## APPENDIX A

## Sampling and Analysis Plan

## King County Department of Natural Resources and Parks Solid Waste Division

## Cedar Hills Regional LandFill

EnVIRONMENTAL CONTROL SYSTEMS MODIFICATIONS PROJECT
CONTRACT No. E00286E12
East Perched Zones Sampling and Analysis Plan
Final

Prepared by
Aspect Consulting, LLC

## [1] King County

Department of
Natural Resources and Parks
Solid Waste Division

| Waste | Resource | Waste |
| :---: | :---: | :---: |
| Prevention | Recovery | Disposal |

## Table of Contents

List of Figures ..... iii
List of Tables ..... iii
Acronyms and Abbreviations ..... iv
1.0 Introduction ..... 1
2.0 Field Sampling Plan ..... 2
2.1 Sampling Locations ..... 2
2.1.1 Groundwater Level Monitoring Locations ..... 2
2.1.2 Groundwater Sampling Network ..... 2
2.1.3 Stream Sampling ..... 3
2.1.4 Leachate Sampling ..... 3
2.1.5 Soil Gas Sampling ..... 3
2.2 Monitoring Parameters ..... 3
2.2.1 Groundwater Monitoring Parameters ..... 3
2.2.2 Surface Water Monitoring Parameters ..... 5
2.2.3 Leachate Monitoring Parameters ..... 5
2.2.4 Soil Gas Monitoring Parameters ..... 5
2.3 Schedule ..... 5
2.4 Groundwater Sampling Procedures ..... 6
2.4.1 Instrument Calibration. ..... 6
2.4.2 Well Inspection ..... 6
2.4.3 Groundwater Level Monitoring ..... 7
2.4.4 Well Redevelopment. ..... 7
2.4.5 Passive Sampler Deployment and Sample Collection ..... 8
2.4.6 Low-Flow Groundwater Sample Collection ..... 9
2.5 Stream Sampling Procedures and Protocols ..... 12
2.5.1 Staff Gage Reading ..... 12
2.5.2 Stream Sampling ..... 12
2.6 Soil Gas Sampling ..... 13
2.6.1 Ideal Sampling Conditions ..... 13
2.6.2 Soil Gas Sampling Procedures ..... 13
2.7 Leachate Sampling ..... 15
2.8 Sample Labeling ..... 15
2.9 Sample Custody ..... 17
2.10 Decontamination and Investigation-Derived Waste Management ..... 18
2.11 Field Documentation and Reporting ..... 18
3.0 Quality Assurance/Quality Control Plan ..... 20
3.1 Analytical Methods ..... 20
3.2 Quality Assurance. ..... 20
3.3 Field Quality Control ..... 21
3.3.1 Trip Blanks ..... 22
3.3.2 Field Blanks ..... 22
3.3.3 Field Duplicates ..... 22
3.3.4 Equipment Rinsate Blanks ..... 22
3.4 Laboratory QA/QC ..... 23
3.5 Data Management and Validation ..... 23
4.0 References ..... 25
ATTACHMENTS
A Field Documentation Records
B Boring Logs for RI Sample Locations

## LIST OF FIGURES

A-1 Cedar Hills Regional Landfill Property Features
A-2 RI/FS Water Level Monitoring Locations
A-3 RI/FS Water Sampling Locations
A-4 RI/FS Water Sampling Locations

## LIST OF TABLES

A-1 List of RI/FS Water Level Monitoring Locations
A-2 List of RI/FS Sampling Locations
A-3 RI/FS Well Screened Intervals
A-4 RI/FS Analytical Methods
A-5 RI/FS Analytical List
A-6 RI/FS Field QC Sampling Matrix

## ACRONYMS AND ABBREVIATIONS

| Aspect | Aspect Consulting, LLC |
| :---: | :---: |
| CHRLF | Cedar Hills Regional Landfill |
| Ecology | Washington State Department of Ecology |
| EPZ | East Perched Zones |
| FS | Feasibility Study |
| FSP | Field Sampling Plan |
| KCEL | King County Environmental Laboratory |
| KCSWD | King County Solid Waste Division |
| $\mu \mathrm{m}$ | micrometer |
| $\mu \mathrm{S} / \mathrm{cm}$ | microsiemens per centimeter |
| $\mathrm{mg} / \mathrm{L}$ | milligrams per liter |
| mL | milliliter |
| mV | millivolt |
| MNA | monitored natural attenuation |
| MTCA | Model Toxics Control Act |
| NTU | nephelometric turbidity unit |
| PDB | passive diffusion bag |
| PVC | polyvinyl chloride |
| QA | quality assurance |
| QAPP | Quality Assurance Project Plan |
| QC | quality control |
| RI | Remedial Investigation |
| RPP | rigid porous polyethylene |
| SAP | Sampling Analysis Plan |
| SKCDPH | Seattle-King County Department of Public Health |
| TDS | total dissolved solids |

TSS
USEPA
USGS
VOC
WAC
total suspended solids
U.S. Environmental Protection Agency
U.S. Geological Survey
volatile organic compound
Washington Administrative Code

### 1.0 INTRODUCTION

In response to compliance requirements in the 2004 Municipal Solid Waste Handling Permit, King County Solid Waste Division (KCSWD) initiated a project to investigate environmental control systems in the pre1986 unlined areas of the Cedar Hills Regional Landfill adjacent to the Main Hill (CHRLF; Figure A1). The report of which is entitled Cedar Hills Regional Landfill Site-Wide Groundwater Wells and Hydrogeologic Report (CH2M HILL and Udaloy, 2004), addressed hydrogeology to resolve questions regarding the East Perched Zones (EPZ) among other portions of the landfill.

A Hydrogeologic Report Addendum (Aspect, 2012a) was developed to fulfill the requirements of the 20092019 MSWH permit, which required an update to the Site-Wide Hydrogeologic Report (CH2M HILL and Udaloy, 2004a) including a more detailed investigation of the EPZ.

Data from the EPZ indicate impacts on groundwater, potentially from landfill gas interactions. As a result of the findings of the Site-Wide Hydrogeologic and Addendum projects, the Washington State Department of Ecology (Ecology) requested that KCSWD implement corrective action for perched saturated zones beneath the EPZ at the CHRLF. Ecology and KCSWD agreed to proceed under the Model Toxics Control Act (MTCA) with an Independent Remedial Action in accordance with Washington Administrative Code (WAC) 173-340-510 and WAC 173-340-515.

This Sampling and Analysis Plan (SAP) has been prepared as part of the CHRLF East Perched Zones Remedial Investigation (RI) /Feasibility Study (FS) Work Plan to meet the requirements of the corrective action issued by the Seattle-King County Department of Public Health (SKCDPH) and entered into by KCSWD under the Environmental Control Systems Modifications (ECSM) Project.

The purpose of this SAP is to ensure that field sample collection, handling, and laboratory analysis conducted during the EPZ RI/FS field activities will generate data to meet project-specific data quality objectives in accordance with the MTCA requirements (WAC 173-340-350). This SAP consists of two major components: a Field Sampling Plan (FSP) defining sampling locations and field protocols and a Quality Assurance Project Plan (QAPP) defining analytical methods, field and laboratory quality assurance (QA) protocols.

Field sampling activities not directly relating to the EPZ RI/FS are not discussed in this SAP. Instead, field sampling activities for these data gaps, which are identified in the CHRLF ECSM Project Data Gaps Report, shall be referenced within a separate report entitled the Field Sampling and Analysis Plan.

### 2.0 FIELD SAMPLING PLAN

### 2.1 Sampling Locations

This section describes the monitoring well network, sampling parameters and analytical methods, sampling protocols, and schedule for the RI/FS field activities.

### 2.1.1 Groundwater Level Monitoring Locations

Groundwater level monitoring will be conducted during the RI to assess the current hydrological model. The groundwater level monitoring network for the RI consists of monitoring wells, groundwater extraction wells, gas probes, and gas extraction wells within the general EPZ area. Table A-1 lists the locations where groundwater level monitoring is required, and Figure A-2 depicts the locations in relation to the EPZ project area.

### 2.1.2 Groundwater Sampling Network

The RI/FS groundwater monitoring network for the RI consists of EPZ perched and regional aquifer monitoring wells, groundwater extraction wells, and select piezometers. Table A-2 lists the locations where groundwater samples will be collected, and Figure A-3 depicts the locations in relation to the EPZ project area. Well construction logs are presented in the Site-Wide Hydrogeologic Report, Volume II (CH2M HILL and Udaloy, 2004) and the East Main Hill Perched Zones Technical Memorandum, Attachment A (Aspect, 2010). RI well and probe boring logs proposed for sampling are presented in Attachment B of this SAP. The geological cross-sections of the site are presented in the main body of the RI/FS Work Plan.

The field sampling will be performed for the purpose of (1) defining the nature and extent of groundwater impacts, (2) providing data to support the evaluation of monitored natural attenuation (MNA), and (3) to support the evaluation of the extraction well system shutdown.

### 2.1.2.1 Passive Diffusion Sampling

Passive diffusion samples will be collected from wells depicted on Figure A-3 on a trial basis to evaluate this technology for future sample collection as compared to conventional low-flow groundwater sampling (see explanation in the Work Plan). Passive diffusion bags (PDBs) are suitable for collection of samples for analysis of volatile organic compounds (VOCs), while rigid porous polyethylene (RPP) samplers are suitable for collection of samples for metals analysis. The deployment of these passive samplers will be 2 weeks prior to the sample collection date to allow for equilibration.

### 2.1.2.2 Low-Flow Groundwater Sampling

Low-flow groundwater samples will be collected from the sampling points summarized in Table A-2. Lowflow groundwater samples will only be collected from EW-9 through EW-13 if there is sufficient water present in these wells at the time of sampling. Historically, these wells have been dry or have not contained sufficient water. There will be some overlap between the low-flow sampling and the passive diffusion sampling because the intent of the conventional, low-flow sample collection is to allow comparison to the passive diffusion sample results.

### 2.1.3 Stream Sampling

Stream samples will be collected from SW-E1, at the downstream end of Stream 3 (Figure A-3). The intent of the stream sampling is to evaluate the concentrations of analytical parameters to evaluate if contaminant plumes within the EPZ are affecting stream conditions.

### 2.1.4 Leachate Sampling

Leachate will be collected from MH-17N (a conveyance manhole) and FS-3 (flow meter vault), if both locations have sufficient flow (Figure A-3). The intent of the leachate sampling is to evaluate the raw leachate from the unlined portion of the Main Hill for comparison to groundwater quality to evaluate if leachate is affecting groundwater conditions.

### 2.1.5 Soil Gas Sampling

Soil gas samples will be collected from existing gas probes, a piezometer, and existing monitoring wells located throughout the project area (Figure A-3). The intent of the soil gas sampling is to determine VOC concentrations in soil gas for evaluation of the vapor intrusion pathway. The data will also be used to evaluate the relative contribution of volatilization from impacted groundwater versus fugitive landfill gas.

### 2.2 Monitoring Parameters

Analytical methods for the laboratory-analyzed parameters for each sample medium are listed in Table A-4 and are discussed from a QA standpoint in Section 3. Table A-5 presents the specific parameters that are the target of this investigation. It is important to follow these tables because there are some variations in the analyte list for the different sample media. Prior to beginning the sampling and analysis program, Tables A4 and A-5 will be provided to the KCEL (laboratory for groundwater, surface water, and leachate samples), and Table A-5 will be provided to Fremont Analytical (laboratory selected for soil gas samples) so that the laboratories can document these project-specific analyte lists.

### 2.2.1 Groundwater Monitoring Parameters

The monitoring parameters required for groundwater samples per WAC 173-351-990, Criteria for Municipal Solid Waste Landfills, are as follows:

- Appendix I - Constituents for Detection Monitoring
- Appendix II - Groundwater Quality Parameters

These are collectively referred to as the RI project-specific analytes. Appendix I parameters include metals and select organic constituents from the VOCs list. Table A-5 contains a complete list of the RI projectspecific organic and inorganic analytes. Mercury is not listed as an Appendix I parameter, but has been added to Table A-5 as it is a routine parameter monitored at the Landfill.

Field parameters, geochemical indicator constituents, and leachate indicator parameters, as defined in WAC 173-351-990, Criteria for Municipal Solid Waste Landfills, Appendix II (Groundwater Quality Parameters), will also be monitored during the RI groundwater monitoring events. These parameters will be analyzed for the following purposes:

- To support source assessment (e.g., use of chloride as an indicator of leachate sources); and
- To evaluate MNA.

The Appendix II list omits several key field parameters and geochemical indicator constituents that are necessary for the evaluation of MNA, and those have been added to Table A-3 in italics. The MNA evaluation will be performed in general accordance with the U.S. Environmental Protection Agency's (USEPA's) Monitored Natural Attenuation of Inorganic Contaminants in Ground Water, Volume 1, Technical Basis for Assessment (2007) and USEPA's Technical Protocol for Evaluating Natural Attenuation of Chlorinated Solvents in Ground Water (1998).

MNA is an approach to remediation that relies on natural processes to attenuate the mass, toxicity, mobility, volume, or concentration of constituents of concern to achieve remediation objectives within an acceptable timeframe. MNA can take place through naturally occurring physical, chemical, and biological processes. However, MNA can be used only at sites with conditions that are favorable. Some MNA parameters are already included as geochemical indicator parameters from Appendix II of WAC 173-351, such as conductivity, pH, alkalinity, chloride, arsenic, dissolved manganese, sulfate, total organic carbon, and dissolved iron.

To evaluate whether the groundwater conditions within the CHRLF EPZ are suitable for MNA and to evaluate if MNA is occurring, a select number of additional field and geochemical parameters have been added to the RI project-specific analyte list including oxidation-reduction potential, dissolved oxygen, nitrate plus nitrite, dissolved ferrous iron, methane, ethene, and ethane. These parameters, coupled with those already included on the project-specific analyte list, will help to assess the following:

- Whether the aquifer is aerobic or anaerobic;
- The presence of anaerobic biological/microbial activity and whether this activity is dissolving metals from the aquifer matrix material;
- The stages of anaerobic degradation process; whether samples are collected from the same groundwater system;
- Whether chlorinated solvent reduction is occurring;
- Whether chlorinated solvents are undergoing biological transformation; and
- Whether organic carbon is available in the aquifer (excluding anthropogenic carbon) for reductive dechlorination.


### 2.2.2 Surface Water Monitoring Parameters

Surface water samples will be analyzed for the same WAC 173-351-990 Appendix I and II analytes as groundwater.

### 2.2.3 Leachate Monitoring Parameters

Leachate samples will be analyzed for select WAC 173-351-990 Appendix I and II analytes as listed in Table A-5. Leachate samples will not be analyzed for the additional MNA parameters.

### 2.2.4 Soil Gas Monitoring Parameters

Soil gas samples will be analyzed for the standard list of VOCs under EPA Method TO-15. Table A-5 presents the EPA Method TO-15 analyte list.

### 2.3 Schedule

The RI field activities will commence within 30 days of Ecology's approval of the RI/FS Work Plan. . Sampling will occur during sequential field activities. Each of the field activities is expected to take approximately 5 to 7 days of field work. Field work will be followed by period of approximately 45 days during which analytical results would be generated; the standard turn-around-time for the KCEL to provide analytical results is 30 calendar days in addition to time for data validation by KCSWD. The total time that the RI data collection period will take is approximately 70 days.

The following table summarizes the main RI activities and the locations where those activities will take place:

| RI Activity | Location |
| :--- | :--- |
| Groundwater level monitoring; stream <br> gaging | Table A-1 locations; SG-4 and SG-5 |
| Deployment of PDB and RPP samplers | MW-102 through MW-104; EW-18 and <br> EW-20; MW-62, MW-30A, MW-47, and <br> EB-6 |
| Collection of PDB and RPP samplers; <br> collection of low-flow groundwater <br> samples | Table A-2 locations |
| Stream sampling and gaging | SW-E1; SG-4 and SG-5 |
| Leachate sampling | MH-17N and FS-3 |
| Soil vapor sampling | EB-6; MW-102 and MW-104; GP-ATC-- <br> 5 and GP-ATC-7; GP-8, GP-56, GP-58, <br> GP-60, GP-62; GP-15 through 20 |

The RI activities will be generally sequenced in the logical order of the intended tasks. For example, groundwater and surface water level measurements will be collected prior to installation of the PDB and RPP samplers in order to obtain the water levels before disturbing the wells during passive sampler installation. The passive diffusion samplers must be allowed to equilibrate for a minimum of 2 weeks. The sample collection activities will begin after the 2-week equilibration period has lapsed and will include the
collection of the passive diffusion samplers, the collection of comparison samples using conventional lowflow sample methodology, the stream sampling and gaging, and the leachate sampling. The soil vapor sampling will be conducted approximately 2 weeks after the groundwater sampling mobilization to allow gas-collection-specific wellhead configurations to be fitted to MW-102, MW-104, and EB-6 and allow enough time for gas equilibration to occur within the closed-off well. However, the exact timing of the soil gas sampling will be weather dependent. The following sections describe the protocols necessary for completing the RI field work.

### 2.4 Groundwater Sampling Procedures

This section summarizes the procedures and protocols that will be followed for the field data collection phase of the RI. Sampling protocols are documented in the Environmental Monitoring Sampling and Analysis Plan for Cedar Hills Regional Landfill (Aspect, 2013; henceforth referred to as the CHRLF Environmental Monitoring SAP). Specific methods to be used in the RI are presented below for the convenience of field crews. Work will be performed under the project-specific Health and Safety Plan.

The field sampling procedures and sample handling protocols for samples collected during the RI will be in accordance with the following:

- CHRLF Environmental Monitoring SAP (Aspect, 2013);
- USEPA low-flow sampling procedures (Puls and Barcelona, 1996, revised 2010);
- U.S. Geological Survey (USGS) passive diffusion sampling protocols (2001); and
- Aspect sampling protocols.

Pertinent portions of these reference documents are summarized in this section of the SAP. Field sampling data will be documented on field report forms (Attachment A) as described in Section 2.11, Field Documentation and Reporting.

### 2.4.1 Instrument Calibration

Prior to initiating any groundwater or surface water sampling, the field instruments for measuring water quality parameters ( pH , conductivity, dissolved oxygen, oxygen-reduction potential, and turbidity) will be checked and calibrated to make sure that all parts are present, working, and clean and that the batteries are fully charged. The instruments will be calibrated per the manufacturers' specifications. Calibration activities will be documented in the field notebook. Instrument calibration should take place each day prior to sampling.

### 2.4.2 Well Inspection

Monitoring wells are protected by secure stick-up monuments that are locked. Expansion well caps are installed in the 2-inch-diameter well casings and provide a watertight seal. Each monitoring well included in the RI field program will be inspected during the first mobilization. Field personnel will take note of the condition of the monument, well cap, well casing, and pump (if present); identify required maintenance
activities; and report the information to the KCSWD lead engineer in accordance with the CHRLF Environmental Monitoring SAP. No groundwater samples will be collected and no passive diffusion samplers will be deployed if the security or integrity of a well or pump equipment is severely compromised. During sample collection, the condition and response of the well to sample purging will be noted for evaluation during the RI and corrective action (if necessary) will be implemented before future sampling events.

### 2.4.3 Groundwater Level Monitoring

Depth-to-groundwater will be measured in the wells and probes listed in Table A-1 and depicted on Figure A-2 using an electric well sounder, graduated to 0.01 foot, according to the following procedure. In some of the gas probes (in particular GP-17, -18, and -19), dedicated tubing inside the probe will need to be temporarily removed in order to collect the water level measurement. Any tubing that is removed will be reinstalled after the reading is collected.

1. Lower the probe through the top of the polyvinyl chloride (PVC) well casing or probe opening until the level alarm sounds.
2. Read the depth-to-water measurement at the top of the PVC casing at the designated mark. If there is no marked reading location, then the depth-to-water measurement will be collected from the north side of the casing.
3. Raise the probe line until the alarm goes silent and lower the line again to collect duplicate and triplicate static water level readings. The depth to the bottom of the monitoring well will also be measured to evaluate siltation of the monitoring well. This measurement can be performed with the level alarm turned off after the static water level has been collected.
4. Record the static water level, date, and time of collection on the field report forms.
5. Decontaminate the water level indicator between wells.

If access to the gas extraction wells is inadequate to obtain a groundwater level reading due to the wellhead configuration, then that well will be skipped and its condition noted on the field report forms.

### 2.4.4 Well Redevelopment

Three of the wells that have been included in the RI sampling program (EB-1, EB-2, and MW-50) have not been sampled in the past. These three wells will need to be redeveloped prior to sampling. Each of these monitoring wells will be developed to remove fine-grained material from inside the well casing and filter pack that may have accumulated overtime and to improve hydraulic communication between the well screen and the surrounding water-bearing formation. The field representative will document well development activities in the field documentation for that well. Well development will be performed by means of a surge block and a 12 -volt submersible pump. During development, the surge block will be surged along the entire length of the well screen. Each well will be developed until visual turbidity is reduced to minimal levels or until a minimum of three well casing volumes of water have been removed.

During RI groundwater sample collection from the EW series wells, the condition and response of the well to sample purging will be evaluated. If during sample collection significant drawdown is encountered and
the determinations in the RI include additional sampling of EW series wells, then rehabilitation will be considered.

### 2.4.5 Passive Sampler Deployment and Sample Collection

PDB and RPP samplers and noncorrosive stainless steel hanging assemblies will be obtained from ALS Environmental. The numbers and types of samplers are presented in the table below and depicted on Figure A-3. VOC samples will be collected using PDBs for monitoring wells MW-102, MW-103, and MW104; detection monitoring wells MW-30A, MW-62, MW-47, and EB-6; and groundwater extraction wells EW-18 and EW-20. Samples collected for metals analysis using an RPP sampler will be collected from detection monitoring wells MW-30A, MW-62, MW-47, and EB-6 and extraction wells EW-20 and EW-25. Therefore, MW-30A, MW-62, MW-47, EB-6, EW-18, and EW-20 will have both a PDB and an RPP sampler installed.

| Wells | Installation of <br> PDB <br> Samplers | Installation of <br> RPP Samplers | Collection of <br> Low-Flow <br> Samples | Comments |
| :---: | :---: | :---: | :---: | :---: |
| MW-102, MW- <br> 103, and MW-104 | 3 | No | No |  |
| EW-20 and EW- <br> 25 | 2 | 2 | No |  |
| MW-62, MW-30A, <br> MW-47, and EB-6 | 4 | 4 | No |  |
| Table A-2 Wells | No | No | Yes | Retrieve PDB and <br> RPP samplers; <br> collect low-flow <br> samples; SW-E1 <br> sampled |
| Total samples | $\mathbf{9}$ | $\mathbf{6}$ | $\mathbf{2 8}$ |  |

Passive sampler deployment

1. Record the static water level in the well at the time of deployment. Collect this reading prior to removing any dedicated pumps (if applicable).
2. Remove dedicated sampling pumps in small diameter monitoring wells that cannot also accommodate the passive sampler. Dedicated pumps that are removed from wells will be kept clean and segregated while out of the wells to prevent contamination. Upon completion of sampling, the removed sample pumps will be decontaminated and reinstalled in the same well from which they were removed.
3. Install the passive samplers in the extraction wells with great care. Pump wiring and electrical submersible pumps are present within the extraction wells. The wiring will have to be adjusted to set the PDB and RPP samplers. If the pump is set too high, the PDB and RPP samplers will not be deployed.
4. Measure the depth to the bottom of the well and compare the measured depth with well construction records to see if sediment has accumulated in the bottom of the well and if the elevation of the passive samplers requires adjusting.
5. Use the plug at the bottom of the sampler, a funnel, and deionized water to fill the passive sampler with laboratory provided deionized water until the water rises and stands at least halfway into the funnel. Remove the funnel and replace the plug. The samplers will then be ready for installation. If the passive samplers were delivered prefilled, they can be installed directly into the wells.
6. The mid-point of the PDB will be placed at the mid-point of the saturated well screen interval using the hanging assemblies. The screen interval depths for the wells receiving passive samplers are provided in Table A-3. The mid-point of the saturated well screen will be determined in the field based on the current water levels. The approximate elevation will be provided to ALS Environmental in order for the samplers to be strung from the hanging assemblies at the appropriate locations. The hanging assembly will hook to the well cap or wellhead assembly.
7. Lower the passive sampler and hanging assembly into the well until the desired depth is reached and attach the hanging assembly to the well cap or wellhead assembly.
8. Allow the passive samplers to equilibrate, undisturbed in the wells for 14 days.

Removal of passive samplers and sample collection
9. Record the static water level in the well.
10. Carefully lift the passive sampler out of the well, minimizing the amount of agitation to the sample. Be careful not to tear or damage the sample bag. Make sure the PDB does not come into contact with potentially contaminated surfaces once it is removed from the well.
11. Detach the sample bag from the hanging assembly and inspect the bag for bio-film buildup that could affect sample results. Note the presence of bio-film on the field report form.
12. Collect the sample by cutting off the corner of the bag and pouring it directly into laboratorysupplied pre-cleaned containers. Samples from the PDBs for VOC analysis are poured into $40-$ milliliter (mL) vials. Once filled and capped, the vials should have no headspace (i.e., no visible air bubbles). Samples from the RPP bags are for metals analysis are poured into preserved poly-bottles.

### 2.4.6 Low-Flow Groundwater Sample Collection

Conventional, low-flow groundwater samples will be collected from the monitoring wells listed in Table A-2 in accordance with the procedures described below, USEPA's low-flow groundwater sampling procedures (Puls and Barcelona, 1996), and the CHRLF Environmental Monitoring SAP (Aspect, 2013). Low-flow sampling will occur after the passive samplers have been removed.

1. Record the static water level in the well. Keep the water level tape suspended in the well so that static water level measurements can be collected during the purge process.
2. Use a QED portable bladder pump with disposable tubing and bladders or the dedicated pump deployed in the well to collect low-flow groundwater samples. Compressed nitrogen will be used for the bladder compression gas, per the CHRLF Environmental Monitoring SAP (Aspect, 2013). New tubing will be used for each well sampled by means of the portable bladder pump.
a. Install any dedicated pumps that were removed for the passive sampler deployment.
b. Install portable or dedicated pumps slowly so that agitation of the well bottom sediment does not occur.
3. Place the pump intake for the portable pump at the center of the saturated section of well screen. Dedicated pumps will be reinstalled at their designated elevations.
4. Purge each well at a low-flow rate 0.1 to 0.5 liter per minute (Puls and Barcelona, 1996). The flow rate should be adjusted to ensure that the water level drawdown during purging is not greater than 4 inches (Puls and Barcelona, 1996).
5. Monitor field parameters (temperature, pH , specific conductivity, dissolved oxygen, oxidationreduction potential, and turbidity) using a YSI meter, flow-through cell, and Hach 2100Q turbidimeter during well purging. These field parameters in addition to the static water level will be recorded at 3 - to 5 -minute intervals throughout well purge until they stabilize. Turbidity is not listed in the ASTM method and will be measured prior to the collection of the groundwater sample. Water temperature will also be monitored but can be affected by air temperature and solar radiation. During well evacuation, the flow rate will be monitored and the total volume of water evacuated will be calculated. Groundwater field parameters will be measured and confirmed as stable before the well is sampled.

Stabilization is defined as three successive readings with the following variations in the parameter values:

- Conductivity $\pm 3 \%$ microsiemens per centimeter ( $\mu \mathrm{S} / \mathrm{cm}$, specific conductance)
- Dissolved oxygen $\pm 10 \%$ milligrams per liter ( $\mathrm{mg} / \mathrm{L}$ ) (or $0.5 \mathrm{mg} / \mathrm{L}$ if readings are less than $1 \mathrm{mg} / \mathrm{L}$ )
- $\mathrm{pH} \pm 0.1$
- Oxidation-reduction potential $\pm 10$ millivolts (mV)
- Turbidity $\pm 10 \%$ nephelometric turbidity units (NTU)
- Turbidity will also be measured directly from the pump discharge point (not from the flow-through cell) immediately the groundwater sample is collected and the reading is recorded.

6. Purge slow-recovery wells and dry and collect the sample within 24 hours after the well has sufficiently recovered if dewatering is the likely outcome of the well purge. At the end of the day, take a second depth-to-water measurement and record it on the field report form.
a. Measure the static water level the following day to confirm the well has sufficiently recovered. Till a cup with enough water to perform field parameter measurements one time, and immediately begin filling the laboratory bottles in the following order:
7. Volatile organics analysis (VOA) bottles (VOCs, methane, ethane, ethene)
8. TOC
9. Chloride and sulfate
10. Bicarbonate and alkalinity
11. Ammonia, nitrate, nitrate + nitrite
12. Metals, total and dissolved
13. Dissolved ferrous iron
14. Total dissolved solids (TDS)
15. Total suspended solids (TSS)
b. Collect only the minimum volume the laboratory requires for each of the above analytes if the well will not produce an adequate volume of water to fill the normal bottle set. Below are the normal and minimum volumes the laboratory typically asks for:

| ANALYTE | NORMAL VOLUME | MINIMUM VOLUME |
| :---: | :---: | :---: |
| VOCs | $4,40 \mathrm{~mL}$ | $1,40 \mathrm{~mL}$ |
| Metals (dissolved) | 500 mL | 150 mL |
| Metals (total) | 500 mL | 200 mL |
| Ammonia/nitrate/nitrite | 250 mL | 50 mL |
| TSS | $1,000 \mathrm{~mL}$ | $1,000 \mathrm{~mL}$ |
| TDS | 500 mL | 150 mL |
| TOC | 125 mL | $1,40 \mathrm{~mL}$ |
| Chloride/sulfate | 125 mL | 50 mL |
| Bicarbonate/Alkalinity | 500 mL | 120 mL |
| Dissolved | $2,40 \mathrm{~mL}$ | $1,40 \mathrm{~mL}$ |
| methanelethene/ethane | 500 mL | 50 mL |
| Dissolved ferrous iron |  |  |

7. Collect groundwater samples at the same low-flow rate after parameter stabilization is reached by directly filling the laboratory-supplied sample containers from the dedicated pump tubing. When the sample bottle is full, replace the lid and fill the next bottle, if required. For dissolved metals samples, an inline 0.1-micrometer ( $\mu \mathrm{m}$ ) filter will be attached to the purge tubing and the dissolved metals bottle will be filled with filtered groundwater. Add an additional label to the dissolved metal sample container that reads, "Field Filtered," indicating to the laboratory which of the two identical sample containers is to be tested for dissolved metals. Additional notes on groundwater sample collection are presented in Table D. 4 of the CHRLF Environmental Monitoring SAP (Aspect, 2013).
8. Collect groundwater quality control (QC) samples (e.g., field duplicates, rinsate blanks, and trip blanks) as per Section 3 and Table A-6 of this SAP.
9. Replace the well cap and monument cap securely. Any damaged or defective well caps or monuments will be noted and reported to KCSWD.

### 2.5 Stream Sampling Procedures and Protocols

### 2.5.1 Staff Gage Reading

Surface water elevations will be monitored by reading the stage height on the staff gages installed at the upstream (SG-4) and downstream (SG-5) ends of Stream 3 (Figure A-2). The measurements in Stream 3 will be taken at the same time as the initial groundwater level measurements so that the stream levels can be compared to the EPZ monitoring well water levels.

Surface water stage readings are recorded at the level of the water on the staff gage to the nearest 0.01 foot. If Stream 3 is dry or the water level is below the staff gage, the water level will be collected from the adjacent piezometer using a water level meter, referencing the survey point at the top of the piezometer. The stage reading is converted to a surface water elevation by adding the stage reading to the surveyed 0.00 -foot mark on the staff gage.

### 2.5.2 Stream Sampling

Stream samples will be collected from SW-E1 (Figure A-3) and handled in accordance with the procedures described below:

1. Measure pH , temperature, conductivity, dissolved oxygen, and turbidity directly within the surface water flow.
2. Collect a water sample of Stream 3 at SW-E1 from approximately mid-depth at mid-channel either using a peristaltic pump and new, dedicated tubing (polyethylene tubing with a short length of silicon tubing through the pump head) or using a laboratory-provided clean, unpreserved vessel to transfer the water into the appropriate sample containers.
a. Minimize the disturbance of sediments in the areas surrounding the surface water monitoring station during sample collection. Dam or block the downstream flow to pool the water so that a clean container or peristaltic pump can be used to fill the sample
containers provided by the laboratory. Materials used to dam or block the flow should be decontaminated prior to use by means of the decontamination procedures specified in this SAP. Damming the stream is only necessary when the flow is low.
b. Wade into the stream, face upstream, and collect the sample from the upstream water to avoid disturbing the sediments. Allow any sediment to pass the collection point prior to collecting a sample. Avoid allowing sediment, algae, weeds, etc., from entering the sample container. Rinse the plastic container twice with the sample water prior to collection of the sample.
3. Pump or pour stream water directly into laboratory-supplied sample containers. Avoid overfilling preserved containers and as this will result in loss of the preservative. Replace the lid securely when the sample bottle is full and fill the next bottle, if required. Filter dissolved metals samples using an inline $0.1-\mu \mathrm{m}$ filter attached to the peristaltic pump tubing. Fill the dissolved metals bottle directly with filtered surface water. Add an additional label to the dissolved metal sample container that reads, "Field Filtered," indicating to the laboratory which of the two identical sample containers is to be tested for dissolved metals.
4. Label and transport stream samples to the laboratory under chain-of-custody per Sections 2.8, 2.9, and 2.11.

### 2.6 Soil Gas Sampling

Soil gas samples will be collected in 6-liter Summa canisters for laboratory analysis of VOCs. The samples will be analyzed according to EPA method TO-15.

### 2.6.1 Ideal Sampling Conditions

The ideal condition for sampling is low and decreasing barometric pressure. Sampling will ideally occur following at least 12 hours of falling barometric pressure, with a drop from peak of at least 0.25 inches mercury.

### 2.6.2 Soil Gas Sampling Procedures

Soil gas sampling will be attempted at the 16 locations indicated on Figure A-4. Two screen intervals will be sampled at locations GP-15 through GP-20. Therefore, up to 22 Summa canister samples will be collected for laboratory analysis. However, GP-ATC-7 could not be located during previous field efforts (Aspect, 2007). Another attempt will be made to locate GP-ATC-7, including the use of a metal detector. If it is not found, that location will not be included in the soil gas sampling program. (Nearby well MW-103 cannot be used for soil gas sampling because its screen is fully submerged.)

The gas probe wellheads are equipped with a suitable port for gas sampling. MW-102, MW-104, and EB-6 will be prepared for gas sampling by removing the downhole pumps (assuming pumps are installed) and
replacing the caps with caps that have a suitable port for LFG sampling. This will be done immediately after groundwater samples have been collected from these wells.

The manifold that is used for soil gas sampling is shown below. The Teflon tube from the wellhead assembly connects to the manifold's left side. The manifold branches to a purge tube that connects to the GEM-2000 and a sample tube that connects to the Summa canister's flow controller.


The following procedure will be used to collect soil gas samples for VOC analysis:

1. Calculate the volume of air in the probe/well/piezometer casing.
2. Calibrate the GEM-2000 multi-gas meter.
3. Check the vacuum in the 6-liter Summa canister. Vacuum should ideally be between 28 and 29 inches, and not less than 25 inches. If vacuum is less than 25 inches, the canister should be replaced with a canister meeting this specification. Remove the brass cap from the canister valve and connect the canister to the 1-hour flow controller. Then connect the sampling manifold (shown above) to the flow controller. Proceed to Step 5 for sampling gas probes with relatively small casing volumes.
4. Pre-purge the monitoring wells, piezometer EB-6, and the gas probes that have relatively large casing volumes by connecting in series a low-flow vacuum pump, a rotameter, and a GEM-2000 to the wellhead tubing. The vacuum pump and rotameter will be sized for a purge rate of one casing volume every 5 to 10 minutes. Start the pump and purge until two casing volumes have been removed. During purging, record methane, carbon dioxide, and oxygen concentrations, LEL, and barometric pressure measured by the GEM-2000 at minimum 1minute intervals. Stop the pump after two casing volumes have been removed, disconnect the wellhead tubing from the pump, and immediately connect the tubing to the sampling manifold.
5. Connect the GEM-2000 and the wellhead tubing to the sampling manifold shown in the diagram above. Open the manifold valve and begin purging with the GEM-2000. The purge rate will be regulated by the flow restrictor in the manifold to approximately $170 \mathrm{ml} / \mathrm{min}$.

During purging, record methane, carbon dioxide, and oxygen concentrations, LEL, and barometric pressure at minimum 20 -second intervals until parameters have stabilized or until a total of three casing volumes have been purged (i.e., one additional casing volume for locations where pre-purging was conducted in Step 4 above, since two casing volumes were previously removed). Stabilization is defined as three readings over a 1-minute period that are within 10 percent of one another.
6. Turn off the GEM-2000 and close the manifold valve.
7. Open the valve on the Summa canister to start sample collection. The sampling rate will be regulated by the flow controller at approximately $100 \mathrm{ml} / \mathrm{min}$.
8. Stop sample collection and close the canister valve when the Summa canister vacuum reaches 5 inches mercury (after approximately one hour).
9. Record the final canister vacuum on the chain of custody and on the Summa canister tag.
10. Disconnect the canister from the manifold and replace the brass cap on the canister valve.

### 2.7 Leachate Sampling

Leachate samples will be collected from MH-17N and FS-3 to characterize raw leachate in the East Main Hill (Figure A-3). Sample collection at each of these stations will be dependent on flow observations at the time of sampling. For manhole $\mathrm{MH}-17 \mathrm{~N}$, the sample location is the leachate line from the north Main Hill that enters the southwest corner of the manhole. At manhole FS-3, the sample location is the influent pipe discharge. The following steps will be taken for the leachate sampling:

1. Measure the static water level in $\mathrm{MH}-17 \mathrm{~N}$ and $\mathrm{FS}-3$ using a water level tape.
2. Monitor the pH and conductivity (measured in specific conductance units) in the manhole and flow meter vault and record the reading(s) on the field report form. Measurement may also be made at the surface prior to sampling (Step 3, below), if downhole measurements are not feasible.
3. Using a peristaltic pump and dedicated tubing or a grab sampler (whichever is most appropriate for the volume of flow and the access point at each leachate sampling location), directly fill the laboratory-supplied sample containers from the dedicated pump tubing. If leachate depths exceed the lift of the peristaltic pump (about 22 ft ), collect the sample using a dedicated Teflon bailer and string. When the sample bottle is full, replace the lid and fill the next bottle, if required. No field filtering of the sample is required because dissolved metals are not a targeted analyte.

### 2.8 Sample Labeling

A label will be placed on each sample container with the following information: name of client (KCSWD), location, date, time, database sample ID, and sampler's initials. PDB, RPP, and low-flow samples collected from the same well will each be considered unique samples and, as such, require unique sample IDs. Sample IDs will be in accordance with KCSWD standards, which consist of an 11- to 13-digit labeling convention:

Examples: W62-151001 = Low-flow groundwater sample from MW-62 on October 1, 2015; GG62151001 = Soil gas sample from GP-62 on October 1, 2015; LE17N151001 = Leachate sample from MH-17N on October 1, 2015.

Digits 1-4: Consist of the medium type and sampling location.

- Medium type is typically the first digit and includes groundwater (W), surface water (S), leachate (L), or gas (G).
- The sampling location is typically a three-digit number (digits 2-4); however, shallow or deep sampling locations are distinguished by " S " or " D ," respectively. If the sampling location is only a two-digit number, the symbol "-" is used for the third digit.

For example, monitoring wells will use just their two- or three-digit number (omitting the MW), whereas extraction wells and piezometers will include an EW or WB before their two-digit number to differentiate the different types of sampling points.

Examples: WB6-151001 = Low-flow groundwater sample from EB-6 on October 1, 2015; EW6151001 = Low-flow groundwater from EW-6 collected on October 1, 2015.

- "VTRP" is used as the media type and sampling location designation for VOC trip blanks.

Example: VTRP151001 = Trip blank collected October 1, 2015
Digits 5-10: Consist of the sampling date.

- Digits 5-6 are the two-digit year.
- Digits 7-8 are the two-digit month.
- Digits 9-10 are the two-digit day.

Digits 11-13: Indicate field QC or other unique samples.

- The symbol "-" is used if the sample is not a field duplicate or a trip blank.
- " D " is used if the sample is a field duplicate.
- " $E$ " is used if the sample is an equipment/rinsate blank.
- " $F$ " is used if the sample is a field blank.
- "PDB" is used for samples collected using PDB samplers.
- "RPP" is used for samples collected using RPP samplers.

Examples: W62-151001RPP = RPP groundwater sample from MW-62 on October 1, 2015;
EW20151001PDB = PDB groundwater sample from EW-20 on October 1, 2015.

Field Filtering: Sample bottles for dissolved metals analysis where the sample was field filtered will need to be identified. "Field Filtered" will be written on all bottles that received field filtration in preparation for dissolved metals analysis.

### 2.9 Sample Custody

Upon collection and labeling, groundwater, surface water, and leachate samples will be placed upright in a cooler. Ice or blue ice will be placed in each cooler to cool samples down to meet sample preservation temperature requirements. Inert cushioning material will be placed in the remaining space of the cooler, as needed, to limit movement and breakage of the sample containers. If the sample coolers are being shipped, not hand carried to the laboratory, the chain-of-custody form will be placed in a waterproof bag taped to the inside lid of the cooler for shipment. Soil gas samples will be shipped back to the laboratory in the same shipping container they were received in. Soil gas samples do not require temperature preservation.

After collection, samples will be maintained in the sampling personnel's custody until formally transferred to the analytical laboratory. For the purposes of this work, custody of the samples will be defined as one of the following:

- In plain view of the field representatives;
- Inside a cooler that is in plain view of the field representative; or
- Inside any locked space such as a cooler, locker, car, or truck to which the field representative has the only immediately available key(s).

A chain-of-custody record provided by the laboratory will be initiated at the time of sampling for all samples collected. The record will be signed by the field representative and others who subsequently take custody of the sample. Couriers or other professional shipping representatives are not required to sign the chain-ofcustody form; however, shipping receipts will be collected and maintained as a part of custody documentation in project files. This procedure tracks the sample status from the point of sample collection to completion of the laboratory analyses. Most importantly, responsibility for sample integrity is placed on a single individual at all times. A copy of the chain-of-custody form with appropriate signatures will be kept by the project manager. This procedure becomes particularly important if the sampling protocol and analytical determination are ever challenged in litigation.

Proper chain-of-custody procedures play a crucial role in enforcement cases. The following are some basic guidelines of legal significance:

- As few people as possible should handle the sample.
- Chain-of-custody records should accompany the sample. The chain-of-custody record should contain the sample number, the date and time the sample was collected, the sample location, analysis required, the name of the person(s) who collected the sample, and the time of sample collection.
- The field team is responsible for the care and custody of the samples collected until custody has been relieved and documented by laboratory personnel. A four-part chain-of-custody document is used. The laboratory keeps the original chain-of-custody documents.

Upon sample receipt, the laboratory will fill out a cooler receipt form to document the sample delivery conditions. A designated sample custodian will accept custody of the shipped samples and will verify that the chain-of-custody form matches the samples received. The laboratory will notify the project manager, as soon as possible, of any issues noted with the sample shipment or custody.

### 2.10 Decontamination and Investigation-Derived Waste Management

Nondisposable sampling equipment (e.g., portable bladder pumps) will be decontaminated between each well. The decontamination sequence consists of a scrub with a nonphosphate (Alconox or Liquinox) solution, followed by a tap water (potable) rinse, and finished with thorough spraying with deionized or distilled water.

Investigation-derived waste water generated during equipment decontamination and monitoring well sampling will be disposed of in the on-site leachate treatment lagoons. Disposable equipment and used personal protective equipment will be placed in plastic trash bags and disposed of at CHRLF.

### 2.11 Field Documentation and Reporting

Inspection and monitoring results will be documented in the field report forms (Attachment A) and in field notebooks. Adequate records will be maintained for each sample collected. The field representative will document pertinent observations and events specific to each activity and specific to each sample collected on field forms (e.g., groundwater sampling form) and in a field notebook, and, when warranted, provide photographic documentation of specific sampling efforts. Field notes will include the following:

- Date, time, weather conditions, project location, and sampler's name
- Sample location, sample type, and sample number
- Description of the field activity
- Sample descriptions and sampling method
- Size, type, and quantity of sample containers
- Field equipment used
- Field parameters

Pertinent observations of the sample condition that are worthy of noting in the field documentation include the following:

- Sample color
- Sedimentation or turbidity
- Oil or sheen
- Separate phase liquids
- Odor
- Effervescence
- Beginning canister vacuum (soil gas samples only)
- Ending canister vacuum (soil gas samples only)

Other information to be included in the field notebook includes the following:

- Reason for sampling
- Problems encountered due to unusual conditions
- Communications with KCSWD, laboratory, or field staff


### 3.0 QUALITY ASSURANCE/QUALITY CONTROL PLAN

A sound quality QA/QC program is essential to obtaining high-quality and well-documented analytical data. The objectives of the QA program include the following:

- Collect, process, and analyze with consistent and appropriate techniques;
- Minimize the number of lost, damaged, or uncollected samples;
- Maintain and document the integrity of the data from sample collection to entry into the data record;
- Obtain comparable data; and
- Obtain reproducible results.


### 3.1 Analytical Methods

As discussed in Section 2.2, "Monitoring Parameters," the RI project-specific analytes for the groundwater and surface water samples are the WAC 173-351-990 Appendix I and II constituents and the additional project-specific MNA parameters methane, nitrate/nitrite, ethane, and ethene. Table A-4 lists the laboratory analytical methods for groundwater, surface water, leachate, and soil gas analyses to be performed during the RI/FS, along with the sample containers, preservation, and analytical holding times for each analysis. It is important to follow this table because there are some variations in the analyte list for the different sample media. This QAPP, including Tables A-4 and A-5, will be provided to the laboratory for its review prior to sample analysis so that the laboratory personnel are aware of the desired analytical methods and projectspecific analyte lists ahead of time.

### 3.2 Quality Assurance

QA is a system of documented checks that validate the reliability of the data set. QA procedures are used to verify that field and laboratory analytical data are within acceptable limits and that the data are accurate, precise, representative, comparable, and complete.

Sampling activities will be traceable to the person collecting the sample and to the specific piece of sampling equipment used to collect that sample. Records (field records, calibration logs, and chain-ofcustody forms) shall be kept so that they are traceable. The following procedures will be used:

- Proper calibration of sampling and field equipment;
- Use of field records to record field measurements and other pertinent information necessary to document what was done;
- Review of field records by the lead engineer to ensure that the procedures are being followed;
- Strict adherence to established field sampling protocols;
- Proper preservation of samples immediately upon collection in the field ( pH to be checked by laboratory);
- Recording of canister vacuum at time of canister closure in field and a subsequent recheck in the laboratory to ensure that no leaks occurred during transport;
- Minimal sample handling after collection; and
- Adherence to chain-of-custody procedures.

Field documents (notebooks, reports, chain-of-custody forms) will be reviewed by the lead scientist or geologist to ensure that the documents are accurate, precise, representative, comparable, and complete.

The laboratory will follow its own standard QA/QC procedures to ensure that the data it provides are accurate, precise, representative, comparable, and complete. Laboratory QA/QC procedures are discussed in the Section 3.4.

### 3.3 Field Quality Control

QC is the system of technical actions that measure the attributes and performance of field activities to control the quality of the data that are collected. The aim is to provide quality data that are adequate, dependable, and economical.

QA/QC samples collected in the field include VOA trip blanks, field blanks, field duplicates, and equipment/rinsate blanks. Table A-6 provides a summary of the type of QC samples and the frequency of collection. QA/QC samples make up roughly 10 percent of all samples collected (with some variation based on the type of QA/QC sample collected, as noted below). The following subsections describe the various field QC samples to be collected and their frequency of collection.

Trip blanks, field blanks, field duplicates, and equipment/rinsate blanks will be collected and submitted to the analytical laboratory to provide a method for assessing the quality of data collected from the monitoring program. The blanks and duplicates will be submitted to the laboratory as blinds, with sample labels that are indistinguishable from the primary samples. Specific QA/QC sample descriptions and collection frequencies are discussed in the following subsections.

### 3.3.1 Trip Blanks

Trip blank samples will be used to monitor possible VOC cross-contamination during sample handling, transport, and storage. Trip blank samples are prepared and supplied by the laboratory using organic-free reagent-grade water in a VOC vial prior to the collection of field samples. The trip blank sample vials are placed with and accompany the VOC samples through the entire transport process. One trip blank will be collected for each sample cooler that contains a VOC sample container. VOA trip blanks will be given a sample ID, in accordance with the labeling protocols, for identification of the primary samples it accompanies. The sample ID will be recorded in the field records and on the chain-of-custody form. Trip blanks will be transported to the laboratory in the same cooler as the VOC samples. VOA trip blanks will be analyzed for the same VOC parameters as the primary samples for the respective media.

In case a target compound is present in a trip blank, results for samples shipped with this trip blank will be evaluated and data qualified accordingly if determined that the results are affected.

### 3.3.2 Field Blanks

Field blanks are analyzed to check for contamination resulting from the sample collection, handling, and transport and from the bottles themselves. At the end of the sampling event, field blanks are prepared in the field by directly filling the sample bottles with deionized water provided by the laboratory and handling the field blank samples in accordance with the primary samples. Field blanks are analyzed for the same sample parameters as the groundwater samples. One field blank per 10 samples will be collected and analyzed. Each field blank will be given a sample ID in accordance with the labeling protocols in order to identify which primary samples it accompanies. The sample ID will be recorded in the field records and on the chain-of-custody form. Field blanks will be analyzed for the same parameters as the primary samples for the respective media.

### 3.3.3 Field Duplicates

During the RI field program, field duplicate samples will be collected at a frequency of 10 percent of the primary samples and will be analyzed for the monitoring well RI project-specific analytes. Field duplicates are collected from the same sample source and device by alternately filling like sample containers for two sample sets until all containers are full. Samples are preserved, stored, and analyzed under identical conditions. The intent of field duplicates is to indicate field and laboratory precision and reproducibility (the degree of contaminant variability within the sample matrix for nonhomogeneous sample matrices). However, the field duplicate sample results include variability introduced during both field sampling and laboratory preparation and analysis, and USEPA data validation guidance provides no specific evaluation criteria for field duplicate samples.

### 3.3.4 Equipment Rinsate Blanks

Equipment rinsate blanks are collected to determine the potential of cross-contamination introduced by non-dedicated equipment (e.g., bladder pump and YSI meter) that is used at multiple sample locations. Deionized water (obtained from the laboratory) is rinsed through the decontaminated sampling equipment
and collected into adequate sample containers for analysis of VOCs. The equipment rinsate blank is then handled in a manner identical to the primary samples collected with that piece of equipment. The blank is then processed, analyzed, and reported as a regular field sample. The rinsate blank collection frequency will be 1 per 20 samples that are collected. When dedicated equipment is used, these blanks are not needed.

### 3.4 Laboratory QA/QC

The laboratories' analytical procedures will meet the requirements specified in the respective analytical methods or approved laboratory standard operating procedures (SOPs), e.g., instrument performance check, initial calibration, calibration check, blanks, surrogate spikes, internal standards, and/or labeled compound spikes. Additional laboratory QA/QC that is related only to Fremont Analytical includes 100 percent laboratory certification of the Summa canister, manifold, and flow controller. Noncertified components will be the new Teflon tubing and the wellhead quick-connect fitting.

Specific laboratory QC analyses required for this project will consist of the following at a minimum:

- Instrument tuning, instrument initial calibration, and calibration verification analyses as required in the analytical methods and the laboratory SOPs;
- Laboratory and/or instrument method blank measurements at a minimum frequency of 5 percent (1 per 20 samples) or in accordance with method requirements, whichever is more frequent; and
- Accuracy and precision measurements will be collected at a minimum frequency of 5 percent (1 per 20 samples) or in accordance with method requirements, whichever is more frequent. In cases where a pair of matrix spike/matrix spike duplicate or matrix spike/laboratory duplicate analyses are not performed on a project sample, a set of laboratory control spike and laboratory control spike duplicate analyses will be performed to provide sufficient measures for analytical precision and accuracy evaluation.

KCEL's data quality objectives are presented in Table E. 1 of the CHRLF Environmental Monitoring SAP (Aspect, 2013). The laboratory's QA officers are responsible for ensuring that the laboratory implements the internal QC and QA procedures detailed in the laboratory's Quality Assurance Manual. The Fremont Analytical methods manual is available on request.

### 3.5 Data Management and Validation

The groundwater and surface water quality data will be validated by KCSWD and managed in KCSWD's project EQUIS database in addition to Aspect's project-specific ACCESS database. The data will be uploaded to Ecology's Environmental Information Management database system only if requested by Ecology. Field and laboratory QC will be validated by KCSWD in accordance with the USEPA National Functional Guidelines for organic and inorganic analyses (USEPA 2008 and 2010, respectively), the laboratory-defined QC limits, and the data validation guidelines detailed in the CHRLF Environmental Monitoring SAP (Aspect, 2013). Data validation is conducted as the data is received for accuracy,
precision, representativeness, completeness, and comparability with regard to the following: sample documentation/custody, holding times, reporting limits, blank/rinsate samples, and surrogate percent recoveries, laboratory duplicates, field duplicates, comparability, and completeness. Section 14.5 of the CHRLF Environmental Monitoring SAP details how the data validation process will be conducted.

### 4.0 REFERENCES

AECOM, 2014, Cedar Hills Regional Landfill Environmental Control Systems Modifications Project Task 215 Field Sampling and Analysis Plan, prepared for King County Department of Natural Resources and Parks Solid Waste Division, October 10, 2014. DRAFT.

Aspect Consulting, LLC (Aspect), 2010, East Main Hill Perched Zones Technical Memorandum, Cedar Hills Regional Landfill, prepared for King County Department of Natural Resources and Parks, Solid Waste Division, October 22, 2010.

Aspect Consulting, LLC (Aspect), 2013, Environmental Monitoring Sampling and Analysis Plan for Cedar Hills Regional Landfill. Prepared for King County Solid Waste Division. Aspect Consulting, LLC, Seattle, WA. December.

CH2M Hill and Udaloy Environmental Services (UES) 2004, Cedar Hills Regional Landfill Site-Wide Hydrogeologic Report, Volumes I, II, and III, prepared for King County Solid Waste Division, May, 2004.

Puls, R.W., and Barcelona, M.J., 1996, Low-Flow (Minimal Drawdown) Ground-Water Sampling Procedures, EPA Ground Water Issue, EPA/540/S-95/504.
U.S. Environmental Protection Agency (USEPA), 1998, Technical Protocol for Evaluating Natural Attenuation of Chlorinated Solvents in Ground Water. EPA/600/R-98/128. U.S. Environmental Protection Agency, Office of Research and Development, September, 1998.

USEPA, 2007, Monitored Natural Attenuation of Inorganic Contaminants in Ground Water. Volume 1: Technical Basis for Assessment. EPA/600/R-07/139. U.S. Environmental Protection Agency, Office of Research and Development, October.

USEPA, 2008, Contract Laboratory Program National Functional Guidelines for Superfund Organic Methods Data Review. USEPA-540-R-08-01. U.S. Environmental Protection Agency, Office of Superfund Remediation and Technical Innovation, June, 2008.

USEPA, 2010, Contract Laboratory Program National Functional Guidelines for Inorganic Superfund Data Review, USEPA 540/R-10/011, U.S. Environmental Protection Agency, Office of Superfund Remediation and Technical Innovation, January, 2010.
U.S. Geological Survey (USGS), 2001, User's Guide for Polyethylene-Based Passive Diffusion Bag Samplers to Obtain Volatile Organic Compound Concentrations in Wells. Part 1: Deployment, Recovery, Data Interpretation, and Quality Control and Assurance. Water-Resources Investigations Report 01-4060.

## Tables

Table A-1 - List of RI/FS Water Level Monitoring Locations

| Monitoring Wells | Gas Probes |  | Groundwater Extraction Wells | Gas Extraction Wells | Surface Water Level Gauge Station |
| :---: | :---: | :---: | :---: | :---: | :---: |
| MW-29 | GP-1A | GP-13A | EW-1 | E-29D | SG-4 |
| MW-30A | GP-1B | GP-13B | EW-2 | E-29C | SG-5 |
| MW-47 | GP-2A | GP-13C | EW-3 | E-29B |  |
| MW-48 | GP-2B | GP-13D | EW-4 | E-29A |  |
| MW-50 | GP-3 | GP-14A | EW-5 | E-61S |  |
| MW-62 | GP-4A | GP-14B | EW-6 | E-61D |  |
| MW-63 | GP-4B | GP-15A | EW-7 | E-62 |  |
| MW-102 | GP-5A | GP-15C | EW-8 | E-63 |  |
| MW-103 | GP-5Ba | GP-15D | EW-9 | E-64 |  |
| MW-104 | GP-5Bb | GP-16A | EW-10 | E-65 |  |
| EB-1 | GP-5Bc | GP-16B | EW-11 |  |  |
| EB-2 | GP-5Bd | GP-16C | EW-12 |  |  |
| EB-3 | GP-6A | GP-17 | EW-13 |  |  |
| EB-4 | GP-6B | GP-18 | EW-14 |  |  |
| EB-5 | GP-6C | GP-19 | EW-15 |  |  |
| EB-5s | GP-6D | GP-45S | EW-16 |  |  |
| EB-6 | GP-6E | GP-45L | EW-17 |  |  |
| EB-7 | GP-6F | GP45d | EW-18 |  |  |
|  | GP-6G | GP-ATC-1S | EW-19 |  |  |
|  | GP-6H | GP-ATC-1D | EW-20 |  |  |
|  | GP-7 | GP-ATC-2 | EW-21 |  |  |
|  | GP-8 | GP-ATC-3S | EW-22 |  |  |
|  | GP-9 | GP-ATC-3D | EW-23 |  |  |
|  | GP-11A | GP-ATC-4 | EW-24 |  |  |
|  | GP-11B | GP-ATC-5S | EW-25 |  |  |
|  | GP-11C | GP-ATC-5D | EW-26 |  |  |
|  | GP-11D | GP-ATC-6S | EW-27 |  |  |
|  | GP-12A | GP-ATC-6D | EW-28 |  |  |
|  | GP-12B | GP-ATC-7 | EW-29 |  |  |
|  | GP-12C | GP-ATC-8S |  |  |  |
|  | GP-12D | GP-ATC-8D |  |  |  |

Bold italics inidicates secondary water level location.

Table A-1

| Location ID | PDB Sample | RPP Sample | Conventional Low Flow or Grab Sample | Soil Gas <br> Samples |
| :---: | :---: | :---: | :---: | :---: |
| Groundwater |  |  |  |  |
| MW-62 | X | X | X |  |
| MW-30A | X | X | X |  |
| MW-47 | X | X | X |  |
| MW-50 |  |  | X |  |
| MW-102 | X |  | X | X |
| MW-103 | X |  | X |  |
| MW-104 | X |  | X | X |
| EB-1 |  |  | X |  |
| EB-2 |  |  | X |  |
| EB-6 | X | X | X | D |
| EW-2 |  |  | X |  |
| EW-6 |  |  | X |  |
| EW-7 |  |  | X |  |
| EW-8 |  |  | X |  |
| EW-9 |  |  | † |  |
| EW-10 |  |  | $\dagger$ |  |
| EW-11 |  |  | + |  |
| EW-12 |  |  | † |  |
| EW-13 |  |  | † |  |
| EW-14 |  |  | X |  |
| EW-15 |  |  | X |  |
| EW-16 |  |  | X |  |
| EW-17 |  |  | X |  |
| EW-18 |  |  | X |  |
| EW-19 |  |  | X |  |
| EW-20 | X | X | X |  |
| EW-21 |  |  | X |  |
| EW-23 |  |  | X |  |
| EW-24 |  |  | X |  |
| EW-25 | X | X | X |  |
| EW-26 |  |  | X |  |
| EW-27 |  |  | X |  |
| EW-29 |  |  | X |  |
| Surface Water |  |  |  |  |
| SW-E1 |  |  | X |  |
| Leachate |  |  |  |  |
| MH-17N \& FS-3 |  |  | X |  |
| Soil Gas |  |  |  |  |
| GP-8 |  |  |  | X |
| GP-15 |  |  |  | S \& M1, M2, or D |
| GP-16 |  |  |  | S \& M or D |
| GP-17 |  |  |  | S \& M or D |
| GP-18 |  |  |  | S \& M or D |
| GP-19 |  |  |  | S \& M or D |
| GP-20 |  |  |  | S \& M or D |
| GP-56 |  |  |  | S |
| GP-58 |  |  |  | S |
| GP-60 |  |  |  | S |
| GP-62 |  |  |  | S |
| GP-ATC-5 |  |  |  | D |
| GP-ATC-7 |  |  |  | D |
| Analytes: | $\begin{gathered} \hline \text { Appendix I } \\ \text { VOCs } \\ \hline \end{gathered}$ | Appendix II Metals | Appendix I \& II Constituents; <br> *MNA parameters | TO-15 VOCs |

Notes:
PDB = Passive diffusion bag sampler
RPP = Rigid Porous Polyethylene sampler
Appendix I \& II constituents are defined in WAC 173-351-990
MNA parameters = ethane and ethene; methane; nitrate/nitrite

* MNA parameters will only be analyzed in groundwater samples.
+ Low-flow groundwater samples will only be collected if sufficient water is present.
$X=$ Sample to be collected. Sample point has only one screened interval.
$S$ = Sample to be collected from the shallow screened interval.
M or $\mathrm{D}=$ Sample to be collected from either the middle or the deep screened interval, whichever exhibits the highest methane. If no methane is detected, select the deep screened interval for sample collection.

M1, M2, or D = Sample to be collected from either one of the two middle or the deep screened interval, whichever exhibits the highest methane concentration. If no methane is detected, select the M2 intermediate screen interval (GP-15C).
$\mathrm{D}=$ Sample to be collected from the deep screened interval.

East Perched Zones RI/FS Work Plan - CHRLF, King County, WA

| Well ID | Top of PVC Elevation (ft) | Total Well <br> Depth (ft bgs) | Top of Screen Depth (ft bgs) | Bottom of Screen Depth (ft bgs) |
| :---: | :---: | :---: | :---: | :---: |
| Groundwater |  |  |  |  |
| MW-29 | 532.92 | 27 | 18 | 27 |
| MW-30A | 568.43 | 35 | 25 | 35 |
| MW-47 | 634.6 | 44 | 23.5 | 43.5 |
| MW-48 | 594.49 | 47 | 37 | 47 |
| MW-50 | 637.02 | 38 | 27.5 | 37.5 |
| MW-62 | 556.21 | 54 | 44 | 54 |
| MW-63 | 515.88 | 17 | 11.5 | 16.5 |
| MW-102 | 552.48 | 49.5 | 34.5 | 49.5 |
| MW-103 | 639.08 | 35 | 25 | 35 |
| MW-104 | 629.68 | 32 | 22 | 32 |
| EB-1 | 532.3 | 22 | 17 | 22 |
| EB-2 | 530.12 | 24 | 13.5 | 23.5 |
| EB-6 | 589.61 | 30 | 20 | 30 |
| EW-2 | 561.56 | 34.8 | 19.3 | 28.6 |
| EW-6 | 582.87 | 59.9 | 45.54 | 54.89 |
| EW-7 | 593.47 | 45.8 | 30.4 | 39.73 |
| EW-8 | 600.38 | 54.5 | 39.2 | 48.4 |
| EW-9 | 602.92 | 46.2 | 31.2 | 40.5 |
| EW-10 | 609.03 | 43.8 | 28.28 | 37.6 |
| EW-11 | 617.6 | 43.5 | 28 | 37.4 |
| EW-12 | 623.25 | 39.8 | 22.5 | 31.8 |
| EW-13 | 633.77 | 39.9 | 24.4 | 33.7 |
| EW-14 | 633.66 | 47.9 | 32.6 | 42 |
| EW-15 | 635.3 | 47.8 | 29.6 | 39 |
| EW-16 | 636.88 | 43.7 | 29.5 | 38.81 |
| EW-17 | 637.27 | 43.5 | 29.5 | 38.9 |
| EW-18 | 639.88 | 43.1 | 27.4 | 36.7 |
| EW-19 | 640 | 44 | 29 | 38.4 |
| EW-20 | 639.03 | 43.2 | 28.7 | 38 |
| EW-21 | 641.04 | 39.5 | 24 | 33.4 |
| EW-22 | 639.71 | 44.1 | 30.5 | 39.8 |
| EW-23 | 640.65 | 44.5 | 30.7 | 40.1 |
| EW-24 | 643.11 | 39 | 24.1 | 33.4 |

Table A-3-RI/FS Well Screened Intervals
East Perched Zones RI/FS Work Plan - CHRLF, King County, WA

| EW-25 | 643.61 | 38.3 | 22.8 | 32 |
| :---: | :---: | :---: | :---: | :---: |
| EW-26 | 642.16 | 36 | 21.1 | 30.5 |
| EW-27 | 640.63 | 36.9 | 21.7 | 31 |
| EW-28 | 640.86 | 22.8 | 7.7 | 17 |
| EW-29 | 638.93 | 23.6 | 8.2 | 17.5 |

Notes:
bgs = below ground surface $\mathrm{ft}=$ feet

East Perched Zones RI/FS Work Plan - CHRLF, King County, WA

| Well ID | Surface Elevation | Total Depth (ft bgs) | Top of Shallow Screen Depth (ft bgs) | Bottom of Shallow <br> Screen Depth (ft bgs) | Top of Mid Screen Depth (ft bgs) | Bottom of Mid Screen Depth (ft bgs) | Top of Deep Screen Depth (ft bgs) | Bottom of Deep Screen Depth (ft bgs) | Notes |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Gas Extraction Wells |  |  |  |  |  |  |  |  |  |
| E-29D |  |  |  |  |  |  |  |  | Boring log has not been located |
| E-29C |  |  |  |  |  |  |  |  | Boring log has not been located |
| E-29B |  |  |  |  |  |  |  |  | Boring log has not been located |
| E-29A |  |  |  |  |  |  |  |  | Boring log has not been located |
| E-61S |  |  |  |  |  |  |  |  | Boring $\log$ has not been located |
| E-61D |  |  |  |  |  |  |  |  | Boring log has not been located |
| E-62 |  |  |  |  |  |  |  |  | Boring log has not been located |
| E-63 | 607.64 | 151 | 22 | 60 | 83 | 103 | 130 | 150 |  |
| E-64 | 615.91 | 146 | 31 | 60 | 80 | 100 | 125 | 145 |  |
| E-65 | 625.35 | 103 | 83 | 102 |  | - |  | - |  |

Table A-3

| Well ID | Top of PVC Elevation <br> (ft) | Total Depth (ft bgs) | Top of Shallow Screen Depth (ft bgs) | Bottom of <br> Shallow <br> Screen <br> Depth (ft <br> bgs) | Top of Mid Screen Depth (ft bgs) | Bottom of Mid Screen Depth (ft bgs) | Top of Deep <br> Screen Depth (ft bgs) | Bottom of Deep Screen Depth (ft bgs) | Notes |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Soil Gas |  |  |  |  |  |  |  |  |  |
| GP-1A | 639.93 | 22.5 | 8 | 12 |  | - | - | - |  |
| GP-1B | 639.82 | 22.5 | 18.5 | 22.5 |  | - |  | - |  |
| GP-2A | 627.03 | 22.5 | 6 | 10 | - | - | - | - |  |
| GP-2B | 627.03 | 22.5 | 18 | 20 |  | - | - | - |  |
| GP-3 | 594.21 | 63 | 5 | 9 | 16 | 20 |  | - | Colocated with MW-48 |
| GP-4A | 605.72 | 24 | 4.5 | 8.5 | - | - | - | - |  |
| GP-4B | 605.85 | 24 | 15.5 | 19.5 |  | - | - | - |  |
| GP-5A | 617.47 | 75 | 6 | 7 | - | - | - | - |  |
| GP-5Ba | 619.33 | 75 | 6 | 7 | - | - | - | - |  |
| GP-5Bb | 619.3 | 75 | 22 | 23 | - | - | - | - |  |
| GP-5BC | 619.31 | 75 | 51 | 52 | - | - | - | - |  |
| GP-5Bd | 619.31 | 75 | 63 | 64 | - | - | - | - |  |
| GP-6A | 634.81 | 203 | 54 | 55 |  | - | - | - |  |
| GP-6B | 634.53 | 203 | 84 | 85 | - | - | - | - |  |
| GP-6C | 634.75 | 203 | 94 | 95 | - | - | - | - |  |
| GP-6D | 634.69 | 203 | 113 | 114 | - | - | - | - |  |
| GP-6E | 634.62 | 203 | 134 | 135 | - | - | - | - |  |
| GP-6F | 634.81 | 203 | 148 | 149 | - | - | - | - |  |
| GP-6G | 634.68 | 203 | 163 | 164 | - | - | - | - |  |
| GP-6H | 634.71 | 203 | 178 | 179 | - | - | - | - |  |
| GP-7 | 640.24 | 58 | 48 | 50 | - | - | - | - |  |
| GP-8 | 642.23 | 60 | 44 | 45 | - | - | - | - |  |
| GP-9 | 644.99 | 70 | 58 | 59 | - | - | - | - |  |
| GP-11A | 566.69 | 100 | 6.5 | 7.5 | - | - | - | - |  |
| GP-11B | 566.71 | 100 | 23.5 | 24.5 | - | - | - | - |  |
| GP-11C | 566.74 | 100 | 54.5 | 55.5 | - | - | - | - |  |
| GP-11D | 566.72 | 100 | 91.5 | 92.5 | - | - | - | - |  |


| Sample <br> Matrix | Analytical Parameter | Analytical Method | Sample Container | No. Containers | Min. Sample Volume, mL | Min. No. Containers | Preservation <br> Requirements | Holding Time |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Water | VOCs | EPA 8260C | 40-mL VOA Vials | 4 | 40, 1 full vial | 1 | $\begin{gathered} 4^{\circ} \mathrm{C} \pm 2^{\circ} \mathrm{C}, \\ \mathrm{HCl} \mathrm{pH}<2 \\ \hline \end{gathered}$ | 14 days for analysis |
|  | Dissolved Metals (freshwater)* | EPA 6020A and EPA 7470 | 500-mL acidwashed HDPE | 1 | 150 | 1 | $\qquad$ | 28 days for $\mathrm{Hg}, 180$ days for other elements |
|  | Total Metals (freshwater)* | EPA 6020A and EPA 7470 | 500-mL acidwashed HDPE | 1 | 200 | 1 | $\begin{gathered} 4^{\circ} \mathrm{C} \pm 2^{\circ} \mathrm{C} ; \mathrm{HNO} \\ \mathrm{pH}<2 \text { in lab } \\ \hline \end{gathered}$ | $\begin{gathered} 28 \text { days for } \mathrm{Hg}, 180 \\ \text { days for other } \\ \text { elements } \\ \hline \end{gathered}$ |
|  | Ammonia, Nitrate-N, <br> Nitrite-N | Kerouel \& Aminot <br> 1997 or SM4500- <br> NH3-G (ammonia); <br> SM4500-NO3-F <br> (nitrate, nitrite) | $\begin{gathered} 250-\mathrm{mL} \mathrm{WM} \\ \text { HDPE } \\ \hline \end{gathered}$ | 1 | 50 | 1 | filter within one day of collection then freeze | 14 days @ -20 ${ }^{\circ} \mathrm{C}$ |
|  | TSS | SM2540D | 1-L WM HDPE | 1 | 1000 | 1 | $4^{\circ} \mathrm{C} \pm 2^{\circ}$ | 7 days |
|  | TDS | SM2540C | $\begin{gathered} \text { 500-mL WM } \\ \text { HDPE } \end{gathered}$ | 1 | 150 | 1 | $4^{\circ} \mathrm{C} \pm 2^{\circ}$ | 7 days |
|  | TOC | SM5310-B | 125-mL amber glass | 1 | 40 | 1 | $\begin{gathered} 4^{\circ} \mathrm{C} \pm 2^{\circ} \mathrm{C}, \mathrm{H} 3 \mathrm{PO} 4 \\ \mathrm{pH}<2 \\ \hline \end{gathered}$ | 28 days |
|  | Chloride, Sulfate | SM4110B | $\begin{gathered} 125-\mathrm{mL} \mathrm{NM} \\ \text { HDPE } \end{gathered}$ | 1 | 50 | 1 | $4^{\circ} \mathrm{C} \pm 2^{\circ}$ | 28 days |
|  | Bicarbonate (HCO3);and Alkalinity (as CaCO3) | $\begin{gathered} \text { SM2320-B or EPA } \\ 310.1 \end{gathered}$ | 500-mL WM HDPE | 1 | 120 | 1 | $4^{\circ} \mathrm{C} \pm 2^{\circ}$ | 14 days |
|  | Dissolved methane; ethene; ethane | RSK 175 (subcontract) | 40-mL VOA Vials | 2 | 40, 1 full vial | 1 | $\begin{gathered} 4^{\circ} \mathrm{C} \pm 2^{\circ} \mathrm{C} ; \\ \text { Na3PO4 or BAK } \end{gathered}$ | 14 days |
|  | Dissolved Ferrous Iron | Hach kit Method 8146 or subcontract | 500-mL HDPE (same bottle as dissolved metals) | 1 | 50 | 1 | $4^{\circ} \mathrm{C} \pm 2^{\circ} \mathrm{C}$, after filtration 0.1micron | ASAP |
| Leachate | VOCs | EPA 8260C | 40-mL VOA Vials | 4 | 40, 1 full vial | 1 | $\begin{gathered} 4^{\circ} \mathrm{C} \pm 2^{\circ} \mathrm{C}, \\ \mathrm{HCl} \mathrm{pH}<2 \\ \hline \end{gathered}$ | 14 days for analysis |
|  | Total Metals (freshwater)* | EPA 6020A and EPA 7471 | 500-mL HDPE | 1 | 200 | 1 | $\begin{gathered} 4^{\circ} \mathrm{C} \pm 2^{\circ} \mathrm{C} \text {; HNO3 } \\ \mathrm{pH}<2 \text { in lab } \\ \hline \end{gathered}$ | 28 days for $\mathrm{Hg}, 180$ days for other elements |
|  | Ammonia, Nitrate-N, <br> Nitrite-N | Kerouel \& Aminot <br> 1997 or SM4500- <br> NH3-G (ammonia); <br> SM4500-NO3-F <br> (nitrate, nitrite) | $\begin{gathered} 250-\mathrm{mL} \mathrm{WM} \\ \text { HDPE } \\ \hline \end{gathered}$ | 1 | 50 | 1 | filter within one day of collection then freeze | 14 days @ -20 ${ }^{\circ} \mathrm{C}$ |
|  | TSS | SM2540D | 1-L WM HDPE | 1 | 1000 | 1 | $4^{\circ} \mathrm{C} \pm 2^{\circ}$ | 7 days |
|  | TDS | SM2540C | $\begin{gathered} \text { 500-mL WM } \\ \text { HDPE } \end{gathered}$ | 1 | 150 | 1 | $4^{\circ} \mathrm{C} \pm 2^{\circ}$ | 7 days |
|  | TOC | SM5310-B | 125-mL amber glass | 1 | 40 | 1 | $\begin{gathered} 4^{\circ} \mathrm{C} \pm 2^{\circ} \mathrm{C}, \mathrm{H} 3 \mathrm{PO} 4 \\ \mathrm{pH}<2 \end{gathered}$ | 28 days |
|  | Chloride, Sulfate | SM4110B | $\begin{gathered} 125-\mathrm{mL} \mathrm{NM} \\ \text { HDPE } \end{gathered}$ | 1 | 50 | 1 | $4^{\circ} \mathrm{C} \pm 2^{\circ}$ | 28 days |
|  | Bicarbonate (HCO3);and <br> Alkalinity (as CaCO3) | SM2320-B or EPA 310.1 | $\begin{gathered} 500-\mathrm{mL} \mathrm{WM} \\ \text { HDPE } \\ \hline \end{gathered}$ | 1 | 120 | 1 | $4^{\circ} \mathrm{C} \pm 2^{\circ}$ | 14 days |
| Soil Gas | vocs | $\begin{aligned} & \hline \text { EPA TO-15 } \\ & \text { (subcontract) } \end{aligned}$ | 6-L Summa Canister | 1 | NA | 1 | - | 30 days |

Notes:

* = WAC 173-351 Appendix I metals list includes antimony, arsenic, barium, beryllium, cadmium, chlomium, cobalt, copper, lead, nickel, selenium, silver, thallium, vanadium, zinc, and nitrate. Also includes select WAC 173-351 Appendix II geochemical parameters including calcium, magnesium, sodium, chloride, potassium, iron, manganese.
ASAP = as soon as possible
BAK = Benzyl ammonium chloride
EPA $=$ U.S. Environmental Protection Agency
HDPE = high-density polyethylene
TDS = total dissolved solids
TOC = total organic carbon
TSS = total suspended solids
VOA $=$ volatile organic analysis
VOC = volatile organic compound

| Volatile Organic Compounds Water \& Leachate Samples |  |  |
| :---: | :---: | :---: |
| CAS | Common Analytical Name | Synonym |
| 67-64-1 | Acetone |  |
| 107-13-1 | Acrylonitrile |  |
| 71-43-2 | Benzene |  |
| 74-97-5 | Bromochloromethane |  |
| 75-27-4 | Bromodichloromethane |  |
| 75-25-2 | Bromoform | Tribromomethane |
| 75-15-0 | Carbon disulfide |  |
| 56-23-5 | Carbon tetrachloride |  |
| 108-90-7 | Chlorobenzene |  |
| 75-00-3 | Chloroethane | Ethyl chloride |
| 67-66-3 | Chloroform | Trichloromethane |
| 124-48-1 | Dibromochloromethane | Chlorodibromomethane |
| 96-12-8 | 1,2-Dibromo-3-chloropropane | DBCP |
| 106-93-4 | 1,2-Dibromoethane | Ethylene dibromide; EDB |
| 95-50-1 | 1,2-Dichlorobenzene | o-Dichlorobenzene; |
| 106-46-7 | 1,4-Dichlorobenzene | p-Dichlorobenzene |
| 110-57-6 | trans-1,4-Dichloro-2-butene |  |
| 75-34-3 | 1,1-Dichloroethane | Ethylidene chloride |
| 107-06-2 | 1,2-Dichloroethane | Ethylene dichloride; EDC |
| 75-35-4 | 1,1-Dichloroethylene | 1,1-Dichloroethene; Vinylidene chloride |
| 156-59-2 | cis-1,2-Dichloroethene | cis-1,2-Dichloroethylene |
| 156-60-5 | trans-1,2-Dichloroethene | trans-1,2-Dichloroethylene |
| 78-87-5 | 1,2-Dichloropropane | Propylene dichloride |
| 10061-01-5 | cis-1,3-Dichloropropene |  |
| 10061-02-6 | trans-1,3-Dichloropropene |  |
| 100-41-4 | Ethylbenzene |  |
| 591-78-6 | 2-Hexanone | Methyl butyl ketone |
| 74-83-9 | Bromomethane | Methyl bromide |
| 74-87-3 | Methyl chloride | Chloromethane |
| 74-95-3 | Dibromomethane | Methylene bromide |
| 75-09-2 | Dichloromethane | Methylene chloride |
| 78-93-3 | 2-Butanone | Methyl ethyl ketone; MEK |
| 74-88-4 | Methyl iodide | lodomethane |
| 108-10-1 | 4-Methyl-2-pentanone | Methyl isobutyl ketone |
| 100-42-5 | Styrene |  |
| 630-20-6 | 1,1,1,2-Tetrachloroethane |  |
| 79-34-5 | 1,1,2,2-Tetrachloroethane |  |
| 127-18-4 | Tetrachloroethylene | Tetrachloroethene; Perchloroethylene |
| 108-88-3 | Toluene |  |
| 71-55-6 | 1,1,1-Trichloroethane | Methyl chloroform |
| 79-00-5 | 1,1,2-Trichloroethane |  |
| 79-01-6 | Trichloroethylene | Trichloroethene |
| 75-69-4 | Trichlorofluoromethane | CFC-11 |
| 96-18-4 | 1,2,3-Trichloropropane |  |
| 108-05-4 | Vinyl acetate |  |
| 75-01-4 | vinyl chloride |  |
| 1330-20-7 | Xylenes |  |


| Inorganics - Water \& Leachate Samples |  |
| :---: | :---: |
| CAS | Metals |
| 7440-36-0 | Antimony |
| 7440-38-2 | Arsenic |
| 7440-39-3 | Barium |
| 7440-41-7 | Beryllium |
| 7440-43-9 | Cadmium |
| 7440-47-3 | Chromium (Total) |
| 7440-48-4 | Cobalt |
| 7440-50-8 | Copper |
| 7439-92-1 | Lead |
| 7439-97-6 | Mercury |
| 7440-02-0 | Nickel |
| 7782-49-2 | Selenium |
| 7440-22-4 | Silver |
| 7440-28-0 | Thallium |
| 7440-62-2 | Vanadium |
| 7440-66-6 | Zinc |
| 14797-55-8 | Nitrate |
|  |  |
|  | Field Parameters |
|  | pH |
|  | Conductivity (measured in specific conductance units) |
|  | Temperature |
|  | Static Water Level |
|  |  |
|  | Geochemical Indicator Parameters |
| 7440-70-2 | Calcium |
| 71-52-3 | Bicarbonate |
| 7439-95-4 | Magnesium |
| 14808-79-8 | Sulfate |
| TSS | Total suspended solids |
| 7440-23-5 | Sodium |
| 16887-00-6 | Chloride |
| 7440-09-7 | Potassium |
|  | Alkalinity |
| 7439-89-6 | Iron (Dissolved) |
| 7439-96-5 | Manganese (Dissolved) |
|  |  |
|  | Leachate Indicators |
| 7664-41-7 | Ammonia |
|  | Total Organic Carbon |
|  | Total Dissolved Solids |
|  |  |
|  | Additional MNA Parameters (GW Only) |
|  | Oxidation Reduction Potential |
|  | Dissolved Oxygen |
| 74-82-8 | Methane |
|  | Nitrate + nitrite |
|  | Ethene and Ethane |

Notes:
Parameters in italics are those for groundwater sample analysis only. They are for monitored natural attenuation evaluation and are not listed in Appendix I or II of WAC 173-351.

| Volatile Organic Compounds - Soil Gas Samples |  |  |
| :---: | :---: | :---: |
| CAS | Common Analytical Name | Synonym |
| 71-55-6 | 1,1,1-Trichloroethane | TCA |
| 79-34-5 | 1,1,2,2-Tetrachloroethane |  |
| 79-00-5 | 1,1,2-Trichloroethane |  |
| 76-13-1 | 1,1,2-Trichlorotrifluoroethane |  |
| 75-34-3 | 1,1-Dichloroethane | 1,1-DCA |
| 75-35-4 | 1,1-Dichloroethene | 1,1-DCE |
| 95-63-6 | 1,2,4-Trimethylbenzene |  |
| 106-93-4 | 1,2-Dibromoethane | Ethylene dibromide; EDB |
| 76-14-2 | $\begin{array}{\|l\|} \hline \text { 1,2-Dichloro-1,1,2,2- } \\ \text { tetrafluoroethane } \\ \hline \end{array}$ | CFC 114 |
| 95-50-1 | 1,2-Dichlorobenzene |  |
| 107-06-2 | 1,2-Dichloroethane |  |
| 78-87-5 | 1,2-Dichloropropane |  |
| 108-67-8 | 1,3,5-Trimethylbenzene |  |
| 106-99-0 | 1,3-Butadiene |  |
| 541-73-1 | 1,3-Dichlorobenzene |  |
| 106-46-7 | 1,4-Dichlorobenzene |  |
| 540-36-3 | 1,4-Difluorobenzene |  |
| 78-93-3 | 2-Butanone | MEK |
| 591-78-6 | 2-Hexanone |  |
| 460-00-4 | 4-Bromofluorobenzene |  |
| 622-96-8 | 4-Ethyltoluene |  |
| 108-10-1 | 4-Methyl-2-pentanone |  |
| 67-64-1 | Acetone |  |
| 71-43-2 | Benzene |  |
| 74-97-5 | Bromochloromethane |  |
| 75-27-4 | Bromodichloromethane |  |
| 75-25-2 | Bromoform |  |
| 74-83-9 | Bromomethane |  |
| 75-15-0 | Carbon Disulfide |  |
| 56-23-5 | Carbon Tetrachloride |  |
| 108-90-7 | Chlorobenzene |  |
| 3114-55-4 | Chlorobenzene-d5 |  |
| 75-00-3 | Chloroethane |  |
| 67-66-3 | Chloroform |  |
| 74-87-3 | Chloromethane |  |
| 110-82-7 | Cyclohexane |  |
| 124-48-1 | Dibromochloromethane |  |
| 75-71-8 | Dichlorodifluoromethane | CFC 12 |
| 74-95-3 | Dichloromethane |  |
| 141-78-6 | Ethyl Acetate |  |
| 100-41-4 | Ethylbenzene |  |
| 1634-04-4 | Methyl tert-Butyl Ether |  |
| 100-42-5 | Styrene |  |
| 127-18-4 | Tetrachloroethene | PCE |
| 109-99-9 | Tetrahydrofuran | THF |
| 108-88-3 | Toluene |  |
| 79-01-6 | Trichloroethene | TCE |
| 75-69-4 | Trichlorofluoromethane | CFC 11 |
| 108-05-4 | Vinyl Acetate |  |
| 75-01-4 | Vinyl Chloride |  |
| 540-59-0 | cis-1,2-Dichloroethene |  |
| 10061-01-5 | cis-1,3-Dichloropropene |  |
| 179601-23-1 | m,p-Xylenes |  |
| 142-82-5 | n-Heptane |  |
| 110-54-3 | n-Hexane |  |


| Volatile Organic Compounds - Soil Gas Samples |  |  |
| :---: | :---: | :---: |
| CAS | Common Analytical Name | Synonym |
| 156-60-5 | trans-1,2-Dichloroethene |  |
| 10061-02-6 | trans-1,3-Dichloropropene |  |
| 526-73-8 | 1,2,3-Trimethylbenzene |  |
| 120-82-1 | 1,2,4-Trichlorobenzene |  |
| 96-12-8 | 1,2-Dibromo-3-chloropropane | DBCP |
| 17060-07-0 | 1,2-Dichloroethane-d4 |  |
| 123-91-1 | 1,4-Dioxane |  |
| 71-36-3 | 1-Butanol |  |
| 540-84-1 | 2,2,4-Trimethylpentane | Isooctane |
| 611-14-3 | 2-Ethyltoluene |  |
| 75-65-0 | 2-Methyl-2-propanol |  |
| 67-63-0 | 2-Propanol |  |
| 107-05-1 | 3-Chloro-1-propene |  |
| 620-14-4 | 3-Ethyltoluene |  |
| 99-87-6 | 4-Isopropyltoluene |  |
| 75-05-8 | Acetonitrile |  |
| 107-02-8 | Acrolein |  |
| 107-13-1 | Acrylonitrile |  |
| 100-44-7 | Benzyl Chloride |  |
| 108-20-3 | Diisopropyl Ether |  |
| 64-17-5 | Ethanol |  |
| 637-92-3 | Ethyl tert-Butyl Ether |  |
| 87-68-3 | Hexachlorobutadiene |  |
| 108-21-4 | Isopropyl Acetate |  |
| 98-82-8 | Isopropylbenzene | Cumene |
| 80-62-6 | Methyl Methacrylate |  |
| 91-20-3 | Naphthalene |  |
| 115-07-1 | Propene |  |
| 2037-26-5 | Toluene-d8 |  |
| 98-83-9 | alpha-Methylstyrene |  |
| 80-56-8 | alpha-Pinene |  |
| 5989-27-5 | d-Limonene |  |
| 123-86-4 | n-Butyl Acetate |  |
| 124-18-5 | n-Decane |  |
| 111-84-2 | n-Nonane |  |
| 111-65-9 | n-Octane |  |
| 103-65-1 | n-Propylbenzene |  |
| 1120-21-4 | n-Undecane |  |
| 135-98-8 | sec-Butylbenzene |  |
| 994-05-8 | tert-Amyl Methyl Ether |  |
| 95-47-6 | o-Xylene |  |


| QC Sample | Approximate <br> Quantity | Analysis |
| :--- | :---: | :---: |
| Water Samples | 3 |  |
| parameters |  |  |
| Field Duplicates | 3 |  |
| Field Blanks | $\sim 10$ | VOCs |
| Trip Blanks | 2 | VOCs |
| Rinsate Blanks |  |  |

## Figures






## ATTACHMENT A

## Field Documentation Records

## Stream Sampling Record

Cedar Hills Regional Landfill 130088


## Passive Diffusion Sampling Record

Cedar Hills Regional Landfill 130088

| Well Number | Screen Interval (ft BGS) | Deployme nt Date | $\left\|\begin{array}{c} \text { Total } \\ \text { Depth } \\ \text { (ft BTOC) } \end{array}\right\|$ | Depth to Water (ft BTOC) | Depth to Middle of PDB (ft BTOC) | Depth to Middle of RPP (ft BTOC) | Collection Date | Collection Time | Depth to Water (ft BTOC) | Temp $\left({ }^{\circ} \mathrm{C}\right)$ | $\begin{gathered} \text { DO } \\ (\mathrm{mg} / \mathrm{L}) \end{gathered}$ | Sp. Cond. ( $\mu \mathrm{S} / \mathrm{cm}$ ) | pH | $\begin{aligned} & \text { ORP } \\ & (\mathrm{mV}) \end{aligned}$ | Observati ons and Comment s |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MW-102 | 34.5-49.5 |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| MW-103 | 25-35 |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| MW-104 | 22-32 |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| EW-18 | 55-65? |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| EW-19 | 30.5-39.8 |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| MW-62 | 44-54 |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| MW-30A | 25-35 |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| MW-47 | 23.50-43.50 |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| EB-6 | 20.45-30.45 |  |  |  |  |  |  |  |  |  |  |  |  |  |  |

## Sampling Event:

Sampler Initials:
Sampling Equipment with IDs:
Decon Equipment:

Disposal of Decon/Purge Water:

Groundwater Sampling Record Cedar Hills Regional Landfill 130088


## Aspect

Gas Well ID:
Sample ID:
Date \& Time $\qquad$
Total Casing Volume (cc):

## Cedar Falls Landfill

## Gas Probe Data Sheet - Cedar Hills Regional Landfill 130088

| Casing Volume Purged | Volume Purged (cc) | Purge Rate ( $\mathrm{ml} / \mathrm{min}$ ) | Purge Time | Purge Time |  | $\mathrm{CH}_{4}$ (\% volume) | $\mathrm{CO}_{2}$ $(\%$ volume) | $\|$$\mathrm{O}_{2}$ <br> $(\%$ volume $)$ | $\begin{gathered} \mathrm{H} 2 \mathrm{~S} \\ (\mathrm{ppm}) \end{gathered}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | 0 | 0 |  | 0 min | 0 sec |  |  |  |  |
|  |  |  |  | min | sec |  |  |  |  |
|  |  |  |  | min | sec |  |  |  |  |
|  |  |  |  | min | sec |  |  |  |  |
|  |  |  |  | min | sec |  |  |  |  |
|  |  |  |  | min | sec |  |  |  |  |
|  |  |  |  | min | sec |  |  |  |  |
|  |  |  |  | min | sec |  |  |  |  |
|  |  |  |  | min | sec |  |  |  |  |
|  |  |  |  | min | sec |  |  |  |  |
|  |  |  |  | min | sec |  |  |  |  |
|  |  |  |  | min | sec |  |  |  |  |
|  |  |  |  | min | sec |  |  |  |  |

Comments:

$\qquad$

Equipment Used: Gem 2000, Water Level Meter

## Attachment B

Boring Logs for RI Sample Locations

LOG OF EXPLORATORY BORING Renumbered as mw-62


Cedar Hills Landfili Adjacent to MW-30A Holt Drilling, Inc. Cable Tool Mike Noll

## BORING of $8 / 91$

BORING NO.
PAGE
PAGE 1 OF 4
REFERENCE ELEV. 554.41' MSL

TOTAL DEPTH
65.50'

2/1/90


DATE COMPLETED
lithologic DESCRIPTION
(4)

G 2
$\left[\begin{array}{ll} \\ G 3\end{array}\right.$

G 4
( (

REMARKS

1) Conductivity of water used during drilling $=165$ to 185 umhos. 2) $G=$ grab samples, collected over a 0.5 to 1 foot interval from bailed drill cuttings; $S B=$ split barrel samples collected using a 2.5 -inch O.D. Sprigs-Hendley core barrel driven using a $300-1 \mathrm{~b}$ hammer. 3) ${ }^{*} \approx$ Muddy water samples, diluted with drilling water. 4) Reference elevation $=$ ground surface.




## LOG OF EXPLORATORY BORING



King County Solid Waste Division Cedar Hills Regional Landfill
Holt Drilling Cable Tool M.D. Noll

| BORING NO. | MW-30A |
| :--- | :---: |
| PAGE | $10 F 2$ |
| REFERENCE ELEV. | 567 |
| TOTAL DEPTH | $\quad$ |
| DATE COMPLETED | $9 / 6 / 89$ |

BORING NO.
NW
REFERENCE ELEV. 567 * DATE COMPLETED $9 / 6 / 89$


## REMARKS

1) Reference elevation is ground surface, based on topography. 2) $C=$ sample of cuttings.


REMARKS

1) Reference elevation is ground surface, based on topography. 2) $C=$ sample of cuttings.

Location Ne corner of Landfill Boundary
Surface Elevation $\qquad$ 633.56

Total Depth 50.0 Feet
Date Completed
31 May 1985

Sweet, Edwards \& Associates, Inc.
BORING LOG

PROJECT CEDAR HILLS SITE DEVELOPMENT PLAN
Page $\qquad$ 2 of

Boring No.
MW-47


Sweet, Edwards \& Associates, Inc.

PROJECT CEDAR HILLS SITE DEVELOPMENT PLAN
Page ${ }^{1}$ of 2

Location East side of landfill
Surface Elevation 637.3 feet a.s.1.
Total Depth 39.5 feet
Date Completed 6/3/85

Boring No. MW-50
Drilling Method odex
Drilled By Kring Drilling Co.
Logged By D.E. Nadler


Sweet, Edwards \& Associates, Inc.
BORING LOG

PROJECT CEDAR HILLS SITE DEVELOPMENT PLAN
Page_ 2 of 2
Boring No.
MW-50


-MONITORING WELL CEDAR HILLS PERCHING INVESTIGATION-REVISED DENSITY.GPJ APRII 13, 2010






# LOG OF EXPLORATORY BORING 



## LOG OF EXPLORATORY BORING

LOG OF EXPLORATORY BORING


## LOG OF EXPLORATORY BORING



LOG OF EXPLORATORY BORING



## REMARKS

SWEET-EDWARDS/EMCON

## LOG OF EXPLORATORY BORING

PROJECT NAME Harper Owes
LOCATION DRILLED BY DRILL METHOD LOGGED BY

Tacoma Pump \& Drill Air Rotary Mike Noll

BORING NO.
PAGE
REFERENCE ELEV.
TOTAL DEPTH
DATE COMPLETED

EB- 6
3 OF 3
587.41' MSL $50.00^{\prime}$ 11/28/90

| Sampling Method and Number | BLOWS/6" |  |  | 嵒1 | LITHO- LOGIC COLUMN | melt. details | LITHOLOGIC DESCRIPTION |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |

(1)


(sher



































| Orill Method | Air Rotary |  |
| :---: | :---: | :---: |
| Boring No. | EW-22A 20 |  |
| roc Elevation | 639.03 ft | Date 9/2/92 |



Total depth drilled $=44.0$ feet


REVISEO





stickup $\quad 0.70 \mathrm{ft}$
Sand Pack $10 \times 20$
Silica Sand
8.0 to 40.7 feet 8.0 to 40.7 feet

|  | Drill Method Boring No. | Air Rotary |  |  |
| :---: | :---: | :---: | :---: | :---: |
|  |  | EW-20A 23 |  |  |
|  |  | 640.65 ft | Date | 8/28/92 |



SILT with gravel (ML); medium gray; dense; alight plasticity; few clay; trace fine gravel and coarse sand; damp (LACUSTRINE)
increase in percent clay content; moderate plasticity
SANDY SILT (ML); medium gray; some very fine sand; wet (LACUSTRINE)
accumulated formation water in borehole overnight

- samples below 38.5 feet wet

Harding Lawson Assoclates
Engineering and Environmental Services
Enginee

Log of Boring and Well Completion
EW-20A
$\underset{\text { APPROVEO }}{\text { Cedar Hills Landfill }}$




stickup

[^0]Log of Boring and Well Completion

EW-29A
(sheet 1 of 2 )




$$
L_{-}^{-}
$$










## LOG OF EXPLORATORY BORING



## LOG OF EXPLORATORY BORING



## LOG OF EXPLORATORY BORING



## LOG OF EXPLORATORY BORING



## LOG OF EXPLORATORYBORING



## LOG OF EXPLORATORYBORING



## LOG OF EXPLORATORY BORING

| OG OF EXPLORATORY BORING |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: |
| PROJECT NAME <br> LOCATION <br> DRILLED BY <br> DRIL METHOD <br> LOGGED BY | Cedar Hills Landfill: Gas Probe Installations <br> T.P.\&D <br> Air Rotary <br> R. J. Rowiand |  |  |  |
|  |  |  | detatl | THOLOGIC |
| (G) |  |  |  |  |
| REMARKS |  |  |  |  |





| PROJECT NAME LOCATION <br> DRMLED BY <br> DRILL METHOD <br> LOGGED BY | Cedar Hills Landill: Gas $P$ <br> Tacoma Pump \& Drill Air Rotary. <br> P. J. Rowland | robe Installations | BORING NO. <br> PAGE <br> REFERENCE ELEV. <br> TOTAL DEPTH <br> DATE COMPLETED | GP-20 <br> 1 OF 2 <br> 3.00' <br> $95.00^{\circ}$ <br> 3/29/88 |
| :---: | :---: | :---: | :---: | :---: |
|  |  | vell details | LIthologic description |  |
|  |  |  | LTTY SAND AND GRA <br> carse sand; medium to co $50 \%$ silt; $10 \%$ cobbles. raded, moderately dense <br> WEATHERED GLACI <br> 12 feet. <br> feet <br> GRAVELIYSILT (GM) arse gravels in a very fine trix, $10 \%$ cobbles/bould e, rounded gravels, satur <br> GLACIAL TILL <br> DESCRIPTION on nex | L <br> TILL <br> Poorly in <br> age. |
|  | ARKS |  |  |  |







Location South of South dormitory

## Surface Elevation

$\qquad$ Total Depth 21 feet
Date Completed $\qquad$
10/7/86
Borlng No. GP-ATC-5
Drilling Method Air Rotary
Drllled By Hayes Well Drilling
Logged By D.E. Mills


Location North of Administration Building
Surface Elevation $\qquad$
Total Depth 22 feet

Date Completed 10/7/86


## APPENDIXB

## Site Analytical Data (as CD)

## APPENDIX C

## Individual Constituent Extent Maps






















## APPENDIX D

## Time Series Concentration Plots for Select Monitoring Wells




## APPENDIX E

## Existing Infrastructure

This Appendix provides subsurface utility maps developed by Aspect ( Aspect, 2010) for subsurface utility lines. The user of these maps is referred to Aspect, 2010 for discussion on data sources and compilation methods for these drawings.





[^0]:    Marding Lawson Assoclates
    Engineering and Environmental Services

