18:45	0.033	-0.03151	-1.161	(-)		14-3705-6085	10-1556-6538	ENVIRON
2		Mar	17	19:30	0.04	-0.02451	-0.8276			04-9304-7933	18-2075-7796	ENVIRON
3		Apr	14	17:15	0.07	0.005494	0.1416			21-0694-5716	12-4502-8479	ENVIRON
4		Jul	13	19:55	0.065	0.0004942	0.01322			17-3624-8523	04-6243-8640	ENVIRON
5			20	19:37	0.105	0.04049	0.8438			16-4492-6137	18-5505-5894	ENVIRON
6		Aug	3	18:20	0.049	-0.01551	-0.4762			09-2291-5121	05-1896-5100	ENVIRON
7			17	17:05	0.054	-0.01051	-0.3079			07-3188-3147	16-9734-3329	ENVIRON
8		Sep	21	17:05	0.024	-0.04051	-1.712	(-)		12-6262-3928	13-9034-6853	ENVIRON
9		Oct	28	16:55	0.023	-0.04151	-1.786	(-)		05-5636-6924	02-7414-4116	ENVIRON
10	2017	Apr	20	17:20	0.105	0.04049	0.8438			05-0815-7437	03-4135-8920	EcoAnalysts
11		Sep	27	17:50	0.045	-0.01951	-0.6236			20-9743-5500	07-1720-0800	EcoAnalysts
12		Dec	28	15:06	0.114	0.04949	0.9862			11-9837-6366	06-0841-7631	EcoAnalysts
13	2018	Jan	23	15:10	0.064	-0.000506	-0.01363			14-9653-2794	04-0773-5227	EcoAnalysts
14		Feb	21	16:50	0.047	-0.01751	-0.5483			15-2546-6673	21-3476-8661	EcoAnalysts
15			28	15:50	0.068	0.003494	0.09136			15-8114-9389	10-5284-4887	EcoAnalysts
16		Mar	6	17:25	0.198	0.1335	1.942	(+)		00-7847-6930	03-9693-3709	EcoAnalysts
17		Jun	1	18:20	0.174	0.1095	1.719	(+)		05-1801-7066	00-0472-0630	EcoAnalysts
18			26	19:10	0.1	0.03549	0.7593			06-7139-6937	00-3402-6456	EcoAnalysts
19		Jul	24	14:29	0.059	-0.005506	-0.1545			16-0134-7952	12-3548-9987	EcoAnalysts
20		Aug	22	17:20	0.076	0.01149	0.284			07-1087-5324	19-8244-5254	EcoAnalysts
21		Oct	3	16:45	0.086	0.02149	0.4981			01-0403-3220	16-6521-7341	EcoAnalysts





**CETIS Test Data Worksheet**

Report Date: 26 Oct-18 10:31 (p 1 of 1)  
 Test Code/ID: 487EA483 / 12-1625-9203

<b>Bivalve Larval Survival and Development Test</b>				<b>EcoAnalysts</b>			
<b>Start Date:</b> 03 Oct-18 16:45 ✓	<b>Species:</b> Mytilus galloprovincialis			<b>Sample Code:</b> 7A5A1C38			
<b>End Date:</b> 05 Oct-18 16:40 ✓	<b>Protocol:</b> EPA/600/R-95/136 (1995)			<b>Sample Source:</b> Reference Toxicant			
<b>Sample Date:</b> 15 May-17	<b>Material:</b> Total Ammonia			<b>Sample Station:</b> P170515.105 ✓			

Conc-mg/L	Code	Rep	Pos	Initial Density	Final Density	# Counted	# Normal	Notes
0	D	1	11	289	277	277	274	OAd on total <sup>pgs</sup> w/A
0	D	2	1	289	294	294	292	
0	D	3	17	289	280	280	275	
0.867 ✓		1	6	289	284	284	279	
0.867		2	2	289	301	301	295	
0.867		3	8	289	303	303	298	
1.46 ✓		1	19	289	324	324	320	
1.46		2	12	289	285	285	280	
1.46		3	14	289	307	307	302	
3.18 ✓		1	4	289	301	301	293	
3.18		2	10	289	264	264	261	
3.18		3	9	289	306	306	295	
6.01 ✓		1	15	289	320	320	289	
6.01		2	13	289	279	279	250	
6.01		3	3	289	301	301	269	
12 ✓		1	16	289	225	225	2	
12		2	21	289	286	286	3	
12		3	5	289	323	323	6	
20.7 ✓		1	20	289	293	293	0	
20.7		2	7	289	305	305	0	
20.7		3	18	289	310	310	0	

# CETIS Summary Report

Report Date: 26 Oct-18 10:35 (p 1 of 1)  
 Test Code/ID: 6336BC4 / 01-0403-3220

## Bivalve Larval Survival and Development Test

EcoAnalysts

Batch ID: 14-2069-0648	Test Type: Development-Survival	Analyst:
Start Date: 03 Oct-18 16:45	Protocol: EPA/600/R-95/136 (1995)	Diluent: Laboratory Seawater
Ending Date: 05 Oct-18 16:40	Species: Mytilus galloprovincialis	Brine: Crystal Sea Marine Mix
Test Length: 48h	Taxon:	Source: Taylor Shellfish
Sample ID: 09-8775-9708	Code: 3AE0045C	Project: Reference Toxicant
Sample Date: 15 May-17	Material: Unionized Ammonia	Source: Reference Toxicant
Receipt Date: 15 May-17	CAS (PC):	Station: P170515.105
Sample Age: 506d 17h	Client: Internal Lab	Age:

### Multiple Comparison Summary

Analysis ID	Endpoint	Comparison Method	✓ NOEL	LOEL	TOEL	TU	PMSD	S
16-6521-7341	Combined Proportion Norma	Dunnett Multiple Comparison Test	0.086	0.138	0.1089		12.9%	1

### Point Estimate Summary

Analysis ID	Endpoint	Point Estimate Method	✓ Level	mg/L	95% LCL	95% UCL	TU	S
16-2351-0172	Combined Proportion Norma	Spearman-Kärber	EC50	0.1056	0.1045	0.1067		1

### Test Acceptability

Analysis ID	Endpoint	Attribute	Test Stat	TAC Limits		Overlap	Decision
				Lower	Upper		
16-6521-7341	Combined Proportion Norma	PMSD	0.1287	<<	0.25	No	Passes Criteria

### Combined Proportion Normal Summary

Conc-mg/L	Code	Count	Mean	95% LCL	95% UCL	Min	Max	Std Err	Std Dev	CV%	%Effect
0	D	3	0.9666	0.8945	1.0000	0.9481	1.0000	0.0168	0.0290	3.00%	0.00%
0.013		3	0.9885	0.9388	1.0000	0.9654	1.0000	0.0115	0.0200	2.02%	-2.27%
0.021		3	0.9896	0.9450	1.0000	0.9689	1.0000	0.0104	0.0180	1.82%	-2.39%
0.046		3	0.9677	0.8287	1.0000	0.9031	1.0000	0.0323	0.0559	5.78%	-0.12%
0.086		3	0.9319	0.7643	1.0000	0.8651	1.0000	0.0390	0.0675	7.24%	3.58%
0.138		3	0.0127	0.0000	0.0306	0.0069	0.0208	0.0042	0.0072	56.77%	98.69%
0.24		3	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000		100.00%

### Combined Proportion Normal Detail

Conc-mg/L	Code	Rep 1	Rep 2	Rep 3
0	D	0.9481	1.0000	0.9516
0.013		0.9654	1.0000	1.0000
0.021		1.0000	0.9689	1.0000
0.046		1.0000	0.9031	1.0000
0.086		1.0000	0.8651	0.9308
0.138		0.0069	0.0104	0.0208
0.24		0.0000	0.0000	0.0000

### Combined Proportion Normal Binomials

Conc-mg/L	Code	Rep 1	Rep 2	Rep 3
0	D	274/289	292/292	275/289
0.013		279/289	295/295	298/298
0.021		320/320	280/289	302/302
0.046		293/293	261/289	295/295
0.086		289/289	250/289	269/289
0.138		2/289	3/289	6/289
0.24		0/289	0/289	0/289

**CETIS Test Data Worksheet**

Report Date: 26 Oct-18 10:35 (p 1 of 1)  
 Test Code/ID: 6336BC4 / 01-0403-3220

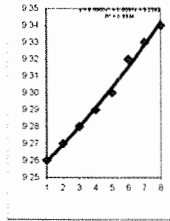
Bivalve Larval Survival and Development Test								EcoAnalysts
Start Date: 03 Oct-18 16:45 ✓		Species: Mytilus galloprovincialis		Sample Code: 3AE0045C				
End Date: 05 Oct-18 16:40 ✓		Protocol: EPA/600/R-95/136 (1995)		Sample Source: Reference Toxicant				
Sample Date: 15 May-17		Material: Unionized Ammonia		Sample Station: P170515.105 ✓				
Conc-mg/L	Code	Rep	Pos	Initial Density	Final Density	# Counted	# Normal	Notes
0	D	1	18	289 ✓	277	277 ✓	274 ✓	
0	D	2	11	289	294	294 ✓	292 ✓	
0	D	3	13	289	280	280 ✓	275 ✓	
0.013 ✓		1	12	289	284	284 ✓	279 ✓	
0.013		2	21	289	301	301 ✓	295 ✓	
0.013		3	3	289	303	303 ✓	298 ✓	
0.021 ✓		1	9	289	324	324 ✓	320 ✓	
0.021		2	15	289	285	285 ✓	280 ✓	
0.021		3	17	289	307	307 ✓	302 ✓	
0.046 ✓		1	16	289	301	301 ✓	293 ✓	
0.046		2	10	289	264	264 ✓	261 ✓	
0.046		3	2	289	306	306 ✓	295 ✓	
0.086 ✓		1	1	289	320	320 ✓	289 ✓	
0.086		2	6	289	279	279 ✓	250 ✓	
0.086		3	8	289	301	301 ✓	269 ✓	
0.138 ✓		1	20	289	225	225 ✓	2 ✓	
0.138		2	5	289	286	286 ✓	3 ✓	
0.138		3	14	289	323	323 ✓	6 ✓	
0.24 ✓		1	19	289	293	293 ✓	0 ✓	
0.24		2	4	289	305	305 ✓	0 ✓	
0.24		3	7	289	310	310 ✓	0 ✓	

# Un-ionized Ammonia Calculator

CLIENT:	Various	Date of Test:	October 3, 2018 ✓
PROJECT:	PG1123, PG1162 ✓	Test Type:	<i>Mytilus gallo</i> ✓
COMMENTS:			

To convert Total Ammonia (mg/L) to Free (un-ionized) Ammonia (mg/L) enter the corresponding total ammonia, salinity, temperature, and pH.

Integer: I-factor	
1	9.26
2	9.27
3	9.28
4	9.29
5	9.30
6	9.32
7	9.33
8	9.34



Sample	Mod NH3T (mg/L)	salinity (ppt)	pH	temp (C)	temp (K)	i-factor	Mod NH3U (mg/L)
1							
2	0 ✓	2.000	10.0	7.5	5.0	278.16	9.2750
3	0.75 ✓	0.867 ✓	28 ✓	7.8 ✓	15.2 ✓	288.36	9.3187
4	1.5 ✓	1.46 ✓	28 ✓	7.8 ✓	15.0 ✓	288.16	9.3187
5	3 ✓	3.18 ✓	28 ✓	7.8 ✓	15.1 ✓	288.26	9.3187
6	6 ✓	6.01 ✓	28 ✓	7.8 ✓	15.0 ✓	288.16	9.3187
7	12 ✓	12 ✓	28 ✓	7.7 ✓	15.1 ✓	288.26	9.3187
8	18 ✓	20.7 ✓	28 ✓	7.7 ✓	15.2 ✓	288.36	9.3187
9							
10							
11							
12							
13							
14							
15							
16							
17							
18							
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45							

### 48 Hour Bivalve Development Reference Toxicant Test

Test ID: <b>PI70515; 105</b>	Replicates: 3	Study Director: <b>B. Hester</b>	Location: <b>Inc. 1</b>				
Dilution Water Batch: <b>FSW100318.02</b>	Organism Batch: <b>TS100218</b>	Associated Test(s): <b>PG1162, PG1123</b>	Organism: <b>Mytilus spp.</b>				
Chamber Size/Type: 30 ml shell vial	Exposure Volume: 10 ml						
Toxicant: Ammonium Chloride:	Lot #: <b>2986C510</b>	Date Prepared: <b>10/3/18</b>	Initials: <b>MK</b>				
Target Concentrations: <b>See spiking worksheet</b>		Quantity of Stock: Target: <b>See spiking worksheet</b>	Quantity of Diluent: Target: <b>200 mL</b>				
See spiking worksheet		Actual: See spiking worksheet	Actual: <b>↓</b>				
<b>0 Hours</b> Date: <b>10/3/18</b> WQ Time: <b>MK</b> Start Time: <b>1645</b> Initials: <b>BH</b>							
<b>STOCK</b>							
	Control	0.75	1.5	3	6	12	18
D.O. (%) (>5.0 mg/L)	7.6	7.8	7.9	8.0	8.0	8.0	8.1
Temperature (16 ± 1°C)	15.5	15.2	15.0	15.1	15.0	15.1	15.2
Salinity (28 ± 1 ppt)	28	28	28	28	28	28	28
pH (7.8 ± 0.5)	7.8	7.8	7.8	7.8	7.8	7.7	7.7
<b>Day 1</b>	Temperature (16 ± 1°C)	<b>15.6°C</b>		<b>10/4 MK</b>			
<b>Final Day</b> Date: <b>10/05/18</b> WQ Time: <b>1440</b> End Time: <b>1640</b> Initials: <b>JL</b>							
<b>STOCK</b>							
	Control	0.75	1.5	3	6	12	18
D.O. (%) (>5.0 mg/L)	8.0	8.1	8.3	8.3	8.3	8.4	8.3
Temperature (16 ± 1°C)	15.3	15.2	14.6	15.2	14.5	15.3	15.2
Salinity (28 ± 1 ppt)	28	28	29	28	29	28	29
pH (7.8 ± 0.5)	7.8	7.8	7.8	7.8	7.8	7.8	7.8

Notes:

### 48 Hour Bivalve Development Reference Toxicant Test

Conc.	Rep	Number Normal	Number Abnormal	Date	Initials
Control	1	274	3	10.25.18	MARH
	2	292	2		
	3	275	5		
0.75	1	279	5		
	2	295	6		
	3	298	5		
1.5	1	320	4		
	2	280	5		
	3	302	5		
3	1	293	8		
	2	261	3		
	3	295	11		
6	1	289	31		
	2	250	29		
	3	269	32		
12	1	2	223		
	2	3	283		
	3	6	317		
18	1	0	293		
	2	0	305		
	3	0	310	↓	↓
<b>Stocking Density</b>					
<b>Rep</b>		<b>Count</b>		<b>Init.</b>	
1		311		MARH	
2		274 R4 315		↓	
3		269 R5 275		↓	
<b>Mean:</b>		289		u	
<b>QA Count Checks:</b>					
#1 conc/rep <u>0/3</u> # normal <u>274</u> # abnormal <u>6</u>	#2 conc/rep <u>1.5/1</u> # normal <u>310</u> # abnormal <u>6</u>	#3 conc/rep <u>12/2</u> # normal <u>5</u> # abnormal <u>280</u>	#4 conc/rep <u>3/1</u> # normal <u>275</u> # abnormal <u>12</u>		
Tech. Init. <u>UB</u>	Tech. Init. <u>UB</u>	Tech. Init. <u>UB</u>	Tech. Init. <u>UB</u>		
Calc. $\frac{274}{280} = 97.9$ $\frac{275}{280} = 98.2$	$\frac{310}{316} = 98.1$ $\frac{320}{324} = 98.8$	$\frac{5}{285} = 1.8$ $\frac{3}{286} = 1.0$	$\frac{275}{287} = 95.8$ $\frac{293}{301} = 97.3$		
QA Check Acceptability: <input checked="" type="checkbox"/> <5% difference in means of QA & orig. counts					

**Ammonia Reference Toxicant  
Spiking Worksheet**

Reference Toxicant ID:

P170575.105

Date Prepared:

10/3/18

Technician Initials:

Mk

# Biv / Echino NH<sub>3</sub> RT

Assumptions in Model

Date:

10/2/2018

Stock ammonia concentration is 9,000 mg/L = 9 mg/mL

Measurement:

9833

Test Solutions			Volume of stock to reach desired concentration		
Measured Concentration	Desired Concentration	Volume	mL stock to increase		
mg/L	mg/L	mL	SALT WATER		
0	0				
0.867	0.75	200			0.023
1.46	1.5	200			0.046
3.18	3	200			0.092
6.01	6	200			0.183
12.0	12	200			0.366
20.7	18	200			0.549

## **APPENDIX B. STATISTICAL COMPARISONS**



## Statistical Results: *Eohaustorius estuarius* Test







Project Name: Anchor Port Gamble 2018 Eohs

Sample: x1  
 Samp ID: SMA-2C-IT6  
 Alias:  
 Replicates: 5  
 Mean: 1  
 SD: 2.236  
 Tr Mean: N/A  
 Trans SD: N/A

Ref Samp: x2  
 Ref ID: CR-21  
 Alias:  
 Replicates: 5  
 Mean: 2  
 SD: 2.739  
 Tr Mean: N/A  
 Trans SD: N/A

Shapiro-Wilk Results:

Residual Mean: 0  
 Residual SD: 4.192  
 SS: 333.903  
 K: 5  
 b: 15.917  
 Alpha Level: 0.05  
 Calculated Value: 0.7588  
 Critical Value: <= 0.842

Normally Distributed: No

Override Option: Not Invoked

Levene's Results:

Test Residual Mean: 4.135  
 Test Residual SD: 3.467  
 Ref. Residual Mean: 6.202  
 Ref. Residual SD: 1.415  
 Deg. of Freedom: 8  
 Alpha Level: 0.1  
 Calculated Value: 1.2344  
 Critical Value: >= 1.860

Variances Homogeneous: Yes

Test Results:

Statistic: Mann-Whitney  
 Balanced Design: Yes  
 Transformation: rank-order

Null: Experimental Hypothesis  
 Alternate: x1 >= x2  
 x1 < x2

Mann-Whitney N1: 5  
 Mann-Whitney N2: 5  
 Degrees of Freedom: 15  
 Experimental Alpha Level: 0.05  
 Calculated Value: 15  
 Critical Value: >= 21.000  
 Accept Null Hypothesis: Yes

Power:  
 Min. Difference for Power:

Replicate Number	Test Data	Trans. Test Data	Reference Data	Trans. Reference Data	Levene's Test Residuals	Levene's Reference Residuals	Mann-Whitney Ranks	Rankits	Shapiro-Wilk Residuals
1	0	4	0	4	2.584	5.168	4		-5.168
2	0	4	5	9	2.584	7.753	4		-5.168
3	0	4	0	4	2.584	5.168	4		-5.168
4	0	4	0	4	2.584	5.168	4		-2.584
5	5	9	5	9	10.337	7.753	4		-2.584
6							4		-2.584
7							4		-2.584
8							9		7.753
9							9		7.753
10							9		10.337

Project Name: Anchor Port Gamble 2018 Eohs

Sample: x1  
 Samp ID: SMA-2C-IT9  
 Alias:  
 Replicates: 5  
 Mean: 4  
 SD: 6.519  
 Tr Mean: N/A  
 Trans SD: N/A

Ref Samp: x2  
 Ref ID: CR-21  
 Alias:  
 Replicates: 5  
 Mean: 2  
 SD: 2.739  
 Tr Mean: N/A  
 Trans SD: N/A

Shapiro-Wilk Results:

Residual Mean: 0  
 Residual SD: 5.765  
 SS: 631.513  
 K: 5  
 b: 22.508  
 Alpha Level: 0.05  
 Calculated Value: 0.8022  
 Critical Value: <= 0.842

Normally Distributed: No

Override Option: Not Invoked

Levene's Results:

Test Residual Mean: 8.57  
 Test Residual SD: 3.999  
 Ref. Residual Mean: 6.202  
 Ref. Residual SD: 1.415  
 Deg. of Freedom: 8  
 Alpha Level: 0.1  
 Calculated Value: 1.2481  
 Critical Value: >= 1.860

Variances Homogeneous: Yes

Test Results:

Statistic: Mann-Whitney  
 Balanced Design: Yes  
 Transformation: rank-order

Null: Experimental Hypothesis  
 Alternate: x1 >= x2  
 x1 < x2

Mann-Whitney N1: 5  
 Mann-Whitney N2: 5  
 Degrees of Freedom:  
 Experimental Alpha Level: 0.05  
 Calculated Value: 11.5  
 Critical Value: >= 21.000  
 Accept Null Hypothesis: Yes

Power:  
 Min. Difference for Power:

Replicate Number	Test Data	Trans. Test Data	Reference Data	Trans. Reference Data	Levene's Test Residuals	Levene's Reference Residuals	Mann-Whitney Ranks	Rankits	Shapiro-Wilk Residuals
1	15	10	0	3.5	15.645	5.168	3.5		-7.141
2	0	3.5	5	8	7.141	7.753	3.5		-7.141
3	5	8	0	3.5	5.779	5.168	3.5		-7.141
4	0	3.5	0	3.5	7.141	5.168	3.5		-5.168
5	0	3.5	5	8	7.141	7.753	3.5		-5.168
6							3.5		-5.168
7							8		5.779
8							8		7.753
9							8		7.753
10							10		15.645

## Statistical Results: *Neanthes arenaceodentata* Test









Project Name: Anchor Port Gamble 2018 Neanthes Dry Weight

Sample: x1  
 Samp ID: SMA-2C-IT6  
 Alias:  
 Replicates: 5  
 Mean: 0.782  
 SD: 0.098  
 Tr Mean: N/A  
 Trans SD: N/A

Ref Samp: x2  
 Ref ID: CR-21  
 Alias:  
 Replicates: 5  
 Mean: 0.735  
 SD: 0.147  
 Tr Mean: N/A  
 Trans SD: N/A

Shapiro-Wilk Results:

Residual Mean: 0  
 Residual SD: 0.081  
 SS: 0.125  
 K: 5  
 b: 0.323  
 Alpha Level: 0.05  
 Calculated Value: 0.8356  
 Critical Value: <= 0.842

Normally Distributed: No

Override Option: Not Invoked

Levene's Results:

Test Residual Mean: 0.07  
 Test Residual SD: 0.06  
 Ref. Residual Mean: 0.103  
 Ref. Residual SD: 0.091  
 Deg. of Freedom: 8  
 Alpha Level: 0.1  
 Calculated Value: 0.6805  
 Critical Value: >= 1.860

Variances Homogeneous: Yes

Test Results:

Statistic: Mann-Whitney  
 Balanced Design: Yes  
 Transformation: rank-order

Null: x1 >= x2  
 Alternate: x1 < x2

Experimental Hypothesis  
 Mann-Whitney N1: 5  
 Mann-Whitney N2: 5  
 Degrees of Freedom:  
 Experimental Alpha Level: 0.05  
 Calculated Value: 9  
 Critical Value: >= 21.000  
 Accept Null Hypothesis: Yes

Power:  
 Min. Difference for Power:

Replicate Number	Test Data	Trans. Test Data	Reference Data	Trans. Reference Data	Levene's Test Residuals	Levene's Reference Residuals	Mann-Whitney Ranks	Rankits	Shapiro-Wilk Residuals
1	0.608	2	0.775	4	0.174	0.04	1		-0.239
2	0.801	5	0.496	1	0.019	0.239	2		-0.174
3	0.839	9	0.802	6	0.057	0.067	3		-0.018
4	0.831	7	0.886	10	0.049	0.151	4		0.019
5	0.832	8	0.717	3	0.05	0.018	5		0.04
6							6		0.049
7							7		0.05
8							8		0.057
9							9		0.067
10							10		0.151















# **Statistical Results: *Mytilus galloprovincialis* Larval Test**



























## **APPENDIX C. CHAIN-OF-CUSTODY LOGS**



Date: 9/6/18

Laboratory: EcoAnalysts  
 Project Name: Port Gamble - OMMP LTM  
 Project Number: 180388-01.01  
 Project Contact: Cheronne Oreiro  
 Phone Number: 206.287.9130  
 Shipment Method: Delivery

Sediment and Field QC

Line	Field Sample ID	Collection Date/Time	Matrix	No. of Containers	Bioassay Archive											Comments		
1	SMA2C-IT9-0-10-180906	9/6/18 0900	SE	1	X													
2	SMA2C-IT2-0-10-180906	0920	SE	1	Y													
3	SMA2C-IT5-0-10-180906	0930	SE	1	Y													
4	SMA2C-IT8-0-10-180906	0940	SE	1	Y													
5	SMA2C-IT4-0-10-180906	0950	SE	1	Y													
6	SMA2C-IT1-0-10-180906	1000	SE	1	Y													
7	SMA2C-IT7-0-10-180916	1010	SE	1	Y													
8			SE															
9			SE															
10			SE															
11			SE															
12			SE															
13			SE															
14			SE															

- 1 See project SAP/QAPP for analyte lists and test methods
- 2 Email sample confirmation report to labdata@anchorqea.com

Additional notes/comments:

Relinquished By: <u>Evan Malczuk</u>	Company: <u>Anchor QEA LLC.</u>	Received By: <u>Lauren Brandkamp</u>	Company: <u>EcoAnalysts</u>
Signature/Printed Name	Date/Time: <u>9/6/18 1047</u>	Signature/Printed Name	Date/Time: <u>9/6/18 1045</u>

Relinquished By:	Company:	Received By:	Company:
Signature/Printed Name	Date/Time	Signature/Printed Name	Date/Time





Date: 9/7/18  
 Laboratory: EcoAnalysts  
 Project Name: Port Gamble - OMMP LTM  
 Project Number: 180388-01.01  
 Project Contact: Cheronne Oreiro  
 Phone Number: 206.287.9130  
 Shipment Method: Delivery

Line	Field Sample ID	Collection Date/Time	Matrix	No. of Containers	Bioassay Archive	Sediment and Field QC										Comments			
						1	2	3	4	5	6	7	8	9	10				
1	SMA1B-IT2-0-10-180907	9/7/18 0845	SE	1	X														
2	SMA1B-IT1-0-10-180907	9/7/18 0855	SE	1	X														
3	SMA2C-IT3-0-10-180907	9/7/18 0935	SE	1	X														
4	SMA2C-IT6-0-10-180907	9/7/18 0940	SE	1	X														
5			SE																
6			SE																
7			SE																
8			SE																
9			SE																
10			SE																
11			SE																
12			SE																
13			SE																
14			SE																

1 See project SAP/QAPP for analyte lists and test methods  
 2 Email sample confirmation report to labdata@anchorqea.com

Additional notes/comments:  
 \_\_\_\_\_  
 \_\_\_\_\_

Relinquished By: <u>EVAN MALCOLM</u> <u>[Signature]</u>	Company: <u>Anchor QEA LLC.</u>	Received By: <u>Lauren Brandkamp</u> <u>[Signature]</u>	Company: <u>EcoAnalysts</u>
Signature/Printed Name	Date/Time: <u>9/7/18 1015</u>	Signature/Printed Name	Date/Time: <u>9/7/18 1015</u>

Relinquished By: _____	Company: _____	Received By: _____	Company: _____
Signature/Printed Name	Date/Time	Signature/Printed Name	Date/Time



Date: 9/17/18  
 Laboratory: EcoAnalysts  
 Project Name: Port Gamble - OMMP LTM  
 Project Number: 180388-01.01  
 Project Contact: Jason Cornetta  
 Phone Number: 206.971.2680  
 Shipment Method: Delivery

Line	Field Sample ID	Collection Date/Time	Matrix	No. of Containers	Larval abnormality/mortality	Sediment and Field QC												Comments							
						1	2	3	4	5	6	7	8	9	10	11	12								
1	SMA1-ST-0-10-Comp-180917	9/17/18 1205	SE	1	X																				
2	SMA1A-IT-0-10-Comp-180917	9/17/18 1510	SE	1	X																				
3	BW-15-0-10-180917	9/17/18 1546	SE	1	X																				
4			SE																						
5			SE																						
6			SE																						
7			SE																						
8			SE																						
9			SE																						
10			SE																						
11			SE																						
12			SE																						
13			SE																						
14			SE																						

- 1 See project SAP/QAPP for analyte lists and test methods
- 2 Email sample confirmation report to labdata@anchorqea.com

Additional notes/comments:

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Relinquished By: <u>Evan Malczyk</u> <u>[Signature]</u>	Company: <u>Anchor QEA LLC.</u>	Received By: <u>Julia Baum</u> <u>[Signature]</u>	Company: <u>Anchor QEA LLC</u>
Signature/Printed Name	Date/Time: <u>9/17/18 1628</u>	Signature/Printed Name	Date/Time: <u>9/17/18 1628</u>

EcoA

Relinquished By:	Company:	Received By:	Company:
Signature/Printed Name	Date/Time	Signature/Printed Name	Date/Time



