**APPENDIX F** Compliance Monitoring and Quality Assurance Project Plan

## **Compliance Monitoring and Quality** Assurance Project Plan (CMP/QAPP)

Quiet Cove Site Anacortes, Washington Ecology Agreed Order No. DE 11346

for

Washington State Department of Ecology on Behalf of Port of Anacortes

August 19, 2019





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File No. 5147-024-07

August 19, 2019

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#### LIST OF ACRONYMS AND ABBREVIATIONS

Acronym/ Abbreviation	Description
ARI	Analytical Resources, Inc.
ASTM	ASTM International
bgs	below ground surface
COC	chain-of-custody
сРАН	carcinogenic polycyclic aromatic hydrocarbons
DQO	data quality objective
Ecology	Washington State Department of Ecology
EDD	electronic data deliverable
EIM	Environmental Information Management system
EPA	United States Environmental Protection Agency
FS	feasibility study
GeoEngineers	GeoEngineers, Inc.
GPS	global positioning system
HASP	Health and Safety Plan
HAZWOPER	Hazardous Waste Operations and Emergency Response
KM	Kaplan-Meier
LCS	laboratory control sample
LCSD	laboratory control sample duplicate
MDL	method detection limit
mg/kg	milligrams per kilogram
MLLW	mean lower low water
MRL	method reporting limit
MS	matrix spike
MSD	matrix spike duplicate
MTCA	Model Toxics Control Act
NAD	North American Datum
NTU	nephelometric turbidity unit
OSHA	Occupational Safety and Health Administration
PAH	polycyclic aromatic hydrocarbon



PARCC	Precision, Accuracy, Representativeness, Completeness, and Comparability
PID	photoionization detector
Port	Port of Anacortes
PPE	personal protective equipment
ppm	part per million
%D	percent difference
%R	percent recovery
PQL	practical quantitation limit
QA	quality assurance
QC	quality control
RI	remedial investigation
RL	reporting limit
RPD	relative percent difference
Site	Quiet Cove Site
SOP	standard operating procedure
SVOC	semi-volatile organic compound
TEF	toxicity equivalency factor
TEQ	toxicity equivalent (refers to concentration basis)
TRL	target reporting limit
VOC	volatile organic compound
VPC	volatile petroleum compound
WAC	Washington Administrative Code



#### **1.0 INTRODUCTION**

This Compliance Monitoring and Quality Assurance Project Plan (CMP/QAPP) is prepared as an appendix to the Interim Action Work Plan (IAWP) for the Quiet Cove Site (Site). The Site is situated along the southeast shoreline of Guemes Channel at 202 O Avenue (at the intersection of 2nd Street and O Avenue) in Anacortes, Washington. The Interim Action is being implemented by the Port of Anacortes (Port) under the regulatory oversight by Department of Ecology (Ecology) and includes remedial excavation and permitted off-site disposal of Site contamination present within the properties owned by the Port. A summary of site conditions including background, history and nature/extent of contamination, and detailed description of planned Interim Action activities are presented in the IAWP. This plan describes compliance monitoring activities and presents quality assurance (QA) and quality control (QC) requirements applicable to Interim Action activities.

The CMP portions of this document were prepared in accordance with Washington Administrative Code (WAC) 173-340-410 (compliance monitoring requirements of Ecology) to describe the protection, performance and confirmation monitoring that will be completed for the Interim Action.

The QAPP portions of this document were prepared following the Ecology's Guidelines for Preparing Quality Assurance Project Plans for Environmental Studies (Ecology, 2004), United States Environmental Protection Agency (EPA) Requirements for Quality Assurance Project Plans (EPA 2001), Guidance for Quality Assurance Project Plans (EPA, 2002), and EPAs National Functional Guidelines for Inorganic and Organic Superfund Methods Data Review (EPA 2017a and 2017b). This CMP/QAPP presents the objectives, procedures, organization, functions, activities, and specific quality assurance/quality control (QA/QC) activities designed to achieve the data quality objectives (DQOs) established for the project. Environmental measurements will be taken to produce data that are scientifically valid, of known and acceptable quality, and meet established objectives. QA/QC procedures will be implemented so that the precision, accuracy, representativeness, completeness, and comparability (PARCC) of the data generated meet the specified DQOs to the maximum extent possible.

#### **2.0 COMPLIANCE MONITORING**

#### 2.1. Protection Monitoring

Protection monitoring will be completed to confirm that human health and the environment are adequately protected during Interim Action construction.

#### 2.1.1. Worker Health and Safety

Interim Action construction activities will be performed in accordance with the requirements of the Washington Industrial Safety and Health Act (RCW 49.17) and the Federal Occupational Safety and Health Act (29 CFR 1910, 1926). These regulations include requirements that workers are to be protected from exposure to contaminants. A site-specific Health and Safety Plan (HASP) is included as an appendix to the IAWP and addresses protection monitoring requirement for GeoEngineers' personnel. The Port's construction contractor (contractor) will be required to prepare and submit a separate HASP for use by the contractor's personnel.



#### 2.1.2. Environmental Protection

Environmental protection measures consisting of Best Management Practices (BMPs) for stormwater, sediment, drainage, and erosion control; dust and noise control; spill prevention and pollution control; and all other controls needed to protect environmental quality will be implemented. Environmental protection measures including installation, inspection and maintenance necessary for stormwater management, control of surface water runoff, and temporary erosion and sediment control measures will be described by the Contractor prior to commencing construction activities. The minimum standards for environmental protection measures that will be implemented are described in the IAWP. If the Port or Ecology determines that the contractor's environmental protection measures are inadequate to meet the intent of applicable regulations, the contractor will be required to implement additional stormwater runoff, erosion control, or spill prevention and control measures to address the deficiencies.

#### **2.2. Performance Monitoring**

Performance monitoring will be conducted to verify that the Interim Action achieves soil remediation levels and/or to document contaminant concentrations that will be left in place. As described in the IAWP, the Interim Action is being implemented to remove the source of Site contamination and cleanup Port owned properties, and therefore any contamination observed at the property boundaries will be left in place and documented so that it can be addressed as part of the final cleanup action for the Site. Performance monitoring activities will include verification soil sampling and analysis as described below.

#### 2.2.1. Verification Soil Sampling and Analysis

Soil verification samples will be collected by GeoEngineers field personnel from the base and/or sidewalls of the remedial excavation as described in the IAWP. Soil samples from the base of the excavation will be collected at a frequency of one sample per 400 square feet. If the area of the base is less than 400 square feet, a minimum of one base sample will be obtained. Sidewall samples will be collected at a frequency of one sample per 20 linear feet of sidewall along the perimeter of the excavation. Sidewall samples will be collected at a depth where field screening evidence of contamination is the highest. If the sidewall does not have field screening evidence of contamination, the sidewall will be collected immediately above the water table (saturated zone). Field personnel will take photos of the sidewall and base soil sample locations to document visual field screening conditions. One duplicate soil sample will be collected per every 20 parent soil samples collected from excavation limits for QA/QC purposes.

Soil samples will be collected by GeoEngineers' field personnel using a clean pair of nitrile gloves and placed in clean laboratory provided containers for chemical analysis. Reusable sampling equipment (if used) will be decontaminated prior to sample collection at each location. Each sample container will be securely capped, labeled, and placed in a cooler with ice immediately upon collection. Samples for volatile analyses (NWTPH-Gx and EPA Method 8260) will be collected as discrete samples at a minimum of 6 inches below the surface of the base or sidewall in accordance with EPA 5035A sampling methods. Samples for non-volatile analysis will be collected from a minimum of 6 inches below the surface and homogenized in a stainless-steel bowl. The field representative will visually classify the soils in accordance with ASTM International (ASTM) Method D 2488 (Standard Practice for Description and Identification of Soils [Visual Manual Procedure]) and record soil descriptions and other relevant field screening details (e.g., staining, debris, odors, etc.) in the field log. Field screening, decontamination, sample container, labeling, and handling procedures are described in Section 5.



Chemical analysis will be performed at an Ecology accredited laboratory—Analytical Resources, Inc. (ARI) of Tukwila, Washington. Chain-of-custody forms will be used to document the transfer of samples during transport and submittal of samples to the laboratory. The following analysis will be performed on each verification soil samples:

- Gasoline-range petroleum hydrocarbons by Ecology Method NWTPH-Gx;
- Diesel- and heavy oil-range petroleum hydrocarbons by Ecology Method NWTPH-Dx;
- Volatile Organic Compounds (VOCs) including benzene, ethylbenzene, toluene and xylenes (BETX), n-hexane, Methyl tert-butyl ether (MTBE), Ethylene Dibromide (EDB) and Ethylene Dichloride (EDC) by EPA Method 8260;
- Polycyclic Aromatic Hydrocarbons (PAHs) by EPA Method 8270D/SIM; and
- Metals including arsenic, cadmium, chromium, lead and mercury by EPA Method 6000/7000 series.

Table 1 summarizes the analytical methods, sample size, containers, preservation and holding times for above mentioned laboratory analysis. Sufficient volume will be collected for each sample to perform each of the listed analysis. Verification soil samples collected to document contaminant conditions that will be left in place will be analyzed at a standard (7 to 10 days) turn-around-time (TAT). Verification soil samples collected to verify compliance with remediation levels will be analyzed at expedited TAT (2 days) to support decision making in the field concerning any additional excavation that may be required to achieve remediation levels. The result of verification soil samples will be compared to soil remediation levels presented in the IAWP. Data validation will be completed as described in Section 6. The results will be submitted to Ecology's Environmental Information Management (EIM) database following data validation.

#### 2.3. Confirmational (Post-Construction) Monitoring

Confirmational monitoring will be performed after Interim Action remedial excavation and restoration activities completed to evaluate changes in groundwater conditions as a result of the removal of source material/contamination from within Port owned properties. Groundwater monitoring will be performed on a quarterly basis for of four consecutive quarters following the Interim Action. Additional monitoring may be performed following the completion of four quarters based on the results and requirements of Ecology. Groundwater monitoring results will be used in conjunction with other existing data (Data Report; an appendix of IAWP) as part of the Remedial Investigation/Feasibility Study (RI/FS) to evaluate the final cleanup plan for the Site.

Groundwater monitoring activities will be completed at wells located along the shoreline and/or downgradient of the Interim Action area. Currently, it is expected that up to 7 existing/newly installed monitoring wells including MW-1 through MW-4, MW-8, MW-9 and MW-13 will be monitored. The exact number and location of the wells that will be monitored will be determined in consultation with Ecology following the completion of remedial excavation. IAWP identifies existing wells that are planned to be decommissioned and reinstalled and new monitoring wells that are planned to be installed as part of the Interim Action. Groundwater monitoring will include water level measurements and groundwater sampling and analysis as described below.

#### 2.3.1. Water Level Measurements

Water level measurements will be obtained at each monitoring well prior to purging and sample collection. All water levels will be measured using an electronic water level indicator and will be recorded to the nearest 0.01 foot. Measurements will be taken from the top of the well casing.

#### 2.3.2. Groundwater Sampling and Analysis

Groundwater samples will be obtained by GeoEngineers field personnel using low-flow/low-turbidity sampling techniques to minimize the suspension of sediment in the samples. The wells will be purged and groundwater samples will be obtained from the wells using a peristaltic or submersible pump and disposable polyethylene tubing. Groundwater will be purged from the wells at a rate of approximately 0.5 liters per minute. A water quality measuring system (YSI Pro or similar) with a flow-through cell will be used to monitor the following water quality parameters during purging. Water quality parameter monitored will include:

- Electrical conductivity (EC);
- Dissolved oxygen (DO);
- Acidity (pH);
- Salinity;
- Total dissolved solids (TDS);
- Turbidity;
- Oxidation-reduction potential (ORP); and
- Temperature.

Samples will be collected from the wells after these parameters vary by less than 10 percent on three consecutive measurements. The stabilized field measurements will be documented in the field log. Following well purging, the flow-through cell will be disconnected and groundwater samples will be collected in laboratory-prepared containers. Both field-filtered and unfiltered samples for metals analysis will be collected. Reusable sampling equipment (if used) will be decontaminated prior to sample collection at each location. One duplicate soil sample will be collected per monitoring event for QA/QC purposes. Each sample container will be securely capped, labeled, and placed in a cooler with ice immediately upon collection. Decontamination, sample container, labeling, and handling procedures are described in Section 5.

Chemical analysis will be performed at an Ecology accredited laboratory – ARI. Chain-of-custody forms will be used to document the transfer of samples during transport and submittal of samples to the laboratory. The following analysis will be performed on each groundwater sample:

- Gasoline-range petroleum hydrocarbons by Ecology Method NWTPH-Gx;
- Diesel- and heavy oil-range petroleum hydrocarbons by Ecology Method NWTPH-Dx;
- Volatile Organic Compounds (VOCs) including benzene, ethylbenzene, toluene and xylenes (BETX), n-hexane, Methyl tert-butyl ether (MTBE), Ethylene Dibromide (EDB) and Ethylene Dichloride (EDC) by EPA Method 8260;



- Metals including arsenic, cadmium, chromium, lead and mercury by EPA Method 6000/7000 series;
- Total alkalinity by SM 2420 B-97;
- Ferrous iron by SM 3500-Fe B-97;
- Nitrate and sulfate by EPA 300.0;
- Dissolved manganese by EPA 6020A; and
- Dissolved methane by EPA RSK-175.

Table 2 summarizes the analytical methods, sample size, containers, preservation and holding times for laboratory analysis. Sufficient water will be collected from each well to perform each of the listed analysis. Groundwater samples will be analyzed on a standard (7-10 days) TAT. The result of groundwater analysis will be compared to groundwater screening levels presented in the IAWP. Data validation will be completed as described in Section 6. The results will be submitted to Ecology's Environmental Information Management (EIM) database following data validation. If additional compliance groundwater monitoring becomes necessary based on the results of the four initial monitoring events, then the sampling frequency and list of groundwater analysis for additional round of monitoring will be determined based on discussions between the Port and Ecology.

#### **3.0 PROJECT MANAGEMENT AND ORGANIZATION**

The project management and organization elements for the Interim Action including the key personnel, roles and responsibilities of the participants and special training/certification are presented in the following sections.

#### **3.1. Project Organization and Responsibilities**

Key individuals and positions providing QA and QC are summarized in the following table. A description of the responsibilities, lines of authority and communication for the key individuals and positions providing QA and QC is presented below.

Project Role	Name and Organization	Contact Information
Project Coordinator	Brad Tesch Port of Anacortes	360.299.1830 brad.tesch@portofanacortes.com 100 Commercial Ave. Anacortes, WA 98221
Technical Project Manager	John Herzog GeoEngineers	206.406.6431 <u>jherzog@geoengineers.com</u> 2101 4 <sup>th</sup> Avenue, Suite 950 Seattle, Washington 98121
Task Manager	Brian Tracy GeoEngineers	206.239.3250 <u>btracy@geoengineers.com</u> 2101 4 <sup>th</sup> Avenue, Suite 950 Seattle, Washington 98121



Project Role	Name and Organization	Contact Information
Field Coordinator/Field Personnel	Nathan Solomon GeoEngineers	206.518.5141 nsolomon@geoengineers.com 2101 4 <sup>th</sup> Avenue, Suite 950 Seattle, Washington 98121
Health and Safety Manger	Mary Lou Sullivan GeoEngineers	253.722.2425 <u>msullivan@geoengineers.com</u> 1101 Fawcett Avenue, Suite 200 Tacoma, Washington 98402
Data Quality Assurance Leader	Mark Lybeer GeoEngineers	206.278.2674 <u>mlybeer@geoengineers.com</u> 2101 4 <sup>th</sup> Avenue, Suite 950 Seattle, Washington 98121
Laboratory Project Manager	Amanda Volgardsen Analytical Resources Inc.	206.695.6220 <u>amanda.volgardsen@arilabs.com</u> 333 9 <sup>th</sup> Avenue North Seattle, WA 98109

#### 3.1.1. Port of Anacortes Project Coordinator

The Port's project coordinator duties consist of implementing the project approach and tasks, overseeing the project team members during performance of project tasks.

#### 3.1.2. Technical Project Manager

The technical project manager is responsible for fulfilling contractual and administrative control of the project. The technical project manager's duties include defining the project approach and tasks, selecting project team members and establishing budgets and schedules.

The technical project manager's duties also include implementing the project approach and tasks, overseeing project team members during performance of project tasks, adhering to and communicating the status of budgets and schedules to the Port project manager, providing technical oversight, and providing overall production and review of project deliverables.

#### 3.1.3. Task Manager

The task manager is responsible for the daily management of project tasks including providing technical direction to the field staff, produces task specific documents and supporting documents, develops schedules and allocates resources for field tasks, coordinates data collection activities to be consistent with information requirements, supervises the compilation of field data and laboratory analytical results, assures that data are correctly and completely reported, implements and oversees field sampling in accordance with project plan and supervises field personnel. Additionally, the task manger coordinates work with on-site subcontractors, verifies that appropriate sampling, testing, and measurement procedures are followed, coordinates the transfer of field data, sample tracking forms, and log books to the technical project manager for data reduction and validation, and participates in QA corrective actions as required.



#### 3.1.4. Field Coordinator

The field coordinator will lead the field sampling effort for the project, serving as the direct point of contact between the task manager, analytical laboratory and subcontractors; and ensures that the appropriate sampling containers, COC forms and field sampling gear including PPE are available. The field coordinator ensures that data collection activities are consistent with information requirements and to assure that field information is correctly and completely reported for the entire duration of the project. The field coordinator will also coordinate appropriate sampling, testing, and measurement procedures and schedule sample delivery/shipment with the analytical laboratory. The field coordinator will transfer field data and sample tracking forms to the project file and data reduction and validation and participate in QA corrective actions as required.

#### 3.1.5. Field Personnel

Field personnel have the primary responsibility for duties involving field data collection and documentation. Technical/field staff are responsible for:

- Understanding and following the IAWP, CMP/QAPP and HASP.
- Checking all equipment and supplies in advance of field operations.
- Ensuring that samples are properly collected, preserved, labeled, packaged, and shipped.
- Ensuring that all field data are carefully recorded in accordance with the IAWP and supporting documents.
- Following COC procedures and SOPs when they are required.

#### 3.1.6. Health and Safety Manager

The health and safety manager will oversee implementation of health and safety programs and verify that work on the project proceeds in accordance with the site-specific HASP.

#### 3.1.7. Quality Assurance Leader

The quality assurance leader will provide oversight required for the completion of sample analyses for the project and verify, in conjunction with the laboratory manager, that the analytical work is proceeding in accordance with internal laboratory standard practices and the QA/QC guidelines for the project. This person will also oversee completion of data validation activities completed for this project. The quality assurance leader maintains independence from the individual(s) generating the data.

#### 3.1.8. Laboratory Project Manager

The laboratory project manager will fulfill the analytical requirements of this project including being responsible for sample analyses using appropriate analytical laboratory methods. The specific procedures to be used for COC transfer, internal calibrations, laboratory analyses, reporting, preventive instrument maintenance, and corrective action will follow standard protocols.

#### **3.2. Special Training Requirements/Certification**

The Superfund Amendments and Reauthorization Act of 1986 required the Secretary of Labor to issue regulations providing health and safety standards and guidelines for workers engaged in hazardous waste



operations. Occupational Safety and Health Administration (OSHA) regulations (29 CFR 1910.120) require training to provide employees with the knowledge and skills necessary to enable them to perform their jobs safely and with minimum risk to their personal health. All sampling personnel will have completed the 40-hour Hazardous Waste Operations and Emergency Response (HAZWOPER) training course and 8-hour refresher courses, as necessary, to meet OSHA regulations.

#### 4.0 DATA QUALITY OBJECTIVES

The primary Data Quality Objectives (DQO) for this Interim Action is to collect environmental sampling data of known, acceptable, and documentable quality. The specific objectives established for the project are:

- Implement the procedures outlined herein for field sampling, sample custody, equipment operation and calibration, laboratory analysis, and data reporting to ensure consistency and thoroughness of data generated.
- Achieve the level of QA/QC required to produce scientifically valid analytical data of known and documented quality. This will be accomplished by establishing criteria for data precision, accuracy, representativeness, completeness, and comparability, and by evaluating project data against these criteria.

#### **4.1. Chemical Quality Objectives**

The sampling design, field procedures, useable laboratory procedures, and QC procedures established for this project were developed to provide defensible data. Specific factors that may affect data usability include quantitative factors (precision, bias, accuracy, completeness, and reporting limits) and qualitative factors such as representativeness and comparability. The specific DQOs associated with these data quality factors are discussed below. Method-specific DQOs for chemical laboratory analyses are presented in Table 3.

#### 4.1.1. Analytical Detection Limits

Analytical methods have quantitative limitations at a given statistical level of confidence that are often expressed as the method detection limit (MDL). Although results reported near the MDL provide insight for contaminant conditions, quality assurance dictates that analytical methods achieve a consistently reliable level of detection known as the practical quantitation limit (PQL), which is typically demonstrated with the lowest point of a linear calibration. The contract laboratory will provide numerical results for all analytes and report them as detected above the PQL or undetected at the PQL.

The PQLs provided by the Ecology-certified laboratory contract laboratory (ARI) are presented in Table 4 for both soil and groundwater. The PQLs presented in Table 4 are considered target reporting limits (TRLs) because several factors may influence final reporting limits. First, moisture and other physical conditions of samples affect detection limits. Second, analytical procedures may require sample dilutions or other practices to quantify a particular analyte at concentrations above the range of the instrument. The effect is that other analytes could be reported as undetected but at a value higher than a specified TRL. Data users must be aware that high non-detect values, although correctly reported, can bias statistical summaries and careful interpretation is required to correctly characterize subsurface conditions. If PQLs are above the established PQLs in Table 4 there may be a need to re-sample depending on the nature of the PQL adjustment.



#### 4.1.2. Precision

Precision is the measure of mutual agreement among replicate or duplicate measurements of an analyte from the same sample and applies to field duplicates (i.e., split samples), replicate analyses, and duplicate spiked environmental samples (matrix spike duplicates). The closer the measured values are to each other, the more precise the measurement process. Precision error may affect data usefulness. Good precision is indicative of relative consistency and comparability between different samples. Precision will be expressed as the relative percent difference (RPD) for spike sample and field duplicate comparisons of various matrices. The RPD is calculated as:

Where:  $\begin{aligned}
RPD(\%) &= \frac{|D_1 - D_2|}{(D_1 + D_2)/2} X 100, \\
D_1 &= Concentration of analyte in primary sample. \\
D_2 &= Concentration of analyte in duplicate sample.
\end{aligned}$ 

The calculation applies to split samples, replicate analyses, duplicate spiked environmental samples (matrix spike duplicates), and laboratory control duplicates. The RPD will be calculated for samples and compared to the applicable criteria. Precision can also be expressed as the percent difference (%D) between replicate analyses. Project RPD goals for all analyses are presented in Table 3, unless the primary and duplicate sample results are less than 5 times the method reporting limit (MRL), in which case RPD goals will not apply for data quality assessment purposes.

#### 4.1.3. Accuracy

Accuracy is a measure of bias in the analytical process. The closer the measurement value is to the true value, the greater the accuracy. Accuracy is typically evaluated by adding a known spike concentration of a target or surrogate compound to a sample prior to analysis. The detected concentration or percent recovery (%R) of the spiked compound reported in the sample provides a quantitative measure of analytical accuracy. Since most environmental data collected represent single points spatially and temporally rather than an average of values, accuracy is generally more important than precision in assessing the data. In general, if %R values are low, non-detect results may be reported for compounds of interest when in fact these compounds are present (i.e., false negative results), and results for detected compounds may be biased low. The reverse is true when %R values are high. In this case, non-detect values are considered accurate, whereas detected values may be higher than true values.

For this project, accuracy will be expressed as the %R of a known surrogate spike, matrix spike, or laboratory control sample (blank spike), concentration:

 $Recovery(\%R) = \frac{Spiked Result - Unspiked Result}{Known Spike Concentration} X 100$ 

Accuracy (%R) criteria for surrogate spikes, matrix spikes, and laboratory control samples (blank spikes) are presented in Table 3.



#### 4.1.4. Representativeness, Completeness, and Comparability

Representativeness expresses the degree to which data accurately and precisely represent the actual site conditions. Representativeness of the data will be evaluated by:

- Comparing actual sampling procedures to those specified in this document.
- Reviewing analytical results for field duplicates to determine the variability in the analytical results.
- Invalidating non-representative data or identifying data to be classified as questionable or qualitative in nature. Only representative data will be used in subsequent data reduction, validation, and reporting activities.

Completeness establishes whether a sufficient amount of valid measurements were obtained to meet project objectives. The number of samples and results expected establishes the comparative basis for completeness. The completeness goal is 90 percent useable data for the samples/analyses planned. If the completeness goal is not achieved, an evaluation will be performed to determine if the data are adequate to meet study objectives.

Comparability expresses the confidence with which one set of data can be compared to another. Although numeric goals do not exist for comparability, a statement on comparability will be prepared to assess overall usefulness of data sets generated during the project, following the evaluation of precision and accuracy.

#### 4.1.5. Holding Times

Holding times are defined as the time between sample collection and extraction, sample collection and analysis, or sample extraction and analysis. Recommended holding times are presented in Tables 1 and 2 for soil and groundwater, respectively. If the analysis of an archived sample is required but the sample exceeds the respective holding time, either discard the sample and collect a new representative sample for analysis and/or consult with Ecology to determine if the sample may still be used.

#### 4.1.6. Quality Control Blank Samples

According to the National Functional Guidelines for Organic Data Review (EPA 2017b), "The purpose of laboratory (or field) blank analysis is to assess the existence and magnitude of contamination resulting from laboratory (or field) activities. The criteria for evaluation of blanks apply to any blank associated with the samples (e.g., method blanks, instrument blanks, trip blanks, and equipment blanks)." Trip blanks are placed with samples during shipment; method blanks are created during sample preparation and follow samples throughout the analysis process.

Analytical results for QC blanks will be interpreted in general accordance with EPA's National Functional Guidelines for Inorganic (EPA 2017a) and Organic Data (EPA 2017b) Review and professional judgment. QC blank samples are discussed further in Section 5.8.2.

#### **5.0 DATA GENERATION AND ACQUISITION**

The data generation and acquisition elements for the CMP/QAPP (as detailed below) address aspects of the project design and implementation including the appropriate methods for measurement and analysis, data collection or generation, data handling, and how QC activities are employed and properly documented.



Sampling methods including field documentation, sampling, and decontamination procedures are also discussed below.

#### 5.1. Surveying

Surveying at the site will be performed referencing vertical datum - North American Vertical Datum of 1988 (NAVD 88) and horizontal datum – North American Datum of 1983 (NAD 83). Mean Lower Low Water (MLLW) for the Site is 0.66 feet above NAVD 88 (i.e. MLLW = NAVD 88 + 0.66). The contractor will perform excavation and post-construction as-built survey to document excavation limits and post-construction site conditions including locations of newly installed monitoring wells.

#### **5.2. Field Screening Procedures**

The potential presence of contamination in soil samples will be evaluated using field screening techniques. Field screening results will be recorded on the field logs and the results will be used as a general guideline to delineate areas of possible contamination. In addition, screening results will be used as a basis for selecting soil samples for chemical analysis. The following screening methods will be used: 1) visual screening; 2) water sheen screening; and 3) headspace vapor screening.

#### 5.2.1.1. Visual Screening

The soil will be observed for unusual color and stains and/or odor indicative of possible contamination. Field personnel will take photos throughout excavation activities to document visual conditions throughout excavation activities.

#### 5.2.1.2. Water Sheen Screening

This is a qualitative field screening method that can help identify the presence or absence of petroleum hydrocarbons. A portion of the soil sample will be placed in a pan containing distilled water. The water surface will be observed for signs of sheen. The following sheen classifications will be used:

Classification	Identifier	Description
No Sheen	(NS)	No visible sheen on the water surface
Slight Sheen	(SS)	Light, colorless, dull sheen; spread is irregular, no rapid; sheen dissipates rapidly
Moderate Sheen	(MS)	Light to heavy sheen; may have some color/iridescence; spread is irregular to flowing, may be rapid; few remaining areas of no sheen on the water surface
Heavy Sheen	(HS)	Heavy sheen with color/iridescence; spread is irregular to flowing, may be rapid; few remaining areas of no sheen on the water surface

#### 5.2.1.3. Headspace Vapor Screening

This is a semi-quantitative field screening method that can help identify the presence or absence of volatile organic compounds (VOCs) in soil samples. A portion of the soil sample will be placed in a resealable plastic bag. The bag will then be sealed capturing air in the bag. The bag is then shaken gently to expose the soil to the air trapped in the bag. The bag will remain closed for approximately 5 minutes at ambient temperature before the headspace vapors are measured. Vapors present within the sample bag's headspace will be measured by inserting the probe of a photoionization detector (PID) through a small opening in the bag, taking care not to clog the probe with soil. The maximum PID reading (in parts per million [ppm]) and the ambient air temperature will be recorded on the field log for each sample. The PID



will be calibrated to 100 ppm isobutylene each day prior to soil sampling. No soil sample used for headspace screening will be submitted to the laboratory for chemical analysis.

#### **5.3. Decontamination Procedures**

Soil samples will be collected using excavation equipment (i.e., backhoe or excavator), hand tools including stainless steel spoons and/or directly from the excavation limits using clean pair of nitrile gloves. Groundwater samples will be collected from monitoring wells using submersible or peristaltic pumps and low-flow sampling procedures.

Reusable sampling equipment that comes in contact with soil or groundwater will be decontaminated before each use. Decontamination procedures for this equipment will consist of the following:

- 1. Washing with a brush and non-phosphate detergent solution (e.g., Liqui-Nox and distilled water);
- 2. Rinsing with distilled water; and
- 3. Wrapping or covering the decontaminated equipment with aluminum foil. Field personnel will limit cross-contamination by changing gloves between sampling locations.

Drilling equipment (auger, soil sampler, direct-push barrel) that comes into contact with soil will be decontaminated before each use. Decontamination procedures for this equipment will consist of the following:

- 1. Washing with pressurized hot water;
- 2. Wash with brush and non-phosphate detergent solution; and
- 3. Rinse with potable water.

Wash water used to decontaminate equipment will be collected and stored on-site in 55-gallon drums.

#### 5.4. Sample Containers, Labeling, Handling and Custody

#### 5.4.1. Sample Containers and Labeling

The Field Coordinator will establish field protocol to manage field sample collection, handling and documentation. Soil and groundwater samples will be placed in appropriate laboratory-prepared containers. Sample containers and preservatives are listed in Tables 1 and 2 for soil and groundwater, respectively.

Sample containers will be labeled with the following information at the time of sample collection:

- Project name and number
- Type of sample preservative used (where applicable)
- Sample name, which will include a reference to date and sampling depth (if applicable)
- Date and time of collection

The sample collection activities will be noted in the field log books. The Field Coordinator will monitor consistency between sample containers/labels, field log books and COC forms.



#### 5.4.2. Sample Storage

Samples will be placed in a cooler with ice after they are collected. The objective of the cold storage will be to attain a sample temperature of 2 to 6 degrees Celsius. Holding times (Tables 1 and 2 for soil and groundwater, respectively) will be observed during sample storage.

#### 5.4.3. Sample Shipment

Samples will be transported and delivered to the analytical laboratory in the sample coolers. The samples will either be transported by field personnel, laboratory personnel or by courier service. The Field Coordinator will ensure that the cooler has been properly secured using clear plastic tape and custody seals.

#### 5.4.4. Chain-of-Custody Records

Field personnel are responsible for the security of samples from the time the samples are collected until the samples have been received by the courier service or laboratory personnel. A COC form will be completed for each group of samples being shipped to the laboratory. Information to be included on the COC form includes:

- Project name and number;
- Sample identification numbers;
- Date and time of sampling;
- Sample matrix (soil and groundwater), preservative, and number of containers for each sample;
- Analyses to be performed;
- Names of sampling personnel;
- Project manager name and contact information including phone number; and
- Shipping information including shipping container number, if applicable.

The original COC form will be signed by a member of the field team. Field personnel will retain copies and place the original and remaining copies in a plastic bag. The plastic bag containing the COC form will be placed in the cooler before sealing the cooler for transport to the laboratory.

#### 5.4.5. Laboratory Custody Procedures

The laboratory will follow their standard operating procedures (SOPs) to document sample handling from time of receipt (sample log-in) to reporting. Documentation will include, at a minimum, the analyst's name or initials, time and date.

#### 5.5. Disposal of Investigation-Derived Materials

#### 5.5.1. Soil and Water Generated during Well Installation/Monitoring

Soil cuttings from borings completed during the post-construction groundwater monitoring well installation, development and purge water removed from the monitoring wells, and decontamination water will be placed in labeled and sealed 55-gallon drums. The drums will be temporarily stored on Site at a secure



location pending receipt of analytical results and off-site disposal at a permitted facility. Each drum will be labeled with the following information:

- Material/media (i.e., soil, water, etc.) contained in the drum;
- Source of the material in the drum (i.e., investigation locations and depths where appropriate);
- Date material was generated; and
- Name and telephone number of GeoEngineers contact person.

#### 5.5.2. Disposition of Incidental Waste

Incidental waste generated during sampling activities includes items such as gloves, plastic sheeting, sample tubing, paper towels and similar expended and discarded field supplies. These materials are considered *de minimis* and will be disposed of in a local trash receptacle or county disposal facility.

#### **5.6. Field Documentation**

The field staff will be responsible for documenting field activities including sampling in an all-weather (e.g. "Rite-in-the-Rain") field notebook and/or on field logs, and by producing a draft technical field report at the end of each day of sampling. The field staff will also be responsible for implementing field QA/QC procedures in accordance with the methods outlined in this document and general good practice sampling protocols. These procedures include recording and documenting relevant and appropriate information regarding project activities, sampling methods and data collected during performance of field activities at each sample location.

The following general guidelines should be followed in documenting fieldwork:

- Documentation will be maintained in a dedicated field notebook and on field forms, as applicable.
- Notebook documentation will be completed in waterproof ink or permanent marker and written errors will be crossed out with a single line.

Field notebooks will include records of pertinent activities completed on site including sampling. Field notebooks will be bound books with sequentially numbered pages. The books will remain in the custody of the Field Coordinator/Personnel until project completion, after which, the books will be kept in the project files. The field notebook and forms will be maintained on a real-time basis and will include, where applicable and appropriate, the following information:

- Date, time of specific activities and weather conditions.
- Names of all personnel on the site, including visitors.
- Specific details regarding sampling activities, including sampling locations, type of sampling, depth, and sample numbers.
- Specific problems and resolutions.
- Identification numbers of monitoring instruments used that day.
- Chain-of-custody details, including sample identification numbers.

A draft field report will be prepared upon completion of field activities each day. Field data that was recorded in the notebooks and field forms will be used to complete the field report. The field report will be used to document construction, sampling, and monitoring activities, sampling and Site personnel, and weather conditions, as well as decisions, corrective actions, and/or modifications to the project plans and procedures discussed in this report. Draft field reports will be reviewed by the Technical Project Manager on a daily basis. The draft field report will be finalized following review by the Task Manager and/or Technical Project Manager on a weekly basis and kept in the project files.

#### **5.7. Analytical Methods**

Samples and QC samples shall be analyzed following the analytical methods listed in Tables 1 and 2 for soil and groundwater, respectively, using laboratory instruments prescribed in the methods. The analytical methods must meet the technical acceptance criteria specified by the method prior to the analysis of environmental samples. Samples that are not analyzed initially (i.e., placed on "hold") will be stored at the laboratory for at least 6 months, and will be disposed of by the laboratory following this period. Samples to be analyzed initially will be analyzed within proper holding times, which are listed in Tables 1 and 2 for soil and groundwater, respectively.

The laboratory is required to comply with their current written standard operating procedures. All laboratory personnel will be responsible for reporting problems that may compromise the quality of the data to the laboratory project manager. A narrative describing the anomaly, the steps taken to identify and correct it and the treatment of the relevant sample batch (i.e., recalculation, reanalysis, re-extraction) will be submitted with the data package.

#### **5.8. Quality Control**

Quality control activities that will be implemented for each sampling, analysis or measurement technique are summarized in Table 3. Formulas for calculating QC statistics are provided in Section 4.1.

The laboratory will maintain and implement documented QA/QC procedures. The laboratory QA/QC program will provide the following:

- Procedures that must be followed for certifying the precision and accuracy of the analytical data generated by the laboratory.
- Documentation of each phase of sample handling, data acquisition, data transfer, report preparation, and report review.
- Accurate and secure storage and retrieval of samples and data.
- Detailed instructions for performing analyses and other activities affecting the quality of analytical data generated by the laboratory.
- Appropriate management-level review and approval of procedures, revisions to procedures, and control of procedures in such a way so that laboratory personnel that require specific procedures have access to them.



#### 5.8.1. Field Quality Control

Field QC samples serve as a control and check mechanism to monitor the consistency of sampling methods and the potential influence of off-Site factors on project samples. Examples of off-Site factors include airborne VOCs and contaminants that may be present in potable water used during drilling activities.

#### 5.8.1.1. Field Duplicates

In addition to replicate analyses performed in the laboratory, field duplicates also serve as measures for precision. Field duplicates measure the precision and consistency of laboratory analytical procedures and methods, as well as the consistency of the sampling techniques used by field personnel. Under ideal field conditions, field duplicates are created by thoroughly mixing a volume of the sample matrix, placing aliquots of the mixed sample in separate containers, and identifying one of the aliquots as the primary sample and the other as the duplicate sample. The frequency at which field duplicate samples will be collected is identified under Section 2.0.

#### 5.8.1.2. Trip Blanks

Trip blanks consist of samples of reagent water that accompany samples to be analyzed for VOCs during sample storage in coolers and transport to the laboratory. They are used to assess potential contamination of samples during collection and transport due to the presence of VOCs in ambient air. Trip blanks will be analyzed on a one per cooler basis containing soil and/or groundwater sample for VOC analysis.

#### 5.8.2. Laboratory Quality Control

Laboratory QC procedures will be evaluated through a formal data quality assessment process. The analytical laboratory will follow standard analytical method procedures that include specified QC monitoring requirements. These requirements will vary by method, but generally include:

- Method blanks
- Internal standards
- Instrument calibrations
- Matrix spike/matrix spike duplicates (MS/MSD)
- Laboratory control samples/laboratory control sample duplicates (LCS/LCSD)
- Laboratory replicates or duplicates
- Surrogate spikes

#### 5.8.2.1. Laboratory Blanks

Laboratory procedures utilize several types of blanks, but the most commonly used blanks for QC monitoring are method blanks. Method blanks are laboratory QC samples that consist of either a soil-like material having undergone a contaminant destruction process, or reagent (contaminant-free) water. Method blanks are extracted and analyzed with each batch of environmental samples undergoing analysis. Method blanks are particularly useful during volatiles analysis since VOCs can be transported in the laboratory through the vapor phase. If a substance is detected in a method blank, then one (or more) of the following occurred:

Sample containers, measurement equipment, and/or analytical instruments were not properly cleaned and contained contaminants.



- Reagents used in the process were contaminated with a substance(s) of interest.
- Volatile substances in ambient laboratory air with high solubility or affinities toward the sample matrix contaminated the samples during preparation or analysis.

It is difficult to determine which of the above scenarios took place if blank contamination occurs. However, it is assumed that the conditions that affected the blanks also likely affected the project samples. If target analytes are detected in method blanks, data validation guidelines assist in determining which substances in project samples are considered "real," and which ones are attributable to the analytical process. Furthermore, the guidelines state, "...there may be instances where little or no contamination was present in the associated blank, but qualification of the sample is deemed necessary. Contamination introduced through dilution water is one example".

#### 5.8.2.2. Calibrations

Several types of instrument calibrations are used, depending on the analytical method, to assess the linearity of the calibration curve and assure that the sample results reflect accurate and precise measurements. The main calibrations used are initial calibrations, daily calibrations, and continuing calibration verification.

#### 5.8.2.3. Matrix Spike/Matrix Spike Duplicates (MS/MSD)

MS/MSD samples are used to assess influences or interferences caused by the physical or chemical properties of the sample itself. For example, extreme pH can affect the results for SVOCs. Or, the presence of a particular compound may interfere with accurate quantitation of another analyte. MS/MSD data is reviewed in combination with other QC monitoring data to determine matrix effects. In some cases, matrix effects cannot be determined due to dilution and/or high levels of related substances in the sample. A matrix spike is evaluated by spiking a project sample with a known amount of one or more of the target analytes, ideally at a concentration that is 5 to 10 times higher than the sample result. A percent recovery is then calculated by subtracting the un-spiked sample result from the spiked sample result, dividing by the known concentration of the spike, and multiplying by 100.

MS/MSD samples will be analyzed at a frequency of one MS/MSD per sample set or batch. The samples for the MS/MSD analyses should be collected from a boring or sampling location that is believed to have only low-level contamination. The MS/MSD sample location will be chosen to be as similar in geologic and contaminated conditions as possible to other samples collected in the batch based on field screening. A sample from an area of low-level contamination is needed because the objective of MS/MSD analyses is to determine the presence of matrix interferences, which can best be achieved with low levels of contaminants. Additional sample volume will be collected for the MS/MSD analyses as required by the laboratory.

#### 5.8.2.4. Laboratory Control Sample/ Laboratory Control Sample Duplicates (LCS/LCSD)

Also known as blank spikes, LCSs are similar to MS samples in that a known amount of one or more of the target analytes are spiked into a prepared sample medium, and a percent recovery of the spiked substances is calculated. The primary difference between LCS and MS samples is that the LCS uses a contaminant-free sample medium. For example, reagent water is typically used for LCS water analyses. The purpose of an LCS is to help assess the overall accuracy and precision of the analytical process including sample preparation, instrument performance, and analyst performance.

#### 5.8.2.5. Laboratory Replicates/Duplicates

Laboratories utilize MS/MSDs, LCS/LCSDs, and/or replicates to assess precision. Replicates are a second analysis of a field-collected environmental sample. Replicates can be split at varying stages of the sample preparation and analysis process; they most commonly consist of a second analysis on the extracted media.

#### 5.8.2.6. Surrogate Spikes

Surrogate spikes are used to verify proper extraction procedures and the accuracy of the analytical instrument. Surrogates are substances with characteristics similar to the target analytes. A known concentration of surrogate is added to the project sample and passed through the instrument, and percent recovery is calculated. Each surrogate used has acceptance limits (i.e., an acceptable range) for percent recovery. If a surrogate recovery is low, sample results may be biased low and depending on the recovery value, a possibility of false negatives may exist. Conversely, when recoveries are above the specified acceptance limits, a possibility of false positives exist, although non-detect results are considered accurate.

#### 5.9. Instrument/Equipment Testing, Inspection, and Maintenance

#### 5.9.1. Field Instrumentation

If field instruments are used, calibration and calibration checks will be performed to facilitate accurate and reliable field measurements. The calibration of the instruments will be checked and adjusted as necessary in general accordance with manufacturers' recommendations. Methods and frequency of calibration checks and instrument maintenance will be based on the type of instrument, stability characteristics, required accuracy, intended use, and environmental conditions.

#### 5.9.2. Laboratory Instrumentation

For chemical analytical testing, calibration procedures will be performed in general accordance with the analytical methods used and the laboratory's Standard Operating Procedures (SOPs). Calibration documentation will be retained at the laboratory for a period of 6 months.

#### 5.10. Laboratory Data Reporting and Deliverables

Laboratories will report data in formatted hardcopy and electronic form to the technical project manager, task manager and QA leader. Upon completion of analyses, the laboratory will prepare electronic deliverables for data packages in accordance with the specifications in the agreed-upon *Special Conditions for Lab Analysis* document. The laboratory will provide electronic data deliverables (EDDs) within 2 business days after GeoEngineers' receipt of printed-copy analytical results, including the appropriate QC documentation. GeoEngineers will establish EDD requirements with the contract laboratory.

Analytical laboratory measurements will be recorded in standard formats that display, at a minimum, the client/field sample identification, the laboratory sample identification, reporting units, analytical methods, analytes tested, analytical results, extraction and analysis dates, quantitation limits, and data qualifiers. Each sample delivery group will be accompanied by sample receipt forms and a case narrative identifying data quality issues.

#### 6.0 DATA REDUCTION AND ASSESSMENT PROCEDURES

The process for generating and checking data, as well as the process for producing reports for field and analytical laboratory data, are summarized in the following sections.



#### 6.1. Data Reduction

Data reduction involves the conversion or transcription of field and analytical data to a useable format. The laboratory personnel will reduce the analytical data for review by the QA leader, task manager and technical project manager. This will involve both hard-copy forms and EDDs. Both forms of data will be compared with each other to verify that the data are reliable and error-free.

#### 6.2. Review of Field Documentation and Laboratory Receipt Information

Documentation of field sampling data will be reviewed periodically for conformance with project QC requirements described in this document. At a minimum, field documentation will be checked for proper documentation of the following:

- Sample collection information (date, time, location, matrices, etc.);
- Field instruments used and calibration data;
- Sample collection protocol;
- Sample containers, preservation and volume;
- Field QC samples collected at the frequency specified;
- Chain-of-custody protocols; and
- Sample shipment information.

Sample receipt forms provided by the laboratory will be reviewed for QC exceptions. The final laboratory data package will describe (in the case narrative) the effects that any identified QC exceptions have on data quality. The laboratory will review transcribed sample collection and receipt information for correctness prior to delivering the final data package.

#### 6.3. Data Verification/Validation

Project decisions, conclusions, and recommendations will be based upon verified (validated) data. The purpose of data verification is to ensure that data used for subsequent evaluations and calculations are scientifically valid, of known and documented quality, and legally defensible. Field data verification will be used to eliminate data not collected or documented in accordance with the protocols specified in the IAWP and this document. Laboratory data verification will be used to eliminate data not obtained using prescribed laboratory procedures.

The QA leader will validate data collected during the supplemental investigation to ensure that the data are valid and usable. At a minimum, a Stage 2B validation will be performed on the Interim Action data in general conformance with EPA functional guidelines for data validation (EPA 2004; and EPA 2008). At a minimum, the following items will be reviewed to verify the data as applicable:

- Documentation that a final review of the data was completed by the laboratory QA coordinator;
- Documentation of analytical and QC methodology;
- Documentation of sample preservation and transport;
- Sample receipt forms and case narratives; and
- The following QC parameters:



- Holding times and sample preservation
- Method blanks
- MS/MSDs
- LCS/LCSDs
- Surrogate spikes
- Duplicates/replicates

When sample analytical data are received from the analytical laboratory, they will undergo a QC review by the QA leader. The accuracy and precision achieved will be compared to the laboratory's analytical control limits. Example control limits are presented in Table 3. Calculations of RPDs will follow standard statistical conventions and formulas as presented in in this document. Additional specifications and professional judgment by the QA leader may be incorporated when appropriate data from specific matrices and field samples are available.

A data quality assessment will be prepared to document the overall quality of the data relative to the DQOs. The major components of the data quality assessment are as follows:

- Data Validation Summary: Summarizes the data validation results for all sample delivery groups by analytical method. The summary identifies any systematic problems, data generation trends, general conditions of the data, and reasons for any data qualification.
- QC Sample Evaluation: Evaluates the results of QC sample analyses, and presents conclusions based on these results regarding the validity of the project data.
- Assessment of DQOs: An assessment of the quality of data measured and generated in terms of accuracy, precision, and completeness relative to objectives established for the project.
- Summary of Data Usability: Summarizes the usability of data, based on the assessment performed in the three preceding steps.

The data quality assessment will help to achieve an acceptable level of confidence in the decisions that are to be made based upon the project data. The project analytical data will be submitted to Ecology's Environmental Information Management (EIM) system within 60 days after the data quality assessment is completed.

#### 6.4. Calculating Chemical Sums

The following guidelines will be used to calculate chemical sums:

- Total benzofluoranthenes represents the sum of detected concentrations of the b, j, and k isomers of benzofluoranthenes (WAC 173-204-562(2)(k)). In some cases, the testing laboratory may report the total benzofluoranthenes concentration rather than concentrations of individual compounds since they may not be able to resolve all three isomers.
- Total cPAHs will be calculated using the toxicity equivalent (TEQ) approach in accordance with WAC 173-340-708(8)(e). Total cPAH TEQs will be calculated using toxicity equivalency factor (TEF) values referenced from Model Toxics Control Act (MTCA) Table 708.2 (WAC 173-340-900). For non-detect results, one-half the PQL will be used in the TEQ calculations.



For the summation of chemical totals, any "U" qualified data, which may be data reported at the PQL, the MDL, or the reporting limit (RL), represent non-detects. For the calculations, no distinction is made between these different types of detection limits, and any "U" qualified data are treated as "non-detects". The following guidelines will be used for reporting and summing non-detects for benzofluoranthenes:

- When all chemicals in a group are non-detect, only the single highest individual chemical quantitation limit in a group will be reported and appropriately qualified.
- If some concentrations were detected and others are not, only the detected concentrations are included in the sum.

Estimated values between the method detection limit and the laboratory reporting limit (i.e. "J" qualified results) will be included in the summation at face value and the sum will also be qualified as estimated with a "J" qualifier. Results that are qualified as estimates with "J" qualifiers through data validation, will also be handled in the same manner.

For calculating total cPAH TEQ, the sum will be calculated using a substitution at one-half the detection limit (i.e., n=1/2). However, using this alternative may result in generated sums that are estimates with unknown bias and precision. Therefore, these estimates will be bounded by reporting sums using a substitution of the detection limit at n=0 and n=1.

#### 7.0 LIMITATIONS

We have prepared this Compliance Monitoring and Quality Assurance Project Plan (CMP/QAPP) for use by the Port of Anacortes during the Interim Action at the Quiet Cove Site located at 202 O Avenue in Anacortes, Washington. Within the limitations of scope, schedule and budget, our services have been executed in accordance with generally accepted environmental science practices in this area at the time this report was prepared. No warranty or other conditions, express or implied, should be understood.

Any electronic form, facsimile or hard copy of the original document (email, text, table, and/or figure), if provided, and any attachments are only a copy of the original document. The original document is stored by GeoEngineers, Inc. and will serve as the official document of record.

#### 8.0 REFERENCES

- U.S. Environmental Protection Agency (EPA) 2001, "EPA Requirements for Quality Assurance Project Plans, EPA QA/R-5" EPA-240/B-01/003, Office of Environmental Information, Washington, DC, dated March 2001.
- U.S. EPA 2002, "Guidance for Quality Assurance Project Plans, EPA QA/G-5," EPA-240/R-02/009, Office of Environmental Information, Washington, DC, dated December 2002.
- U.S. EPA 2017a, "National Functional Guidelines for Inorganic Superfund Methods Data Review." OLEM 9355.0-135, EPA 540-R-2017-001, Office of Superfund Remediation and Technology Innovation (OSRTI), Washington, DC, dated January 2017.



U.S. EPA 2017b "National Functional Guidelines for Organic Superfund Methods Data Review." OLEM 9355.0-136, EPA 540-R-2017-002, Office of Superfund Remediation and Technology Innovation (OSRTI), Washington, DC, dated January 2017.

Washington State Department of Ecology (Ecology), 2004, "Guidelines for Preparing Quality Assurance Project Plans for Environmental Studies," dated July 2004 and revised December 2016.



# Table 1 Soil Sample Test Methods, Sample Size, Containers, Preservation and Holding Times

Quiet Cove Site

Anacortes, Washington

Laboratory Analysis	Analytical Method	Minimum Sample Size	Sample Container	Sample Preservation	Holding Time <sup>1</sup>
Gasoline-Range Hydrocarbons	NWTPH-Gx	5 g	Two 40mL glass vial (VOA)	Cool ≤6 °C, 5mL MeOH	14 days to extraction/analysis
Diesel- and Oil-Range Hydrocarbons	NWTPH-Dx	100 g	8-oz amber glass WM with Teflon-lined lid	Cool ≤6 ° C	14 days to extraction/analysis
VOCs (BTEX, n-Hexane, MTBE, EDB and EDC)	EPA 8260	5 g	Three 40mL glass vial (VOA)	Cool ≤6°C Two VOAs - Sodium Bisulfate One VOA - Methanol	14 days to extraction/analysis
PAHs	EPA 8270/SIM	100 g	8-oz glass WM with Teflon-lined lid	Cool ≤6 ° C	14 days to extraction, 40 days from extraction to analysis
Metals (As, Cd, Cr, Pb and Hg)	EPA 6010/6020/ 7470/7471	100 g	4-oz glass WM with Teflon-lined lid	Cool ≤6°C	180 days/28 days for Mercury

PAHs = Polycyclic Aromatic Hydrocarbons

SIM = selected ion mode

g = gram mL = milliliter

oz. = ounce

WM = wide mouth

#### Notes:

<sup>1</sup> Holding times are based on elapsed time from date of collection.

NWTPH = Northwest total petroleum hydrocarbons

Gx = gasoline-range extended

Dx = diesel-range extended

EPA = Environmental Protection Agency

VOC = volatile organic compound

BTEX = Benzene, Toluene, Ethylbenzene, Xylenes

MTBE = Methyl tert-butyl ether

EDB = Ethylene Dibromide

EDC = Ethylene Dichloride



## Table 2

Groundwater Sample Test Methods, Sample Size, Containers, Preservation and Holding Times

Quiet Cove Site

Anacortes, Washington

Laboratory Analysis	Analytical Method	Minimum Sample Size	Sample Container	Sample Preservation	Holding Time <sup>1</sup>
Gasoline-Range Hydrocarbons	NWTPH-Gx	40 mL	Two 40mL glass vial (VOA)	Cool $\leq$ 6 C, HCl to pH < 2	14 days to extraction/analysis
Diesel- and Oil-Range Hydrocarbons	NWTPH-Dx	500 mL	Two 500 mL amber glass with Teflon-lined lid	Cool ≤6 C	7 days to extraction 40 days from extraction to analysis
VOCs (BTEX, n-Hexane, MTBE, EDB and EDC)	EPA 8260	40 mL	Three 40mL glass vial (VOA)	Cool ≤6 C, HCl to pH < 2	14 days to extraction/analysis
PAHs	EPA 8270/SIM	500 mL	Two 500mL amber glass with Teflon-lined lid	Cool ≤6 ° C	7 days to extraction, 40 days from extraction to analysis
Metals (As, Cd, Cr, Pb and Hg)	EPA 6010/6020/200.8 7470/7471	100mL mL	500mL HDPE bottle	Cool ≤6 C, HNO <sub>3</sub> - pH<2 (Dissolved metals preserved after filtration)	180 days/28 days for Mercury
Total Alkalinity	SM 2420	250 mL	One 500 mL HDPE bottle	Cool ≤6 C, No Headspace	14 days to extraction/analysis
Ferrous Iron	SM 3500-Fe	250 mL	One 250 mL amber glass with Teflon-lined lid	Cool ≤6 ° C HCl preserved	24-hrs to extraction/analysis
Sulfate/Nitrate	EPA 300.0	50 mL	One 500 mL HDPE	Cool ≤6°C No Headspace	28 days to extraction/analysis SO4, 48 hours to extraction/analysis NO3
Dissolved Manganese	EPA 6020A	100 mL	One 500 mL HDPE	Cool ≤6°C HNO <sub>3</sub> preservative if field filtered, unpreserved if not field filtered	6 months to extraction/analysis
Dissolved Methane	EPA RSK-175	40 mL	Two 40mL glass vial (VOA)	Cool ≤6°C	14 days to extraction/analysis

#### Notes:

<sup>1</sup>Holding times are based on elapsed time from date of collection.

NWTPH = Northwest total petroleum hydrocarbons

Gx = gasoline-range extended

Dx = diesel-range extended

PAHs = Polycyclic Aromatic Hydrocarbons SIM = selected ion mode



EPA = Environmental Protection Agency VOC = volatile organic compound BTEX = Benzene, Toluene, Ethylbenzene, Xylenes MTBE = Methyl tert-butyl ether EDB = Ethylene Dibromide EDC = Ethylene Dichloride L = liter mL = milliliter HCl = hydrocloric acid HDPE = high density polyethylene HNO<sub>3</sub> = nitric acid



## Table 3

### Measurement Quality Objectives Quiet Cove Site Anacortes, Washington

Laboratory	Sample	ry Control e (LCS) mits <sup>1,2</sup>		pike (MS) mits <sup>1,2</sup>	-	tandard (SS) nits <sup>1,2,3</sup>	or Lab D	nte Samples Duplicate mits <sup>4</sup> (%)	Sam	uplicate iples mits <sup>4</sup> (%)
Analysis	Soil	Water	Soil	Water	Soil	Water (Default liq-liq Extraction)	Soil	Water	Soil	Water
Gasoline-Range Hydrocarbons	70-121	72-128	28-162	72-128	74-152	72-133	≤30	≤30	≤30	≤30
Diesel- and Heavy Oil-Range Hydrocarbons	63-120	56-120	63-120	56-120	50 - 150	50 - 150	≤30	≤30	≤30	≤30
VOCs (BTEX, n-Hexane, MTBE, EDB and EDC)	Laboratory Specific Control Limits <sup>1</sup>	80 - 149 (EDC-d4) 80 - 120 (All Other Surrogates)	80 - 129 (EDC-d4) 80 - 120 (All Other Surrogates)	≤30	≤30	≤30	≤30			
PAHs	Laboratory Specific Control Limits <sup>1</sup>	Laboratory Specific Control Limits <sup>1</sup>	Laboratory Specific Control Limits <sup>1</sup>	≤30	≤30	≤30	≤30			
Metals (As, Cd, Cr, Pb and Hg)	80 - 120	80 - 120	75 - 125	75 - 125		-	≤20	≤20	≤20	≤20
Total Alkalinity	90-110	90-110					≤20	≤20	≤20	≤20
Ferrous Iron		90-110		75-125				≤20		≤20
Sulfate/Nitrate		90-110		75-125				≤20		≤20
Dissolved Manganese		80-120		75-125				≤20		≤20
Dissolved Methane		80-120		80-120		88-120 (Ethene) 72-122 (Propane)		≤30		≤30

#### Notes:

<sup>1</sup>Compound-specific ranges will be provided by the laboratory when contracted.

<sup>2</sup>Percent recovery limits are expressed as ranges based on laboratory control limits. Limits will vary for individual analytes.

<sup>3</sup>Individual surrogate recoveries are compound-specific.

<sup>4</sup>RPD control limits are only applicable if the primary and duplicate sample concentrations are greater than 5 times the method reporting limit (MRL). For results less than 5 times the MRL, the difference between the primary and duplicate samples must be less than 2X the MRL for soil and 1X the MRL for waters.

EPA = U.S. Environmental Protection Agency

VOCs = Volatile Organic Compounds

BTEX = Benzene, Toluene, Ethylbenzene, Xylenes

MTBE = Methyl tert-butyl ether

EDB = Ethylene Dibromide

EDC = Ethylene Dichloride

PAHs = Polycyclic Aromatic Hydrocarbons

As = Arsenic; Cd = Cadmium; Cr = Chromium (total); Pb = Lead; Hg = Mercury

RL = Reporting Limit

RPD = Relative percent difference

MS = Matrix spike



## Table 4

#### Method Analysis and Target Reporting Limits for Soil and Groundwater

Quiet Cove Site

Anacortes, Washington

				Practical Quantitation Limit <sup>2</sup> (PQL)		
Laboratory	CAS	Analytical		Groundwater		
Analysis	Number <sup>1</sup>	Method	(mg/kg)	(µg∕L)		
Petroleum Hydrocarbons			1	1		
Gasoline-Range	8006-61-9	NWTPH-Gx	5	100		
Diesel-Range	68334-30-5	NWTPH-DX	5	100		
Heavy Oil-Range	30109	NWTPH-DX	10	200		
Volatile Organic Compounds (VOCs)			-	1		
Benzene	71-43-2	EPA 8260	0.05	0.2		
Ethylbenzene	100-41-4	EPA 8260	0.05	0.2		
Toluene	108-88-3	EPA 8260	0.05	0.2		
Xylenes	1330-20-7	EPA 8260	0.05	0.2		
n-Hexane	110-54-3	EPA 8260	0.001	0.2		
Methyl t-Butyl Ether (MTBE)	1634-04-4	EPA 8260	0.001	0.5		
Ethylene Dibromide (EDB)	106-93-4	EPA 8260	0.001	0.2		
Ethylene Dichloride (EDC)	107-06-2	EPA 8260	0.001	0.2		
Non-carcinogenic Polycyclic Aromatic Hydro	carbons (PAHs)					
1-Methylnaphthalene	90-12-0	EPA 8270-SIM	0.0067	-		
2-Methylnaphthalene	91-57-6	EPA 8270-SIM	0.0067			
Acenaphthene	83-32-9	EPA 8270-SIM	0.0067			
Acenaphthylene	208-96-8	EPA 8270-SIM	0.0067			
Anthracene	120-12-7	EPA 8270-SIM	0.0067			
Benzo[g,h,i]perylene	191-24-2	EPA 8270-SIM	0.0067	-		
Fluoranthene	206-44-0	EPA 8270-SIM	0.0067	-		
Fluorene	86-73-7	EPA 8270-SIM	0.0067	-		
Naphthalene	91-20-3	EPA 8270-SIM	0.0067	-		
Phenanthrene	85-01-8	EPA 8270-SIM	0.0067	-		
Pyrene	129-00-0	EPA 8270-SIM	0.0067			
Carcinogenic PAHs (cPAHs)	<b>II</b>					
Benzo[a]anthracene	56-55-3	EPA 8270-SIM	0.0067	-		
Benzo[a]pyrene	50-32-8	EPA 8270-SIM	0.0067			
Benzo[b]fluoranthene	205-99-2	EPA 8270-SIM	0.0067			
Benzo[k]fluoranthene	207-08-9	EPA 8270-SIM	0.0067			
Chrysene	218-01-9	EPA 8270-SIM	0.0067			
Dibenz[a,h]anthracene	53-70-3	EPA 8270-SIM	0.0067			
Indeno[1,2,3-c,d]pyrene	193-39-5	EPA 8270-SIM	0.0067			
Metals	100 00 0		0.0007			
Arsenic (As)	7440-38-2	EPA 6020	5.0	0.5		
Cadmium (Cd)	7440-38-2	EPA 6020	0.20	0.10		
Chromium (Cr) (total)	7440-43-9	EPA 6020	0.20	1.0		
Lead (Pb)	7439-92-1	EPA 6020	2	0.1		
Mercury (Hg)	7439-92-1 7439-97-6	EPA 6020 EPA 7470A/7471A	0.05	0.1		

#### Notes:

<sup>1</sup>Chemical abstract service (CAS) registry number.

<sup>2</sup> Practical Quantitation Limit (PQL) values from ARI of Tukwila, Washington for the purposes of the RI/FS Work Plan.

mg/kg = milligram per kilogram

µg/L = microgram per liter

