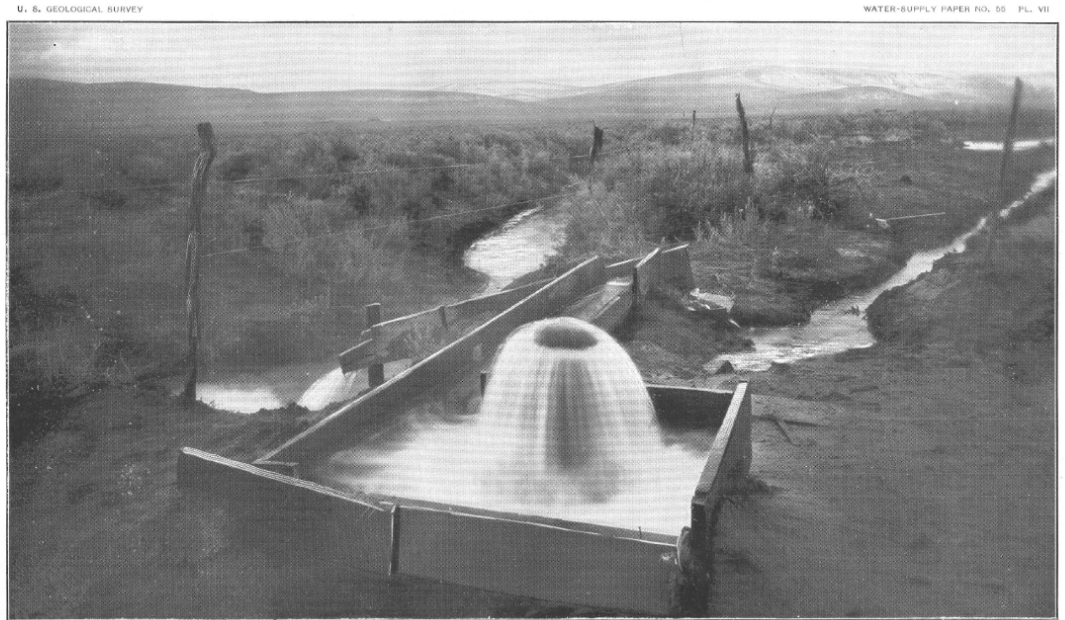


# Quality Assurance Project Plan

## City of Moxee ASR Feasibility Study



GANO WELL, MOXEE VALLEY.

October 2023

State of Washington Department of Ecology  
Office of Columbia River  
Agreement No. WRYBIP-2123-Moxeec-00036

Ecology Publication No. 23-12-015  
Aspect Project No. 190623

## Publication Information

---

Each study conducted by the Washington State Department of Ecology (Ecology) must have an approved Quality Assurance Project Plan (QAPP). The QAPP describes the objectives of the study and the procedures to be followed to achieve those objectives.

This QAPP was prepared by a licensed hydrogeologist. A signed and stamped copy of the report is available upon request. This QAPP is available via Ecology's publication database and upon request. The Ecology publication number for this QAPP is 23-12-015. This QAPP is valid through August 31, 2028.

Field data collected for this project will be uploaded to Ecology's Environmental Information Management (EIM) database under Study ID: WRYBIP-2123-Moxeec-00036.

## Contact Information

---

The authors for this QAPP can be contacted as follows:

Silas Sleeper, GIT, Aspect Consulting, (206) 453-6058, [ssleeper@aspectconsulting.com](mailto:ssleeper@aspectconsulting.com)

Tyson Carlson, LHG, Aspect Consulting, (509) 895-5923, [tcarlson@aspectconsulting.com](mailto:tcarlson@aspectconsulting.com)

For more information contact:

Scott Tarbutton, LHG  
QAPP Coordinator – Office of Columbia River  
4601 N Monroe St, Spokane, WA 99205  
Phone: (509) 867-6534

Washington State Department of Ecology – <https://Ecology.wa.gov>

- Headquarters, Olympia 360-407-6000
- Northwest Regional Office, Bellevue 425-649-7000
- Southwest Regional Office, Olympia 360-407-6300
- Central Regional Office, Union Gap 509-575-2490
- Eastern Regional Office, Spokane 509-329-3400

**COVER PHOTO:** G.O. Smith 1901 Geology and Water Resources of A Portion of Yakima County WA, USGS No. 55.

*Any use of product or firm names in this publication is for descriptive purposes only and does not imply endorsement by the author or the Department of Ecology.*

## ADA Accessibility

---

The Department of Ecology is committed to providing people with disabilities access to information and 504 and 508 of the Rehabilitation Act, and Washington State Policy #188.

**To request an ADA accommodation, contact Ecology by phone at 509-454-4241 or email at [tim.poppleton@ecy.wa.gov](mailto:tim.poppleton@ecy.wa.gov). For Washington Relay Service or TTY call 711 or 877-833-6341. Visit Ecology's website for more information.**

# Quality Assurance Project Plan

## City of Moxee ASR Feasibility Study

October 2023

Approved by:

Signature: 	Date: 10/12/2023
Ian Lauer, LG, Project Geologist, Aspect Consulting, LLC, Author	
Signature: 	Date: 10/12/2023
Tyson Carlson, LHG, Principal Hydrogeologist, Aspect Consulting, LLC, Project Manager	
Signature: 	Date: 10/13/2023
Justin Bellamy, PE, Principal Engineer, HLA Engineering and Surveying Inc., City Engineer	
Signature: 	Date: 10/13/2023
Jeff Burkett, Public Works City of Moxee, Client	
Signature: 	Date: 10/16/2023
Scott Tarbutton, LHG, Office of Columbia River Quality Assurance Coordinator	
Signature: 	Date: 10/16/2023
McKenna Murray, Office of Columbia River Project Manager	
Signature: 	Date: 10/12/2023
Justin Doty, Anatek, Project Manager	

# 1.0 Table of Contents

<b>1.0</b>	<b>Table of Contents</b> .....	<b>i</b>
	List of Figures .....	iv
	List of Tables .....	iv
<b>2.0</b>	<b>Abstract</b> .....	<b>1</b>
<b>3.0</b>	<b>Background</b> .....	<b>2</b>
3.1	Introduction and Problem Statement.....	2
3.2	Study Area and Surroundings.....	3
3.3	Water Quality Impairment Studies .....	20
3.4	Effectiveness Monitoring Studies .....	20
<b>4.0</b>	<b>Project Description</b> .....	<b>21</b>
4.1	Project Goals .....	21
4.2	Project Objectives .....	21
4.3	Information Needed and Sources .....	21
4.4	Tasks Required .....	21
4.5	Systematic Planning Process .....	22
<b>5.0</b>	<b>Organization and Schedule</b> .....	<b>23</b>
5.1	Key Individuals and Their Responsibilities .....	23
5.2	Special Training and Certifications.....	23
5.3	Organization Chart .....	24
5.4	Proposed Project Schedule .....	24
5.5	Budget and Funding .....	24
<b>6.0</b>	<b>Quality Objectives</b> .....	<b>25</b>
6.1	Data Quality Objectives .....	25
6.2	Measurement Quality Objectives .....	25
6.3	Acceptance Criteria for Quality of Existing Data.....	36
6.4	Model Quality Objectives.....	36
<b>7.0</b>	<b>Study Design</b> .....	<b>37</b>
7.1	Study Boundaries .....	37

7.2	Field Data Collection .....	37
7.3	Modeling and Analysis Design.....	39
7.4	Assumptions of Study Design.....	39
7.5	Possible Challenges and Contingencies .....	39
<b>8.0</b>	<b>Field Procedures.....</b>	<b>40</b>
8.1	Invasive Species Evaluation.....	40
8.2	Measurement and Sampling Procedures.....	40
8.3	Containers, Preservation Methods, Holding Times .....	43
8.4	Equipment Decontamination.....	45
8.5	Sample ID .....	45
8.6	Chain of Custody .....	45
8.7	Field Log Requirements.....	46
8.8	Other Activities.....	47
<b>9.0</b>	<b>Laboratory Procedures .....</b>	<b>48</b>
9.1	Lab Procedures Table .....	48
9.2	Sample Preparation Method(s).....	51
9.3	Special Method Requirements.....	52
9.4	Laboratories Accredited for Methods.....	52
<b>10.0</b>	<b>Quality Control Procedures .....</b>	<b>53</b>
10.1	Field and Laboratory Quality Control.....	53
10.2	Corrective Action Processes.....	54
<b>11.0</b>	<b>Data Management Procedures .....</b>	<b>55</b>
11.1	Data Recording and Reporting Requirements .....	55
11.2	Laboratory Data Package Requirements.....	55
11.3	Electronic Transfer Requirements .....	55
11.4	Data Upload Procedures .....	55
11.5	Model Information Management .....	55
<b>12.0</b>	<b>Audits and Reports.....</b>	<b>56</b>
12.1	Audits .....	56
12.2	Responsible Personnel .....	56
12.3	Frequency and Distribution of Reports.....	56

12.4	Responsibility for Reports .....	56
<b>13.0</b>	<b>Data Verification .....</b>	<b>57</b>
13.1	Field Data Verification, Requirements, and Responsibilities .....	57
13.2	Laboratory Data Verification .....	58
13.3	Validation Requirements, if Necessary .....	58
13.4	Model Quality Assessment.....	59
<b>14.0</b>	<b>Data Quality (Usability) Assessment .....</b>	<b>59</b>
14.1	Process for Determining Project Objectives were Met .....	59
14.2	Treatment of Nondetects.....	59
14.3	Data Analysis and Presentation Methods.....	59
14.4	Sampling Design Evaluation .....	59
14.5	Documentation of Assessment .....	59
<b>15.0</b>	<b>References .....</b>	<b>60</b>
<b>16.0</b>	<b>Appendices .....</b>	<b>62</b>
	Appendix A. Laboratory Accreditations .....	62
	Appendix B. Well Logs.....	62
	Appendix C. Aspect Field Data Sheets .....	62
	Appendix D. Glossaries, Acronyms, and Abbreviations.....	63

## List of Figures

Figure 1. Study Area Map .....	4
Figure 2. NE-SW Cross Section of Moxee Valley.....	6

## List of Tables

Table 1. Aquifer Test Results for the Ellensburg Formation.....	9
Table 2. Water Quality Data Available from DOH SENTRY Database.....	10
Table 3. Water Quality Data Available from Ecology EIM Database.....	11
Table 4. Groundwater and Drinking Water Regulatory Limits.....	14
Table 5. Organization of Project Staff and Resonsibilities.....	23
Table 6. Tentative Project Schedule .....	24
Table 7. Field Method MQOs and Field Equipment Information .....	26
Table 8. Laboratory MQOs of Water Samples .....	27
Table 9. Water Quality Sampling and Groundwater Level Monitoring Schedule .....	37
Table 10. City of Moxee Well Attributes Summary .....	38
Table 11. Field Parameter Stabilization Criteria.....	43
Table 12. Containers, Preservation Methods, and Holding Times... ..	43
Table 13. Lab Procedures.....	48

## 2.0 Abstract

The proposed Aquifer Storage and Recovery (ASR) program is being considered as a component of the City of Moxee's (City's) long-term water supply strategy of developing a surface water source to offset declining groundwater supplies while also improving seasonal groundwater flow to the Yakima River.

The feasibility study implementation plan (referred to herein as the Study) will assess the technical, operational, regulatory, and cost requirements to implement a future ASR project in the City's municipal water system. Tasks have been designed to address key components required in an ASR reservoir permit application as outlined in Washington Administrative Code (WAC) 173-157-110. As such, tasks under this Study include:

1. Developing a hydrogeologic conceptual model detailing the target aquifer system;
2. Assess source water availability, legal framework, and water rights to implement project;
3. Evaluating existing infrastructure and establishing targets for injection, storage, and recovery;
4. Assessing water quality characteristics of potential source water (e.g., canal water) and the target aquifer to evaluate compliance with groundwater standards and antidegradation policy, as described in WAC 173-200; and
5. Developing treatment requirements and alternatives for injected water (if needed).

A large part of this Study will rely on existing information and build off past efforts funded by the Yakima Basin Integrated Plan (YBIP) Groundwater Storage Subcommittee and Ecology. However, based on a review of all the past work, State of Washington Department of Ecology (Ecology) and the City determined that additional information needs to be collected under this Study to better understand source water quality, aquifer water quality and aquifer characteristics (Aspect, 2022). Key data collection tasks and schedule are identified below by section of this QAPP:

- Section 3.2.3: Provides a description of the water quality constituents to be evaluated;
- Section 4.4: Presents the details of the tasks to be completed, in sequential order;
- Section 5: Outlines the project schedule and team;
- Section 7.2: Describes water quality sampling locations and frequency (sampling schedule); and
- Section 8.2: Details the water quality sampling and well/aquifer testing procedures.



## 3.0 Background

The Ellensburg Formation aquifer is currently the City's only available water supply source. As a component of its long-term water supply strategy, the City is evaluating development of an ASR program to offset declining water levels in the lower Ellensburg Formation aquifer (Aspect, 2022). The City's proposed ASR project has the potential to address multiple goals of the YBIP, including expanding instream and out of stream uses and a Total Water Supply Available (TWSA)-positive outcome.

### 3.1 Introduction and Problem Statement

The goal of the proposed Study is to address key components required in an ASR reservoir permit application as outlined in WAC 173-157-110. Much of the information required for an ASR reservoir permit application was documented through past efforts funded by Ecology and the YBIP Groundwater Subcommittee. However, specific data gaps and proposed data collection were documented in a technical memorandum (herein referred to as the Data Gap Memo; Aspect, 2022) warranting additional data collection and analyses under this Study.

Following Ecology's review of the technical memo, specific project objectives were identified:

- Develop a hydrogeologic conceptual model to evaluate ASR feasibility and address informational requirements of Chapter 173-157-120 WAC;
- Assess source water availability, legal framework, and water rights to implement ASR in accordance with Chapter 173-157-130 and -140 WAC;
- Perform an engineering evaluation and determine the feasibility of incorporating ASR operations into the City's municipal water system;
- Assess water quality in the target aquifer and source water to identify constituents of concern, water quality compatibility, and compliance with:
  - Groundwater quality standards and antidegradation policy (Chapter 173-200 WAC);
  - Surface water treatment (Chapter 246-290 portions of Part 6);
  - Drinking water standards (Chapter 246-290-310); and
  - Source approval (Chapter 246-290-130).
- Identify the additional information requirements of WAC 173-157 that are not addressed in this Study.

The purpose of this QAPP is to describe the project objectives and procedures to achieve the goals for bullet 4 above. A future QAPP is necessary to address additional data collection (well and aquifer testing, additional water quality sampling, and surface water treatment) for the ASR program. This QAPP addresses the following elements:

- Study design;
- Data and measurement quality objectives;

- Field and laboratory procedures;
- Quality control procedures;
- Data verification and validation protocols;
- Data management procedures; and
- Reporting.

The objective of this assessment is to:

1. Collect water quality samples from both source water supply and groundwater to perform geochemical analysis of water compatibility and assess treatment.
2. Collect one round of groundwater level measurements taken at all relevant wells to produce a consistent groundwater elevation map and confirm the City's SCADA system function.

The QAPP follows the recommended guidelines from Ecology's *Guidelines for Preparing Quality Assurance Project Plans for Environmental Studies* (Ecology, 2004; updated 2016) to conduct water level and water quality analyses effectively and accurately as part of the Study.

The Study and development of this QAPP are funded under the YBIP Groundwater Subcommittee (Agreement No. WRYBIP-2123-Moxeec-00036) between the City and Ecology. Aspect and HLA Engineering and Surveying (HLA) are under contract to the City to prepare this QAPP and complete the Study.

## **3.2 Study Area and Surroundings**

The Study is within the Moxee Valley, located in Yakima County, Washington, as shown on Figure 1.

The City's water system serves a population between 3,000 and 4,000 people. Three groundwater wells (Well Nos. 1, 3, and 4) are active sources to the City's water system, Well No. 2 is maintained as an emergency backup source. Within the Moxee Valley there are three main sources of surface water within the Study area: (1) the Yakima River; (2) the Selah-Moxee Irrigation Canal system; and (3) the Roza Irrigation Canal system. Surface water is the primary source of irrigation water and groundwater serves as the primary source of drinking water in the Study area. Figure 1 shows the Moxee Valley, the City's Urban Growth Area, City water supply wells, irrigation canals, and the Yakima River.

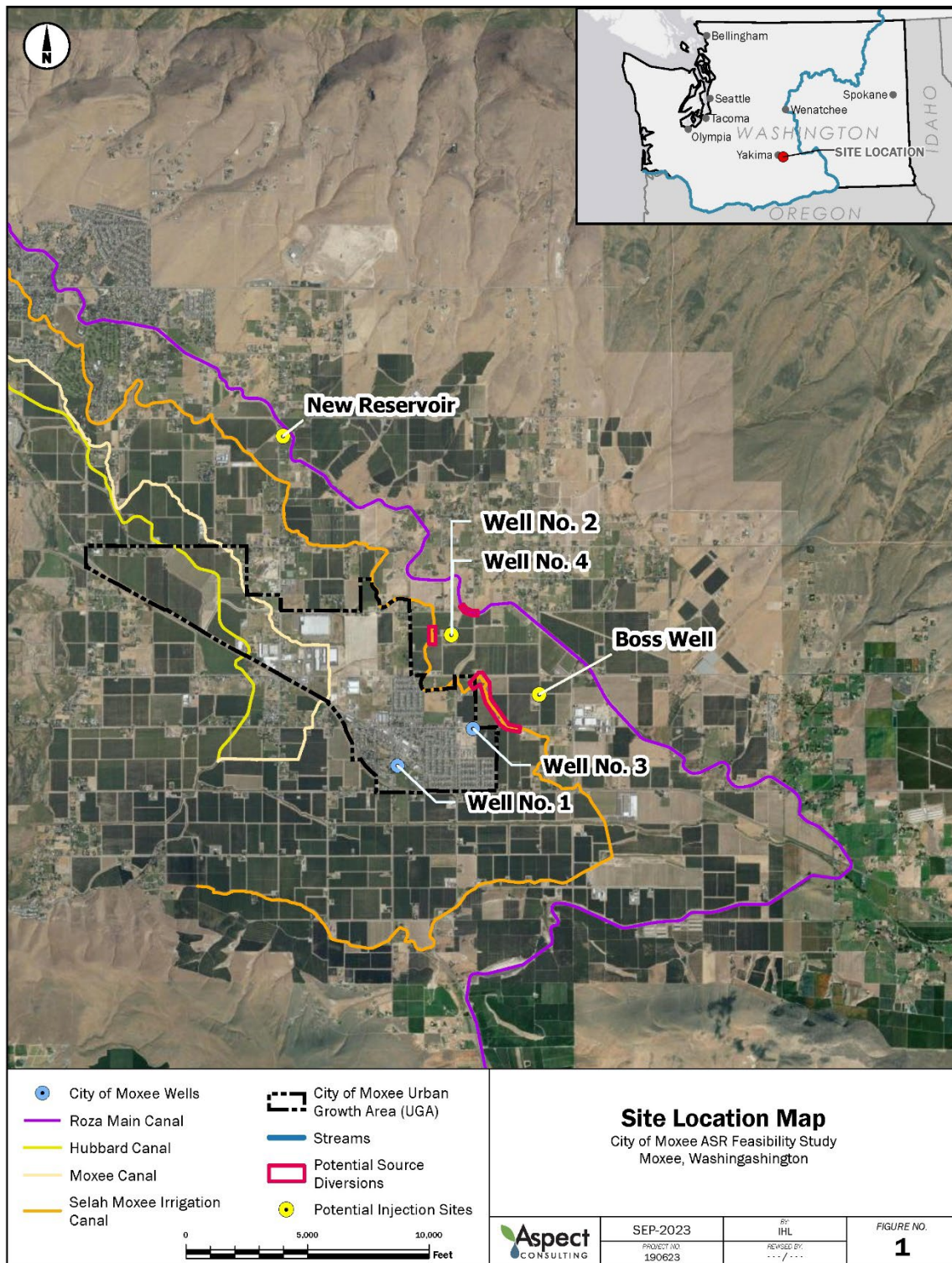


Figure 1. City of Moxee ASR Site Location Map

The Moxee Valley lies within the Yakima Fold and Thrust Belt (YFTB), a broad region of east-west compression and clockwise plate rotation that has created several northwest and west trending anticlines and thrust faults, and northwest and north trending regional strike slip faults. Pending development of the project-specific hydrogeologic conceptual model, within the YFTB, the Study area is bounded to the:

- West by the Yakima River;
- East by an unconformity between the Ellensburg Formation and Columbia River Basalt Group (CRBG);
- North by Yakima Ridge anticline; and
- South by the Ahtanum-Moxee syncline (Jones et al., 2006).

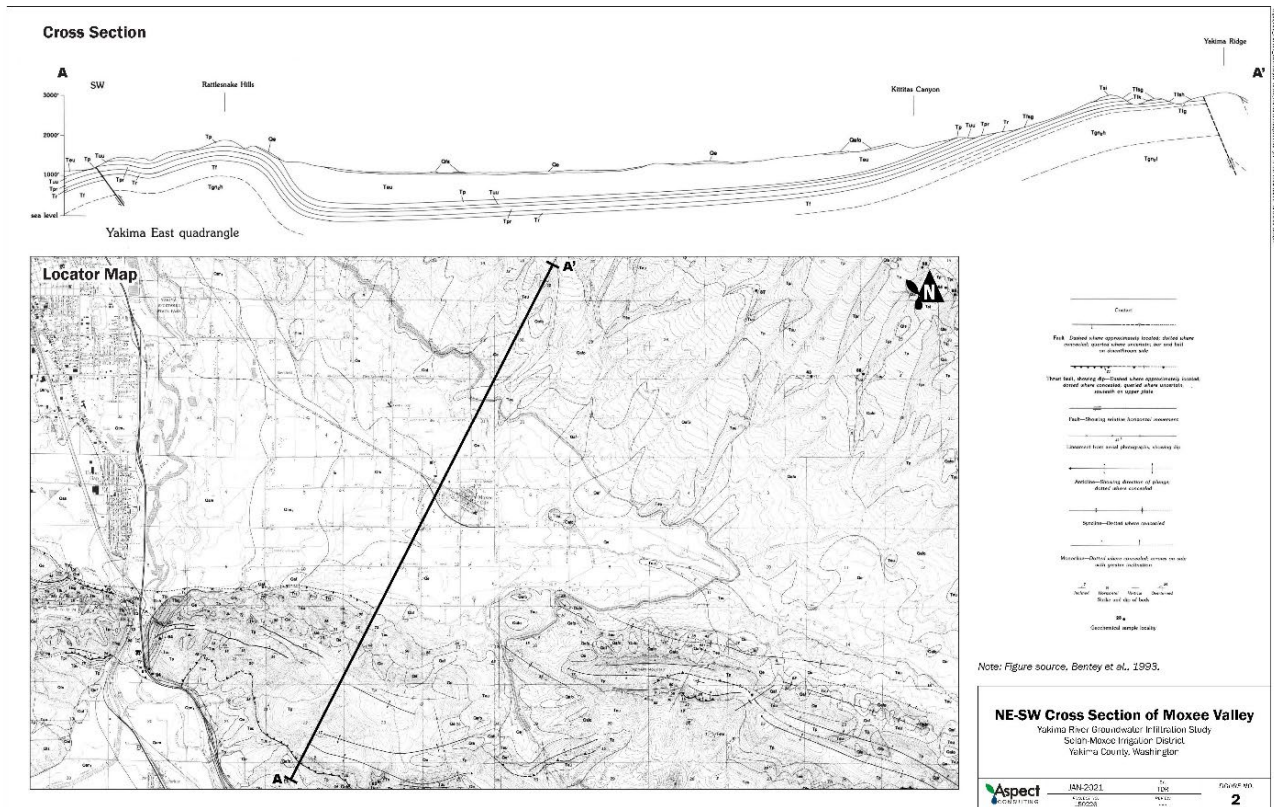
A generalized cross section of the Moxee Valley as depicted by Bentley et al. (1993) is provided as Figure 2.

The City relies on groundwater supply from the Upper Undifferentiated Ellensburg Formation (Ellensburg Formation). The Ellensburg Formation was deposited as a vertically stratified semi-consolidated to consolidated sedimentary units that overly the CRBG. All City wells are completed across water bearing zones of the Ellensburg Formation and as such, the Ellensburg Aquifer is the target aquifer for ASR.

The Ellensburg Aquifer is a semi-confined to confined multi-layered aquifer system. The City's wells are completed in a portion of the Ellensburg Aquifer that appears semi-confined ( $S = 2.4 \times 10^{-4}$ ) based on estimation of storativity ( $S$ ) from Well No. 4 pumping test data (Aspect, 2020), which is consistent with the range of estimates ( $2 \times 10^{-3}$  to  $7 \times 10^{-4}$ ) by others compiled in Vaccaro et al., 2009.

### **3.2.1 History of Study Area**

G.O. Smith (1901) first described the hydrogeologic conditions (folds and confining units) and water bearing units of the Ellensburg Formation that resulted in artesian conditions in the Moxee Valley. G.O. Smith (1901) documented several Ellensburg Aquifer wells completed from 525 to 1,026 feet below ground surface (bgs) that capture water from singular to multiple water bearing units with a depth to top of unit from 515 to 1,020 feet bgs. The wells had a potentiometric surface of 30 to 115 feet (Clark Well Nos 2 and 3) of water above ground surface flowing 0.5 to 2.00 cubic feet per second (cfs). Some City wells (e.g., Well No. 2) have experienced water level declines of approximately 70 feet since time of construction in the



**Figure 2. NE-SW Cross Section of Moxee Valley adapted from Bentley et al, 1993**

1980s. The City’s Well No. 2 is located in the same section as Clark Well Nos. 2 and 3. Offsetting groundwater level decline is the impetus for ASR as a future water supply strategy, using available groundwater storage capacity.

The ASR project proposes use of Yakima River water as the source water to offset the decline in groundwater levels and provide additional supply to the City. The City is considering using water that is either diverted from the Selah-Moxee Irrigation District (SMID) or the Roza Irrigation canal systems, which may require treatment to drinking water standards (Chapter 246-290-310) prior to injection. Three potential points of diversion (two from the SMID Canal and one from the Roza Main Canal) were identified based on proximity to existing wells as shown on Figure 1.

The Study is part of the YBIP which was developed between 2009 and 2012 in response to growing concerns about water availability in the Yakima River Basin. This is a 30-year resiliency plan that aims to protect fish habitat and improve water availability and reliability. The YBIP is currently being implemented in phases through funds from the state and federal governments. This Study falls under the groundwater storage element of the YBIP and aims to address municipal water reliability and instream flow augmentation.

### 3.2.2 Summary of Previous Studies and Existing Data

The City's proposed ASR plan has been identified by the YBIP's Groundwater Subcommittee as a potential method to improve groundwater supplies within the Yakima River Basin. Therefore, the subcommittee elected to fund the Study, which includes a Data Gap Memo (Aspect, 2022) detailing the existing geology, hydrology, and water chemistry data and gaps in the data that will need to be addressed during the project. The Data Gaps Memo identified that the existing data is generally of sufficient quality to support the Study, with minimal additional improvements.

Key findings include:

- The existing hydrogeologic data and reports provide a solid foundational knowledge for assessing the hydrogeologic setting and viability of the local Ellensburg Formation aquifer near the City for ASR once synthesis of additional existing and new data is completed for the FS.
- The condition of the City's wells are documented, and completion information is sufficient to support analysis required for the FS. Additionally, many neighboring well logs exist with sufficient completion information to inform additional hydrologic interpretation. Opportunistically, additional exploration and/or testing of existing wells may be incorporated into the conceptual model developed under the FS.
- A survey of City wellhead conditions and approximate elevations is needed to determine absolute groundwater elevations across the aquifer near the City.
- Manual water level data collection is needed for all City production wells, as allowed by wellhead access, to validate past automated measurements and correct data to groundwater elevations.
- Site-specific data provided from the City's SCADA data and recent pumping tests at Well 4 provide local validation (and support future updates) of the regional numerical model and aquifer parameters. Additional processing of SCADA data may be required to utilize higher frequency measurements. This will allow for estimation of aquifer performance during ASR as part of the FS.
- Water quality data exists for all sources and for the City's wells but has a variable analyte list and temporal coverage. A complete set of water quality samples will need to be collected for the proposed source waters and storage aquifer in order to assess chemical compatibility, treatment needs, and regulatory compliance. Sampling will be performed for all potential water sources consistent with WAC 246-290-310 and 173-200.

The Yakima River Basin has been the focus of multiple existing efforts by federal, state, tribal, and local groups, including the YBIP workgroups, to characterize and plan for long term sustainability of the surface and groundwater resources within the watershed. This includes:

- A series of conceptual and numerical modeling reports prepared for the City of Yakima during development of its ASR program within the Ellensburg Formation aquifer of the Ahtanum-Moxee subbasin. (Golder, 2001, 2002, 2007, and 2014).
- Previous studies that assess the suitability of areas within the Yakima Basin for shallow aquifer recharge and aquifer storage and recovery (Aspect, 2021; Gibson and Campana, 2018; Sleeper, 2020).
- Development of the hydrogeologic framework and characterization of the geochemistry of various aquifer systems within the Columbia River Basalt Group (CRBG; Whiteman et al., 1994).
- Mapping of the extent and depth to the top of basalt and interbedded hydrogeologic units, Yakima River Basin aquifer system (Jones and Watkins, 2008).
- A characterization of six sedimentary sub-basins, their hydrogeologic framework, and aquifers parameters within the Yakima River Basin (Jones et al., 2006).
- Development of a hydrogeologic framework, geologic characterization, geochemistry, and groundwater trends, and aquifer parameters for aquifers of the Yakima River Basin (Vaccaro et al., 2009).
- Numerical modeling of the Yakima River Basin Aquifer System (Vaccaro et al., 2009; Ely et. al., 2011).
- Analysis of ambient groundwater quality in the Moxee Valley Surficial Aquifer (Ecology, 2007).

Results from these studies include geologic framework and estimates of aquifer parameters, including storage coefficients, transmissivities, and groundwater velocities of the associated aquifer systems. Many of these reports focus on the larger areal extents than the Study Area. Additional site-specific evaluation of aquifer characteristics, and water level and water quality data, is necessary to characterize the target aquifer.

A data gap memorandum (Aspect, 2023) outlines available existing data and hydrogeologic reports and data needs pertaining to aquifer parameters, groundwater levels, groundwater quality, and surface water quality. Key findings from the data gap analysis are highlighted below.

### **Aquifer Parameters**

Aquifer and well testing reports are available for City Well No. 4 (Aspect, 2020); this report was completed to document well construction and testing. Pumping test results for the well were used to determine parameters for the Ellensburg Formation, which was screened from 750 to 1,104 feet below ground surface (bgs) at Well 4. In addition, detailed mapping and testing results collected for the USGS CPRAS and Yakima Models yield general aquifer parameters for the Ellensburg Formation and surrounding aquifers. In addition, detailed aquifer testing and

modeling of the Ellensburg Formation aquifer were completed as part of the City of Yakima ASR program development. Results documented in these reports are summarized in Table 1.

**Table 1. Aquifer Test Results for Ellensburg Formation**

Study Area	Study / Aquifer Test	Hydraulic Conductivity (ft/d)	Transmissivity (ft <sup>2</sup> /d)	Storativity (unitless)
City of Moxee	City Well No. 4 <sup>1</sup>	-	38,600	2.4 x 10 <sup>-4</sup>
Regional	Vaccaro et al (2009) <sup>2</sup>	72	-	2 x 10 <sup>-3</sup> to 7 x 10 <sup>-4</sup>
Modeled	Ely (2011) <sup>3</sup>	12.9 to 69.2	-	4.62 x 10 <sup>-5</sup>
City of Yakima	Golder (2009)	7.5	-	7 x 10 <sup>-4</sup>

**Notes:**

1. Aspect, 2020
2. Average value, K ranged from 0.01 to 2,265 ft/d
3. Model-calibrated results

Ecology’s Environmental Information Management System (EIM) records 73 wells in the area, 42 of which were completed at depths associated with the Ellensburg Formation and had associated well logs; however, little additional well testing data was yielded from these logs and no studies with aquifer testing data were identified.

**Groundwater Levels**

The City has had a Supervisory Control And Data Acquisition (SCADA) system in place since 2015 to continuously monitor and record water levels and pumping rates at each of its water supply wells. Periodic water level data from 2015 to 2022 has been retrieved from the wells, but they only report depth of water above the sensor. Data has been corrected for elevations reported in well logs and pump set depth. However, field verification of SCADA water level readings, and refinement of measuring point data will need to be performed to improve accuracy to the level required for modelling.

The USGS National Water Information System (NWIS) and Ecology’s EIM database provide publicly available groundwater monitoring within the Yakima Basin. The EIM systems identified eight wells near the City with groundwater level measurements spanning a discontinuous period from 1977 through 2021. These data were entered into the EIM database as part of data compilation efforts completed for the Ecology’s Columbia River Groundwater Database (CRGWDB) study.

Well water level data were also queried from the USGS NWIS database. The NWIS database contained over 300 groundwater wells within an approximate 48-square-mile area surrounding the City, with periods of record spanning from 1890 to 2008 (discontinuous). Twenty-two of these wells are listed as completed within the Ellensburg Formation, of which five have two or



more recorded water levels. Several of these are mapped directly with hydrographs via a USGS monitoring website (Keys, 2008).

### Groundwater Quality

The City conducts water quality sampling at each of its groundwater wells to comply with DOH drinking water source requirements. Although these water quality data are useful in characterizing ambient groundwater, the chemical analyses completed per DOH requirements do not include several constituents and field parameters important for assessing geochemical compatibility with treated surface water (e.g., silica, sulfide, and oxidation-reduction potential [ORP]) and only report total metals concentrations rather than total and dissolved concentration components. Additionally, water quality analyses completed under DOH requirements are reported only to the State Reporting Limit (SRL) as opposed to the Method Detection Limit (MDL) and, consequently, water quality results obtained from DOH records are often qualitatively reported as “less than” the SRL rather than reporting the measured concentration.

The DOH SENTRY database contains results for routine compliance sampling for each of the City’s wells. Table 2 identifies the periods of record of various analyte suites for the City’s wells from the DOH database.

**Table 2. Water Quality Data Available from DOH SENTRY Database**

Analyte Suite / Test Panel	Period of Record <sup>1</sup>	Note
Inorganic Constituents	1985 – 2022 <sup>2</sup>	As, Ag, Ba, Be, Ca, Cd, Cl, Cr, Cu, Cyanide, Fe, Fl, Hg, Mg, Mn, Na, Ni, NO <sub>3</sub> , Pb, Sb, Se, SO <sub>4</sub> , Th, Zn, Color, Sp. Cond., Hardness, Turbidity
Synthetic Organic Compounds	1991 - 2022 <sup>3</sup>	Analytical suites vary annually between insecticides, pesticides, and soil fumigants
Volatile Organic Compounds	1988 - 2022	Results for various temporal resolutions from each well
Radionuclides	2001 - 2021	Results for various temporal resolutions from each well

**Notes:**

1. Not all wells span full periods of record
2. Not all constituents span whole record, full constituents list first recorded in 2003.

In addition to the City’s data, the EIM database listed an ambient groundwater quality study (ID KSIN0002), which measured water quality of 20 wells in the Moxee area in 2006. The study reported field parameters of temperature, pH, conductivity, and dissolved oxygen (DO), and a limited analytical suite including bacteria, total dissolved solids, iron, magnesium, chloride, nitrate, phosphorous. Multiple samples were reported with bacterial contamination, but no other exceedances were reported.

### Source Water Quality

Water quality data are limited for the source waters being considered (SMID and Roza irrigation canals). USGS NWIS database includes water quality results for all three potential sources, but they vary spatially and temporally. From 1970 to 2004, USGS collected samples from the Yakima

River at various points along the central Yakima Valley, including physical parameters, major inorganics, nutrients, stable isotopes, and microbiological samples. Sites on the Roza main canal were sampled from 1986 and 2004, including physical parameters, major inorganics, nutrients, and stable isotopes. A single sample was available from the SMID Canal from 2000, which included physical parameters, major inorganics, organics, pesticides, nutrients, and perfluorooctanoic acid (PFOA). The samples for the Roza irrigation canals were collected at approximately the same location 5.5 miles northwest of the City, while the SMID Canal was sampled 1 mile north of the City and immediately adjacent to the City’s Well Nos. 2 and 4 (Figure 1). These data are summarized in Table 3 below.

**Table 3. Water Quality Data Available from Ecology EIM Database**

Source <sup>1</sup>	Period of Record	Number of Samples	Analyte Suites <sup>2</sup>
Yakima River near Selah Gap (12487000)	1985 – 2022	4	Physical parameters, inorganic constituents (IOC), silica (2004), and stable isotopes (1985)
Yakima River near Yakima (12500005)	1987	2	Physical parameters, IOCs, silica (2004), and stable isotopes (1985)
Yakima River near Terrace Heights	1970-1977	98	Physical parameters, IOCs, silica (1977), bacterial
Union Gap Canal (463716120291800)	2012	3	Volatile organic compounds (VOC)
Roza Canal at N 33rd (12485003)	1986-2004	2	Physical parameters, IOCs, stable isotopes (1986), silica (2004)
Selah-Moxee Canal (463411120223900)	2000	1	Physical parameters, IOCs, VOCs, pesticides and herbicides

**Notes:**

1. The table does not list outfalls or irrigation drains
2. Not all constituents span the whole record for any given source.

Following the planned water quality assessment and review from Ecology’s Water Quality Program and DOH Regional Engineer, feedback from Ecology’s Water Resources Program and Office of Columbia River will be necessary to inform a future QAPP to address additional data collection (well and aquifer testing, additional water quality sampling, and surface water treatment).

The City will collect a limited set of data, described in this QAPP, to supplement existing datasets. Specifically, this assessment will:

1. Collect water quality samples from both source water supply and groundwater to perform geochemical analysis of water compatibility and assess treatment.

2. Collect one round of groundwater level measurements taken at all relevant wells to produce a consistent groundwater elevation map and confirm the City’s SCADA system function.

### 3.2.3 Parameters of Interest

The water quality analytes were selected to evaluate the potential for water quality impacts related to ASR and compliance with Washington State Groundwater Quality Standards (Chapter 173-200 WAC) and Drinking Water Standards (Chapter 246-290 WAC). The source water supply has limited existing data and will be analyzed for a full analyte suite, whereas the target aquifer has been regularly sampled to DOH drinking water quality standards by the City and will only be sampled with a limited analyte set of field parameters, general chemistry, and bacteria. The following sections describe the water quality analytes selected for this water quality assessment. The schedule for monitoring these constituents during the Study is presented in Section 7.2.

#### Field Parameters

Field parameters will be measured to provide independent corroboration of laboratory results, and to analyze constituents that have short hold times and can be reliably measured in the field. Field parameters also include measurements to develop groundwater elevation contour maps. These include:

- Electrical conductivity
- Dissolved Oxygen (DO)
- Oxidation Reduction Potential (ORP)
- pH
- Temperature
- Turbidity
- Continuous and discrete groundwater depth-to-water
- Groundwater level measuring point elevation

#### General Chemistry

The general chemistry suite includes inorganic constituents and conventional water quality parameters. Groundwater and surface water samples will be analyzed for this suite of constituents in both the dissolved (field-filtered to 45 microns) and total fractions. Geochemical analysis will evaluate chemical compatibility of native groundwater and surface water, and monitor for potential chemical reactions of the recharge water with aquifer material (mineral dissolution and precipitation) during aquifer storage. This analytical suite will also inform source treatment requirements in the context of Chapter 173-200 WAC (Groundwater Quality Standards) and WAC 246-290-310 (Drinking Water standards). Constituents will include:

Alkalinity	Silica	Lead
Bicarbonate	Arsenic	Magnesium
Chloride	Antimony	Manganese

Total Dissolved Solids (TDS)	Aluminum	Mercury
Total Suspended Solids (TSS)	Barium	Nickel
Total and Dissolved (DOC) Organic Carbon (TOC)	Beryllium	Potassium
Phosphorus	Cadmium	Selenium
Bromide	Calcium	Silver
Fluoride	Chromium	Sodium
Nitrate-N	Copper	Thallium
Nitrite-N	Iron	Uranium
Sulfate	Zinc	

### **Volatile and Semivolatile Organic Compounds**

As described in Section 3.2.2, baseline characterization was completed for the native groundwater in the target storage aquifer (Ellensburg Formation aquifer at City water supply wells). As required by the Washington State Department of Health (DOH), the City has three decades of groundwater quality data including both volatile organic compounds (VOCs) and synthetic organic compounds (SOCs). Over the period of record (1990-present) neither SOCs nor VOCs were detected in the City’s water supply wells that are completed in the source aquifer.

The Study will evaluate potential surface water sources (e.g., SMID Canal and Roza Main Canal) for both VOCs and SOCs. Therefore, measurement of VOCs and SVOCs is necessary to accurately assess surface water quality.

### **Herbicides and Pesticides**

The City has evaluated herbicides and pesticides for DOH drinking water compliance. Over the period of record (1990-present) neither herbicides or pesticides were detected in the City’s water supply wells that are completed in the source aquifer.

The Study will evaluate potential surface water sources (e.g., SMID Canal and Roza Main Canal) for both herbicides and pesticides. Therefore, herbicides and pesticides will be measured at potential surface water sources as part of this Study. This will include the analytes specified in U.S. Environmental Protection Agency (EPA) Methods:

- Chlorinated Pesticides
- Chlorinated Acid Herbicides
- Pesticides as carbamates
- Herbicides – diquat, paraquat, endothall, and glyphosate

## Bacteriological Constituents

The Study will evaluate bacteriological constituents (total coliform and E. Coli) in native groundwater and potential surface water sources (e.g., SMID Canal and Roza Main Canal) to determine baseline conditions. The Study will evaluate the following constituents:

- E. coli (presence/absence)
- Total coliforms (plate count)

## Radionuclides

Radionuclides were detected in groundwater at City wells. In addition, radionuclides have not been analyzed for potential surface water sources. Thus, the Study will evaluate the following radionuclides in potential surface water sources:

- Radium 226 + Radium 228
- Gross Alpha radiation
- Gross Beta radiation

## Per- and polyfluoroalkyl substances (PFAS)

PFAS were not detected in groundwater at City wells. PFAS have not been analyzed for potential surface water sources. Thus, the Study will evaluate the following PFAS in potential surface water sources:

- Perfluorooctanoic acid (PFOA)
- Perfluorooctane sulfonate (PFOS)
- Perfluorohexane sulfonate (PFHxS)
- Perflourononanoic acid (PFNA)
- Perfluorobutane sulfonate (PFBS)

### 3.2.4 Regulatory Criteria or Standards

The introduction of recharge water to the Ellensburg Formation aquifer is subject to the Antidegradation Rule and the numerical groundwater quality standards (GWQS) defined in Groundwater Quality Standards (Chapter 173-200 WAC). Table 4 presents the regulatory criteria by analyte method that will be considered during the project.

**Table 4. Groundwater and Drinking Water Regulatory Limits**

Analyte	Unit	WAC 173-200-040	WAC 246-290 Primary Drinking Water Standard	WAC 246-290 Secondary Drinking Water Standard
Field Parameters				
Specific conductance	uS/cm			700
Turbidity	NTU		5*	

Analyte	Unit	WAC 173-200-040	WAC 246-290 Primary Drinking Water Standard	WAC 246-290 Secondary Drinking Water Standard
pH	SU	6.5-8.5		6.5-8.5
EPA 200.7 and 200.8 (General Chemistry)				
Aluminum	ug/L			
Antimony	ug/L		6	
Arsenic	ug/L	0.05	10	
Barium	ug/L	1,000	2,000	
Beryllium	ug/L		4	
Cadmium	ug/L	10	5	
Calcium	ug/L			
Chromium	ug/L	50	100	
Copper	ug/L	1,000	1,300	
Iron	ug/L	300		300
Lead	ug/L	50	15	
Magnesium	ug/L			
Manganese	ug/L	50		50
Nickel	ug/L			
Potassium	ug/L			
Selenium	ug/L	10	50	
Silica (SiO <sub>2</sub> )	ug/L			
Silver	ug/L	50		100
Sodium	ug/L		20	
Thallium	ug/L		2	
Uranium	Ug/L		30	
Zinc	ug/L	5,000		5,000
EPA 245.7 (General Chemistry)				
Mercury	ug/L	2	2	
EPA 300.0 (General Chemistry)				
Bromide	mg/L			
Chloride	mg/L	250		250
Fluoride	mg/L	4	4	<u>2</u>
Sulfate	mg/L	250		250
Nitrate as Nitrogen	mg/L	10	10	
Nitrite as Nitrogen	mg/L		1	
EPA 335.1 / SM4500 CN-G (General Chemistry)				
Cyanide, Total	ug/L		200	
SM2320B (General Chemistry)				
Alkalinity as Carbonate	mg/L			
Bicarbonate Ion	mg/L			
SM2540C (General Chemistry)				
Total Dissolved Solids	mg/L	500		500

Analyte	Unit	WAC 173-200-040	WAC 246-290 Primary Drinking Water Standard	WAC 246-290 Secondary Drinking Water Standard
SM2540D (General Chemistry)				
Total Suspended Solids	mg/L			
SM5310B (General Chemistry)				
Total Organic Carbon	mg/L			
Dissolved Organic Carbon	mg/L			
SM 4500-P F (General Chemistry)				
Phosphorus	mg/L			
EPA 524.3 (VOCs and SVOCs)				
1,1,1,2-Tetrachloroethane	ug/L			
1,1,1-Trichloroethane	ug/L	200	200	
1,1,2,2-Tetrachloroethane	ug/L			
1,1,2-Trichloroethane	ug/L		5	
1,1-Dichloroethane	ug/L	1		
1,1-Dichloroethylene	ug/L		7	
1,1-Dichloropropene	ug/L			
1,2,3-Trichlorobenzene	ug/L			
1,2,3-Trichloropropane	ug/L			
1,2,4-Trichlorobenzene	ug/L		70	
1,2,4-Trimethylbenzene	ug/L			
Dibromochloropropane (DBCP)	ug/L		0.2	
1,2-Dibromoethane (EDB)	ug/L	0.001	0.05	
1,2-Dichlorobenzene	ug/L		600	
1,2-Dichloroethane (EDC)	ug/L	0.5	5	
1,2-Dichloropropane	ug/L	0.6	5	
1,3,5-Trimethylbenzene	ug/L			
1,3-Dichlorobenzene	ug/L			
1,3-Dichloropropane	ug/L			
1,3-Dichloropropene	ug/L	0.2		
1,4-Dichlorobenzene	ug/L	4	75	
2,2-Dichloropropane	ug/L			
2-Chlorotoluene	ug/L			
4-Bromofluorobenzene	ug/L			
4-Chlorotoluene	ug/L			
Acetone	ug/L			
Benzene	ug/L	1	5	
Bromobenzene	ug/L			
Bromochloromethane	ug/L			
Bromodichloromethane	ug/L	0.3	0	
Bromoform	ug/L	5	0	
Bromomethane	ug/L			

Analyte	Unit	WAC 173-200-040	WAC 246-290 Primary Drinking Water Standard	WAC 246-290 Secondary Drinking Water Standard
Carbon Tetrachloride	ug/L	0.3	5	
Chlorobenzene	ug/L		100	
Chloroethane	ug/L			
Chloroform	ug/L	7	70	
Chloromethane	ug/L			
cis-1,2-Dichloroethylene (DCE)	ug/L		70	
1,3-Dichloropropene	ug/L	0.2		
Dibromochloromethane	ug/L	0.5	60	
Dibromomethane	ug/L			
Dichlorodifluoromethane	ug/L			
Ethylbenzene	ug/L		700	
Hexachlorobutadiene	ug/L			
Isopropylbenzene	ug/L			
Methyl tert-butyl ether (MTBE)	ug/L			
Methylene Chloride	ug/L	5	5	
Naphthalene	ug/L			
n-Butylbenzene	ug/L			
n-Propylbenzene	ug/L			
Xylenes (Total)(o-, m-, p-)	ug/L		10,000	
p-Isopropyltoluene	ug/L			
sec-Butylbenzene	ug/L			
Styrene	ug/L		100	
tert-Butylbenzene	ug/L			
Tetrachloroethane (PCE)	ug/L	0.8	5	
Toluene	ug/L		1,000	
trans-1,2-Dichloroethylene	ug/L		100	
Trichloroethane	ug/L		5	
Trichloroethene (TCE)	ug/L	3	5	
Trichlorofluoromethane	ug/L			
Total trihalomethanes (TTHM)	ug/L		80	
Vinyl Chloride	ug/L	0.02	2	
<b>EPA 525.2 (VOCs and SVOCs)</b>				
Alachlor	ug/L		2	
Atrazine	ug/L		3	
Benzo(a)pyrene	ug/L	0.008	0.2	
Bis(2-ethylhexyl) adipate	ug/L		400	
Bis(2-ethylhexyl) phthalate	ug/L	6	6	
Bromacil	ug/L			
Butachlor	ug/L			



Analyte	Unit	WAC 173-200-040	WAC 246-290 Primary Drinking Water Standard	WAC 246-290 Secondary Drinking Water Standard
Fluorene	ug/L			
Hexachlorobenzene	ug/L	0.05	1	
Hexachlorocyclopentadiene	ug/L		50	
Metolachlor	ug/L			
Metribuzin	ug/L			
Propachlor	ug/L			
Simazine	ug/L		4	
<b>EPA 505 (Herbicides and Pesticides)</b>				
gamma-BHC (Lindane)	ug/L	0.06		
Aldrin	ug/L	0.005		
Endrin	ug/L	0.2	2	
DDT (DDE, DDD, DDT)	ug/L	0.3		
Dieldrin	ug/L	0.005		
Heptachlor	ug/L	0.02	0.4	
Heptachlor epoxide	ug/L	0.009	0.2	
lindane	ug/L	0.06	0.2	
Methoxychlor	ug/L	100	40	
PCB polychlorinated biphenyls	ug/L	0.01	0.5	
Chlordane	ug/L	0.06	2	
Toxaphene	ug/L	0.08	3	
<b>EPA 515.4 (Herbicides and Pesticides)</b>				
2,4-D	ug/L	100	70	
2,4-DB	ug/L			
3,5-Dichlorobenzoic acid	ug/L			
Acifluorfen	ug/L			
Chloramben	ug/L			
Chlorthal (Dacthal)	ug/L			
Dalapon	ug/L		200	
Dicamba	ug/L			
Dichloroprop	ug/L			
Dinoseb	ug/L		7	
Pentachlorophenol	ug/L		1	
Picloram	ug/L		500	
Silvex	ug/L	10	50	
<b>EPA 531.2 (Herbicides and Pesticides)</b>				
3-Hydroxycarbofuran	ug/L			
Aldicarb	ug/L			
Aldicarb Sulfoxide	ug/L			
Aldoxycarb	ug/L			
Carbaryl	ug/L			

Analyte	Unit	WAC 173-200-040	WAC 246-290 Primary Drinking Water Standard	WAC 246-290 Secondary Drinking Water Standard
Carbofuran	ug/L		40	
Methiocarb	ug/L			
Methomyl	ug/L			
Oxamyl	ug/L		260	
Propoxur	ug/L			
EPA 547 (Pesticides and Herbicides)				
Glyphosate	ug/L		700	
EPA 548.1 (Pesticides and Herbicides)				
Endothall	ug/L		100	
EPA 549.2 (Pesticides and Herbicides)				
Diquat	ug/L		0.2	
EPA 1613 (Pesticides and Herbicides)				
Dioxin [2,3,7,8-TCDD]	ug/L	0.0000007	30	
SM9221B (Bacteriological)				
Fecal Coliform	MPN/100mL			
SM9223B (Bacteriological)				
E. coli	MPN/100mL			
Total Coliform	MPN/100mL	1/100		
EPA 900 (Radionuclides)				
Gross Alpha	pCi/l	15	15	
Gross Beta	pCi/l	50	4*	
EPA 903/904 (Radionuclides)				
Radium-226	pCi/l	3		
Radium-226+228	pCi/l	5	5	
EPA 533 (PFOAs)				
PFOA	ng/L		10	
PFOS	ug/L		15	
PFHxS	ug/L		65	
PFNA	ug/L		9	
PFBS	ug/L		345	

**Notes:**

ug/L – Micrograms per liter

mg/L – Milligrams per liter

uS/cm – microsiemens per centimeter

SU. – standard units

\* drinking water limit for turbidity is based on a treatment technique in lieu of a Maximum Contaminant Level, where unfiltered surface water cannot exceed 5 NTU (WAC 246-290-632).

### **3.3 Water Quality Impairment Studies**

Not applicable.

### **3.4 Effectiveness Monitoring Studies**

Not applicable.

## 4.0 Project Description

### 4.1 Project Goals

The overall project goal is to assess the potential for ASR to augment existing water supplies and meet future water demands within the City’s water service area. This phase of the project focuses specifically on understanding the water quality conditions of the source water and the aquifer planned for reservoir storage to support geochemical evaluation. Tasks have been designed to determine water quality characteristics.

### 4.2 Project Objectives

The objectives of the Study include:

- Refine the hydrogeologic conceptual model to evaluate ASR feasibility and address informational requirements of Chapter 173-157-120 WAC;
- Assessment of source water quality;
- Assessment of groundwater quality in the target aquifer; and
- Collection of groundwater level measurements in the target aquifer.

### 4.3 Information Needed and Sources

Manual water level measurements are needed to corroborate and correct existing water level measurements (documented in the City’s SCADA systems). This information will be used to normalize past measurements (relative to sea level) which will allow a better assessment of groundwater trends and aquifer conditions (e.g., groundwater flow direction and velocity, and available storage volumes of the target aquifer). Pressure transducers will also be deployed as needed to continuously monitor groundwater levels.

Water quality data is also needed from potential surface water sources and the target aquifer. Previous water quality data collected by the City (as part of DOH compliance) will be compiled, along with the data that is the subject of this QAPP. Additional data collected by the City under its own funding in 2023 will also be compiled, but appropriately caveated since it will be collected outside the scope of this QAPP.

Additional details on field data collection for the Study are provided in Section 7.2.

### 4.4 Tasks Required

Study objectives require completing the following tasks.

#### **Task 1: Water Quality Sampling and Analyses**

The task will determine source water quality and background water quality in the target aquifer, in accordance with Ecology guidelines.

**Task 1.1: Water Quality Sampling.** This task includes sampling of potential surface water sources (e.g., SMID Canal and Roza Main Canal) and groundwater. Source water quality data will be used to determine water treatment requirements for municipal and ASR uses. Groundwater sampling will be collected from at least two and up to four City wells during a single sampling event to assess spatial variability of water quality within the target aquifer. During both groundwater and surface water sampling, field water quality parameters (i.e., pH, specific conductance, temperature, ORP, dissolved oxygen, and turbidity) will be collected.

### **Task 2: Development of a Conceptual Hydrogeologic Model**

The task is to refine the existing conceptual hydrogeologic model to evaluate the feasibility of implementing ASR and address information requirements of Chapter 173-157-120 WAC.

**Task 2.1: Water Level Measurements.** This task includes taking water levels in all the City wells to evaluate groundwater trends and aquifer conditions (e.g., groundwater flow direction and velocity, and available storage volumes of the target aquifer). Groundwater level measurements will be compared to historical SCADA data to evaluate long-term groundwater trends as documented in City wells.

It is recognized that additional testing and data collection may be required to satisfy certain information requirements of Chapter 173-157 WAC, which may be beyond the scope of this phase of the Study and will be identified for data collection in subsequent project phases.

## **4.5 Systematic Planning Process**

Finalization of this QAPP is adequate systematic planning for the project.

## 5.0 Organization and Schedule

### 5.1 Key Individuals and Their Responsibilities

Table 5 shows the responsibilities of those who will be involved in this project.

**Table 5. Organization of Project Staff and Responsibilities**

Staff	Title	Responsibilities
Scott Tarbutton Office of Columbia River Phone: (509) 867-6534	OCR Quality Assurance Coordinator	Provides internal review of the QAPP and approves the final QAPP
McKenna Murray Office of Columbia River Phone: (509) 823-0996	OCR Project Manager	Provides oversight for the Study and Ecology Grant. Clarifies scope of the project. Provides review of the QAPP.
John Kirk Water Resources Program Phone: (509) 457-7146	Hydrogeologist	Provides technical oversight and review of the study, provides technical and permitting support
Jeff Burkett City of Moxee Phone: (509) 575-8851	Public Works	Reviews the draft and final QAPP and project deliverables, submittals for the Ecology Grant
Justin Bellamy HLA Engineering Phone: (509) 966-7000	City Engineer	HLA Project Manager, Completes the Engineering Evaluation of Water System
Tyson Carlson Aspect Consulting Phone: (509) 895-5923	Principal Investigator and Project Manager	Co-author of QAPP, Aspect Project Manager, approach development, data analysis, QA/QC
Jason Shira Aspect Consulting Phone: (206) 838-5843	Senior Hydrogeologist	Technical oversight data analysis
Silas Sleeper Aspect Consulting Phone: (206) 453-6058	Field Geologist	Co-author of QAPP. Collects data and records field information.
Ian Lauer Aspect Consulting Phone: (509) 888-1527	Field Geologist	Plans/schedules field dates/logistics. Procures equipment. Collects data and records field information.
Lea Beard Aspect Consulting Phone (206) 780-7749	Data Scientist	Reviews and uploads EIM data.
Giles Hamilton LabTest (509) 575-3999	Laboratory Manager	Prepares laboratory reports, conducts laboratory QA/QC.
Justin Doty Anatek Labs, Inc. (208) 883-2839	Project Manager	Prepares laboratory reports
Todd Taruscio Anatek Labs, Inc. (208) 883-2839	Laboratory Manager	Prepares laboratory reports, conducts laboratory QA/QC.

### 5.2 Special Training and Certifications

A hydrogeologist licensed in the State of Washington will perform all analysis and interpretation of field data and provide oversight of hydrogeologic data collection. All field staff involved in this project will have either the relevant experience in the required standard operating procedures (SOPs) or be trained by more senior field staff or the project manager who have the required

experience. The experienced staff will then lead the field data collection and oversee/mentor less-experienced staff.

### 5.3 Organization Chart

Not applicable – See Table 5.

### 5.4 Proposed Project Schedule

Table 6 below provides the anticipated project schedule proposed under this project.

**Table 6. Tentative Project Schedule**

<b>Task</b>	<b>Completion Date</b>	<b>Note</b>
Final QAPP	September 2023	--
Groundwater and Surface Water Quality Testing	October 2023	Task will commence at the start of the irrigation season
Submit Draft Feasibility Analysis Report	September 2024	--
Receive Ecology Comments	October / November 2024	
Database uploaded to EIM	December 2024	--
Complete Final Report	December 2024	Following receipt and discussion of Ecology comments on the draft report.

### 5.5 Budget and Funding

The City has received funding from Ecology’s Office of Columbia River and the YBIP Groundwater Subcommittee (Agreement No. WRYBIP-2123-Moxeec-00036) to conduct the Study and all tasks as described in Section 4.4. Aspect and HLA are under contract to the City to prepare this QAPP and complete the Study. This work builds upon two Moxee Valley shallow aquifer recharge studies (Aspect, 2020; Sleeper, 2020) that were completed with previous YBIP Groundwater Subcommittee funding.

## 6.0 Quality Objectives

### 6.1 Data Quality Objectives <sup>1</sup>

The main data quality objective (DQO) for this Study is to collect water quality samples from potential surface water and groundwater sites, as well as measure (periodic and continuous) water levels from City wells shown on Figure 1. These analyses will use common methodologies to evaluate water quality and groundwater flow direction that meet the measurement quality objectives (MQOs) described below.

### 6.2 Measurement Quality Objectives

Measurement Quality Objectives (MQOs) are statements of the precision, bias, and lower measurement limits necessary to meet the Study objectives. Precision and bias together express data accuracy, whereas other considerations include the representativeness, completeness, and comparability of the data.

The field investigation will be conducted to measure water levels, collect representative water samples for analyses, and measure water quality field parameters. The MQOs for the field investigation are described by the analytical methods and field equipment used to collect measurements, and the standard operating procedures employed to make descriptions in the field.

#### 6.2.1 Targets for Precision, Bias, and Sensitivity

The data collection instrumentation will meet the MQOs listed in Table 7, and the groundwater samples will be analyzed using standard methods that meet the MQOs listed in Table 8.

---

<sup>1</sup> DQO can also refer to **Decision** Quality Objectives. The need to identify Decision Quality Objectives during the planning phase of a project is less common. For projects that do lead to important decisions, DQOs are often expressed as tolerable limits on the probability or chance (risk) of the collected data leading to an erroneous decision. And for projects that intend to estimate present or future conditions, DQOs are often expressed in terms of acceptable uncertainty (e.g., width of an uncertainty band or interval) associated with a point estimate at a desired level of statistical confidence.



**Table 7. Field Method MQOs and Field Equipment Information**

Parameter	Equipment /Method	Bias (median)	Precision Field Duplicates (median)	Equipment Information			Expected Range
				Accuracy	Resolution	Range	
<b>Air Monitoring</b>							
Temperature	Van Essen Baro-Diver	--	--	0.1°C	0.01°C	-10 to 50°C	-7 to 31°C
Barometric Pressure	Van Essen Baro-Diver	--	--	0.016 ft-H <sub>2</sub> O	0.003 ft-H <sub>2</sub> O	--	29 to 33 ft-H <sub>2</sub> O
<b>Groundwater Level Measurements</b>							
Temperature	Van Essen TD-Diver	--	--	0.1°C	0.01°C	0 to 50°C	1 to 25°C
Pressure	Van Essen TD-Diver	--	--	0.016 ft-H <sub>2</sub> O	0.007 ft-H <sub>2</sub> O	max 330 ft-H <sub>2</sub> O	20 to 200 ft-H <sub>2</sub> O
	Weiss	--	--	0.5% Full Scale	0.01 PSI	Max 200 PSI	
Depth to Water Table	Electronic Water level	--	--	0.05 ft	0.01 ft	--	250 to 750 ft
Wellhead Position (GPS)	Arrow Gold+ GNSS Receiver	--	--	0.3 feet	0.01 ft	--	--
<b>Field Water Quality Parameters</b>							
pH	AquaTroll 500	--	--	0.1 SU	0.01 SU	0 to 14 SU	6.5 to 8.5 SU
Specific conductivity		--	--	±0.5% + 1 uS/cm	0.1 uS/cm	0 to 350,000 uS/cm	150 to 500 uS/cm
Dissolved oxygen		--	--	± 0.1mg/L	0.01 mg/L	0 to 20 mg/L	0 to 10 mg/L
Oxidation-Reduction Potential		--	--	±5 mV	0.1 mV	-1400 to +1400 mV	-300 to +300 mV
Temperature		--	--	±0.1°C	0.01°C	-5 to 50°C	1 to 25°C

**Notes:** mV = millivolts; ft H<sub>2</sub>O = feet of water; PSI = pounds per square inch; SU = standard units; uS/cm = microsiemens per centimeter; mg/L = milligrams per liter; °C = temperature in Celsius

**Table 8. Laboratory MQOs of Water Samples**

Analytical Method	Analyte	Method Detection Limit	Method Reporting Limit	Units	Field Dup. (RPD)	Matrix Spike (%Rec)	Matrix Spike (RPD)	Blank Spike (LCS % Rec)	Blank Spike (RPD)
<b>General Chemistry, Inorganics in Drinking Water</b>									
EPA 300.0	Bromide	0.0130	0.100	mg/L	-	90-110	20	90-110	20
EPA 300.0	Chloride	0.0280	0.100	mg/L	-	90-110	20	90-110	20
EPA 300.0	Fluoride	0.0140	0.100	mg/L	-	90-110	20	90-110	20
EPA 300.0	Nitrate/N	0.0180	0.100	mg/L	20	90-110	20	90-110	20
EPA 300.0	Nitrite/N	0.0180	0.100	mg/L	-	90-110	20	90-110	20
EPA 300.0	Sulfate	0.0170	0.100	mg/L	-	90-110	20	90-110	20
SM 2320 B	Alkalinity	2.00	2.00	mg/L	-	85-115	20	85-115	20
SM 2320 B	Bicarbonate	2.00	2.00	mg/L	-	-	-	-	-
SM 2540 C	TDS	43.6	50.0	mg/L	10	80-120	20	80-120	20
SM 2540 D	TSS	1.00	1.00	mg/L	10	-	-	-	-
SM 4500-P F	Total P	0.00698	0.0100	mg/L	-	80-120	25	80-120	25
SM 5310 B	DOC	0.100	0.500	mg/L	-	70-130	30	80-120	20
SM 5310 B	TOC	0.0600	0.100	mg/L	-	70-130	30	85-115	15
<b>Metals by ICP in Drinking Water</b>									
EPA 200.7	Aluminum	0.00800	0.0100	mg/L	-	70-130	20	85-115	-
EPA 200.7	Calcium	0.0182	0.100	mg/L	20	70-130	20	85-115	20
EPA 200.7	Dissolved Aluminum	0.00700	0.0100	mg/L	-	70-130	20	85-115	-
EPA 200.7	Dissolved Calcium	0.0173	0.100	mg/L	-	70-130	20	85-115	-
EPA 200.7	Dissolved Iron	0.00720	0.0100	mg/L	-	70-130	20	85-115	-
EPA 200.7	Dissolved Magnesium	0.0154	0.100	mg/L	-	70-130	20	85-115	-
EPA 200.7	Dissolved Potassium	0.0521	0.500	mg/L	-	70-130	20	85-115	20
EPA 200.7	Dissolved Silicon	0.0435	0.100	mg/L	-	70-130	20	85-115	-
EPA 200.7	Dissolved Sodium	0.0124	0.100	mg/L	-	70-130	20	85-115	-
EPA 200.7	Iron	0.00720	0.0100	mg/L	20	70-130	20	85-115	20
EPA 200.7	Magnesium	0.0154	0.100	mg/L	20	70-130	20	85-115	20
EPA 200.7	Potassium	0.0521	0.500	mg/L	20	70-130	20	85-115	20
EPA 200.7	Silica (as SiO <sub>2</sub> )	0.0930	0.214	mg/L	-	-	-	-	-
EPA 200.7	Silicon	0.100	0.100	mg/L	20	70-130	20	85-115	20
EPA 200.7	Sodium	0.0124	0.100	mg/L	20	70-130	20	85-115	20
EPA 200.7	Diss. Silica (as SiO <sub>2</sub> )	0.0930	0.214	mg/L	-	-	-	-	-
<b>Metals by ICP-MS in Drinking Water</b>									
EPA 200.8	Antimony	0.000330	0.00100	mg/L	-	70-130	20	85-115	-
EPA 200.8	Arsenic	0.000830	0.00100	mg/L	-	70-130	20	85-115	-
EPA 200.8	Barium	0.0000800	0.000130	mg/L	-	70-130	20	85-115	-
EPA 200.8	Beryllium	0.000150	0.000300	mg/L	-	70-130	20	85-115	-
EPA 200.8	Cadmium	0.000132	0.00100	mg/L	-	70-130	20	85-115	-
EPA 200.8	Chromium	0.000990	0.00100	mg/L	-	70-130	20	85-115	-
EPA 200.8	Copper	0.000267	0.00100	mg/L	-	70-130	20	85-115	-
EPA 200.8	Dissolved Antimony	0.000330	0.00100	mg/L	-	70-130	20	85-115	-
EPA 200.8	Dissolved Arsenic	0.000830	0.00100	mg/L	-	70-130	20	85-115	-
EPA 200.8	Dissolved Barium	0.000130	0.00100	mg/L	-	70-130	20	85-115	-

Analytical Method	Analyte	Method Detection Limit	Method Reporting Limit	Units	Field Dup. (RPD)	Matrix Spike (%Rec)	Matrix Spike (RPD)	Blank Spike (LCS % Rec)	Blank Spike (RPD)
EPA 200.8	Dissolved Beryllium	0.000150	0.00100	mg/L	-	70-130	20	85-115	-
EPA 200.8	Dissolved Cadmium	0.000132	0.00100	mg/L	20	70-130	20	85-115	-
EPA 200.8	Dissolved Chromium	0.000990	0.00100	mg/L	-	70-130	20	85-115	-
EPA 200.8	Dissolved Copper	0.000267	0.00100	mg/L	-	70-130	20	85-115	-
EPA 200.8	Dissolved Lead	0.000430	0.00100	mg/L	-	70-130	20	85-115	-
EPA 200.8	Dissolved Manganese	0.000110	0.00100	mg/L	-	70-130	20	85-115	-
EPA 200.8	Dissolved Nickel	0.000430	0.00100	mg/L	-	70-130	20	85-115	-
EPA 200.8	Dissolved Selenium	0.000460	0.00100	mg/L	-	70-130	20	85-115	-
EPA 200.8	Dissolved Silver	0.000270	0.00100	mg/L	-	70-130	20	85-115	-
EPA 200.8	Dissolved Thallium	0.000160	0.00100	mg/L	-	70-130	20	85-115	-
EPA 200.8	Dissolved Uranium	0.000290	0.00100	mg/L	-	70-130	20	85-115	-
EPA 200.8	Dissolved Zinc	0.000760	0.00100	mg/L	-	70-130	20	85-115	-
EPA 200.8	Lead	0.000430	0.00100	mg/L	-	70-130	20	85-115	-
EPA 200.8	Manganese	0.000110	0.00100	mg/L	-	70-130	20	85-115	-
EPA 200.8	Nickel	0.000430	0.00100	mg/L	-	70-130	20	85-115	-
EPA 200.8	Selenium	0.000460	0.00100	mg/L	-	70-130	20	85-115	-
EPA 200.8	Silver	0.000270	0.00100	mg/L	-	70-130	20	85-115	-
EPA 200.8	Thallium	0.000160	0.00100	mg/L	-	70-130	20	85-115	-
EPA 200.8	Uranium	0.000290	0.00100	mg/L	-	70-130	20	85-115	-
EPA 200.8	Zinc	0.000760	0.00100	mg/L	-	70-130	20	85-115	-
<b>Mercury in Water</b>									
EPA 245.7	Dissolved Mercury	0.000200	0.00100	ug/L	18	63-111	18	76-113	18
EPA 245.7	Mercury	0.000200	0.00100	ug/L	18	63-111	18	63-113	-
<b>Semivolatiles in Drinking Water</b>									
EPA 505	gamma-BHC (Lindane)	0.00320	0.0200	ug/L	25	65-135	25	70-130	20
EPA 505	Heptachlor	0.00360	0.0400	ug/L	25	65-135	25	70-130	20
EPA 505	Aldrin	0.00480	0.100	ug/L	25	65-135	25	70-130	20
EPA 505	Heptachlor epoxide	0.00160	0.0200	ug/L	25	65-135	25	70-130	20
EPA 505	4,4'-DDE	0.00180	0.100	ug/L	25	65-135	25	70-130	20
EPA 505	Dieldrin	0.00170	0.100	ug/L	25	65-135	25	70-130	20
EPA 505	Endrin	0.00240	0.0100	ug/L	25	65-135	25	70-130	20
EPA 505	4,4'-DDD	0.00210	0.100	ug/L	25	65-135	25	70-130	20
EPA 505	4,4'-DDT	0.00520	0.100	ug/L	25	65-135	25	70-130	20
EPA 505	Methoxychlor	0.00460	0.100	ug/L	25	65-135	25	70-130	20
EPA 505	Aroclor 1232 (PCB-1232)	0.100	0.500	ug/L	25	65-135	25	70-130	20
EPA 505	Aroclor 1242 (PCB-1242)	0.100	0.300	ug/L	25	65-135	25	70-130	20
EPA 505	Aroclor 1248 (PCB-1248)	0.100	0.100	ug/L	25	65-135	25	70-130	20
EPA 505	Aroclor 1254 (PCB-1254)	0.100	0.100	ug/L	25	65-135	25	70-130	20
EPA 505	Aroclor 1260 (PCB-1260)	0.0375	0.200	ug/L	25	65-135	25	70-130	20
EPA 505	PCBs	0.0950	0.500	ug/L	25	65-135	25	70-130	20
EPA 505	Chlordane	0.0715	0.200	ug/L	25	65-135	25	70-130	20
EPA 505	Toxaphene	0.227	1.00	ug/L	25	65-135	25	70-130	20
EPA 515.4	Dalapon	0.531	1.00	ug/L	20	70-130	30	70-130	20
EPA 515.4	Dicamba	0.0710	0.200	ug/L	20	70-130	30	70-130	20
EPA 515.4	Dichloroprop	0.260	0.500	ug/L	20	70-130	30	70-130	20

Analytical Method	Analyte	Method Detection Limit	Method Reporting Limit	Units	Field Dup. (RPD)	Matrix Spike (%Rec)	Matrix Spike (RPD)	Blank Spike (LCS %Rec)	Blank Spike (RPD)
EPA 515.4	2,4-D	0.0330	0.100	ug/L	20	70-130	30	70-130	20
EPA 515.4	Pentachlorophenol	0.00900	0.0400	ug/L	20	70-130	30	70-130	20
EPA 515.4	2,4,5-TP (Silvex)	0.0350	0.200	ug/L	20	70-130	30	70-130	20
EPA 515.4	2,4-DB	0.240	1.00	ug/L	20	70-130	30	70-130	20
EPA 515.4	Dinoseb	0.0680	0.200	ug/L	20	70-130	30	70-130	20
EPA 515.4	Picloram	0.0480	0.100	ug/L	20	70-130	30	70-130	20
EPA 515.4	3,5-Dichlorobenzoic Acid	0.156	0.500	ug/L	20	70-130	30	70-130	20
EPA 515.4	Chloramben	0.0490	0.200	ug/L	20	70-130	30	70-130	20
EPA 515.4	Acifluorfen	0.322	1.00	ug/L	20	70-130	30	70-130	20
EPA 525.2	Alachlor	0.0550	0.200	ug/L	30	20-130	30	20-130	25
EPA 525.2	Atrazine	0.0670	0.100	ug/L	30	20-130	30	20-130	25
EPA 525.2	Benzo[a]pyrene	0.0100	0.0200	ug/L	30	20-130	30	20-130	25
EPA 525.2	bis(2-Ethylhexyl)phthalate	0.127	0.600	ug/L	30	20-150	30	20-150	25
EPA 525.2	bis-2(ethylhexyl)adipate	0.0690	0.600	ug/L	30	20-150	30	20-150	25
EPA 525.2	Bromacil	0.0500	0.100	ug/L	30	20-130	30	20-130	25
EPA 525.2	Butachlor	0.0590	0.100	ug/L	30	20-130	30	20-130	25
EPA 525.2	Fluorene	0.0350	0.200	ug/L	30	20-130	30	20-130	25
EPA 525.2	gamma-BHC (Lindane)	0.0152	0.0400	ug/L	30	20-130	30	20-130	25
EPA 525.2	Hexachlorobenzene	0.0370	0.100	ug/L	30	20-130	30	20-130	25
EPA 525.2	Hexachlorocyclopentadiene	0.0410	0.100	ug/L	30	20-130	30	20-130	25
EPA 525.2	Methoxychlor	0.0480	0.200	ug/L	30	20-130	30	20-130	25
EPA 525.2	Metribuzin	0.0570	0.100	ug/L	30	20-130	30	20-130	25
EPA 525.2	Propachlor	0.0540	0.100	ug/L	30	20-130	30	20-130	25
EPA 525.2	Simazine	0.0630	0.0700	ug/L	30	20-130	30	20-130	25
EPA 549.2	Diquat	0.208	0.400	ug/L	20	70-130	25	70-130	20
SM 6251 B	Monochloroacetic acid	0.437	2.00	ug/L	20	70-130	20	70-130	20
SM 6251 B	Monobromoacetic acid	0.272	1.00	ug/L	20	70-130	20	70-130	20
SM 6251 B	Dichloroacetic acid	0.374	1.00	ug/L	20	70-130	20	70-130	20
SM 6251 B	Trichloroacetic acid	0.483	1.00	ug/L	20	70-130	20	70-130	20
SM 6251 B	Bromochloroacetic acid (BCAA)	0.191	1.00	ug/L	20	70-130	20	70-130	20
SM 6251 B	Dibromoacetic acid	0.275	1.00	ug/L	20	70-130	20	70-130	20
SM 6251 B	Total HAA5	0.500	1.00	ug/L	20	70-130	20	70-130	20

**Volatiles in Drinking Water**

EPA 524.3	1,3-Dichloropropene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Total Trihalomethane	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Benzene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Bromochloromethane	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Bromodichloromethane	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Bromoform	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Bromomethane	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Carbon Tetrachloride	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Chlorobenzene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Chloroform	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Chloromethane	0.100	0.500	ug/L	-	70-130	30	70-130	20

Analytical Method	Analyte	Method Detection Limit	Method Reporting Limit	Units	Field Dup. (RPD)	Matrix Spike (%Rec)	Matrix Spike (RPD)	Blank Spike (LCS % Rec)	Blank Spike (RPD)
EPA 524.3	cis-1,2-dichloroethene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	cis-1,3-Dichloropropene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	1,2-Dibromo-3-chloropropane (DBCP)	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	1,2-Dibromoethane (EDB)	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	1,2-Dichlorobenzene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	1,3-Dichlorobenzene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	1,4-Dichlorobenzene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Dichlorodifluoromethane	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	1,1-Dichloroethane	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	1,2-Dichloroethane	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	1,1-Dichloroethene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	trans-1,2-Dichloroethene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	1,2-Dichloropropane	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	trans-1,3-Dichloropropene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Ethylbenzene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Hexachlorobutadiene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Isopropylbenzene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Methylene chloride	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Naphthalene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Styrene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	1,1,1,2-Tetrachloroethane	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	1,1,2,2-Tetrachloroethane	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Tetrachloroethene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Toluene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	1,2,4-Trichlorobenzene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	1,1,1-Trichloroethane	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	1,1,2-Trichloroethane	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Trichloroethene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Trichlorofluoromethane	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	1,2,3-Trichloropropane	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Vinyl Chloride	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	m+p-Xylene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	o-Xylene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Total Xylene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	1,1-dichloropropene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	1,2,3-Trichlorobenzene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	1,2,4-Trimethylbenzene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Chloroethane	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	1,3,5-Trimethylbenzene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	2,2-Dichloropropane	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	1,3-Dichloropropane	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	2-Chlorotoluene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	4-Chlorotoluene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Bromobenzene	0.100	0.500	ug/L	-	70-130	30	70-130	20

Analytical Method	Analyte	Method Detection Limit	Method Reporting Limit	Units	Field Dup. (RPD)	Matrix Spike (%Rec)	Matrix Spike (RPD)	Blank Spike (LCS % Rec)	Blank Spike (RPD)
EPA 524.3	Dibromochloromethane	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Dibromomethane	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	methyl-t-butyl ether	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	(MTBE) n-Butylbenzene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	n-Propylbenzene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	p-isopropyltoluene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	sec-Butylbenzene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	tert-Butylbenzene	0.100	0.500	ug/L	-	70-130	30	70-130	20
EPA 524.3	Acetone	0.500	2.50	ug/L	-	70-130	30	70-130	20
EPA 524.3	MTBE-d3	-	-	Surr.	-	70-130	-	-	-
EPA 524.3	4-Bromofluorobenzene	-	-	Surr.	-	70-130	-	-	-
EPA 524.3	1,2-Dichlorobenzene-d4	-	-	Surr.	-	70-130	-	-	-

**Notes:** Dup. = Duplicate Sample, RPD = relative percent difference, LCS = laboratory control sample, %Rec = percent recovered, Surr. = Surrogate

### Water Quality Analyses

The MQOs for the water quality analyses are summarized above in Table 8, including samples, laboratory blanks, and duplicates. Water quality sampling will be performed using industry-standard procedures to minimize bias and maximize precision. One field duplicate and data validation (DV) sample will be collected during each sampling event (Section 7, Table 9). All sampling equipment will be decontaminated before and after completion of sampling activities. Additional quality control procedures are detailed in Section 10.

Anatek Labs, Inc. (Anatek) and LabTest are accredited by Ecology for all analytical procedures performed for this project and by the National Environmental Laboratory Accreditation Program (NELAP) for a comprehensive analytical laboratory accreditation. LabTest will perform nitrate, nitrite, and bacteriological analyses and Anatek will perform all remaining analyses. The laboratories are responsible for ensuring that all procedures performed comply with all requirements specified in the accreditation programs, laboratory quality assurance (QA) manuals, individual analytical methods, and this QAPP. Copies of the lab accreditation for Anatek and LabTest are included as Appendix A.

The quality and usability of data collected will be determined, based on the outcomes of data verification and validation, and expressed as data quality indicators (DQIs): precision, accuracy (bias), representativeness, comparability, completeness, and sensitivity. The DQIs routinely obtained by the laboratory for the analytical procedures performed for this project are considered adequate. The definitions of the DQIs are presented as follows:

#### 6.2.1.1 Precision

Precision is defined as the degree of agreement between or among independent, similar, or repeated measurements. Precision is a measure of variability in the results of replicate measurements due to random error. Precision is usually assessed by analyzing duplicate field

measurements and random error is imparted by the variation in field procedures. Therefore, field sampling precision is addressed by collection of replicate measurements.

Precision is also expressed in terms of analytical variability. For this investigation, analytical variability will be measured as the relative percent difference (RPD) or coefficient of variation between analytical laboratory duplicates and between the matrix spike (MS) and matrix spike duplicate (MSD) analyses. Precision will be calculated as the RPD as follows:

$$RPD (\%) = 100 \times \frac{|S - D|}{(S + D)/2}$$

where:

S = analyte concentration in a sample  
D = analyte concentration in a duplicate sample

The resultant RPD will be compared with criteria established by this QAPP in Table 8, and deviations from these criteria will be reported. If the QAPP criteria are not met, the laboratory will supply a justification of why the limits were exceeded and implement the appropriate corrective actions. The RPD will be evaluated during data review and validation. The data reviewer will note deviations from the specified limits and will comment on the effect of the deviations on the reported data.

#### **6.2.1.2 Bias**

Bias is the difference between the sample mean and the true value. It will be measured as the percent recoveries of MS and MSD, organic surrogate compounds, and the laboratory control sample (LCS). Additional potential bias will be assessed using calibration standards and blank samples (e.g., method blanks), which are detailed in Section 7, Table 9 and Section 10. In cases where accuracy is determined from spiked samples, accuracy will be expressed as the percent recovery. The closer these values are to 100 percent, the more accurate the data.

**Surrogate recovery will be calculated as follows:**

$$\text{Recovery (\%)} = \frac{MC}{SC} \times 100$$

where:

SC = spiked concentration  
MC = measured concentration

**MS percent recovery will be calculated as follows:**

$$\text{Recovery (\%)} = \frac{MC - USC}{SC} \times 100$$

where:

SC = spiked concentration  
MC = measured concentration  
USC = unspiked sample concentration

**MSD percent recovery will be calculated as follows:**

$$\text{Recovery (\%)} = \frac{MDC - USC}{SC} \times 100$$

where:

SC = spiked concentration  
MDC = measured duplicate spike concentration  
USC = unspiked sample concentration

and

$$\text{RPD (\%)} = \frac{MC - MDC}{(MC + MDC)/2} \times 100,$$

where:

RPD = relative percent difference.

Field staff will minimize bias in the field measurements by strictly following equipment calibration and measurement protocols. Potential sources of field bias in measurements include measurement procedure, inability to measure all forms of the parameter of interest, and calibration problems. Table 7 presents the bias data quality objectives for pressure transducer and temperature sensor data for instrument QC checks.

The resultant percent recoveries will be compared with criteria established by this QAPP in Table 8, and deviations from these criteria will be reported (and in laboratory limits for RPD reported by the lab in individual reports). If the objective criteria are not met, the laboratory will supply a justification of why the limits were exceeded and implement the appropriate corrective actions. Percent recoveries will be evaluated during data review and validation, and the data reviewer will comment on the effect of the deviations on the reported data.

### **Groundwater Level Monitoring**

The MQOs for the groundwater level monitoring of supply wells are as follows:



- Obtain horizontal well locations within 2-meter (6.5 feet) accuracy;
- Obtain the elevation (if not already obtained) of the wellhead or water level reference point relative to ground surface;
- Obtain ground surface elevations within a 3-foot accuracy (using GPS measurements, with elevations cross-referenced with a 10-meter digital elevation model available from the Washington State Department of Natural Resources);
- Obtain groundwater level measurements within a 0.1-foot accuracy. Measurements are recorded to  $\pm 0.01$  foot and are accurate to  $\pm 0.05$  foot per 100 feet (Jelinski et al., 2015); and
- Continuous measurement of groundwater levels is conducted using a pressure transducer with an onboard datalogger. Measurement of barometric pressure is necessary to correct measured water level data for the effects of changes in atmospheric pressure. Calibration and maintenance of pressure transducers are provided by the manufacturer and should be consulted. Table 7 provides accuracy and resolution for Van Essen Baro- and TD-Diver typically used for long-term deployments.

A description of the water level monitoring techniques that will be used to obtain the MQOs for the water level measurements and well locations is provided in the *Field Procedures* section (Section 8.2).

### **6.2.1.3 Sensitivity**

Sensitivity will be determined by reviewing Method Reporting Limits (MRLs). MRLs will be set low enough to allow meaningful comparisons with screening criteria to the extent possible, taking into account matrix effects. The laboratory will be directed to report compounds detected above the Method Detection Limit (MDL) and positively identified below the MRL as estimated (J flag).

Sensitivity is also a measure of the capability of the field method and instrument used to detect a change. It is described by its range, accuracy, and resolution. This is usually reported for each instrument by the manufacturer. Examples of this information are provided in Table 7.

## **6.2.2 Targets for Comparability, Representativeness, and Completeness**

### **6.2.2.1 Comparability**

Comparability is the degree to which the data can be compared to historical data, reference values (such as background), and reference materials. This will be achieved through the use of standard operating procedures (SOPs) to collect field measurements and samples, training of field staff, field data-collection similarities (location, duration, time of year, weather conditions, etc.), instrumentation sensitivity, EPA-approved methods to analyze samples, and consistent units to report analytical results. Data comparability also depends on data quality. Data of unknown quality cannot be compared.

### 6.2.2.2 Representativeness

Representativeness is the degree to which sample results represent the system under Study. This component is generally considered during the design phase of a program. This program will use the results of all analyses to evaluate the data in terms of its intended use. Typically, a combination of continuous measurements, spot measurements, and historical data is needed to represent the expected variability of spatial and temporal conditions.

Representativeness of the measurements and samples will be ensured during the collection process by: (1) employing proper decontamination procedures, (2) thorough purging of the well and ensuring stability of field parameters prior to collecting groundwater samples (Section 6.3), and (3) use of continuous monitoring equipment for groundwater level monitoring. The representativeness of analytical results will be determined by evaluating hold times, sample preservation, and blank contamination (e.g., trip blanks). Samples with expired hold times, improper preservation, or contamination may not be representative.

### 6.2.2.3 Completeness

**Completeness will be calculated as follows:**

$$\text{Completeness (\%)} = \frac{V}{P} \times 100$$

where:

- V = number of valid measurements  
P = number of planned measurements

Valid and invalid data (i.e., data qualified with the R flag [rejected]) will be identified during data validation. The completeness target for the Study is 100 percent of water quality samples. However, problems occasionally arise during data collection. A completeness of 95 percent is acceptable for discrete measurements. In general, the project is designed to accommodate some data loss and still meet project goals and objectives.

For continuous deployed measurements, additional variables can negatively impact completeness, including vandalism/theft/tampering, equipment failure, unacceptable fouling or drift, and unpredictable hydrologic events (steep drops in water level between visits). For these reasons, a completeness of 80 percent is acceptable for continuous measurements. Given these difficulties, redundancy is an important component when designing studies with continuous data collection, particularly at important boundary conditions and within the most critical areas. If completeness targets are not achieved, then a determination will be made as to whether the data that were successfully collected are sufficient to meet project needs. This will depend on a number of factors, such as the needs of the analysis framework, and the times and locations where data were lost. If successfully collected data are not sufficient, then one or a combination of the following approaches will be used:

1. Estimate missing data values from existing data, if this can be done with reasonable confidence;

2. Conduct targeted additional sampling to fill data gaps; and
3. Recollect all or a portion of data.

If completeness targets are not met, the study report will analyze the effect of the incomplete data on meeting the study objectives, account for data completeness (or incompleteness) in any data analyses, and document data completeness and its consequences in any study reports.

### **6.3 Acceptance Criteria for Quality of Existing Data**

Existing groundwater quality data was collected in the study area under YBIP Groundwater Subcommittee funding (Sleeper, 2020). A QAPP was prepared to support that 2020 study and the data collected follows the same measurement quality objectives discussed in Section 6.2 of this QAPP. The City also conducts water quality sampling at each of its groundwater wells to comply with DOH drinking water source requirements, but no Ecology-approved QAPP was prepared for this work.

### **6.4 Model Quality Objectives**

Not applicable.

## 7.0 Study Design

The study-design is a non-randomized study design. Sampling locations and analytical suites are preselected based on opportunistically available locations and the study objectives. A narrative of the overall study objective is provided in Section 4. This section provides the details of the data collection and analysis.

### 7.1 Study Boundaries

The study area is shown on Figure 1. Overall, this Study considers the performance of the target storage aquifer over the conceptual boundary shown on Figure 1. However, the data collection activities for the Study will not extend beyond the footprint of the Moxee Valley.

### 7.2 Field Data Collection

#### 7.2.1 Sampling Locations and Frequency

Water quality sampling and water level measurements will occur according to the schedule shown in Table 9. The analyte suite is described in Sections 3.2.3 and 6.2 and will be sampled according to the quality objectives described in Section 6. The key considerations for the sampling schedule are outlined in the following sections.

**Table 9. Water Quality Sampling and Groundwater Level Monitoring Schedule**

Anticipated Scheduling Date	Surface Water Sources (SMID Canal and Roza Main Canal)						City Owned Groundwater Wells (Wells 1, 2, 3, and 4)
Fall 2023	Gen Chem	SVOC	Herbicide and & Pesticides	Bact.	Radionuclide	PFAS	General Chemistry, PFAS
Fall 2023							Depth to Water – Manual Pressure - Continuous

**Notes:**

Field parameters will be measured during every sampling event.

One field duplicate and data validation (DV) sample will be collected during each sampling event. The DV sample for a trip blank will include the VOC, general chemistry, and bacteria sample suites (note that no MS/MSD analyses will be completed for bacteria).

#### Water Quality Sampling Schedule

To characterize ambient water quality conditions in the target aquifer, water quality samples will be collected from four spatially distributed production wells during a single sampling event to assess spatial variability of water quality within the target aquifer.

The locations of City Wells are presented in Figure 1 and well construction details are included in Table 10 and well logs are included in Appendix B.

Within 1-2 days of the groundwater sampling all source water samples will be collected during a single sampling event to obtain a snapshot of water quality delivered by the potential source water canals.

**Table 10. City of Moxee Well Attributes Summary**

City Well No.	Ground Surface Elevation (ft amsl) <sup>a</sup>	Well Depth (feet)	Open Interval (ft amsl)	Initial Static Water Level <sup>b</sup> (ft amsl)	Current Static Water Level <sup>c, d</sup> (ft amsl)
1	1,049	1,326	-176 to -231	1,065	1,060
2	1,177	978	411 to 207	1,155	1,085
3	1090	783	Unknown	Unknown	1,094
4	1,177	1,110	302 to 78	1,078	1,078

**Notes:**

a - Approximate elevation obtained from Google Earth.

b - Initial static water level measurement dates: Well 1 (January 1943), Well 2 (March 1983), Well 3 (January 2015), Well 4 (January 2021)

c - Current static water level measurement dates: Well 1 (January 2022), Well 2 (March 2022), Well 3 (January 2022), Well 4 (January 2022)

d - Well 1 is currently artesian

amsl = above mean sea level; ft = feet

## 7.2.2 Field Parameters and Laboratory Analytes to be Measured

Field parameters will be measured using an AquaTroll 500 multimeter, as described in Section 8.2, to provide independent corroboration of laboratory results, and to analyze constituents that have short hold times and can be reliably measured in the field. These include:

- Electrical conductivity
- Dissolved Oxygen
- ORP
- pH
- Temperature
- Turbidity

In addition to manual measurements of the above constituents during sampling events measurements will be collected until values are stable, as described in Section 8.2.

Groundwater depth-to-water measurements will be conducted using an electronic water level indicator as discussed in Section 8. A dedicated pressure transducer will be installed in the subject wells to collect continuous groundwater level measurements.

Laboratory analytes to be measured from water quality sampling throughout the Study are listed above in Section 6.

## **7.3 Modeling and Analysis Design**

### **7.3.1 Analytical Framework**

Data analysis will include evaluating water quality and groundwater levels, following these key considerations:

- Groundwater level trends in City wells will be determined using historical and contemporary groundwater level data.
- Comparison to applicable regulatory criteria summarized in Section 3.2.4.

### **7.3.2 Model Setup and Data Needs**

Not Applicable.

## **7.4 Assumptions of Study Design**

The Study assumes that existing water quality and groundwater level data are of sufficient quality to compare with contemporary data collected under this QAPP.

## **7.5 Possible Challenges and Contingencies**

### **7.5.1 Logistical Problems**

Logistical problems that interfere with measurement collection may occur during field work. These problems include:

1. Inability to access source water and groundwater measurement locations;
2. Inability to install pressure transducers into City wells;
3. Inability to retrieve data from the City's SCADA system;
4. Data quality retrieved from the City's SCADA system does not meet this QAPP MQOs; and
5. Water quality samples meeting hold times and temperature criteria when shipping samples to laboratory for analysis.

### **7.5.2 Practical Constraints**

No practical constraints have been identified for this study.

### **7.5.3 Schedule Limitations**

Schedule limitations include iterative QAPP review and approval and sampling during the irrigation season (about April 1 through October 31) while the canals are fully charged and operational. No other limitations have been currently identified but could potentially arise from unforeseen circumstances.

## 8.0 Field Procedures

### 8.1 Invasive Species Evaluation

Field staff will follow EPA's SOP EAP070, on minimizing the spread of invasive species (Ecology, 2023). At the end of each field visit, field staff will clean field gear in accordance with the SOP for minimizing the spread of invasive species for areas of both moderate and extreme concern.

Field staff will minimize the spread of invasive species after conducting field work by:

Inspecting and cleaning all equipment by removing any visible soil, vegetation, vertebrates, invertebrates, plants, algae, or sediment. If necessary, a scrub brush will be used and then rinsed with clean water either from the site or brought for that purpose. The process will be continued until all equipment is clean.

Draining all water in samplers or other equipment that may harbor water from the site. This step will take place before leaving the sampling site or at an interim site. If cleaning after leaving the sampling site, field staff will ensure that no debris will leave the equipment and potentially spread invasive species during transit or cleaning.

Established Ecology procedures will be followed if an unexpected contamination incident occurs.

### 8.2 Measurement and Sampling Procedures

The procedures used in this Study are typical for hydrogeologic investigations. SOPs to be followed include the following: *Standard Operating Procedure for Manually Obtaining Surface Water Samples* (Ecology, 2006), *Manual Well-Depth and Depth-to-Water Measurements* (Ecology, 2018a), *Standard Operating Procedures to Minimize the Spread of Invasive Species* (Ecology, 2018b), *General Sampling Procedure, Office of Drinking Water* (DOH, 2003), *Use of Submersible Pressure Transducers During Groundwater Studies* (Ecology, 2019), *Purging and Sampling Monitoring Wells for General Chemistry Parameters* (Ecology, 2018c).

#### 8.2.1 Well Location Survey

The horizontal location of the well will be determined using a Trimble GPS. Care will be taken to collect a GPS location with a greater horizontal accuracy than 6.5 feet, as discussed in the *Quality Objectives* section (Section 6). The ground surface elevation will also be determined based on the Trimble GPS and shall have a vertical accuracy of equal to, or better than, 3 feet.

#### 8.2.2 Groundwater Level Monitoring

Manual groundwater levels will be measured at the City's four wells with either an electronic water level indicator or pressure gage. The manual water level measurements will be used to convert the City's SCADA data to depth to water and a common datum (elevation above mean sea level).

Automated water level data will be obtained from pressure transducers reporting to the City's SCADA system. These transducers are vented to the atmosphere, allowing measurement of gaged

(submergence) pressure. Data obtained from the City's SCADA system will be examined for inconsistencies and suspect data flagged for evaluation.

Water levels should be collected using an electrical water level meter with engineer's scale accurate to a hundredth of a foot (0.01 feet). Shut-in pressures should be collected using a pressure gage with an appropriate pressure range and accurate to less than 5 percent of the full range. A permanent measuring point (MP) will be made from which all depth-to-water measurements are taken at each well to ensure data comparability. An MP will be established, or the existing MP will be used if already established.

Establish a permanent measuring point (MP) via the method below:

1. MPs are normally established on the top rim of the actual well casing; this position is commonly referred to as "top of casing" (TOC). Locate the MP at a convenient place from which to measure the water level. If the TOC is level, collect the measurement from the north edge.
2. Clearly mark the MP. The MP must be as permanent as possible and be clearly visible and easily located. The MP may be marked using a permanent black marker, bright colored paint stick, or with a notch filed into the TOC.
3. Describe the position of the MP clearly in the field-data sheets.
4. The MP height is established in reference to a land surface datum (LSD). The LSD is generally chosen to be approximately equivalent to the average altitude of the ground surface around the well.
5. Measure the height of the MP in feet relative to the LSD. Generally, MPs are established to the nearest 0.1 feet using a pocket tape to measure the distance from the MP to the LSD. Note that values for measuring points that lie below land surface should be preceded by a minus sign (-). Record the height of the MP and the date it was established.
6. MPs and the LSD may change over time, the distance between the two should be checked whenever there have been activities, such as land development that could have affected either the MP or LSD at the site. Such changes must be measured as accurately as possible, documented and dated in field-data sheets, and in any database(s) into which the water-level data are entered.

All subsequent water level measurements should be referenced to the established MP. The MP value will be used to convert measurements into values that are relative to land surface.

After a permanent MP is established for each well, continue sampling using the following process:

1. Open the top of the well and note any popping sounds that would indicate pressure buildup, any odors, and the condition of the well head.
2. If there is a pressure transducer attached to the well cap carefully note the initial position of the cap (mark cap position on casing with permanent marker). If the well was airtight, wait a few minutes for the water level to return to equilibrium with atmospheric pressure.



3. Turn the water level meter on and slowly lower the probe into the well until it makes a tone indicating contact with the water level. To confirm contact with the distinct water boundary, slowly raise and lower the electric-tape probe in and out of the water column. If necessary, adjust the sensitivity setting of the meter to provide a “crisp” indication of the water surface. Measure the depth to water against the MP and mark down the date and time the reading was made.
4. At the precise location the indicator shows contact with the water surface, pinch the tape between your fingernails at the MP. Read the depth-to-water.
5. Repeat measurement to ensure that the water level is stable (not rising or falling over time).
6. When the probe is pulled back up, make a note of any mud, staining, or anything else on the tip. Before moving on to the next well, decontaminate the probe with a brush or paper towel, then rinse with distilled water and 10 percent bleach.

On occasion, condensation on the interior casing wall and probe can prematurely trigger the electric-tape indicator giving a false positive reading. In this situation it can help to center the tape in the well casing above the water level and lightly shake the tape to remove the excess water on the probe.

### **8.2.3 Atmospheric Pressure Monitoring**

A barometric pressure transducer and datalogger will be deployed within City limits. Data from this transducer will be used to assess the effects of barometric pressure on water level measurements in City wells. Barometric efficiency can affect the representativeness of water level measurements from vented and unvented transducers (Spane, 2002). Corrections for barometric efficiency of wells will be made, as appropriate.

### **8.2.4 Groundwater and Source Water Sampling**

Groundwater quality samples from City Wells will be collected in general accordance with Ecology (2018c) and DOH (2003) when using existing turbine pumps. Groundwater samples will be collected from the existing sample port at City Wells during operation of the existing pump, prior to any type of water storage or chlorine feed. The well will be purged for a minimum of 10 minutes (or three well volumes) prior to the collection of the groundwater samples or until the water quality parameters stabilize. If necessary, groundwater quality samples will be collected during using low-flow groundwater sampling techniques via a bladder pump.

Field water quality parameters (temperature, pH, specific conductivity, ORP, dissolved oxygen, and turbidity) will be monitored from each well at approximately 3- to 5-minute intervals throughout well purging using an Aqua Troll 500 and flow-through cell plumbed into the sampling port. Water quality parameters will be considered stable when three successive measurements indicate that the parameters fall within the stabilization criteria established in Standard Operating Procedure EAP099 Purging and Sampling Monitoring Wells for General Chemistry Parameters (Ecology, 2018) and shown in Table 11 below. Once the water quality parameters have stabilized, the groundwater quality samples shall be collected from the respective sampling port.

**Table 11. Field Parameter Stabilization Criteria**

Parameter	Value	Units
pH	±0.1	SU
Specific Conductance	±10.0	uS/cm
Dissolved Oxygen	+ 0.05 for values < 1 + 0.2 for values > 1	mg/L
Temperature	±0.1	Degrees Celsius
ORP	±10	millivolts

Source water samples will be collected from the canal bank of the surface water body. Field water quality parameters (temperature, pH, specific conductivity, ORP, dissolved oxygen, and turbidity) will be obtained using an AquaTroll 500 water quality probe. Surface water samples will be collected as a grab sample either by directly dipping the laboratory-supplied sample bottle through the water column, or by pumping water with a peristaltic pump directly into the laboratory-supplied sample bottle, if the canal is too shallow to collect a sample without disturbing the canal bottom.

All samples collected for dissolved metals will be field filtered. Sample will be collected after pumping three filter volumes through filter cartridge. A minimum of one surface water sample will be collected for each site and submitted to the laboratory for analysis.

### 8.3 Containers, Preservation Methods, Holding Times

The sample bottles and respective preservatives for each sample will be provided by the laboratory and filled accordingly. A description of the sample bottles, preservatives and analytical methods are provided in Table 12.

New latex gloves will be worn at all times during the collection of the water quality parameters and samples and switched between locations. Samples for dissolved metal analyses shall be filtered with a 0.45-micron pore-size filter. All bottles shall be clearly labeled with a unique sample name, location name, date, time, and preservative. Samples shall be stored in a cooler at 4 degrees Celsius (°C) and delivered to the laboratory under standard chain-of-custody protocols, within the hold times provided in Table 12.

**Table 12. Containers, Preservation Methods, and Holding Times**

Parameter	Container	Preservative	Holding Time
<b>General Chemistry / Water Quality Parameters (all metals and Dissolved fractions)</b>			
Alkalinity (mg/L)	250 mL Plastic	Unpreserved	14 days
Bicarbonate (mg/L)	1 L Plastic		28 days
Chloride (mg/L)			7 days
TDS (mg/L)			
TSS (mg/L)			
Total Organic Carbon (mg/L)	1 L Plastic	H2SO4	28 days

Parameter	Container	Preservative	Holding Time	
Phosphorous, Total (mg/L)	1 L Plastic	Unpreserved	48 hours	
Bromide (mg/L)				
Fluoride (mg/L)				
Nitrate-N (mg/L)			28 days	
Nitrite-N (mg/L)				
Sulfate (mg/L)				
Silica (silicon) (µg/L)	1 L Plastic	HNO3	6 months	
Arsenic (µg/L)	1 L Plastic	HNO3	6 months	
Antimony (µg/L)				
Aluminum (µg/L)				
Barium (µg/L)				
Beryllium (µg/L)				
Cadmium (µg/L)				
Calcium (µg/L)				
Chromium (µg/L)				
Copper (µg/L)				
Iron (µg/L)				
Lead (µg/L)				
Magnesium (µg/L)				
Manganese (µg/L)				
Mercury (ug/L)				28 days
Nickel (µg/L)				6 months
Potassium (µg/L)				
Selenium (µg/L)				
Silver (µg/L)				
Sodium (µg/L)				
Thallium (µg/L)				
<b>Volatile Organic Compounds (VOCs)</b>				
All VOCs	40 mL VOA	Na2S2O3	14 Days	
<b>Synthetic Organic Compounds (SOCs)</b>				
SOCs Measured Via EPA Methods 508.1 and 525.2	1 L Amber	HCl + Na2SO3	14 Days	
SOCs Measured Via EPA Method 515.4	250 mL Amber	Na2SO3	14 Days	
<b>Herbicides and Pesticides</b>				

Parameter	Container	Preservative	Holding Time
Chlorinated Pesticides	1 L Amber	HCl + Na <sub>2</sub> SO <sub>3</sub>	14 Days
Chlorinated Acid Herbicides	G, Amber, Teflon-Lined Cap	<6°C	14 days until extraction, 21 days after extraction
Pesticides as carbamates	60 mL glass container	30mL/L of C <sub>2</sub> H <sub>3</sub> ClO <sub>2</sub> , 80mg/L of Na <sub>2</sub> S <sub>2</sub> O <sub>3</sub> . <sup>1</sup> Cool 4°C	28 Days
Herbicides – diquat	G, Amber, Teflon-Lined Cap	100mg/L of Na <sub>2</sub> S <sub>2</sub> O <sub>3</sub> , 4°C	14 days until extraction, 21 days after extraction
Herbicides – endothall	G, Amber, Teflon-Lined Cap	4°C	14 days until extraction, 21 days after extraction
Herbicides – glyphosate	Glass Container	100mg/L Na <sub>2</sub> S <sub>2</sub> O <sub>3</sub> , 4°C	14 Days
<b>Bacteriological (LabTest)</b>			
E. coli	250 mL sterile plastic	Na <sub>2</sub> S <sub>2</sub> O <sub>3</sub>	30 hours
Total Coliform			

1. After the addition of C<sub>2</sub>H<sub>3</sub>ClO<sub>2</sub> and Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub>, seal and shake sample bottle for 1 min prior to storage.

## 8.4 Equipment Decontamination

Water samples are collected from dedicated sampling equipment or directly into laboratory provided containers to prevent cross-contamination. All sampling equipment will be decontaminated before and after completion of all sampling activities. Sampling equipment will be decontaminated with an industry standard, phosphorous-free detergent and brush or paper towel, then rinsed with distilled water.

## 8.5 Sample ID

All bottles shall be clearly labeled with a unique sample name, location name, date, time, and preservative. Samples shall be stored in a cooler at 4°C and delivered to the laboratory under standard chain-of-custody protocols, within the hold times provided in Table 12.

## 8.6 Chain of Custody

After collection, samples will be maintained in Aspect's custody until formally transferred to the analytical laboratory. For purposes of this work, custody of the samples will be defined as follows:

- In plain view of the field representatives
- Inside a cooler that is in plain view of the field representative

- Inside any locked space, such as a cooler, locker, car, or truck to which the field representative has the only immediately available key(s)

A chain-of-custody record provided by the laboratory will be initiated at the time of sampling for all samples collected. The record will be signed by the field representative and others who subsequently take custody of the samples. Couriers or other professional shipping representatives are not required to sign the chain-of-custody form; however, shipping receipts will be collected and maintained as a part of custody documentation in the project files. A copy of the chain-of-custody form with appropriate signatures will be maintained in Aspect's files and included as an appendix to the project report.

## 8.7 Field Log Requirements

During the collection of any field samples accompanying field documentation must be made clearly stating:

- Name and location of project
- Field personnel
- Sequence of events
- Any changes or deviations from the QAPP or SOPs
- Environmental conditions
- Date, time, location, ID, unique sample name, and description of each sample
- Field instrument calibration procedures
- Field measurement results
- Identity of QC samples collected
- Unusual circumstances that might affect interpretation of results

For this Study, data collected in the field will be contained in a field log (a binder backed by electronic scans of documents) that will consist of field notes (freehand notes) and Aspect field data sheets (Appendix C).

Field notes should be bound, waterproof notebooks with prenumbered pages (Rite in the Rain®). Permanent, waterproof ink should be used for all entries. Corrections should be made with single-line strikethroughs, initials, and date of correction. Use of white-out or correction fluid is not permitted.

While conducting field work, the field hydrogeologist or technician (Section 5) will document general pertinent observations and events in waterproof field notes and, when warranted, provide photographic documentation of specific sampling efforts. Data collected during the sample collection procedures will be recorded on standard Aspect field data sheets (Appendix D). Field notes will include a description of each field activity, sample descriptions, and associated details, such as the date, time, and field conditions. The laboratory chain-of-custody forms will be filled out before leaving the site. Upon completion of a field task, the field

personnel will then scan field notes and Aspect field data sheets into computer files and provide the original versions to the Aspect Project Manager. Copies of Aspect field data sheet and laboratory chain of custody are provided in Appendix D.

## **8.8 Other Activities**

Not Applicable.

## 9.0 Laboratory Procedures

### 9.1 Lab Procedures Table

Table 13 presents the lab procedures for each analyte including the sample matrix, number of samples, expected range of results, reporting limit, and analytical method.

**Table 13. Lab Procedures**

Analytical Method	Analyte	Sample Matrix	Number of Samples <sup>1</sup>	Expected Range of Results	Method Reporting Limit	Units
<b>General Chemistry, Inorganics in Drinking Water</b>						
EPA 300.0	Bromide	Water	8	0.02-0.2	0.100	mg/L
EPA 300.0	Chloride	Water	8	45279	0.100	mg/L
EPA 300.0	Fluoride	Water	8	1.7-3.6	0.100	mg/L
EPA 300.0	Nitrate/N	Water	8	0.02-2.3	0.100	mg/L
EPA 300.0	Nitrite/N	Water	8	0.02-2.3	0.100	mg/L
EPA 300.0	Sulfate	Water	8	26-32	0.100	mg/L
SM 2320 B	Alkalinity	Water	8	138-144	2.00	mg/L
SM 2320 B	Bicarbonate	Water	8	130-142	2.00	mg/L
SM 2540 C	TDS	Water	8	250-335	50.0	mg/L
SM 2540 D	TSS	Water	8	<5-5	1.00	mg/L
SM 4500-P F	Total P	Water	8	0.01-1.75	0.0100	mg/L
SM 5310 B	DOC	Water	8	<0.5	0.500	mg/L
SM 5310 B	TOC	Water	8	0.5-0.61	0.100	mg/L
<b>Metals by ICP in Drinking Water (All metals are total)</b>						
EPA 200.7	Aluminum	Water	8	3-17	0.0100	mg/L
EPA 200.7	Calcium	Water	8	2400-9900	0.100	mg/L
EPA 200.7	Iron	Water	8	8-550	0.0100	mg/L
EPA 200.7	Magnesium	Water	8	530-6230	0.100	mg/L
EPA 200.7	Potassium	Water	8	8200- 12500	0.500	mg/L
EPA 200.7	Silica (as SiO <sub>2</sub> )	Water	8	55000- 64000	0.214	mg/L
EPA 200.7	Sodium	Water	8	59500- 80000	0.100	mg/L
<b>Metals by ICP-MS in Drinking Water</b>						
EPA 200.8	Antimony	Water	8	0.02-0.08	0.00100	mg/L
EPA 200.8	Arsenic	Water	8	0.1-1.7	0.00100	mg/L
EPA 200.8	Barium	Water	8	7.7-20	0.000130	mg/L
EPA 200.8	Beryllium	Water	8	<0.3	0.000300	mg/L
EPA 200.8	Cadmium	Water	8	<0.4	0.00100	mg/L
EPA 200.8	Chromium	Water	8	<2.1	0.00100	mg/L
EPA 200.8	Copper	Water	8	<2.1	0.00100	mg/L
EPA 200.8	Lead	Water	8	2-50	0.00100	mg/L
EPA 200.8	Manganese	Water	8	0.9-2.1	0.00100	mg/L
EPA 200.8	Nickel	Water	8	0.9-18	0.00100	mg/L

Analytical Method	Analyte	Sample Matrix	Number of Samples <sup>1</sup>	Expected Range of Results	Method Reporting Limit	Units
EPA 200.8	Selenium	Water	8	0.1-0.3	0.00100	mg/L
EPA 200.8	Silver	Water	8	56000-66300	0.00100	mg/L
EPA 200.8	Thallium	Water	8	0.009-0.07	0.00100	mg/L
EPA 200.8	Uranium	Water	8	Unknown	0.00100	mg/L
EPA 200.8	Zinc	Water	8	Unknown	0.00100	mg/L
<b>Mercury in Water</b>						
EPA 245.7	Mercury	Water	8	Unknown	0.00100	ug/L
<b>Semivolatiles in Drinking Water</b>						
EPA 505	gamma-BHC (Lindane)	Water	3	<RL	0.0200	ug/L
EPA 505	Heptachlor	Water	3	<RL	0.0400	ug/L
EPA 505	Aldrin	Water	3	<RL	0.100	ug/L
EPA 505	Heptachlor epoxide	Water	3	<RL	0.0200	ug/L
EPA 505	4,4'-DDE	Water	3	<RL	0.100	ug/L
EPA 505	Dieldrin	Water	3	<RL	0.100	ug/L
EPA 505	Endrin	Water	3	<RL	0.0100	ug/L
EPA 505	4,4'-DDD	Water	3	<RL	0.100	ug/L
EPA 505	4,4'-DDT	Water	3	<RL	0.100	ug/L
EPA 505	Methoxychlor	Water	3	<RL	0.100	ug/L
EPA 505	Aroclor 1232 (PCB-1232)	Water	3	<RL	0.500	ug/L
EPA 505	Aroclor 1242 (PCB-1242)	Water	3	<RL	0.300	ug/L
EPA 505	Aroclor 1248 (PCB-1248)	Water	3	<RL	0.100	ug/L
EPA 505	Aroclor 1254 (PCB-1254)	Water	3	<RL	0.100	ug/L
EPA 505	Aroclor 1260 (PCB-1260)	Water	3	<RL	0.200	ug/L
EPA 505	PCBs	Water	3	<RL	0.500	ug/L
EPA 505	Chlordane	Water	3	<RL	0.200	ug/L
EPA 505	Toxaphene	Water	3	<RL	1.00	ug/L
EPA 515.4	Dalapon	Water	3	<RL	1.00	ug/L
EPA 515.4	Dicamba	Water	3	<RL	0.200	ug/L
EPA 515.4	Dichloroprop	Water	3	<RL	0.500	ug/L
EPA 515.4	2,4-D	Water	3	<RL	0.100	ug/L
EPA 515.4	Pentachlorophenol	Water	3	<RL	0.0400	ug/L
EPA 515.4	2,4,5-TP (Silvex)	Water	3	<RL	0.200	ug/L
EPA 515.4	2,4-DB	Water	3	<RL	1.00	ug/L
EPA 515.4	Dinoseb	Water	3	<RL	0.200	ug/L
EPA 515.4	Picloram	Water	3	<RL	0.100	ug/L
EPA 515.4	3,5-Dichlorobenzoic Acid	Water	3	<RL	0.500	ug/L
EPA 515.4	Chloramben	Water	3	<RL	0.200	ug/L
EPA 515.4	Acifluorofen	Water	3	<RL	1.00	ug/L
EPA 525.2	Alachlor	Water	3	<RL	0.200	ug/L
EPA 525.2	Atrazine	Water	3	<RL	0.100	ug/L
EPA 525.2	Benzo[a]pyrene	Water	3	<RL	0.0200	ug/L
EPA 525.2	bis(2-Ethylhexyl)phthalate	Water	3	<RL	0.600	ug/L
EPA 525.2	bis-2(ethylhexyl)adipate	Water	3	<RL	0.600	ug/L
EPA 525.2	Bromacil	Water	3	<RL	0.100	ug/L
EPA 525.2	Butachlor	Water	3	<RL	0.100	ug/L



Analytical Method	Analyte	Sample Matrix	Number of Samples <sup>1</sup>	Expected Range of Results	Method Reporting Limit	Units
EPA 525.2	Fluorene	Water	3	<RL	0.200	ug/L
EPA 525.2	gamma-BHC (Lindane)	Water	3	<RL	0.0400	ug/L
EPA 525.2	Hexachlorobenzene	Water	3	<RL	0.100	ug/L
EPA 525.2	Hexachlorocyclopentadiene	Water	3	<RL	0.100	ug/L
EPA 525.2	Methoxychlor	Water	3	<RL	0.200	ug/L
EPA 525.2	Metribuzin	Water	3	<RL	0.100	ug/L
EPA 525.2	Propachlor	Water	3	<RL	0.100	ug/L
EPA 525.2	Simazine	Water	3	<RL	0.0700	ug/L
EPA 549.2	Diquat	Water	3	<RL	0.400	ug/L
SM 6251 B	Monochloroacetic acid	Water	3	<RL	2.00	ug/L
SM 6251 B	Monobromoacetic acid	Water	3	<RL	1.00	ug/L
SM 6251 B	Dichloroacetic acid	Water	3	<RL	1.00	ug/L
SM 6251 B	Trichloroacetic acid	Water	3	<RL	1.00	ug/L
SM 6251 B	Bromochloroacetic acid (BCAA)	Water	3	<RL	1.00	ug/L
SM 6251 B	Dibromoacetic acid	Water	3	<RL	1.00	ug/L
SM 6251 B	Total HAA5	Water	3	<RL	1.00	ug/L

#### Volatiles in Drinking Water

EPA 524.3	1,3-Dichloropropene	Water	3	<RL	0.500	ug/L
EPA 524.3	Total Trihalomethane	Water	3	<RL	0.500	ug/L
EPA 524.3	Benzene	Water	3	<RL	0.500	ug/L
EPA 524.3	Bromochloromethane	Water	3	<RL	0.500	ug/L
EPA 524.3	Bromodichloromethane	Water	3	<RL	0.500	ug/L
EPA 524.3	Bromoform	Water	3	<RL	0.500	ug/L
EPA 524.3	Bromomethane	Water	3	<RL	0.500	ug/L
EPA 524.3	Carbon Tetrachloride	Water	3	<RL	0.500	ug/L
EPA 524.3	Chlorobenzene	Water	3	<RL	0.500	ug/L
EPA 524.3	Chloroform	Water	3	<RL	0.500	ug/L
EPA 524.3	Chloromethane	Water	3	<RL	0.500	ug/L
EPA 524.3	cis-1,2-dichloroethene	Water	3	<RL	0.500	ug/L
EPA 524.3	cis-1,3-Dichloropropene	Water	3	<RL	0.500	ug/L
EPA 524.3	1,2-Dibromo-3-chloropropane (DBCP)	Water	3	<RL	0.500	ug/L
EPA 524.3	1,2-Dibromoethane (EDB)	Water	3	<RL	0.500	ug/L
EPA 524.3	1,2-Dichlorobenzene	Water	3	<RL	0.500	ug/L
EPA 524.3	1,3-Dichlorobenzene	Water	3	<RL	0.500	ug/L
EPA 524.3	1,4-Dichlorobenzene	Water	3	<RL	0.500	ug/L
EPA 524.3	Dichlorodifluoromethane	Water	3	<RL	0.500	ug/L
EPA 524.3	1,1-Dichloroethane	Water	3	<RL	0.500	ug/L
EPA 524.3	1,2-Dichloroethane	Water	3	<RL	0.500	ug/L
EPA 524.3	1,1-Dichloroethene	Water	3	<RL	0.500	ug/L
EPA 524.3	trans-1,2-Dichloroethene	Water	3	<RL	0.500	ug/L
EPA 524.3	1,2-Dichloropropane	Water	3	<RL	0.500	ug/L
EPA 524.3	trans-1,3-Dichloropropene	Water	3	<RL	0.500	ug/L
EPA 524.3	Ethylbenzene	Water	3	<RL	0.500	ug/L
EPA 524.3	Hexachlorobutadiene	Water	3	<RL	0.500	ug/L

Analytical Method	Analyte	Sample Matrix	Number of Samples <sup>1</sup>	Expected Range of Results	Method Reporting Limit	Units
EPA 524.3	Isopropylbenzene	Water	3	<RL	0.500	ug/L
EPA 524.3	Methylene chloride	Water	3	<RL	0.500	ug/L
EPA 524.3	Naphthalene	Water	3	<RL	0.500	ug/L
EPA 524.3	Styrene	Water	3	<RL	0.500	ug/L
EPA 524.3	1,1,1,2-Tetrachloroethane	Water	3	<RL	0.500	ug/L
EPA 524.3	1,1,2,2-Tetrachloroethane	Water	3	<RL	0.500	ug/L
EPA 524.3	Tetrachloroethene	Water	3	<RL	0.500	ug/L
EPA 524.3	Toluene	Water	3	<RL	0.500	ug/L
EPA 524.3	1,2,4-Trichlorobenzene	Water	3	<RL	0.500	ug/L
EPA 524.3	1,1,1-Trichloroethane	Water	3	<RL	0.500	ug/L
EPA 524.3	1,1,2-Trichloroethane	Water	3	<RL	0.500	ug/L
EPA 524.3	Trichloroethene	Water	3	<RL	0.500	ug/L
EPA 524.3	Trichlorofluoromethane	Water	3	<RL	0.500	ug/L
EPA 524.3	1,2,3-Trichloropropane	Water	3	<RL	0.500	ug/L
EPA 524.3	Vinyl Chloride	Water	3	<RL	0.500	ug/L
EPA 524.3	m+p-Xylene	Water	3	<RL	0.500	ug/L
EPA 524.3	o-Xylene	Water	3	<RL	0.500	ug/L
EPA 524.3	Total Xylene	Water	3	<RL	0.500	ug/L
EPA 524.3	1,1-dichloropropene	Water	3	<RL	0.500	ug/L
EPA 524.3	1,2,3-Trichlorobenzene	Water	3	<RL	0.500	ug/L
EPA 524.3	1,2,4-Trimethylbenzene	Water	3	<RL	0.500	ug/L
EPA 524.3	Chloroethane	Water	3	<RL	0.500	ug/L
EPA 524.3	1,3,5-Trimethylbenzene	Water	3	<RL	0.500	ug/L
EPA 524.3	2,2-Dichloropropane	Water	3	<RL	0.500	ug/L
EPA 524.3	1,3-Dichloropropane	Water	3	<RL	0.500	ug/L
EPA 524.3	2-Chlorotoluene	Water	3	<RL	0.500	ug/L
EPA 524.3	4-Chlorotoluene	Water	3	<RL	0.500	ug/L
EPA 524.3	Bromobenzene	Water	3	<RL	0.500	ug/L
EPA 524.3	Dibromochloromethane	Water	3	<RL	0.500	ug/L
EPA 524.3	Dibromomethane	Water	3	<RL	0.500	ug/L
EPA 524.3	methyl-t-butyl ether (MTBE)	Water	3	<RL	0.500	ug/L
EPA 524.3	n-Butylbenzene	Water	3	<RL	0.500	ug/L
EPA 524.3	n-Propylbenzene	Water	3	<RL	0.500	ug/L
EPA 524.3	p-isopropyltoluene	Water	3	<RL	0.500	ug/L
EPA 524.3	sec-Butylbenzene	Water	3	<RL	0.500	ug/L
EPA 524.3	tert-Butylbenzene	Water	3	<RL	0.500	ug/L
EPA 524.3	Acetone	Water	3	<RL	2.50	ug/L
EPA 524.3	MTBE-d3	Water	3	<RL		Surr.
EPA 524.3	4-Bromofluorobenzene	Water	3	<RL		Surr.
EPA 524.3	1,2-Dichlorobenzene-d4	Water	3	<RL		Surr.

**Note:**

1. See Section 7.2.1 and Table 9 for sampling schedule.

## 9.2 Sample Preparation Method(s)

Samples will be prepared and extracted by an accredited lab in accordance with industry standards and analytical methods. The selected laboratory is discussed in Section 9.4.

### **9.3 Special Method Requirements**

Not applicable.

### **9.4 Laboratories Accredited for Methods**

Analysis of water quality samples will be performed by Anatek of Moscow, Idaho or their Spokane, Washington office, with the exception of bacteriological, nitrate, and nitrite analysis. Anatek is accredited by Ecology for analysis of all parameters included in this project (see Appendix A).

Contact information for the laboratory is:

Anatek Labs, Inc  
1282 Alturas Dr  
Moscow, ID

Project Manager: Justin Doty  
Phone: 208 883 2839  
Email: [Justin@anateklabs.com](mailto:Justin@anateklabs.com)

Bacteriological, nitrate, and nitrite analysis will be performed by LabTest of Yakima, Washington, to minimize holding times for analysis. LabTest is accredited by Ecology for these analysis (see Appendix A).

Contact information for the laboratory is:

LabTest  
201 East D Street  
Yakima, WA

Lab Supervisor: Giles Hamilton  
Phone: 509-575-3999  
Email: [vws155@gmail.com](mailto:vws155@gmail.com)

## 10.0 Quality Control Procedures

Implementing QC procedures provides the information needed to assess the quality of the data that is collected. These procedures also help identify problems or issues associated with data collection or data analysis while the project is underway.

### 10.1 Field and Laboratory Quality Control

Standard EPA Level II procedures will be followed by the laboratory for one standard check, method blank, analytical duplicate, and matrix spike per laboratory batch (typically 10 to 20, as accommodated by laboratory autosampling equipment and sample backlog). Field procedures will follow standard guidelines and SOPs for the relevant field activity. As detailed below, data validation samples will be collected at a minimum of every 10 samples collected.

#### Data Validation Samples

Field quality control (QC) is accomplished through the analysis of controlled data validation (DV) samples that are introduced to the laboratory from the field. Field duplicates and trip blanks will be collected and submitted to the investigation laboratory to provide a means of assessing the quality of data resulting from the field sampling program.

#### Trip Blank

Trip blank samples will be used to monitor any possible cross-contamination that occurs during the transport of VOCs and samples. Trip blank samples are prepared by the laboratory using organic-free reagent-grade water into a VOA vial prior to the collection of field samples. Two vials per trip blank sample are placed with and accompany the VOCs samples through the entire transport process. Trip blank samples will be prepared and analyzed only for VOCs.

#### Field Duplicates

Field duplicate samples are used to check for sampling and analysis reproducibility. Field duplicate samples will be collected at a frequency of 10 percent of the field samples for every matrix and analytical method.

A set of DV samples will be collected for at least every 10 water samples collected. The DV sample set will include the following for calculation of DV parameters and acceptance criteria, and Section 9 for description of lab procedures):

- A MS/MSD
- A “blind” field duplicate (i.e., not indicated to the lab as a field duplicate)
- Trip blanks (for VOCs, bacteria, and inorganic constituent suites)

Except for the trip blank, the chemical analysis of DV samples will include the entire list of chemical analytes (Section 6). The trip blank will include only analysis of VOCs. The blind field duplicate will be labeled in a manner that does not indicate its true sample location, and the MS/MSD will be labeled, as such, for laboratory processing.

## 10.2 Corrective Action Processes

The laboratory will follow the analytical method for corrective action procedures when the sample results do not meet the QC acceptance criteria. The laboratory will notify the Aspect hydrogeologist that submitted the samples and include a narrative in the laboratory report when following the analytical method corrective action procedure results in a sample result not meeting the QC acceptance criteria. Findings will be reviewed by the Aspect project manager. QC results may indicate problems with data during the course of the project. Corrective action processes (such as recalibration) will be used if:

- Activities are inconsistent with the QAPP
- Field instruments yield unusual results
- Results do not meet MQOs or performance expectations
- If some other unforeseen problem arises

## **11.0 Data Management Procedures**

### **11.1 Data Recording and Reporting Requirements**

Field technicians will record all field data in a water-resistant field notebook, electronic data forms, or Aspect standard field data sheet. Before leaving each site, staff will check field notebooks, data sheets, or electronic data forms for missing or improbable measurements. Field technicians will enter field-generated data into spreadsheets or a project database as soon as practical after they return from the field. For data collected electronically, data will be backed up on servers when staff return from the field. Raw data files will be stored separate from processed data files.

The Aspect field hydrogeologist and field technician will check data entry against the field notebook data for errors and omissions. The hydrogeologist will notify the Aspect project manager of missing or unusual data.

All final spreadsheet files, paper field notes, and final products created as part of the data collection and data QA process will be kept with the project data files.

Data will be uploaded to Ecology's EIM database as described in Section 11.4.

### **11.2 Laboratory Data Package Requirements**

All continuous and laboratory data will be stored in a project database that includes station location information and data QA information. This database will facilitate summarization and graphical analysis of the data.

### **11.3 Electronic Transfer Requirements**

The lab will provide an EPA Level II data package as a pdf and an electronic data deliverable (EDD).

### **11.4 Data Upload Procedures**

Following completion of the QC procedures described in Section 10 and the DV procedures described in Section 8.2, all quality assured data will be formatted and uploaded to Ecology's EIM database by an Aspect data scientist. The EIM study ID will be *WRYBIP-2123-Moxeec-00036*.

### **11.5 Model Information Management**

Not applicable.

## **12.0 Audits and Reports**

### **12.1 Audits**

Field technicians will be required to review this QAPP prior to each monitoring event and to maintain a copy of the QAPP and its appendices in the field. Field technicians may be audited at any time by appropriate project manager or the Aspect data manager (Section 5) to ensure that field work is being completed according to this QAPP, work plan, and published SOPs. Projects that involve complex data analysis may be audited by the appropriate project manager or other personnel familiar with the analysis procedures.

### **12.2 Responsible Personnel**

Personnel responsible for the audits are as follows:

- Field audit: Aspect Project Manager
- Field consistency review: experienced (at least 3 years) staff (senior hydrogeologist or project manager)
- Data analysis: Aspect hydrogeologists (field, senior, and principal, as required for specific analysis)

Personnel assigned to these roles are listed in Table 2.

### **12.3 Frequency and Distribution of Reports**

Results of the field data collection, data quality assessment, and any data analysis will be documented in the final ASR Feasibility Study Report. The final report will be distributed to all other stakeholders involved or interested in the Study as determined by the City and Ecology.

Data analysis documentation may be accomplished in one document at the end of the project or in stages during different phases of the project. For complex projects, the project team may elect to write separate reports on the data collected, QA/QC, and model scenarios. For this project, the data analysis documentation will be included in the Water Quality Evaluation section (and appendices) of the final ASR Feasibility Study Report.

Field and Laboratory Data will be entered into EIM when data collection is complete.

### **12.4 Responsibility for Reports**

The Aspect Project Manager is responsible for verifying data completeness and usability before the data are used in the technical report and entered into Environmental Information Management (EIM) database. The Aspect Project Manager is also responsible for writing the final technical report, unless an alternate author is agreed upon and documented at the start of the project.

The Aspect Project Manager is responsible for assigning a peer reviewer with the appropriate expertise for the technical report. Depending on the type of final report, there may be an internal

and external review process. The peer reviewer is responsible for working with the report author to resolve or clarify any issues with the report.

## **13.0 Data Verification**

Data verification is the process of evaluating the completeness, correctness, and conformance/compliance of a specific data set against the method, procedural, or contractual requirements.

### **13.1 Field Data Verification, Requirements, and Responsibilities**

Field notebooks, data sheets, and electronic information storage will be checked for missing or improbable measurements, and initial data will be verified before leaving each site. This process involves checking the data sheet (written or electronic) for omissions or outliers. If measurement data are missing or a measurement is determined to be an outlier, the measurement will be flagged in the data sheet and repeated if possible. The field hydrogeologist or field technician is responsible for in-field data verification.

Upon returning from the field, data are either manually entered (data recorded on paper) or downloaded from instruments and then uploaded into the appropriate database or project folder (see Section 11: Data Management Section). Manually entered data will be verified/checked by a staff member who did not enter the data. Downloaded electronic data files will also be checked for completeness and appropriate metadata (such as file name, time code).

Following data entry verification, raw field measurement data will undergo a quality analysis verification process to evaluate the performance of the sensors. Field measurement data may be adjusted for bias or drift (increasing bias over time) based on the results of fouling, field, or standards checks following general USGS guidelines (Wagner, 2007) and this process:

#### **Review Discrete Field QC Checks**

The field check of instrumentation will consist of a manual measurement for water levels, and measurement of water quality standards in the field (checks with water quality standards will be completed separate from calibration events). Review of the field checks will consist of the following:

1. Review post check data for field QC instrument check (water quality and water level), reject data as appropriate.
2. Assign a criteria to the field check values indicating either acceptance, rejection, or qualification of the data and assign a data flag detailing the reason to rejection or qualify data based on the post-check.

#### **Review/Adjust Time Series (Continuous) Data**

1. Plot compensated pressure data converted to depth-to-water time series with field checks.



2. Reject data based on deployment/retrieval times, site visit disruption, blatant fouling events, and sensor/equipment failure.
3. Review sensor offsets for recalibration. Flag any potential chronic drift or bias issues specific to the instrument.
4. If applicable, review fouling check and make drift adjustment, if necessary. In some situations, an event fouling adjustment may be warranted based on abrupt changes in groundwater levels, barometric pressure, etc.
5. Review residuals from both field checks and post-checks, together referred to as QC checks. Adjust data, as appropriate, using a weight-of-evidence approach. Give the most weight to checks rated excellent, then good, and then fair. Do not use field checks rated poor. Potential data adjustments include:
  - a. **Bias** – Data are adjusted by the average difference between the QC checks and deployed instrument. Majority of QC checks must show bias to use this method.
  - b. **Regression** – Data adjusted using regression, typically linear, between QC checks and deployed instrument. This accounts for both a slope and bias adjustment. The regression must have at least five data points and an  $R^2$  value of  $>0.95$  to use for adjustment. Do not extrapolate regressions beyond the range of the QC checks.
  - c. **Calibration/Sensor Drift** – Data adjusted using linear regression with time from calibration or deployment to post-check or retrieval. Majority of QC checks, particularly post-checks, must confirm pattern of drift.
6. Typically, choose the adjustment that results in the smallest residuals and bias between the adjusted values and QC checks. Best professional judgement and visual review are necessary to confirm adjustment.
7. If the evidence is weak, or inconclusive, do not adjust the data.

It will be noted in the final report if any data is adjusted. Data adjustment must be performed or reviewed by an Aspect Project Manager, or personnel, with the appropriate training and experience in processing raw sensor data.

## 13.2 Laboratory Data Verification

The lab will provide an EPA Level II data package. Additional laboratory data validation (check batch QC) will be conducted by Aspect's project data scientist (Table 2). Laboratory validation results will be summarized on the laboratory reports, and Aspect's validation results will be summarized in the final report. An Aspect hydrogeologist will verify the validated laboratory results.

## 13.3 Validation Requirements, if Necessary

Not applicable.

## **13.4 Model Quality Assessment**

Not applicable.

# **14.0 Data Quality (Usability) Assessment**

## **14.1 Process for Determining Project Objectives were Met**

The Aspect Project Manager will assess all data (qualified and unqualified), results or verification, compliance with MQOs, and the overall quality of the data set to provide a final determination regarding usability in the context of the project-specific goals and objectives. The final report will document whether the final, acceptable-quality data set meets the needs of the project (allows desired conclusions/decisions to be made with the desired level of certainty).

## **14.2 Treatment of Nondetects**

Nondetects will be reported as the MRL for that analyte with the appropriate flag (“<”) indicating it as a nondetect.

## **14.3 Data Analysis and Presentation Methods**

Data found to be of acceptable quality for project objectives will be analyzed before being summarized. Any relevant and interesting data analysis will be presented in the final report using a combination of tables and plots of various kinds, such as time-series plots, histograms, and box plots.

The report will contain a summary table of chemistry; figures of continuous data (water level hydrographs, potentiometric maps, etc.); discussion of results pertaining to each sample location; and a map of study area. Additionally, a conceptual hydrogeologic model will be included showing a cross section of the target aquifer in relation to the City Wells and Yakima River.

## **14.4 Sampling Design Evaluation**

The Aspect Project Manager will decide whether the data package meets the MQOs and the criteria for completeness, representativeness, and comparability. If so, the sampling design will be considered effective. If the sampling design is found ineffective, the approach will be modified in accordance with Ecology, and/or the Study will be halted for redesign.

## **14.5 Documentation of Assessment**

In the final report, the Aspect Project Manager will include a summary and detailed description of the data quality assessment and model quality evaluation findings. This summary is usually included in the Data Quality section of reports. The final report will also provide results of the data analysis, uncertainty analysis, and margin of safety.

## 15.0 References

- Aspect Consulting, LLC, 2022. City of Moxee Aquifer Storage and Recovery: Assessment of Existing Data
- Jelinski, J., Clayton, C.S., and Fulford, J.M., 2015, Accuracy testing of electric groundwater-level measurement tapes: U.S. Geological Survey Open-File Report 2014–1236, 27 p.
- Jones, M.A., Vaccaro, J.J., and Watkins, A.M., 2006, Hydrogeologic framework of sedimentary deposits in six structural basins, Yakima River Basin, Washington: U.S. Geological Survey Scientific Investigations Report 2006-5116, 24 p., 6 pls.
- Sleeper, S.S., 2020, A Geochemical Assessment of Potential Groundwater Storage Locations within the Yakima River Basin, 1372, <https://digitalcommons.cwu.edu/etd/1372>.
- Spane, F. A., 2002. Considering Barometric Pressure in Groundwater Flow Investigations. J. Water Resources Research, Vol. 38, No. 6. Published June 18, 2002.
- U.S. environmental Protection Agency (EPA,) 1995, Technical Guidance Manual for Hydrogeologic Investigations and Groundwater Monitoring, Chapter 4: Slug and Pumping Tests, February 1995.
- Wagner, R.J., Kimbrough, R.A., and Turney, G.L., 2007, Quality-assurance plan for water-quality activities in the U.S. Geological Survey Washington Water Science Center: U.S. Geological Survey Open-File Report 2007–1307, 48 p.
- Washington State Department of Ecology (Ecology), 2004, Guidelines for Preparing Quality Assurance Project Plans for Environmental Studies, Publication No. 04-03-030, July 2004.
- Washington State Department of Ecology (Ecology), 2006, Environmental Assessment Program – Standard Operating Procedure for Manually Obtaining Surface Water Samples, October 24, 2006.
- Washington State Department of Ecology (Ecology), 2018a, Standard Operating Procedures for Manual Well-Depth and Depth-to-Water Measurements Version 1.2, EAP052 Pub. No. 18-03-215, Prepared by Pamela Marti, Environmental Assessment Program, Washington State Department of Ecology.
- Washington State Department of Ecology (Ecology), 2018b, Standard Operating Procedures to Minimize the Spread of Invasive Species, Version 2.2, EAP070 Pub. No. 18-03-201, Prepared by Parsons, J., Hallock, D., Seiders, K., Ward, B., Coffin, C., Newell, E., Deligeannis, C., and Welch K., Environmental Assessment Program, Washington State Department of Ecology.
- Washington State Department of Ecology (Ecology), 2018c, Standard Operating Procedure EAP099, Version 1.2: [Standard Operating Procedure EAP099, Version 1.0: Purging and Sampling Monitoring Wells for General Chemistry Parameters](#). Washington State Department of Ecology, Olympia.  
<https://apps.ecology.wa.gov/publications/SummaryPages/1803214.html>

Washington State Department of Ecology (Ecology), 2019, Standard Operating Procedure EAP074, Version 1.2: Use of Submersible Pressure Transducers During Groundwater Studies. Washington State Department of Ecology, Olympia.  
<https://fortress.wa.gov/ecy/publications/SummaryPages/1903205.html>.

Washington State Department of Ecology (Ecology), 2020, Aquifer Test Procedures, Publication No. 20-11-093, October 2020.

Washington State Department of Health (DOH), 2003, General Sampling Procedure, Office of Drinking Water No. 331-219, July 2003.

## **16.0 Appendices**

**Appendix A. Laboratory Accreditations**

**Appendix B. Well Logs**

**Appendix C. Aspect Field Data Sheets**

## Appendix D. Glossaries, Acronyms, and Abbreviations

### Glossary of General Terms

**Ambient:** Background or away from point sources of contamination. Surrounding environmental condition.

**Clean Water Act:** A federal act passed in 1972 that contains provisions to restore and maintain the quality of the nation's waters. Section 303(d) of the Clean Water Act establishes the TMDL program.

**Conductivity:** A measure of water's ability to conduct an electrical current. Conductivity is related to the concentration and charge of dissolved ions in water.

**Dissolved oxygen (DO):** A measure of the amount of oxygen dissolved in water.

**Dilution factor:** The relative proportion of effluent to stream (receiving water) flows occurring at the edge of a mixing zone during critical discharge conditions as authorized in accordance with the state's mixing zone regulations at WAC 173-201A-100.

<http://apps.leg.wa.gov/WAC/default.aspx?cite=173-201A-020>

**Effluent:** An outflowing of water from a natural body of water or from a human-made structure. For example, the treated outflow from a wastewater treatment plant.

**Fecal coliform (FC):** That portion of the coliform group of bacteria which is present in intestinal tracts and feces of warm-blooded animals as detected by the product of acid or gas from lactose in a suitable culture medium within 24 hours at 44.5 plus or minus 0.2 degrees Celsius. Fecal coliform bacteria are "indicator" organisms that suggest the possible presence of disease-causing organisms. Concentrations are measured in colony forming units per 100 milliliters of water (cfu/100 mL).

**Nutrient:** Substance such as carbon, nitrogen, and phosphorus used by organisms to live and grow. Too many nutrients in the water can promote algal blooms and rob the water of oxygen vital to aquatic organisms.

**pH:** A measure of the acidity or alkalinity of water. A low pH value (0 to 7) indicates that an acidic condition is present, while a high pH (7 to 14) indicates a basic or alkaline condition. A pH of 7 is considered to be neutral. Since the pH scale is logarithmic, a water sample with a pH of 8 is ten times more basic than one with a pH of 7.

**Surface waters of the state:** Lakes, rivers, ponds, streams, inland waters, salt waters, wetlands

**Total suspended solids (TSS):** Portion of solids retained by a filter.

**Turbidity:** A measure of water clarity. High levels of turbidity can have a negative impact on aquatic life.

## Acronyms and Abbreviations

Aspect	Aspect Consulting, LLC
ASR	Aquifer Storage and Recovery
Anatek	Anatek Labs, Inc
City	City of Moxee
Commerce	State of Washington Department of Commerce
DBPs	Disinfection Byproducts
DO	Dissolved oxygen
DOC	Dissolved organic carbon
DOH	Department of Health
DQI	data quality indicator
DQO	data quality objective
DV	design verification
ECBID	East Columbia Basin Irrigation District
EDD	Electronic Data Deliverable
e.g.	For example
Ecology	Washington State Department of Ecology
EIM	Environmental Information Management database
EPA	U.S. Environmental Protection Agency
et al.	And others
FC	Fecal coliform
GIS	Geographic Information System software
GPS	Global Positioning System
GWMA	Groundwater Management Area
GWQS	Groundwater Quality Standards
HAAs	Haloacetic Acids
i.e.	In other words
LCS	laboratory control sample
MDL	minimum detection limit
MQO	measurement quality objective
MRL	minimum reporting limit

MS	matrix spike
MSD	matrix spike duplicate
NELAP	National Environmental Laboratory Accreditation Program
NTR	National Toxics Rule
OCR	Office of Columbia River
PFAS	per- and polyfluoroalkyl substances
QA	quality assurance
QAPP	Quality Assurance Project Plan
QC	quality control
Reclamation	U.S. Bureau of Reclamation
RPD	relative percent difference
RSD	relative standard deviation
SAP	Sampling Analysis Plain
SCADA	Supervisory Control and Data Acquisition
SOP	Standard operating procedures
Study	Feasibility Study
THMs	Trihalomethanes
TDS	total dissolved solids
TOC	total organic carbon
TSS	total suspended solids
USFS	United States Forest Service
USGS	U.S. Geological Survey
VOA	volatile organic analysis
VOCs	volatile organic compounds
WAC	Washington Administrative Code
WRIA	Water Resource Inventory Area

### **Units of Measurement**

°C	degrees centigrade
cfs	cubic feet per second
cfu	colony forming units
cms	cubic meters per second, a unit of flow



dw	dry weight
ft	feet
g	gram, a unit of mass
gpm	gallons per minute
kcfs	1,000 cubic feet per second
km	kilometer, a unit of length equal to 1,000 meters
L/s	liters per second (0.03531 cubic foot per second)
m	meter
mg	milligram
mgd	million gallons per day
mg/L	milligrams per liter (parts per million)
mL	milliliter
NTU	nephelometric turbidity units
s.u.	standard units
ug/L	micrograms per liter (parts per billion)
uS/cm	microsiemens per centimeter, a unit of conductivity

## Quality Assurance Glossary

**Accreditation:** A certification process for laboratories, designed to evaluate and document a lab’s ability to perform analytical methods and produce acceptable data (Kammin, 2010). For Ecology, it is defined according to WAC 173-50-040: “Formal recognition by [Ecology] that an environmental laboratory is capable of producing accurate and defensible analytical data.”

**Accuracy:** The degree to which a measured value agrees with the true value of the measured property. USEPA recommends that this term not be used, and that the terms *precision* and *bias* be used to convey the information associated with the term *accuracy* (USEPA, 2014).

**Analyte:** An element, ion, compound, or chemical moiety (pH, alkalinity) which is to be determined. The definition can be expanded to include organisms, e.g., fecal coliform, Klebsiella (Kammin, 2010).

**Bias:** Discrepancy between the expected value of an estimator and the population parameter being estimated (Gilbert, 1987; USEPA, 2014).

**Blank:** A synthetic sample, free of the analyte(s) of interest. For example, in water analysis, pure water is used for the blank. In chemical analysis, a blank is used to estimate the analytical response to all factors other than the analyte in the sample. In general, blanks are used to assess possible contamination or inadvertent introduction of analyte during various stages of the sampling and analytical process (USGS, 1998).

**Calibration:** The process of establishing the relationship between the response of a measurement system and the concentration of the parameter being measured (Ecology, 2004).

**Check standard:** A substance or reference material obtained from a source independent from the source of the calibration standard; used to assess bias for an analytical method. This is an obsolete term, and its use is highly discouraged. See Calibration Verification Standards, Lab Control Samples (LCS), Certified Reference Materials (CRM), and/or spiked blanks. These are all check standards but should be referred to by their actual designator, e.g., CRM, LCS (Kammin, 2010; Ecology, 2004).

**Comparability:** The degree to which different methods, data sets and/or decisions agree or can be represented as similar; a data quality indicator (USEPA, 2014; USEPA, 2020).

**Completeness:** The amount of valid data obtained from a project compared to the planned amount. Usually expressed as a percentage. A data quality indicator (USEPA, 2014; USEPA 2020).

**Continuing Calibration Verification Standard (CCV):** A quality control (QC) sample analyzed with samples to check for acceptable bias in the measurement system. The CCV is usually a midpoint calibration standard that is re-run at an established frequency during the course of an analytical run (Kammin, 2010).

**Control chart:** A graphical representation of quality control results demonstrating the performance of an aspect of a measurement system (Kammin, 2010; Ecology 2004).

**Control limits:** Statistical warning and action limits calculated based on control charts. Warning limits are generally set at +/- 2 standard deviations from the mean, action limits at +/- 3 standard deviations from the mean (Kammin, 2010).

**Data integrity:** A qualitative DQI that evaluates the extent to which a data set contains data that is misrepresented, falsified, or deliberately misleading (Kammin, 2010).

**Data quality indicators (DQI):** Commonly used measures of acceptability for environmental data. The principal DQIs are precision, bias, representativeness, comparability, completeness, sensitivity, and integrity (USEPA, 2006).

**Data quality objectives (DQO):** Qualitative and quantitative statements derived from systematic planning processes that clarify study objectives, define the appropriate type of data, and specify tolerable levels of potential decision errors that will be used as the basis for establishing the quality and quantity of data needed to support decisions (USEPA, 2006).

**Data set:** A grouping of samples organized by date, time, analyte, etc. (Kammin, 2010).

**Data validation:** The process of determining that the data satisfy the requirements as defined by the data user (USEPA, 2020). There are various levels of data validation (USEPA, 2009).

**Data verification:** Examination of a data set for errors or omissions, and assessment of the Data Quality Indicators related to that data set for compliance with acceptance criteria (MQOs). Verification is a detailed quality review of a data set (Ecology, 2004).

**Detection limit (limit of detection):** The concentration or amount of an analyte which can be determined to a specified level of certainty to be greater than zero (Ecology, 2004).

**Duplicate samples:** Two samples taken from and representative of the same population, and carried through and steps of the sampling and analytical procedures in an identical manner. Duplicate samples are used to assess variability of all method activities including sampling and analysis (USEPA, 2014).

**Field blank:** A blank used to obtain information on contamination introduced during sample collection, storage, and transport (Ecology, 2004).

**Initial Calibration Verification Standard (ICV):** A QC sample prepared independently of calibration standards and analyzed along with the samples to check for acceptable bias in the measurement system. The ICV is analyzed prior to the analysis of any samples (Kammin, 2010).

**Laboratory Control Sample (LCS)/LCS duplicate:** A sample of known composition prepared using contaminant-free water or an inert solid that is spiked with analytes of interest at the midpoint of the calibration curve or at the level of concern. It is prepared and analyzed in the same batch of regular samples using the same sample preparation method, reagents, and analytical methods employed for regular samples. Monitors a lab's performance for bias and precision (USEPA, 2014).

**Matrix spike/Matrix spike duplicate:** A QC sample prepared by adding a known amount of the target analyte(s) to an aliquot of a sample to check for bias and precision errors due to interference or matrix effects (Ecology, 2004).

**Measurement Quality Objectives (MQOs):** Performance or acceptance criteria for individual data quality indicators, usually including precision, bias, sensitivity, completeness, comparability, and representativeness (USEPA, 2006).

**Measurement result:** A value obtained by performing the procedure described in a method (Ecology, 2004).

**Method:** A formalized group of procedures and techniques for performing an activity (e.g., sampling, chemical analysis, data analysis), systematically presented in the order in which they are to be executed (USEPA, 2001).

**Method blank:** A blank prepared to represent the sample matrix, prepared and analyzed with a batch of samples. A method blank will contain all reagents used in the preparation of a sample, and the same preparation process is used for the method blank and samples (Ecology, 2004; Kammin, 2010).

**Method Detection Limit (MDL):** The minimum measured concentration of a substance that can be reported with 99% confidence that the measured concentration is distinguishable from

method blank results (USEPA, 2016). MDL is a measure of the capability of an analytical method of distinguished samples that do not contain a specific analyte from a sample that contains a low concentration of the analyte (USEPA, 2020).

**Minimum level:** Either the sample concentration equivalent to the lowest calibration point in a method or a multiple of the method detection limit (MDL), whichever is higher. For the purposes of NPDES compliance monitoring, EPA considers the following terms to be synonymous: “quantitation limit,” “reporting limit,” and “minimum level” (40 CFR 136).

**Parameter:** A specified characteristic of a population or sample. Also, an analyte or grouping of analytes. Benzene and nitrate + nitrite are all parameters (Kammin, 2010; Ecology, 2004).

**Population:** The hypothetical set of all possible observations of the type being investigated (Ecology, 2004).

**Precision:** The extent of random variability among replicate measurements of the same property; a data quality indicator (USGS, 1998).

**Quality assurance (QA):** A set of activities designed to establish and document the reliability and usability of measurement data (Kammin, 2010).

**Quality Assurance Project Plan (QAPP):** A document that describes the objectives of a project, and the processes and activities necessary to develop data that will support those objectives (Kammin, 2010; Ecology, 2004).

**Quality control (QC):** The routine application of measurement and statistical procedures to assess the accuracy of measurement data (Ecology, 2004).

**Relative Percent Difference (RPD):** RPD is commonly used to evaluate precision. The following formula is used:

$$RPD = [Abs(a-b)/((a + b)/2)] * 100\%$$

where “Abs()” is absolute value and a and b are results for the two replicate samples. RPD can be used only with 2 values. Percent Relative Standard Deviation is (%RSD) is used if there are results for more than 2 replicate samples (Ecology, 2004).

**Relative Standard Deviation (RSD):** A statistic used to evaluate precision in environmental analysis. It is determined in the following manner:

$$RSD = (100\% * s)/x$$

where s is the sample standard deviation and x is the mean of results from more than two replicate samples (Kammin, 2010).

**Replicate samples:** Two or more samples taken from the environment at the same time and place, using the same protocols. Replicates are used to estimate the random variability of the material sampled (USGS, 1998).

**Reporting level:** Unless specified otherwise by a regulatory authority or in a discharge permit, results for analytes that meet the identification criteria (i.e., rules for determining qualitative presence/absence of an analyte) are reported down to the concentration of the minimum level established by the laboratory through calibration of the instrument. EPA considers the terms “reporting limit,” “quantitation limit,” and “minimum level” to be synonymous (40 CFR 136).

**Representativeness:** The degree to which a sample reflects the population from which it is taken; a data quality indicator (USGS, 1998).

**Sample (field):** A portion of a population (environmental entity) that is measured and assumed to represent the entire population (USGS, 1998).

**Sample (statistical):** A finite part or subset of a statistical population (USEPA, 1992).

**Sensitivity:** In general, denotes the rate at which the analytical response (e.g., absorbance, volume, meter reading) varies with the concentration of the parameter being determined. In a specialized sense, it has the same meaning as the detection limit (Ecology, 2004).

**Spiked blank:** A specified amount of reagent blank fortified with a known mass of the target analyte(s); usually used to assess the recovery efficiency of the method (USEPA, 2014).

**Spiked sample:** A sample prepared by adding a known mass of target analyte(s) to a specified amount of matrix sample for which an independent estimate of target analyte(s) concentration is available. Spiked samples can be used to determine the effect of the matrix on a method’s recovery efficiency (USEPA, 2014).

**Split sample:** A discrete sample subdivided into portions, usually duplicates (Kammin, 2010).

**Standard Operating Procedure (SOP):** A document which describes in detail a reproducible and repeatable organized activity (Kammin, 2010).

**Surrogate:** For environmental chemistry, a surrogate is a substance with properties similar to those of the target analyte(s). Surrogates are unlikely to be native to environmental samples. They are added to environmental samples for quality control purposes, to track extraction efficiency and/or measure analyte recovery. Deuterated organic compounds are examples of surrogates commonly used in organic compound analysis (Kammin, 2010).

**Systematic planning:** A step-wise process which develops a clear description of the goals and objectives of a project, and produces decisions on the type, quantity, and quality of data that will be needed to meet those goals and objectives. The DQO process is a specialized type of systematic planning (USEPA, 2006).

## References for QA Glossary

40 CFR 136. Title 40 Code of Federal Regulations, Part 136: Guidelines Establishing Test Procedures for the Analysis of Pollutants. Available at: <https://www.ecfr.gov/cgi-bin/text-idx?SID=3cf9acace214b7af340ea8f6919a7c39&mc=true&node=pt40.25.136&rgn=div5> (accessed 26 Feb. 2020).

- Ecology, 2004. Guidance for the Preparation of Quality Assurance Project Plans for Environmental Studies. Washington State Department of Ecology, Olympia, WA. Available at: <https://fortress.wa.gov/ecy/publications/SummaryPages/0403030.html> (accessed 6 Mar. 2020).
- Gilbert, R.O., 1987. Statistical Methods for Environmental Pollution Monitoring. Van Nostrand Reinhold, New York, NY.
- Kammin, W., 2010. Definition developed or extensively edited by William Kammin, 2010. Washington State Department of Ecology, Olympia, WA.
- USEPA, 1992. Guidelines for exposure assessment. U.S. Environmental Protection Agency, Risk Assessment Forum, Washington, D.C. EPA/600/Z-92/001. Available at: [https://www.epa.gov/sites/production/files/2014-11/documents/guidelines\\_exp\\_assessment.pdf](https://www.epa.gov/sites/production/files/2014-11/documents/guidelines_exp_assessment.pdf) (accessed 26 Feb. 2020).
- USEPA, 2001. EPA Requirements for Quality Assurance Project Plans, EPA QA/R-5. U.S. Environmental Protection Agency, Washington, DC. EPA/240/B-01/003. Available at: <https://www.epa.gov/quality/epa-qar-5-epa-requirements-quality-assurance-project-plans> (accessed 26 Feb. 2020).
- USEPA, 2006. Guidance on Systematic Planning Using the Data Quality Objectives Process EPA QA/G-4. U.S. Environmental Protection Agency, Washington, DC. Available at: <https://www.epa.gov/sites/production/files/2015-06/documents/g4-final.pdf> (accessed 26 Feb. 2020).
- USEPA, 2009. Guidance for Labeling Externally Validated Laboratory Analytical Data for Superfund Use, OSWER No. 9200.1-85, EPA 540-R-08-005. U.S. Environmental Protection Agency, Washington, DC. Available at: <https://www.epa.gov/nscep>.
- USEPA, 2014. Compendium: Project Quality Assurance and Quality Control: Chapter 1. U.S. Environmental Protection Agency, Washington, DC. SW-846 Update V. Available at: [https://www.epa.gov/sites/production/files/2015-10/documents/chap1\\_1.pdf](https://www.epa.gov/sites/production/files/2015-10/documents/chap1_1.pdf) (accessed 26 Feb. 2020).
- USEPA, 2016. Definition and Procedure for the Determination of the Method Detection Limit, Revision 2. EPA 821-R-16-006. U.S. Environmental Protection Agency, Washington, DC. Available at: [https://www.epa.gov/sites/production/files/2016-12/documents/mdl-procedure\\_rev2\\_12-13-2016.pdf](https://www.epa.gov/sites/production/files/2016-12/documents/mdl-procedure_rev2_12-13-2016.pdf) (accessed 6 Mar. 2020).
- USEPA, 2020. Glossary: Environmental Sampling and Analytical Methods (ESAM) Program. U.S. Environmental Protection Agency, Washington, DC. Available at: <https://www.epa.gov/esam/glossary> (accessed 26 Feb. 2020).
- USGS, 1998. Principles and Practices for Quality Assurance and Quality Control. Open-File Report 98-636. U.S. Geological Survey, Reston, VA. Available at: <https://pubs.usgs.gov/of/1998/ofr98-636/> (accessed 26 Feb. 2020).

WAC 173-50-040. Title 173 Washington Administrative Code. Accreditation of Environmental Laboratories: Definitions. Available at:  
<https://apps.leg.wa.gov/WAC/default.aspx?cite=173-50-040> (accessed 26 Feb. 2020).

## **APPENDIX A**

### **Laboratory Accreditations**



The State of  
Department



Washington  
of Ecology

**Anatek Labs, Inc - Moscow**  
**Moscow, ID**

has complied with provisions set forth in Chapter 173-50 WAC and is hereby recognized by the Department of Ecology as an ACCREDITED LABORATORY for the analytical parameters listed on the accompanying Scope of Accreditation. This certificate is effective March 21, 2023 and shall expire March 20, 2024.

Witnessed under my hand on June 27, 2023

Rebecca Wood  
Lab Accreditation Unit Supervisor

Laboratory ID  
**C595**

# WASHINGTON STATE DEPARTMENT OF ECOLOGY

## ENVIRONMENTAL LABORATORY ACCREDITATION PROGRAM

### SCOPE OF ACCREDITATION

#### Anatek Labs, Inc - Moscow

#### Moscow, ID

is accredited for the analytes listed below using the methods indicated. Full accreditation is granted unless stated otherwise in a note. EPA is the U.S. Environmental Protection Agency. SM is "Standard Methods for the Examination of Water and Wastewater." SM refers to EPA approved method versions. ASTM is the American Society for Testing and Materials. USGS is the U.S. Geological Survey. AOAC is the Association of Official Analytical Chemists. Other references are described in notes.

Matrix/Analyte	Method	Notes
<b>Drinking Water</b>		
Turbidity	EPA 180.1_2_1993	1,8
Chloride	EPA 300.0_2.1_1993	1
Fluoride	EPA 300.0_2.1_1993	1,8
Nitrate	EPA 300.0_2.1_1993	1,8
Nitrate + Nitrite	EPA 300.0_2.1_1993	1
Nitrite	EPA 300.0_2.1_1993	1,8,9
Orthophosphate	EPA 300.0_2.1_1993	1
Sulfate	EPA 300.0_2.1_1993	1
Perchlorate	EPA 331.0_1.0_2005	1
Cyanide, Total	EPA 335.4_1_1993	1,8
Color	SM 2120 B-2011	1,9
Alkalinity	SM 2320 B-2011	1
Hardness (calc.)	SM 2340 B-2011	1
Specific Conductance	SM 2510 B-2011	1
Solids, Total Dissolved	SM 2540 C-2011	1
pH	SM 4500-H+ B-2011	1,6
Nitrate	SM 4500-NO <sub>3</sub> <sup>-</sup> F-2011	1,8
Nitrate + Nitrite	SM 4500-NO <sub>3</sub> <sup>-</sup> F-2011	1
Nitrite	SM 4500-NO <sub>3</sub> <sup>-</sup> F-2011	1,8
Total Organic Carbon	SM 5310 B-2011	1
Anionic Surfactants (MBAS)	SM 5540 C-2011	1
Aluminum	EPA 200.7_4.4_1994	1
Barium	EPA 200.7_4.4_1994	1
Beryllium	EPA 200.7_4.4_1994	1
Cadmium	EPA 200.7_4.4_1994	1
Calcium	EPA 200.7_4.4_1994	1

Anatek Labs, Inc - Moscow

<b>Matrix/Analyte</b>	<b>Method</b>	<b>Notes</b>
<b>Drinking Water</b>		
Chromium	EPA 200.7_4.4_1994	1
Copper	EPA 200.7_4.4_1994	1
Iron	EPA 200.7_4.4_1994	1
Magnesium	EPA 200.7_4.4_1994	1
Manganese	EPA 200.7_4.4_1994	1
Nickel	EPA 200.7_4.4_1994	1,8
Potassium	EPA 200.7_4.4_1994	1
Sodium	EPA 200.7_4.4_1994	1,8
Aluminum	EPA 200.8_5.4_1994	1
Antimony	EPA 200.8_5.4_1994	1,8
Arsenic	EPA 200.8_5.4_1994	1,8
Barium	EPA 200.8_5.4_1994	1,8
Beryllium	EPA 200.8_5.4_1994	1,8
Cadmium	EPA 200.8_5.4_1994	1,8
Chromium	EPA 200.8_5.4_1994	1,8
Copper	EPA 200.8_5.4_1994	1,8
Lead	EPA 200.8_5.4_1994	1,8
Manganese	EPA 200.8_5.4_1994	1
Mercury	EPA 200.8_5.4_1994	1,8
Nickel	EPA 200.8_5.4_1994	1,8
Selenium	EPA 200.8_5.4_1994	1,8
Silver	EPA 200.8_5.4_1994	1
Thallium	EPA 200.8_5.4_1994	1,8
Total Uranium	EPA 200.8_5.4_1994	1,8
Zinc	EPA 200.8_5.4_1994	1
1,2,3-Trichloropropane	EPA 504.1_1.1_1995	1
1,2-Dibromo-3-chloropropane (DBCP)	EPA 504.1_1.1_1995	1,8
1,2-Dibromoethane (EDB, Ethylene dibromide)	EPA 504.1_1.1_1995	1,8
Aldrin	EPA 505_2.1_1995	1
Aroclor-1016 (PCB-1016)	EPA 505_2.1_1995	1,8
Aroclor-1221 (PCB-1221)	EPA 505_2.1_1995	1,8
Aroclor-1232 (PCB-1232)	EPA 505_2.1_1995	1,8
Aroclor-1242 (PCB-1242)	EPA 505_2.1_1995	1,8
Aroclor-1248 (PCB-1248)	EPA 505_2.1_1995	1,8
Aroclor-1254 (PCB-1254)	EPA 505_2.1_1995	1,8
Aroclor-1260 (PCB-1260)	EPA 505_2.1_1995	1,8
Chlordane (tech.)	EPA 505_2.1_1995	1,8

Anatek Labs, Inc - Moscow

Matrix/Analyte	Method	Notes
<b>Drinking Water</b>		
Dieldrin	EPA 505_2.1_1995	1
Endrin	EPA 505_2.1_1995	1,8
gamma-BHC (Lindane, gamma-Hexachlorocyclohexane)	EPA 505_2.1_1995	1,8
Heptachlor	EPA 505_2.1_1995	1,8
Heptachlor epoxide	EPA 505_2.1_1995	1,8
Methoxychlor	EPA 505_2.1_1995	1,8
Toxaphene (Chlorinated camphene)	EPA 505_2.1_1995	1,8
2,4,5-T	EPA 515.4_1_2000	1
2,4-D	EPA 515.4_1_2000	1,8
2,4-DB	EPA 515.4_1_2000	1
3,5-Dichlorobenzoic acid	EPA 515.4_1_2000	
Acifluorfen	EPA 515.4_1_2000	1
Bentazon	EPA 515.4_1_2000	
Chloramben	EPA 515.4_1_2000	
Dacthal Acid Metabolites	EPA 515.4_1_2000	1
Dalapon	EPA 515.4_1_2000	1,8
Dicamba	EPA 515.4_1_2000	1
Dichloroprop (Dichlorprop)	EPA 515.4_1_2000	1
Dinoseb (2-sec-butyl-4,6-dinitrophenol, DNBP)	EPA 515.4_1_2000	1,8
Pentachlorophenol	EPA 515.4_1_2000	1,8
Picloram	EPA 515.4_1_2000	1,8
Silvex (2,4,5-TP)	EPA 515.4_1_2000	1,8
3-Hydroxycarbofuran	EPA 531.2_1_2001	1
Aldicarb (Temik)	EPA 531.2_1_2001	1
Aldicarb sulfone	EPA 531.2_1_2001	1
Aldicarb sulfoxide	EPA 531.2_1_2001	1
Carbaryl (Sevin)	EPA 531.2_1_2001	1
Carbofuran (Furaden)	EPA 531.2_1_2001	1,8
Methiocarb (Mesurol)	EPA 531.2_1_2001	1
Methomyl (Lannate)	EPA 531.2_1_2001	1
Oxamyl	EPA 531.2_1_2001	1,8
Propoxur (Baygon)	EPA 531.2_1_2001	1
Glyphosate	EPA 547_1990	1,8
Diquat	EPA 549.2_1_1997	1,8
Bromoacetic acid (MBAA, BAA)	SM 6251 B-05	1,8
Bromochloroacetic acid (BCAA)	SM 6251 B-05	1
Chloroacetic acid (MCAA, CAA)	SM 6251 B-05	1,8

Anatek Labs, Inc - Moscow

Matrix/Analyte	Method	Notes
<b>Drinking Water</b>		
Dibromoacetic acid (DBAA)	SM 6251 B-05	1,8
Dichloroacetic acid (DCAA)	SM 6251 B-05	1,8
Total haloacetic acids (HAA5)	SM 6251 B-05	1,8
Trichloroacetic acid (TCAA)	SM 6251 B-05	1,8
1,1,1,2-Tetrachloroethane	EPA 524.3_1.0_2009	1
1,1,1-Trichloroethane	EPA 524.3_1.0_2009	1
1,1,2,2-Tetrachloroethane	EPA 524.3_1.0_2009	1
1,1,2-Trichloroethane	EPA 524.3_1.0_2009	1
1,1-Dichloroethylene	EPA 524.3_1.0_2009	1
1,1-Dichloropropene	EPA 524.3_1.0_2009	1
1,2,3-Trichlorobenzene	EPA 524.3_1.0_2009	1
1,2,3-Trichloropropane	EPA 524.3_1.0_2009	1
1,2,4-Trichlorobenzene	EPA 524.3_1.0_2009	1
1,2,4-Trimethylbenzene	EPA 524.3_1.0_2009	1
1,2-Dichlorobenzene	EPA 524.3_1.0_2009	1
1,2-Dichloroethane (Ethylene dichloride)	EPA 524.3_1.0_2009	1
1,2-Dichloropropane	EPA 524.3_1.0_2009	1
1,3,5-Trimethylbenzene	EPA 524.3_1.0_2009	1
1,3-Dichlorobenzene	EPA 524.3_1.0_2009	1
1,3-Dichloropropane	EPA 524.3_1.0_2009	1
1,4-Dichlorobenzene	EPA 524.3_1.0_2009	1
2,2-Dichloropropane	EPA 524.3_1.0_2009	
2-Chlorotoluene	EPA 524.3_1.0_2009	1
4-Chlorotoluene	EPA 524.3_1.0_2009	1
4-Isopropyltoluene (p-Cymene)	EPA 524.3_1.0_2009	1
Benzene	EPA 524.3_1.0_2009	1
Bromobenzene	EPA 524.3_1.0_2009	1
Bromochloromethane	EPA 524.3_1.0_2009	1
Bromodichloromethane	EPA 524.3_1.0_2009	1,8
Bromoform	EPA 524.3_1.0_2009	1,8
Carbon disulfide	EPA 524.3_1.0_2009	1
Carbon tetrachloride	EPA 524.3_1.0_2009	1
Chlorobenzene	EPA 524.3_1.0_2009	1
Chlorodibromomethane	EPA 524.3_1.0_2009	1,8
Chloroethane (Ethyl chloride)	EPA 524.3_1.0_2009	
Chloroform	EPA 524.3_1.0_2009	1,8
cis-1,2-Dichloroethylene	EPA 524.3_1.0_2009	1

Anatek Labs, Inc - Moscow

Matrix/Analyte	Method	Notes
<b>Drinking Water</b>		
cis-1,3-Dichloropropene	EPA 524.3_1.0_2009	1
Dibromomethane	EPA 524.3_1.0_2009	1
Dichlorodifluoromethane (Freon-12)	EPA 524.3_1.0_2009	1
Diethyl ether	EPA 524.3_1.0_2009	1
Ethylbenzene	EPA 524.3_1.0_2009	1
Hexachlorobutadiene	EPA 524.3_1.0_2009	1
Iodomethane (Methyl iodide)	EPA 524.3_1.0_2009	1
Isopropylbenzene	EPA 524.3_1.0_2009	1
m+p-xylene	EPA 524.3_1.0_2009	1
Methyl bromide (Bromomethane)	EPA 524.3_1.0_2009	1
Methyl chloride (Chloromethane)	EPA 524.3_1.0_2009	1
Methyl tert-butyl ether (MTBE)	EPA 524.3_1.0_2009	1
Methylene chloride (Dichloromethane)	EPA 524.3_1.0_2009	1
Naphthalene	EPA 524.3_1.0_2009	1
n-Butylbenzene	EPA 524.3_1.0_2009	1
n-Propylbenzene	EPA 524.3_1.0_2009	1
o-Xylene	EPA 524.3_1.0_2009	1
sec-Butylbenzene	EPA 524.3_1.0_2009	1
Styrene	EPA 524.3_1.0_2009	1
tert-Butylbenzene	EPA 524.3_1.0_2009	1
Tetrachloroethylene (Perchloroethylene)	EPA 524.3_1.0_2009	1
Toluene	EPA 524.3_1.0_2009	1
Total Trihalomethanes	EPA 524.3_1.0_2009	1,8
trans-1,2-Dichloroethylene	EPA 524.3_1.0_2009	1
trans-1,3-Dichloropropylene	EPA 524.3_1.0_2009	1
Trichloroethene (Trichloroethylene)	EPA 524.3_1.0_2009	1
Trichlorofluoromethane (Freon 11)	EPA 524.3_1.0_2009	1
Vinyl chloride	EPA 524.3_1.0_2009	1,8
Xylene (total)	EPA 524.3_1.0_2009	1
2,4-Dinitrotoluene (2,4-DNT)	EPA 525.2_2_1995	1
2,6-Dinitrotoluene (2,6-DNT)	EPA 525.2_2_1995	1
4,4'-DDD	EPA 525.2_2_1995	1
4,4'-DDE	EPA 525.2_2_1995	1
4,4'-DDT	EPA 525.2_2_1995	1
Acenaphthylene	EPA 525.2_2_1995	1
Acetochlor	EPA 525.2_2_1995	1
Alachlor	EPA 525.2_2_1995	1,8

Anatek Labs, Inc - Moscow

<b>Matrix/Analyte</b>	<b>Method</b>	<b>Notes</b>
<b>Drinking Water</b>		
alpha-BHC (alpha-Hexachlorocyclohexane)	EPA 525.2_2_1995	1
alpha-Chlordane	EPA 525.2_2_1995	1
Anthracene	EPA 525.2_2_1995	1
Atrazine	EPA 525.2_2_1995	1,8
Benzo(a)anthracene	EPA 525.2_2_1995	1
Benzo(a)pyrene	EPA 525.2_2_1995	1
Benzo(g,h,i)perylene	EPA 525.2_2_1995	1
Benzo(k)fluoranthene	EPA 525.2_2_1995	1
Benzo[b]fluoranthene	EPA 525.2_2_1995	1
beta-BHC (beta-Hexachlorocyclohexane)	EPA 525.2_2_1995	1
bis(2-Ethylhexyl) phthalate (DEHP)	EPA 525.2_2_1995	1,8
Bromacil	EPA 525.2_2_1995	1
Butachlor	EPA 525.2_2_1995	1
Butyl benzyl phthalate	EPA 525.2_2_1995	1,8
Chrysene	EPA 525.2_2_1995	1
Cyanazine	EPA 525.2_2_1995	1
delta-BHC	EPA 525.2_2_1995	1
Di(2-ethylhexyl)adipate	EPA 525.2_2_1995	1,8
Diazinon	EPA 525.2_2_1995	1
Dibenz(a,h) anthracene	EPA 525.2_2_1995	1
Diethyl phthalate	EPA 525.2_2_1995	1,8
Dimethyl phthalate	EPA 525.2_2_1995	1,8
Di-n-butyl phthalate	EPA 525.2_2_1995	1,8
EPTC (Eptam, s-ethyl-dipropyl thio carbamate)	EPA 525.2_2_1995	1
Fluorene	EPA 525.2_2_1995	1
gamma-Chlordane	EPA 525.2_2_1995	1
Hexachlorobenzene	EPA 525.2_2_1995	1,8
Hexachlorocyclopentadiene	EPA 525.2_2_1995	1,8
Indeno(1,2,3-cd) pyrene	EPA 525.2_2_1995	1
Metolachlor	EPA 525.2_2_1995	1
Metribuzin	EPA 525.2_2_1995	1
Molinate	EPA 525.2_2_1995	1
Phenanthrene	EPA 525.2_2_1995	1
Prometon	EPA 525.2_2_1995	1
Propachlor (Ramrod)	EPA 525.2_2_1995	1
Pyrene	EPA 525.2_2_1995	1
Simazine	EPA 525.2_2_1995	1,8

Anatek Labs, Inc - Moscow

Matrix/Analyte	Method	Notes
<b>Drinking Water</b>		
Terbacil	EPA 525.2_2_1995	1
trans-Nonachlor	EPA 525.2_2_1995	1
11-Chloroeicosafluoro-3-oxaundecane-1-sulfonic acid (11-CI-PF3OUdS)	EPA 533	1
1H,1H,2H,2H,-Perfluorodecanesulfonic acid (8:2 FTS)	EPA 533	1
1H,1H,2H,2H,-Perfluorooctanesulfonic acid (6:2 FTS)	EPA 533	1
1H,1H,2H,2H-Perfluorohexanesulfonic acid (4:2 FTS)	EPA 533	1
4,8-Dioxa-3H-perfluorononanoic acid (ADONA)	EPA 533	1
9-Chlorohexadecafluoro-3-oxanonane-1-sulfonic acid (9-CI-PF3ONS)	EPA 533	1
Hexafluoropropylene oxide dimer acid (HFPO-DA)	EPA 533	1
Nonafluoro-3,6-dioxaheptanoic acid (NFDHA)	EPA 533	1
Perfluoro(2-ethoxyethane)sulfonic acid (PFEEESA)	EPA 533	1
Perfluoro-3-methoxypropanoic acid (PFMPA)	EPA 533	1
Perfluoro-4-methoxybutanoic acid (PFMBA)	EPA 533	1
Perfluorobutane sulfonic acid (PFBS)	EPA 533	1
Perfluorobutanoic acid (PFBA)	EPA 533	1
Perfluorodecanoic acid (PFDA)	EPA 533	1
Perfluorododecanoic acid (PFDoA)	EPA 533	1
Perfluoroheptane sulfonic acid (PFHpS)	EPA 533	1
Perfluoroheptanoic acid (PFHpA)	EPA 533	1
Perfluorohexane sulfonic acid (PFHxS)	EPA 533	1
Perfluorohexanoic acid (PFHxA)	EPA 533	1
Perfluorononanoic acid (PFNA)	EPA 533	1
Perfluorooctane sulfonic acid (PFOS)	EPA 533	1
Perfluorooctanoic acid (PFOA)	EPA 533	1
Perfluoropentane sulfonic acid (PFPeS)	EPA 533	1
Perfluoropentanoic acid (PFPeA)	EPA 533	1
Perfluoroundecanoic acid (PFUnA)	EPA 533	1
11-Chloroeicosafluoro-3-oxaundecane-1-sulfonic acid (11-CI-PF3OUdS)	EPA 537.1 revision 2 (3/20)	10
4,8-Dioxa-3H-perfluorononanoic acid (ADONA)	EPA 537.1 revision 2 (3/20)	10
9-Chlorohexadecafluoro-3-oxanonane-1-sulfonic acid (9-CI-PF3ONS)	EPA 537.1 revision 2 (3/20)	10
Hexafluoropropylene oxide dimer acid (HFPO-DA)	EPA 537.1 revision 2 (3/20)	10
N-Ethylperfluorooctane sulfonamido acetic acid (NEtFOSAA)	EPA 537.1 revision 2 (3/20)	10
N-Methylperfluorooctane sulfonamido acetic acid (NMeFOSAA)	EPA 537.1 revision 2 (3/20)	10
Perfluorobutane sulfonic acid (PFBS)	EPA 537.1 revision 2 (3/20)	10
Perfluorodecanoic acid (PFDA)	EPA 537.1 revision 2 (3/20)	10
Perfluorododecanoic acid (PFDoA)	EPA 537.1 revision 2 (3/20)	10
Perfluoroheptanoic acid (PFHpA)	EPA 537.1 revision 2 (3/20)	10



Anatek Labs, Inc - Moscow

Matrix/Analyte	Method	Notes
<b>Drinking Water</b>		
Perfluorohexane sulfonic acid (PFHxS)	EPA 537.1 revision 2 (3/20)	10
Perfluorohexanoic acid (PFHxA)	EPA 537.1 revision 2 (3/20)	10
Perfluorononanoic acid (PFNA)	EPA 537.1 revision 2 (3/20)	10
Perfluorooctane sulfonic acid (PFOS)	EPA 537.1 revision 2 (3/20)	10
Perfluorooctanoic acid (PFOA)	EPA 537.1 revision 2 (3/20)	10
Perfluorotetradecanoic acid (PFTeDA)	EPA 537.1 revision 2 (3/20)	10
Perfluorotridecanoic acid (PFTrDA)	EPA 537.1 revision 2 (3/20)	10
Perfluoroundecanoic acid (PFUnA)	EPA 537.1 revision 2 (3/20)	10
N-Ethylperfluorooctane sulfonamido acetic acid (NEtFOSAA)	EPA 537_1.1_2009	1
N-Methylperfluorooctane sulfonamido acetic acid (NMeFOSAA)	EPA 537_1.1_2009	1
Perfluorobutane sulfonic acid (PFBS)	EPA 537_1.1_2009	1
Perfluorodecanoic acid (PFDA)	EPA 537_1.1_2009	1
Perfluorododecanoic acid (PFDoA)	EPA 537_1.1_2009	1
Perfluoroheptanoic acid (PFHPA)	EPA 537_1.1_2009	1
Perfluorohexane sulfonic acid (PFHxS)	EPA 537_1.1_2009	1
Perfluorohexanoic acid (PFHxA)	EPA 537_1.1_2009	1
Perfluorononanoic acid (PFNA)	EPA 537_1.1_2009	1
Perfluorooctane sulfonic acid (PFOS)	EPA 537_1.1_2009	1
Perfluorooctanoic acid (PFOA)	EPA 537_1.1_2009	1
Perfluorotetradecanoic acid (PFTeDA)	EPA 537_1.1_2009	1
Perfluorotridecanoic acid (PFTrDA)	EPA 537_1.1_2009	1
Perfluoroundecanoic acid (PFUnA)	EPA 537_1.1_2009	1
Endothall	EPA 548.1_1_1992	1,8
Total coli/E.coli - detect	SM 9223 B Colilert 180 (PA)	8,9
<b>Non-Potable Water</b>		
n-Hexane Extractable Material (O&G)	EPA 1664B -10 (HEM)	1
Turbidity	EPA 180.1_2_1993	1
Chloride	EPA 300.0_2.1_1993	1
Fluoride	EPA 300.0_2.1_1993	1
Nitrate	EPA 300.0_2.1_1993	1
Nitrate + Nitrite	EPA 300.0_2.1_1993	1
Nitrite	EPA 300.0_2.1_1993	1
Orthophosphate	EPA 300.0_2.1_1993	1
Sulfate	EPA 300.0_2.1_1993	1
Perchlorate	EPA 331.0_1.0_2005	7
Cyanide, Total	EPA 335.4_1_1993	1
Phenolics, Total	EPA 420.1_1978	1,9

Anatek Labs, Inc - Moscow

Matrix/Analyte	Method	Notes
<b>Non-Potable Water</b>		
Perchlorate	EPA 6850-07	1
Alkalinity	SM 2320 B-2011	1
Hardness (calc.)	SM 2340 B-2011	1
Specific Conductance	SM 2510 B-2011	1
Solids, Total Dissolved	SM 2540 C-2011	1
Solids, Total Suspended	SM 2540 D-2011	1,9
Cyanide, Weak Acid Dissociable	SM 4500 CN <sup>-</sup> I-2011	
Cyanides, Amenable to Chlorination	SM 4500-CN <sup>-</sup> G-2011	9
pH	SM 4500-H+ B-2011	1,6
Ammonia	SM 4500-NH3 G-2011	1
Nitrate	SM 4500-NO3 <sup>-</sup> F-2011	1
Nitrate + Nitrite	SM 4500-NO3 <sup>-</sup> F-2011	1
Nitrite	SM 4500-NO3 <sup>-</sup> F-2011	1
Nitrogen, Total Kjeldahl	SM 4500-Norg C-2011	1
Orthophosphate	SM 4500-P F-2011	1
Phosphorus, total	SM 4500-P F-2011	1
Total Organic Carbon	SM 5310 B-2011	1
Phenolics, Total	SM 5530 D-2010	1,9
Mercury	EPA 1631 E-02	1
Aluminum	EPA 200.7_4.4_1994	1
Barium	EPA 200.7_4.4_1994	1
Beryllium	EPA 200.7_4.4_1994	1
Boron	EPA 200.7_4.4_1994	1
Cadmium	EPA 200.7_4.4_1994	1
Calcium	EPA 200.7_4.4_1994	1
Chromium	EPA 200.7_4.4_1994	1
Cobalt	EPA 200.7_4.4_1994	1
Copper	EPA 200.7_4.4_1994	1
Iron	EPA 200.7_4.4_1994	1
Lead	EPA 200.7_4.4_1994	1
Magnesium	EPA 200.7_4.4_1994	1
Manganese	EPA 200.7_4.4_1994	1
Molybdenum	EPA 200.7_4.4_1994	1
Nickel	EPA 200.7_4.4_1994	1
Sodium	EPA 200.7_4.4_1994	1
Titanium	EPA 200.7_4.4_1994	1
Vanadium	EPA 200.7_4.4_1994	1

Anatek Labs, Inc - Moscow

<b>Matrix/Analyte</b>	<b>Method</b>	<b>Notes</b>
<b>Non-Potable Water</b>		
Zinc	EPA 200.7_4.4_1994	1
Aluminum	EPA 200.8_5.4_1994	1
Antimony	EPA 200.8_5.4_1994	1
Arsenic	EPA 200.8_5.4_1994	1
Barium	EPA 200.8_5.4_1994	1
Beryllium	EPA 200.8_5.4_1994	1,9
Cadmium	EPA 200.8_5.4_1994	1
Chromium	EPA 200.8_5.4_1994	1
Cobalt	EPA 200.8_5.4_1994	1
Copper	EPA 200.8_5.4_1994	1
Lead	EPA 200.8_5.4_1994	1
Manganese	EPA 200.8_5.4_1994	1
Molybdenum	EPA 200.8_5.4_1994	1
Nickel	EPA 200.8_5.4_1994	1
Selenium	EPA 200.8_5.4_1994	1
Silver	EPA 200.8_5.4_1994	1
Strontium	EPA 200.8_5.4_1994	
Thallium	EPA 200.8_5.4_1994	1
Tin	EPA 200.8_5.4_1994	1
Titanium	EPA 200.8_5.4_1994	1
Total Uranium	EPA 200.8_5.4_1994	1
Vanadium	EPA 200.8_5.4_1994	1
Zinc	EPA 200.8_5.4_1994	1
Mercury	EPA 245.7_2005	1
4,4'-DDD	EPA 608.3	1
4,4'-DDE	EPA 608.3	1
4,4'-DDT	EPA 608.3	1
Aldrin	EPA 608.3	1
alpha-BHC (alpha-Hexachlorocyclohexane)	EPA 608.3	1
Aroclor-1016 (PCB-1016)	EPA 608.3	1
Aroclor-1221 (PCB-1221)	EPA 608.3	1
Aroclor-1232 (PCB-1232)	EPA 608.3	1
Aroclor-1242 (PCB-1242)	EPA 608.3	1
Aroclor-1248 (PCB-1248)	EPA 608.3	1
Aroclor-1254 (PCB-1254)	EPA 608.3	1
Aroclor-1260 (PCB-1260)	EPA 608.3	1
beta-BHC (beta-Hexachlorocyclohexane)	EPA 608.3	1

Anatek Labs, Inc - Moscow

<b>Matrix/Analyte</b>	<b>Method</b>	<b>Notes</b>
<b>Non-Potable Water</b>		
Chlordane (tech.)	EPA 608.3	1
delta-BHC	EPA 608.3	1
Dieldrin	EPA 608.3	1
Endosulfan I	EPA 608.3	1
Endosulfan II	EPA 608.3	1
Endosulfan sulfate	EPA 608.3	1
Endrin	EPA 608.3	1
Endrin aldehyde	EPA 608.3	1
Endrin ketone	EPA 608.3	1
gamma-BHC (Lindane, gamma-Hexachlorocyclohexane)	EPA 608.3	1
Heptachlor	EPA 608.3	1
Heptachlor epoxide	EPA 608.3	1
Methoxychlor	EPA 608.3	1
Toxaphene (Chlorinated camphene)	EPA 608.3	1
1,1,1,2-Tetrachloroethane	EPA 624.1	1
1,1,1-Trichloroethane	EPA 624.1	1
1,1,2,2-Tetrachloroethane	EPA 624.1	
1,1,2-Trichloroethane	EPA 624.1	1
1,1-Dichloroethane	EPA 624.1	1
1,1-Dichloroethylene	EPA 624.1	1
1,1-Dichloropropene	EPA 624.1	1
1,2,3-Trichlorobenzene	EPA 624.1	1
1,2,3-Trichloropropane	EPA 624.1	1
1,2,4-Trimethylbenzene	EPA 624.1	1
1,2-Dibromo-3-chloropropane (DBCP)	EPA 624.1	1
1,2-Dibromoethane (EDB, Ethylene dibromide)	EPA 624.1	1
1,2-Dichlorobenzene	EPA 624.1	1
1,2-Dichloroethane (Ethylene dichloride)	EPA 624.1	1
1,2-Dichloropropane	EPA 624.1	1
1,3,5-Trimethylbenzene	EPA 624.1	1
1,3-Dichlorobenzene	EPA 624.1	1
1,3-Dichloropropane	EPA 624.1	1
1,4-Dichlorobenzene	EPA 624.1	1
2,2-Dichloropropane	EPA 624.1	1
2-Butanone (Methyl ethyl ketone, MEK)	EPA 624.1	
2-Chloroethyl vinyl ether	EPA 624.1	1
2-Chlorotoluene	EPA 624.1	1

Anatek Labs, Inc - Moscow

<b>Matrix/Analyte</b>	<b>Method</b>	<b>Notes</b>
<b>Non-Potable Water</b>		
2-Hexanone	EPA 624.1	
4-Chlorotoluene	EPA 624.1	1
4-Isopropyltoluene (p-Cymene)	EPA 624.1	1
4-Methyl-2-pentanone (MIBK)	EPA 624.1	
Acrolein (Propenal)	EPA 624.1	1
Acrylonitrile	EPA 624.1	1
Benzene	EPA 624.1	1
Bromobenzene	EPA 624.1	1
Bromochloromethane	EPA 624.1	1
Bromodichloromethane	EPA 624.1	1
Bromoform	EPA 624.1	1
Carbon disulfide	EPA 624.1	
Carbon tetrachloride	EPA 624.1	1
Chlorobenzene	EPA 624.1	1
Chlorodibromomethane	EPA 624.1	1
Chloroethane (Ethyl chloride)	EPA 624.1	1
Chloroform	EPA 624.1	1
cis-1,2-Dichloroethylene	EPA 624.1	1
cis-1,3-Dichloropropene	EPA 624.1	1
Dibromomethane	EPA 624.1	1
Diethyl ether	EPA 624.1	1
Ethylbenzene	EPA 624.1	1
Iodomethane (Methyl iodide)	EPA 624.1	1
Isopropylbenzene	EPA 624.1	1
m+p-xylene	EPA 624.1	1
Methyl bromide (Bromomethane)	EPA 624.1	1
Methyl chloride (Chloromethane)	EPA 624.1	1
Methyl tert-butyl ether (MTBE)	EPA 624.1	1
Methylene chloride (Dichloromethane)	EPA 624.1	1
n-Butylbenzene	EPA 624.1	1
n-Propylbenzene	EPA 624.1	1
o-Xylene	EPA 624.1	1
sec-Butylbenzene	EPA 624.1	1
Styrene	EPA 624.1	1
tert-Butylbenzene	EPA 624.1	
Tetrachloroethylene (Perchloroethylene)	EPA 624.1	1
Toluene	EPA 624.1	1

Anatek Labs, Inc - Moscow

Matrix/Analyte	Method	Notes
<b>Non-Potable Water</b>		
trans-1,2-Dichloroethylene	EPA 624.1	1
trans-1,3-Dichloropropylene	EPA 624.1	1
trans-1,4-Dichloro-2-butene	EPA 624.1	1
Trichloroethene (Trichloroethylene)	EPA 624.1	1
Trichlorofluoromethane (Freon 11)	EPA 624.1	1
Vinyl chloride	EPA 624.1	1
Xylene (total)	EPA 624.1	1
1,2,4-Trichlorobenzene	EPA 625.1	1
1,2-Dinitrobenzene	EPA 625.1	
1,2-Diphenylhydrazine	EPA 625.1	1
1,3-Dinitrobenzene (1,3-DNB)	EPA 625.1	
1-Methylnaphthalene	EPA 625.1	
2,2'-Oxybis(1-chloropropane)	EPA 625.1	1
2,3,4,6-Tetrachlorophenol	EPA 625.1	1
2,3,5,6-Tetrachlorophenol	EPA 625.1	
2,4,5-Trichlorophenol	EPA 625.1	1
2,4,6-Trichlorophenol	EPA 625.1	1
2,4-Dichlorophenol	EPA 625.1	1
2,4-Dimethylphenol	EPA 625.1	1
2,4-Dinitrophenol	EPA 625.1	1
2,4-Dinitrotoluene (2,4-DNT)	EPA 625.1	1
2,6-Dinitrotoluene (2,6-DNT)	EPA 625.1	1
2-Chloronaphthalene	EPA 625.1	1
2-Chlorophenol	EPA 625.1	1
2-Methylnaphthalene	EPA 625.1	1
2-Methylphenol (o-Cresol)	EPA 625.1	
2-Nitroaniline	EPA 625.1	1
2-Nitrophenol	EPA 625.1	1
3,3'-Dichlorobenzidine	EPA 625.1	1
3-Nitroaniline	EPA 625.1	1
4,6-Dinitro-2-methylphenol	EPA 625.1	1
4-Bromophenyl phenyl ether (BDE-3)	EPA 625.1	1
4-Chloro-3-methylphenol	EPA 625.1	1
4-Chloroaniline	EPA 625.1	1
4-Chlorophenyl phenylether	EPA 625.1	1
4-Methylphenol (p-Cresol)	EPA 625.1	
4-Nitroaniline	EPA 625.1	1

Anatek Labs, Inc - Moscow

Matrix/Analyte	Method	Notes
<b>Non-Potable Water</b>		
4-Nitrophenol	EPA 625.1	1
Acenaphthene	EPA 625.1	1
Acenaphthylene	EPA 625.1	1
Aniline	EPA 625.1	
Anthracene	EPA 625.1	1
Benzidine	EPA 625.1	1
Benzo(a)anthracene	EPA 625.1	1
Benzo(a)pyrene	EPA 625.1	1
Benzo(g,h,i)perylene	EPA 625.1	1
Benzo(k)fluoranthene	EPA 625.1	1
Benzo[b]fluoranthene	EPA 625.1	1
Benzyl alcohol	EPA 625.1	1
bis(2-Chloroethoxy)methane	EPA 625.1	1
bis(2-Chloroethyl) ether	EPA 625.1	1
bis(2-Ethylhexyl) phthalate (DEHP)	EPA 625.1	1
Butyl benzyl phthalate	EPA 625.1	1
Carbazole	EPA 625.1	
Chrysene	EPA 625.1	1
Dibenz(a,h) anthracene	EPA 625.1	1
Diethyl phthalate	EPA 625.1	1
Dimethyl phthalate	EPA 625.1	1
Di-n-butyl phthalate	EPA 625.1	1
Di-n-octyl phthalate	EPA 625.1	1
Fluoranthene	EPA 625.1	1
Fluorene	EPA 625.1	1
Hexachlorobenzene	EPA 625.1	1
Hexachlorobutadiene	EPA 625.1	1
Hexachlorocyclopentadiene	EPA 625.1	1
Hexachloroethane	EPA 625.1	1
Indeno(1,2,3-cd) pyrene	EPA 625.1	1
Isophorone	EPA 625.1	1
m+p Cresol	EPA 625.1	
Naphthalene	EPA 625.1	1
n-Decane	EPA 625.1	1
Nitrobenzene	EPA 625.1	1
N-Nitrosodimethylamine	EPA 625.1	1
N-Nitroso-di-n-propylamine	EPA 625.1	1

Anatek Labs, Inc - Moscow

Matrix/Analyte	Method	Notes
<b>Non-Potable Water</b>		
N-Nitrosodiphenylamine	EPA 625.1	1
n-Octadecane	EPA 625.1	1
Pentachlorophenol	EPA 625.1	1
Phenanthrene	EPA 625.1	1
Phenol	EPA 625.1	1
Pyrene	EPA 625.1	1
Pyridine	EPA 625.1	
Fecal coliform-count	SM 9221 B+E1+C (LTB/BGB/EC-MPN)	
Total coliforms-count	SM 9221 B+E1+C (LTB/BGB/EC-MPN)	
E.coli-count	SM 9221 B+F+C (LTB/BGB/EC Mug-MPN)	
Total coliforms-count	SM 9221 B+F+C (LTB/BGB/EC Mug-MPN)	
<b>Solid and Chemical Materials</b>		
Chloride	EPA 300.0_2.1_1993	1
Fluoride	EPA 300.0_2.1_1993	1
Nitrate	EPA 300.0_2.1_1993	1
Nitrite	EPA 300.0_2.1_1993	1
Orthophosphate	EPA 300.0_2.1_1993	1
Sulfate	EPA 300.0_2.1_1993	1
Cyanide, Total	EPA 335.4_1_1993	
Cyanide, Total	EPA 9012 B-04	1
pH	EPA 9045 D_2004	1
Cyanide, Total	SM 4500-CN <sup>-</sup> E-2011	1
Cyanides, Amenable to Chlorination	SM 4500-CN <sup>-</sup> G-2011	4
Ammonia	SM 4500-NH3 G-2011	1,9
Nitrate (calc.)	SM 4500-NO3 <sup>-</sup> F-2011	1
Nitrogen, Total Kjeldahl	SM 4500-Norg C-2011	1
Phosphorus, total	SM 4500-P F-2011	1,4
Aluminum	EPA 6010D_(7/14)	1
Barium	EPA 6010D_(7/14)	1
Beryllium	EPA 6010D_(7/14)	1
Boron	EPA 6010D_(7/14)	1,4
Cadmium	EPA 6010D_(7/14)	1
Calcium	EPA 6010D_(7/14)	1
Chromium	EPA 6010D_(7/14)	1
Cobalt	EPA 6010D_(7/14)	1,4
Copper	EPA 6010D_(7/14)	1
Iron	EPA 6010D_(7/14)	1



Anatek Labs, Inc - Moscow

Matrix/Analyte	Method	Notes
<b>Solid and Chemical Materials</b>		
Magnesium	EPA 6010D_(7/14)	1
Manganese	EPA 6010D_(7/14)	1
Molybdenum	EPA 6010D_(7/14)	1
Nickel	EPA 6010D_(7/14)	1
Sodium	EPA 6010D_(7/14)	1
Vanadium	EPA 6010D_(7/14)	1
Zinc	EPA 6010D_(7/14)	1
Aluminum	EPA 6020B_(7/14)	1
Antimony	EPA 6020B_(7/14)	1
Arsenic	EPA 6020B_(7/14)	1
Barium	EPA 6020B_(7/14)	1
Beryllium	EPA 6020B_(7/14)	1
Boron	EPA 6020B_(7/14)	1
Cadmium	EPA 6020B_(7/14)	1
Chromium	EPA 6020B_(7/14)	1
Cobalt	EPA 6020B_(7/14)	1
Copper	EPA 6020B_(7/14)	1
Iron	EPA 6020B_(7/14)	1
Lead	EPA 6020B_(7/14)	1
Magnesium	EPA 6020B_(7/14)	1
Manganese	EPA 6020B_(7/14)	1
Mercury	EPA 6020B_(7/14)	1
Molybdenum	EPA 6020B_(7/14)	1
Nickel	EPA 6020B_(7/14)	1
Potassium	EPA 6020B_(7/14)	1,4
Selenium	EPA 6020B_(7/14)	1
Sodium	EPA 6020B_(7/14)	1
Strontium	EPA 6020B_(7/14)	1
Thallium	EPA 6020B_(7/14)	1
Tin	EPA 6020B_(7/14)	1,4
Titanium	EPA 6020B_(7/14)	1
Vanadium	EPA 6020B_(7/14)	1
Zinc	EPA 6020B_(7/14)	1
Methamphetamine	ALI SOP 602	1,5
4,4'-DDD	EPA 8081B_(2/07)	1
4,4'-DDE	EPA 8081B_(2/07)	1
4,4'-DDT	EPA 8081B_(2/07)	1

Anatek Labs, Inc - Moscow

Matrix/Analyte	Method	Notes
<b>Solid and Chemical Materials</b>		
Aldrin	EPA 8081B_(2/07)	1
alpha-BHC (alpha-Hexachlorocyclohexane)	EPA 8081B_(2/07)	1
beta-BHC (beta-Hexachlorocyclohexane)	EPA 8081B_(2/07)	1
Chlordane (tech.)	EPA 8081B_(2/07)	1
Dacthal (DCPA)	EPA 8081B_(2/07)	1
delta-BHC	EPA 8081B_(2/07)	1
Dieldrin	EPA 8081B_(2/07)	1
Endosulfan I	EPA 8081B_(2/07)	1
Endosulfan II	EPA 8081B_(2/07)	1
Endosulfan sulfate	EPA 8081B_(2/07)	1
Endrin	EPA 8081B_(2/07)	1
Endrin aldehyde	EPA 8081B_(2/07)	1
Endrin ketone	EPA 8081B_(2/07)	1
gamma-BHC (Lindane, gamma-Hexachlorocyclohexane)	EPA 8081B_(2/07)	1
Heptachlor	EPA 8081B_(2/07)	1
Heptachlor epoxide	EPA 8081B_(2/07)	1
Methoxychlor	EPA 8081B_(2/07)	1
Toxaphene (Chlorinated camphene)	EPA 8081B_(2/07)	1
Aroclor-1016 (PCB-1016)	EPA 8082A_(2/07)	1
Aroclor-1221 (PCB-1221)	EPA 8082A_(2/07)	1
Aroclor-1232 (PCB-1232)	EPA 8082A_(2/07)	1
Aroclor-1242 (PCB-1242)	EPA 8082A_(2/07)	1
Aroclor-1248 (PCB-1248)	EPA 8082A_(2/07)	1
Aroclor-1254 (PCB-1254)	EPA 8082A_(2/07)	1
Aroclor-1260 (PCB-1260)	EPA 8082A_(2/07)	1
Azinphos-ethyl (Ethyl guthion)	EPA 8141B_2_(2/07)	1,4
Azinphos-methyl (Guthion)	EPA 8141B_2_(2/07)	1
Bolstar (Sulprofos)	EPA 8141B_2_(2/07)	1,4
Carbophenothion	EPA 8141B_2_(2/07)	1
Chlorfenvinphos	EPA 8141B_2_(2/07)	1
Chlorpyrifos	EPA 8141B_2_(2/07)	1
Coumaphos	EPA 8141B_2_(2/07)	1,4
Demeton	EPA 8141B_2_(2/07)	
Demeton-o	EPA 8141B_2_(2/07)	1
Demeton-s	EPA 8141B_2_(2/07)	1,4
Diazinon	EPA 8141B_2_(2/07)	1,4
Dichlorovos (DDVP, Dichlorvos)	EPA 8141B_2_(2/07)	1

Anatek Labs, Inc - Moscow

Matrix/Analyte	Method	Notes
<b>Solid and Chemical Materials</b>		
Dimethoate	EPA 8141B_2_(2/07)	1
Dioxathion	EPA 8141B_2_(2/07)	
Disulfoton	EPA 8141B_2_(2/07)	1
EPN	EPA 8141B_2_(2/07)	1
Ethion	EPA 8141B_2_(2/07)	1,4
Ethoprop	EPA 8141B_2_(2/07)	1
Famphur	EPA 8141B_2_(2/07)	1
Fensulfothion	EPA 8141B_2_(2/07)	1,4
Fenthion	EPA 8141B_2_(2/07)	1
Malathion	EPA 8141B_2_(2/07)	1
Merphos	EPA 8141B_2_(2/07)	1,4
Methyl parathion (Parathion, methyl)	EPA 8141B_2_(2/07)	1
Mevinphos	EPA 8141B_2_(2/07)	1,4
Monocrotophos	EPA 8141B_2_(2/07)	1,4
Naled	EPA 8141B_2_(2/07)	1
Parathion, ethyl	EPA 8141B_2_(2/07)	1
Phorate	EPA 8141B_2_(2/07)	1
Phosmet (Imidan)	EPA 8141B_2_(2/07)	1,4
Ronnel	EPA 8141B_2_(2/07)	1
Sulfotepp	EPA 8141B_2_(2/07)	1
Terbufos	EPA 8141B_2_(2/07)	1
Tetrachlorvinphos (Stirophos, Gardona)	EPA 8141B_2_(2/07)	1
Tetraethyl pyrophosphate (TEPP)	EPA 8141B_2_(2/07)	1
Thionazin (Zinophos)	EPA 8141B_2_(2/07)	1,4
Tokuthion (Prothiophos)	EPA 8141B_2_(2/07)	1,4
Trichloronate	EPA 8141B_2_(2/07)	1,4
2,4,5-T	EPA 8151A_(1/98)	1
2,4-D	EPA 8151A_(1/98)	1
2,4-DB	EPA 8151A_(1/98)	1
Acifluorfen	EPA 8151A_(1/98)	
Bentazon	EPA 8151A_(1/98)	
Chloramben	EPA 8151A_(1/98)	
Dacthal (DCPA)	EPA 8151A_(1/98)	
Dalapon	EPA 8151A_(1/98)	1
DCPA di acid degradate	EPA 8151A_(1/98)	
Dicamba	EPA 8151A_(1/98)	1
Dichloroprop (Dichlorprop)	EPA 8151A_(1/98)	1

Anatek Labs, Inc - Moscow

Matrix/Analyte	Method	Notes
<b>Solid and Chemical Materials</b>		
Dinoseb (2-sec-butyl-4,6-dinitrophenol, DNBP)	EPA 8151A_(1/98)	1
MCPA	EPA 8151A_(1/98)	1
MCPP	EPA 8151A_(1/98)	
Pentachlorophenol	EPA 8151A_(1/98)	1
Picloram	EPA 8151A_(1/98)	
Silvex (2,4,5-TP)	EPA 8151A_(1/98)	1
1,3,5-Trinitrobenzene (1,3,5-TNB)	EPA 8330B_(10/06)	4
1,3-Dinitrobenzene (1,3-DNB)	EPA 8330B_(10/06)	4
2,4,6-Trinitrobenzene	EPA 8330B_(10/06)	4
2,4,6-Trinitrotoluene (2,4,6-TNT)	EPA 8330B_(10/06)	4
2,4-Dinitrotoluene (2,4-DNT)	EPA 8330B_(10/06)	4
2,6-Dinitrotoluene (2,6-DNT)	EPA 8330B_(10/06)	4
2-Amino-4,6-dinitrotoluene (2-am-dnt)	EPA 8330B_(10/06)	4
2-Nitrotoluene	EPA 8330B_(10/06)	4
3-Nitrotoluene	EPA 8330B_(10/06)	4
4-Amino-2,6-dinitrotoluene (4-am-dnt)	EPA 8330B_(10/06)	4
4-Nitrotoluene	EPA 8330B_(10/06)	4
Methyl-2,4,6-trinitrophenylnitramine (tetryl)	EPA 8330B_(10/06)	4
Nitrobenzene	EPA 8330B_(10/06)	4
Octahydro-1,3,5,7-tetranitro-1,3,5,7-tetrazocine (HMX)	EPA 8330B_(10/06)	4
RDX (hexahydro-1,3,5-trinitro-1,3,5-triazine)	EPA 8330B_(10/06)	4
Tetryl (methyl-2,4,6-trinitrophenylnitramine)	EPA 8330B_(10/06)	4
1,1,1,2-Tetrachloroethane	EPA 8260D_4_(6/18)	1
1,1,1-Trichloroethane	EPA 8260D_4_(6/18)	1
1,1,2,2-Tetrachloroethane	EPA 8260D_4_(6/18)	1
1,1,2-Trichloroethane	EPA 8260D_4_(6/18)	1
1,1-Dichloroethane	EPA 8260D_4_(6/18)	1
1,1-Dichloroethylene	EPA 8260D_4_(6/18)	1
1,1-Dichloropropene	EPA 8260D_4_(6/18)	1
1,2,3-Trichlorobenzene	EPA 8260D_4_(6/18)	1
1,2,3-Trichloropropane	EPA 8260D_4_(6/18)	1
1,2,4-Trichlorobenzene	EPA 8260D_4_(6/18)	1
1,2,4-Trimethylbenzene	EPA 8260D_4_(6/18)	1
1,2-Dibromo-3-chloropropane (DBCP)	EPA 8260D_4_(6/18)	1
1,2-Dibromoethane (EDB, Ethylene dibromide)	EPA 8260D_4_(6/18)	1
1,2-Dichlorobenzene	EPA 8260D_4_(6/18)	1
1,2-Dichloroethane (Ethylene dichloride)	EPA 8260D_4_(6/18)	1

Anatek Labs, Inc - Moscow

Matrix/Analyte	Method	Notes
<b>Solid and Chemical Materials</b>		
1,2-Dichloropropane	EPA 8260D_4_(6/18)	1
1,3,5-Trimethylbenzene	EPA 8260D_4_(6/18)	1
1,3-Dichlorobenzene	EPA 8260D_4_(6/18)	1
1,3-Dichloropropane	EPA 8260D_4_(6/18)	1
1,4-Dichlorobenzene	EPA 8260D_4_(6/18)	1
2,2-Dichloropropane	EPA 8260D_4_(6/18)	1
2-Butanone (Methyl ethyl ketone, MEK)	EPA 8260D_4_(6/18)	1
2-Chlorotoluene	EPA 8260D_4_(6/18)	1
2-Hexanone	EPA 8260D_4_(6/18)	1
4-Chlorotoluene	EPA 8260D_4_(6/18)	1
4-Isopropyltoluene (p-Cymene)	EPA 8260D_4_(6/18)	1,4
4-Methyl-2-pentanone (MIBK)	EPA 8260D_4_(6/18)	1
Acetone	EPA 8260D_4_(6/18)	1
Acrylonitrile	EPA 8260D_4_(6/18)	1
Benzene	EPA 8260D_4_(6/18)	1
Bromobenzene	EPA 8260D_4_(6/18)	1
Bromochloromethane	EPA 8260D_4_(6/18)	1
Bromodichloromethane	EPA 8260D_4_(6/18)	1
Bromoform	EPA 8260D_4_(6/18)	1
Carbon disulfide	EPA 8260D_4_(6/18)	1
Carbon tetrachloride	EPA 8260D_4_(6/18)	1
Chlorobenzene	EPA 8260D_4_(6/18)	1
Chlorodibromomethane	EPA 8260D_4_(6/18)	1,4
Chlorodifluoromethane (Freon-22)	EPA 8260D_4_(6/18)	
Chloroethane (Ethyl chloride)	EPA 8260D_4_(6/18)	1
Chloroform	EPA 8260D_4_(6/18)	1
cis-1,2-Dichloroethylene	EPA 8260D_4_(6/18)	1
cis-1,3-Dichloropropene	EPA 8260D_4_(6/18)	1
Dibromochloropropane	EPA 8260D_4_(6/18)	
Dibromomethane	EPA 8260D_4_(6/18)	1
Dichlorodifluoromethane (Freon-12)	EPA 8260D_4_(6/18)	1
Diethyl ether	EPA 8260D_4_(6/18)	1,4
Ethylbenzene	EPA 8260D_4_(6/18)	1
Hexachlorobutadiene	EPA 8260D_4_(6/18)	1
Iodomethane (Methyl iodide)	EPA 8260D_4_(6/18)	1,4
Isopropylbenzene	EPA 8260D_4_(6/18)	1
m+p-xylene	EPA 8260D_4_(6/18)	1

Anatek Labs, Inc - Moscow

Matrix/Analyte	Method	Notes
<b>Solid and Chemical Materials</b>		
Methyl bromide (Bromomethane)	EPA 8260D_4_(6/18)	1
Methyl chloride (Chloromethane)	EPA 8260D_4_(6/18)	1
Methyl tert-butyl ether (MTBE)	EPA 8260D_4_(6/18)	1
Methylene chloride (Dichloromethane)	EPA 8260D_4_(6/18)	1
Naphthalene	EPA 8260D_4_(6/18)	1
n-Butylbenzene	EPA 8260D_4_(6/18)	1
n-Propylbenzene	EPA 8260D_4_(6/18)	1
o-Xylene	EPA 8260D_4_(6/18)	1
p-Xylene	EPA 8260D_4_(6/18)	1
sec-Butylbenzene	EPA 8260D_4_(6/18)	1
Styrene	EPA 8260D_4_(6/18)	1
tert-Butylbenzene	EPA 8260D_4_(6/18)	1
Tetrachloroethylene (Perchloroethylene)	EPA 8260D_4_(6/18)	1
Toluene	EPA 8260D_4_(6/18)	1
trans-1,2-Dichloroethylene	EPA 8260D_4_(6/18)	1
trans-1,3-Dichloropropylene	EPA 8260D_4_(6/18)	1
trans-1,4-Dichloro-2-butene	EPA 8260D_4_(6/18)	1,4
Trichloroethene (Trichloroethylene)	EPA 8260D_4_(6/18)	1
Trichlorofluoromethane (Freon 11)	EPA 8260D_4_(6/18)	1
Vinyl chloride	EPA 8260D_4_(6/18)	1
Xylene (total)	EPA 8260D_4_(6/18)	1
1,2,4-Trichlorobenzene	EPA 8270E_6_(6/18)	1
1,2-Dichlorobenzene	EPA 8270E_6_(6/18)	1
1,2-Dinitrobenzene	EPA 8270E_6_(6/18)	
1,2-Diphenylhydrazine	EPA 8270E_6_(6/18)	1,4
1,3,5-Trinitrobenzene (1,3,5-TNB)	EPA 8270E_6_(6/18)	
1,3-Dichlorobenzene	EPA 8270E_6_(6/18)	1
1,3-Dinitrobenzene (1,3-DNB)	EPA 8270E_6_(6/18)	
1,4-Dichlorobenzene	EPA 8270E_6_(6/18)	
1,4-Dinitrobenzene	EPA 8270E_6_(6/18)	1
1-Methylnaphthalene	EPA 8270E_6_(6/18)	
2,2'-Oxybis(1-chloropropane)	EPA 8270E_6_(6/18)	
2,3,4,6-Tetrachlorophenol	EPA 8270E_6_(6/18)	1
2,3,5,6-Tetrachlorophenol	EPA 8270E_6_(6/18)	
2,4,5-Trichlorophenol	EPA 8270E_6_(6/18)	1
2,4,6-Trichlorophenol	EPA 8270E_6_(6/18)	1
2,4-Dichlorophenol	EPA 8270E_6_(6/18)	1

Anatek Labs, Inc - Moscow

Matrix/Analyte	Method	Notes
<b>Solid and Chemical Materials</b>		
2,4-Dimethylphenol	EPA 8270E_6_(6/18)	1
2,4-Dinitrophenol	EPA 8270E_6_(6/18)	1
2,6-Dinitrotoluene (2,6-DNT)	EPA 8270E_6_(6/18)	1
2-Chloronaphthalene	EPA 8270E_6_(6/18)	1
2-Chlorophenol	EPA 8270E_6_(6/18)	1
2-Methylnaphthalene	EPA 8270E_6_(6/18)	1
2-Methylphenol (o-Cresol)	EPA 8270E_6_(6/18)	1
2-Naphthylamine	EPA 8270E_6_(6/18)	
2-Nitroaniline	EPA 8270E_6_(6/18)	1
2-Nitrophenol	EPA 8270E_6_(6/18)	1
3,3'-Dichlorobenzidine	EPA 8270E_6_(6/18)	1
3-Nitroaniline	EPA 8270E_6_(6/18)	1
4,6-Dinitro-2-methylphenol	EPA 8270E_6_(6/18)	1
4-Bromophenyl phenyl ether (BDE-3)	EPA 8270E_6_(6/18)	1
4-Chloro-3-methylphenol	EPA 8270E_6_(6/18)	1
4-Chloroaniline	EPA 8270E_6_(6/18)	1
4-Chlorophenyl phenylether	EPA 8270E_6_(6/18)	1
4-Methylphenol (p-Cresol)	EPA 8270E_6_(6/18)	1,4
4-Nitroaniline	EPA 8270E_6_(6/18)	1
4-Nitrophenol	EPA 8270E_6_(6/18)	1
Acenaphthene	EPA 8270E_6_(6/18)	1
Acenaphthylene	EPA 8270E_6_(6/18)	1
Aniline	EPA 8270E_6_(6/18)	1
Anthracene	EPA 8270E_6_(6/18)	1
Azinphos-methyl (Guthion)	EPA 8270E_6_(6/18)	1,3,4
Benzidine	EPA 8270E_6_(6/18)	1
Benzo(a)anthracene	EPA 8270E_6_(6/18)	1
Benzo(a)pyrene	EPA 8270E_6_(6/18)	1
Benzo(g,h,i)perylene	EPA 8270E_6_(6/18)	1
Benzo(k)fluoranthene	EPA 8270E_6_(6/18)	1
Benzo[b]fluoranthene	EPA 8270E_6_(6/18)	1
Benzyl alcohol	EPA 8270E_6_(6/18)	1
beta-BHC (beta-Hexachlorocyclohexane)	EPA 8270E_6_(6/18)	3
bis(2-Chloroethoxy)methane	EPA 8270E_6_(6/18)	1
bis(2-Chloroethyl) ether	EPA 8270E_6_(6/18)	1
Butyl benzyl phthalate	EPA 8270E_6_(6/18)	1
Carbazole	EPA 8270E_6_(6/18)	1

Anatek Labs, Inc - Moscow


<b>Matrix/Analyte</b>	<b>Method</b>	<b>Notes</b>
<b>Solid and Chemical Materials</b>		
Carbophenothion	EPA 8270E_6_(6/18)	1
Chlorpyrifos	EPA 8270E_6_(6/18)	1,3
Chrysene	EPA 8270E_6_(6/18)	1
Demeton-s	EPA 8270E_6_(6/18)	1
Di(2-ethylhexyl)adipate	EPA 8270E_6_(6/18)	
Di(2-ethylhexyl)phthalate	EPA 8270E_6_(6/18)	1
Dibenz(a,h) anthracene	EPA 8270E_6_(6/18)	1
Dibenzofuran	EPA 8270E_6_(6/18)	1
Diethyl phthalate	EPA 8270E_6_(6/18)	1
Dimethoate	EPA 8270E_6_(6/18)	1,3
Dimethyl phthalate	EPA 8270E_6_(6/18)	1
Di-n-butyl phthalate	EPA 8270E_6_(6/18)	1
Di-n-octyl phthalate	EPA 8270E_6_(6/18)	1
Disulfoton	EPA 8270E_6_(6/18)	1,3
EPN	EPA 8270E_6_(6/18)	1
Famphur	EPA 8270E_6_(6/18)	1
Fenthion	EPA 8270E_6_(6/18)	1,3
Fluoranthene	EPA 8270E_6_(6/18)	1
Fluorene	EPA 8270E_6_(6/18)	1
Hexachlorobenzene	EPA 8270E_6_(6/18)	1
Hexachlorobutadiene	EPA 8270E_6_(6/18)	1
Hexachlorocyclopentadiene	EPA 8270E_6_(6/18)	1
Hexachloroethane	EPA 8270E_6_(6/18)	1
Indeno(1,2,3-cd) pyrene	EPA 8270E_6_(6/18)	1
Isophorone	EPA 8270E_6_(6/18)	1
m,p Cresol	EPA 8270E_6_(6/18)	1
Malathion	EPA 8270E_6_(6/18)	1,3
Methyl parathion (Parathion, methyl)	EPA 8270E_6_(6/18)	1,3
Mevinphos	EPA 8270E_6_(6/18)	1,3
Naled	EPA 8270E_6_(6/18)	1,3
Naphthalene	EPA 8270E_6_(6/18)	1
Nitrobenzene	EPA 8270E_6_(6/18)	1
n-Nitrosodimethylamine	EPA 8270E_6_(6/18)	1
N-Nitroso-di-n-propylamine	EPA 8270E_6_(6/18)	1
n-Nitrosodiphenylamine	EPA 8270E_6_(6/18)	1
Parathion	EPA 8270E_6_(6/18)	3
Pentachlorophenol	EPA 8270E_6_(6/18)	1



Matrix/Analyte	Method	Notes
<b>Solid and Chemical Materials</b>		
Phenanthrene	EPA 8270E_6_(6/18)	1
Phenol	EPA 8270E_6_(6/18)	1
Phorate	EPA 8270E_6_(6/18)	1,3
Pyrene	EPA 8270E_6_(6/18)	1
Pyridine	EPA 8270E_6_(6/18)	1
Sulfotepp	EPA 8270E_6_(6/18)	1,3
Terbufos	EPA 8270E_6_(6/18)	1,3
Tetrachlorvinphos (Stirophos, Gardona)	EPA 8270E_6_(6/18)	1,3
Diesel range organics (DRO)	WDOE NWTPH-Dx_(1997)	1,2
Gasoline range organics (GRO)	WDOE NWTPH-Gx_(1997)	1,2

**Accredited Parameter Note Detail**

(1) Accreditation is based in part on recognition of Florida Department of Health NELAP accreditation. (2) Analytical Methods for Petroleum Hydrocarbons, Publication No. ECY 97-602, June 1997. (3) Tributyl phosphate used for internal standard and normal 8270 surrogates used. (4) Accreditation is limited to water only. (5) Anatek Labs, Inc. SOP for determination of methamphetamine by HPLC-MS. (6) Approved for compliance testing only when holding time is met.(7) Method not approved for NPDES testing. (8) Accreditation based in part on recognition of Idaho Department of Health and Welfare accreditation. (9) Provisional accreditation pending submittal of acceptable Proficiency Testing (PT) results (WAC 173-50-110).(10) Interim accreditation pending the successful completion of an on-site audit to verify method capabilities (WAC 173-50-100).



07/03/2023

Authentication Signature  
 Rebecca Wood, Lab Accreditation Unit Supervisor

Date

The State of  
Department



Washington  
of Ecology

**LabTest**  
**Yakima, WA**

has complied with provisions set forth in Chapter 173-50 WAC and is hereby recognized by the Department of Ecology as an ACCREDITED LABORATORY for the analytical parameters listed on the accompanying Scope of Accreditation. This certificate is effective July 14, 2022 and shall expire July 13, 2023.

Witnessed under my hand on August 2, 2022

Rebecca Wood  
Lab Accreditation Unit Supervisor

Laboratory ID  
**C1008**

# WASHINGTON STATE DEPARTMENT OF ECOLOGY

## ENVIRONMENTAL LABORATORY ACCREDITATION PROGRAM

### SCOPE OF ACCREDITATION

#### LabTest

#### Yakima, WA

is accredited for the analytes listed below using the methods indicated. Full accreditation is granted unless stated otherwise in a note. EPA is the U.S. Environmental Protection Agency. SM is "Standard Methods for the Examination of Water and Wastewater." SM refers to EPA approved method versions. ASTM is the American Society for Testing and Materials. USGS is the U.S. Geological Survey. AOAC is the Association of Official Analytical Chemists. Other references are described in notes.

Matrix/Analyte	Method	Notes
<b>Drinking Water</b>		
Sulfate	ASTM D516-90	
pH	EPA 150.1_1982	1
Turbidity	EPA 180.1_2_1993	
Cyanide, Total	EPA 335.4_1_1993	
Nitrate	EPA 353.2_2_1993	
Nitrite	EPA 353.2_2_1993	1
Color	SM 2120 B-2011	
Hardness (calc.)	SM 2340 B-2011	
Specific Conductance	SM 2510 B-2011	
Chloride	SM 4500-Cl <sup>-</sup> E-2011	
Thallium	EPA 200.9 Rev 2.2 (1994)	2,3
Calcium	SM 3111 B-2011	2,3
Iron	SM 3111 B-2011	2,3
Magnesium	SM 3111 B-2011	2,3
Sodium	SM 3111 B-2011	2,3
Zinc	SM 3111 B-2011	2,3
Antimony	SM 3113 B-2010	
Arsenic	SM 3113 B-2010	
Barium	SM 3113 B-2010	
Beryllium	SM 3113 B-2010	
Cadmium	SM 3113 B-2010	
Chromium	SM 3113 B-2010	
Copper	SM 3113 B-2010	
Lead	SM 3113 B-2010	
Manganese	SM 3113 B-2010	
Nickel	SM 3113 B-2010	

LabTest

Matrix/Analyte	Method	Notes
<b>Drinking Water</b>		
Selenium	SM 3113 B-2010	2,3
Silver	SM 3113 B-2010	
Fecal coliform-count	SM 9222 D (mFC)-06	
Total coli/E.coli - detect	SM 9223 B Colilert® 24 (PA)	
<b>Non-Potable Water</b>		
Sulfate	ASTM D516-90	
Turbidity	EPA 180.1_2_1993	
Cyanide, Total	EPA 335.4_1_1993	
Ammonia	EPA 350.1_2_1993	1
Nitrate	EPA 353.2_2_1993	
Nitrate + Nitrite	EPA 353.2_2_1993	
Nitrite	EPA 353.2_2_1993	1
Specific Conductance	SM 2510 B-2011	1
Chloride	SM 4500-Cl <sup>-</sup> E-2011	
Biochemical Oxygen Demand (BOD)	SM 5210 B-2011	
Thallium	EPA 200.9 Rev 2.2 (1994)	2
Calcium	SM 3111 B-2011	2
Iron	SM 3111 B-2011	2
Magnesium	SM 3111 B-2011	2
Sodium	SM 3111 B-2011	2
Zinc	SM 3111 B-2011	2
Antimony	SM 3113 B-2010	
Arsenic	SM 3113 B-2010	
Barium	SM 3113 B-2010	
Beryllium	SM 3113 B-2010	
Cadmium	SM 3113 B-2010	
Chromium	SM 3113 B-2010	
Copper	SM 3113 B-2010	
Lead	SM 3113 B-2010	
Manganese	SM 3113 B-2010	
Nickel	SM 3113 B-2010	
Selenium	SM 3113 B-2010	2
Silver	SM 3113 B-2010	
Fecal coliform-count	SM 9222 D (mFC)-06	

---

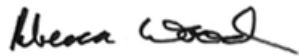
**Matrix/Analyte**

**Method**

**Notes**

**Accredited Parameter Note Detail**

(1) Provisional accreditation pending submittal of acceptable Proficiency Testing (PT) results (WAC 173-50-110).(2) Provisional status pending the submission of an acceptable corrective action plan in response to the 2019 audit findings (3) Provisional status for Drinking Water Parameters must be resolved within 90 days of the scope effective date.



08/02/2022

---

Authentication Signature

---

Date

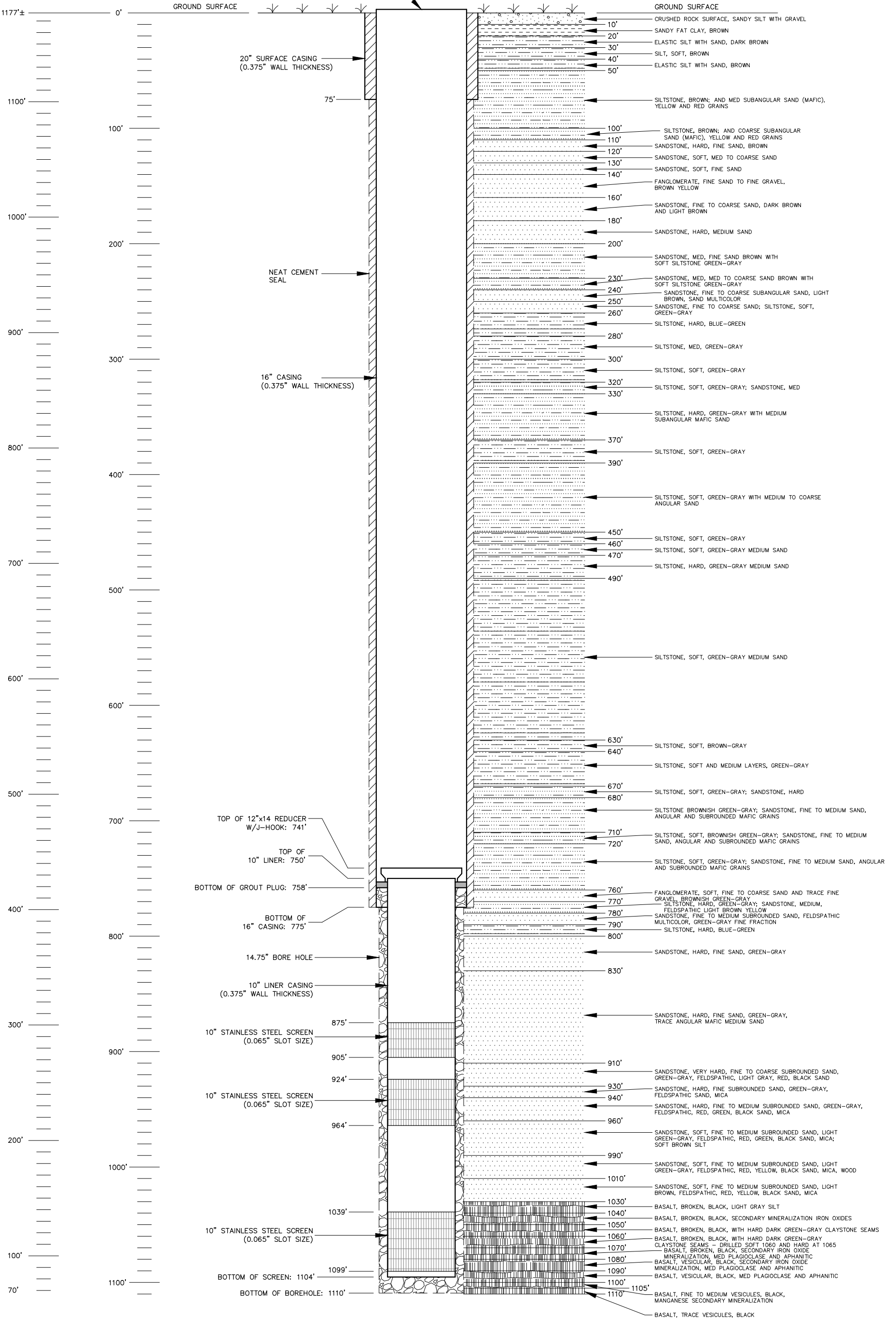
Rebecca Wood, Lab Accreditation Unit Supervisor

# **APPENDIX B**

## **Well Logs**

ELEVATION

DEPTH



NOTE: WELL CONSTRUCTION DETAILS AND FORMATION INFORMATION FROM WELL LOG DATED AUGUST 25, 2020.



2803 River Road  
Yakima, WA 98902  
509.966.7000  
Fax 509.965.3800  
www.hlacivil.com

19218	10-20-20
FILE NAMES:	
DRAWING: 19218.dwg	
DESIGNED BY:	JLB
ENTERED BY:	AJH

**CITY OF MOXEE**  
WELL NO. 4  
WELL CONSTRUCTION SCHEMATIC

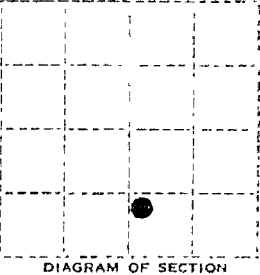
The Dep. The Department of Ecology does NOT Warranty the Data and/or the Information on this Well Report.

STATE OF WASHINGTON  
DEPARTMENT OF CONSERVATION  
AND DEVELOPMENT

WELL LOG No. Appl. 626  
Date January, 1943 Cert. 297A

Record by \_\_\_\_\_  
Source Driller's record

Location: State of WASHINGTON  
County Yakima  
Area \_\_\_\_\_  
Map Sec. 1, T. 12 N., R. 19 E.  
Town of Moxee City N. R. E.  
W.



Drilling Co. G. D. Hall & Associates  
Address Yakima, Washington  
Method of Drilling \_\_\_\_\_ Date \_\_\_\_\_ 19\_\_\_\_  
Owner Town of Moxee City,  
Address \_\_\_\_\_  
Land surface datum \_\_\_\_\_ ft. above  
below

CORRELATION	MATERIAL	THICKNESS (feet)	DEPTH (feet)
-------------	----------	------------------	--------------

Transcribe driller's terminology literally but paraphrase as necessary, in parentheses. If material water-bearing so state and record static level if reported. Give depths in feet below land surface datum unless otherwise indicated. Correlate with stratigraphic column if feasible. Following top of materials, list all casings, perforations, screens, etc.

See attached sheet

Pump test:

Dim: 1326' x 10"  
SWL: flowing  
DD: \_\_\_\_\_  
Meas. Dis: 875 g.p.m. Cert. For 300  
Shut-in pressure at g.s. 37 # per sq. in.  
Controlled by valve  
Casing: 10" from 0 to 344'  
8" " 334' " 789'  
6.5 " 765' " 1326'  
Perf: 12 - 1/8" x 1" per ft.

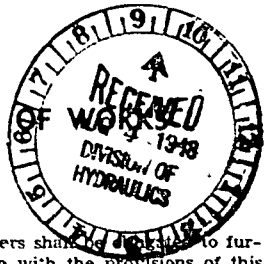


A 626

V Top soil	35	35
- Cement gravel	34	69
- Brown clay - wet	51	120
- Hard Brown clay	83	203
- Blue Clay - wet	72	275
- Hard Blue shale	5	280
- Blue Clay - wet	30	310
- Hard blue shale	15	325
- Hard blue clay	54	379
- Sandstone	31	410
- Loose sand	18	428
- Hard shale	7	435
- Coarse sand	26	461
- Clay	4	465
- Sand	15	480
- Sandstone	6	486
- Blue clay - wet	38	524
- Hard shale	45	569
- Sand	11	580
- Blue clay - wet	65	645
- Sandstone	23	668
- Blue clay	22	690
- Sandstone	8	698
- Clay - wet	65	763
- Sand	12	775
- Blue sand	8	783
- Sand and gravel	52	835
- Sand	22	857
- Blue clay	18	875
- Sand and clay	45	920
- Sandstone	70	990
- Blue clay	31	1021
- Brown sandstone	6	1027
- Packed sand	45	1072
- Shale	15	1087
- Blue clay	47	1134
- Sand and clay	4	1138
- Blue clay	40	1178
- Gray sandstone	22	1200
- Blue shale	23	1223
- Sandstone	53	1276
- Sand	50	1326

The Dep. The Department of Ecology does NOT Warranty the Data and/or the Information on this Well Report.

# RECORD BY WELL DRILLER OR OTHER CONSTRUCTOR FOR WITHDRAWAL OF GROUND WATER



Under Permit No. G. W. 727

("The well driller or other constructor of works for the withdrawal of public ground waters shall be required to furnish the permittee a certified record of the factual information necessary to show compliance with the provisions of this section." Sec. 8, Chap. 263, Laws of 1945.)

- The Town of Morse City, Washington  
(Name and address of owner of well or other works for withdrawal of water)
- Nature of works from which water is withdrawn Well  
(Well, tunnel, or infiltration trench)
- Name or number of works (if any) #1
- Date on which work on well or other structure was started January 1942
- Date on which work was completed January 1943
- If work on well or other structure was abandoned, give date  
and reason for abandonment

### 7. DESCRIPTION OF WORKS

(a) WELL: Depth 1326 ft. Diameter 10 in. or ft. Dug or drilled To be drilled  
Flowing or pump well Flowing

IF PUMP WELL. Type and size of pump is \_\_\_\_\_  
 Type and size of motor or engine is \_\_\_\_\_  
 Depth from ground surface to water level before pumping \_\_\_\_\_ feet  
 After continuous operation for at least four hours, the measured discharge of the pump is \_\_\_\_\_ g.p.m., and the drawdown of water level is \_\_\_\_\_ feet

Date of test \_\_\_\_\_  
 IF FLOWING WELL: Measured discharge 875 g.p.m., on January 6, 1943  
(Date)  
 Shut-in pressure at ground surface 37 lbs. per sq. in. on January 6, 1943  
(Date)  
 Water is controlled by valve  
(Cap, valve, etc.)

CASING: (Give diameter, commercial specifications and depth below ground surface of each casing size.)

<u>10</u> in. diameter	<u>Standard weight steel</u>	from <u>0</u> to <u>344</u> ft.
<u>8</u> in. diameter	" " "	from <u>334</u> to <u>789</u> ft.
<u>6.5</u> in. diameter	" " "	from <u>765</u> to <u>1326</u> ft.
in. diameter		from . . . to . . . ft.

Describe and show depth of shoe, plug, adapter, liner or other details:

Perforated casing or screens:

<u>12 - 1/4" x 1" per foot</u>	from <u>1225</u> to <u>1280</u> ft.
<small>(Number per foot and size of perforations or describe screen)</small>	
	from . . . to . . . ft.
	from . . . to . . . ft.
	from . . . to . . . ft.
	from . . . to . . . ft.



The Department of Ecology does NOT Warranty the Data and/or the Information on this Well Report.

Flowing

STATE OF WASHINGTON  
DEPARTMENT OF CONSERVATION  
AND DEVELOPMENT

WELL LOG

No. Decla #414

Date Jan., 19 43

Cert. #499-D

Record by Lester J. LaCoursier

Source G. W. Decla. Claim

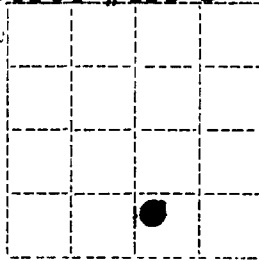
Location: State of WASHINGTON

County Yakima

~~xxx~~ Within limits of town

~~xxx~~ of Moxee City

SW 1/4 SE 1/4 sec. 1 T. 12 N., R. 19 E.



Drilling Co. \_\_\_\_\_

Address \_\_\_\_\_

Method of Drilling drilled Date Jan. 6 19 43

Owner Moxee City

Address Moxee City, Wash.

Land surface, datum \_\_\_\_\_ ft. <sup>above</sup> <sub>below</sub>

CORRE-LATION	MATERIAL	THICKNESS (feet)	DEPTH (feet)
--------------	----------	------------------	--------------

(Transcribe driller's terminology literally but paraphrase as necessary, in parentheses If material water-bearing, so state and record static level if reported Give depths in feet below land-surface datum unless otherwise indicated Correlate with stratigraphic column, if feasible Following log of materials, list all casings, perforations, screens, etc )

	<u>See following page</u>		
<u>Pump Test:</u>			
	<u>Dim: 1326' x 12"</u>		
	<u>Discharge: 875 g.p.m.</u>		
	<u>Shut-in pressure: 37 lbs./ sq. in.</u>		
	<u>Control: Valve</u>		
<u>Casing:</u>			
	<u>12" dia. Schedule 40 from 0' to 129'</u>		
	<u>10" " " " " 0' to 344'</u>		
	<u>8" " " " " 334' to 789'</u>		
	<u>6 5/8" " " " " 765' to 1320'</u>		
	<u>Perforations: 1/4 x 1" per foot from 1225' to 1280'.</u>		

Turn up \_\_\_\_\_

Sheet \_\_\_\_\_ of \_\_\_\_\_ sheets

LOG OF WELL. (Describe each stratum or formation clearly, indicate if water bearing, and type thickness and depth as indicated.)

MOORE CITY

MATERIAL	Thickness (Feet)	Depth to Bottom (Feet)
Top Soil	35	35
Cement-Gravel	34	69
Brown Clay	206	275
Blue Clay	104	379
Sandstone	31	410
Loose sand	18	428
Hard Shale	7	435
Coarse Sand	26	461
Clay	4	465
Sand	15	480
Sandstone	6	486
Blue Clay	38	524
Hard Shale	45	569
Sand	11	580
Blue Clay	65	645
Sandstone	23	668
Blue Clay	22	690
Sandstone	8	698
Clay	65	763
Sand	12	775
Blue Sand	8	783
Sand and Gravel	52	835
Sand	22	857
Blue Clay	18	875
Sand and Clay	45	920
Sandstone	70	990
Blue Clay	31	1021
Brown Sandstone	6	1027
Packed Sand	45	1072
Shale	15	1087
Blue Clay	47	1134
Sand and Clay	4	1138
Blue Clay	40	1178
Grey Sandstone	22	1200
Blue Shale	23	1223
Sandstone	53	1276
Sand	50	1326

(b) INFORMATION TAKEN: Covered or open.

Dimensions: Length ..... ft. Minimum depth ..... ft. Maximum depth ..... ft.

Bottom width ..... ft. Discharge ..... gpm. Date of year.

The Department of Ecology does NOT Warranty the Data and/or the Information on this Well Report.

# WATER WELL REPORT

STATE OF WASHINGTON

Application No. \_\_\_\_\_  
 Permit No. G4-27813P

(1) **OWNER:** Name City of Moxee Address PO Box 248, Moxee, WA 98936  
 LOCATION OF WELL: County Yakima — NE 1/4 SW 1/4 Sec 31 T 13 N, R 20E W.M.  
 Bearing and distance from section or subdivision corner \_\_\_\_\_

(3) **PROPOSED USE:** Domestic  Industrial  Municipal   
 Irrigation  Test Well  Other

(4) **TYPE OF WORK:** Owner's number of well 2  
 (if more than one).....  
 New well  Method: Dug  Bored   
 Deepened  Cable  Driven   
 Reconditioned  Rotary  Jetted

(5) **DIMENSIONS:** See Attached Exhibit A  
 Diameter of well \_\_\_\_\_ inches.  
 Drilled 978 ft. Depth of completed well 978 ft.

(6) **CONSTRUCTION DETAILS:** See Attached Exhibit A  
 Casing installed: \_\_\_\_\_ " Diam. from \_\_\_\_\_ ft. to \_\_\_\_\_ ft.  
 Threaded  \_\_\_\_\_ " Diam. from \_\_\_\_\_ ft. to \_\_\_\_\_ ft.  
 Welded  \_\_\_\_\_ " Diam. from \_\_\_\_\_ ft. to \_\_\_\_\_ ft.

Perforations: Yes  No   
 Type of perforator used \_\_\_\_\_  
 SIZE of perforations \_\_\_\_\_ in. by \_\_\_\_\_ in.  
 \_\_\_\_\_ perforations from \_\_\_\_\_ ft. to \_\_\_\_\_ ft.  
 \_\_\_\_\_ perforations from \_\_\_\_\_ ft. to \_\_\_\_\_ ft.  
 \_\_\_\_\_ perforations from \_\_\_\_\_ ft. to \_\_\_\_\_ ft.

Screens: Yes  No  See Attached Exhibit A  
 Manufacturer's Name \_\_\_\_\_  
 Type \_\_\_\_\_ Model No. \_\_\_\_\_  
 Diam. \_\_\_\_\_ Slot size \_\_\_\_\_ from \_\_\_\_\_ ft. to \_\_\_\_\_ ft.  
 Diam. \_\_\_\_\_ Slot size \_\_\_\_\_ from \_\_\_\_\_ ft. to \_\_\_\_\_ ft.

Gravel packed: Yes  No  See Attached  
 Size of gravel: \_\_\_\_\_  
 Gravel placed from \_\_\_\_\_ ft. to \_\_\_\_\_ ft.

Surface seal: Yes  No  To what depth? 772 ft.  
 Material used in seal pressure grouted  
 Did any strata contain unusable water? Yes  No   
 Type of water? sulfur Depth of strata 440  
 Method of sealing strata off cased & pressure grouted

(7) **PUMP:** Manufacturer's Name \_\_\_\_\_  
 Type: \_\_\_\_\_ H.P. \_\_\_\_\_

(8) **WATER LEVELS:** Land-surface elevation \_\_\_\_\_ ft.  
 above mean sea level.....  
 Static level 22 ft. below top of well Date \_\_\_\_\_  
 Artesian pressure \_\_\_\_\_ lbs. per square inch Date \_\_\_\_\_  
 Artesian water is controlled by \_\_\_\_\_ (Cap. valve, etc.)

(9) **WELL TESTS:** Drawdown is amount water level is lowered below static level  
 Was a pump test made? Yes  No  If yes, by whom? Valley Pump  
 Yield: 1600 gal./min. with 189 ft. drawdown after 24 hrs.  
 " " " " " "  
 " " " " " "  
 Recovery data (time taken as zero when pump turned off) (water level measured from well top to water level)  

Time	Water Level	Time	Water Level	Time	Water Level

 Date of test \_\_\_\_\_  
 Bailor test \_\_\_\_\_ gal./min. with \_\_\_\_\_ ft. drawdown after \_\_\_\_\_ hrs.  
 Artesian flow \_\_\_\_\_ g.p.m. Date \_\_\_\_\_  
 Temperature of water \_\_\_\_\_ Was a chemical analysis made? Yes  No

(10) **WELL LOG:**  
 Formation: Describe by color, character, size of material and structure, and show thickness of aquifers and the kind and nature of the material in each stratum penetrated, with at least one entry for each change of formation.

MATERIAL	FROM	TO
brown dirt, sand & small gravel	0	4
large gravel 3"-8" size w/sand & dirt mixed	4	5
soft sandy pea gravel w/sm amt lt brown clay	5	15
soft brown sand w/sand mixed	15	53
dense dark brown hard sticky clay	53	63
brown dense hard clay (sticky) some gravel	63	85
brown soft sand pea gravel w/brown clay	85	150
pea gravel & coarse sand slightly cemented together (soft formation)	150	220
small gravel w/sand & brown clay	220	235
green soft clay & coarse sand formation changed to more green clay, less coarse sand	235	262
green clay (soft) & fine silt	262	284
<del>silt</del> slab of sandstone or conglomerate	284	365
green clay soft some pea gravel	365	395
hard black slate stone some cracks & seams, water bearing est. 150-175 gpm d	395	432
blue soft clay w/silt & pea gravel, drilled ft./min. w/5000 lbs	432	478
blue soft clay & sand	478	560
green soft clay & sm gravel	560	582
soft green clay w/course sand & some pea gravel mixed	582	660
greenish gray soft clay & sand	660	755
tan colored hard sticky clay	755	770

continued on Page 2  
 Work started 12-7, 1982, Completed 3-28, 1983

**WELL DRILLER'S STATEMENT:**  
 This well was drilled under my jurisdiction and this report is true to the best of my knowledge and belief.  
 NAME Leach Well Drilling, Inc. (Type or print)  
 Address 821 W. Broadway, Moses Lake, WA 98837  
 [Signature] Richard L. Seel (Well Driller)  
 License No. 0276 Date 5-9, 1983

The Department of Ecology does NOT Warrant the Data and/or the Information on this Well Report.



Map No. R. 20 E. ~~W~~

Town. ~~Moxee~~, Wash X

Company. No. T. 


 Sec.

Farm. E. S. Wiseman

Authority. N. C. Jannsen

Elevation. 1150'

Collector. W. Warren 12 N. 


 6

Confidential. No

Date 1908 NE  $\frac{1}{4}$ , NW  $\frac{1}{4}$

No.	Strata	Thickness		Depth	
		Feet	In.	Feet	In.
1.	Alluvium Soil and gravel Ringold and/or Ellensburg formation	18		18	
2.	Sandstone	20		38	
3.	Sandstone and shale	60		98	
4.	Sandstone	120		218	
5.	Shale	40		258	
6.	Sandstone	229		487	
7.	Shale	190		677	
8.	Gravel	2		679	
9.	Shale	31		710	
10.	Sandstone	53		763	
11.	Shale, flow	65		828	
12.	Sandstone	100		928	
	5" well				

JBR

County.

**YAKIMA**

Index No. Y.4706.36





Moxee, Washington Well 4

From	To	Material
0	10	crushed rock surface, sandy silt with gravel
10	20	sandy fat clay, brown
20	30	elastic silt with sand, dark brown
30	40	silt, soft, brown
40	50	elastic silt with sand, brown
50	100	siltstone, brown; and med subangular sand (mafic), yellow and red grains
100	110	siltstone, brown; and coarse subangular sand (mafic), yellow and red grains
110	120	sandstone, hard, fine sand, brown
120	130	sandstone, soft, med to coarse sand
130	140	sandstone, soft, fine sand
140	160	fanglomerate, fine sand to fine gravel, brown-yellow
160	180	sandstone, fint to coarse sand, dark brown and light brown
180	200	sandstone, hard, medium sand
200	230	sandstone, med, fine sand brown with soft siltstone green-gray
230	240	sandstone, med, med to coarse sand brown with soft siltstone green-gray
240	250	sandstone, fine to coarse subangular sand, light brown, sand multicolor

Moxee, Washington Well 4

From	To	Material
250	260	sanstone, fine to coarse sand; siltstone, soft, green-gray
260	280	siltstone, hard, blue-green
280	300	siltstone, med, green-gray
300	320	siltstone, soft, green-gray
320	330	siltstone, soft, green-gray; sandstone, med
330	370	siltstone, hard, green-gray with medium subangular mafic sand
370	390	siltstone, soft, green-gray
390	450	siltstone, soft, green-gray with medium to coarse angular sand
450	460	siltstone, soft, green gray
460	470	siltstone, soft, green gray medium sand
470	490	siltstone, hard, green gray medium sand
490	630	siltstone, soft, green gray medium sand
630	640	siltstone, soft, brown gray
640	670	siltstone, soft and medium layers, green gray
670	680	siltstone, soft, green gray; sandstone, hard
680	710	siltstone brownish green gray; sandstone, fine to medium sand, angular and subrounded mafic grains

Moxee, Washington Well 4

From	To	Material
710	720	siltstone, soft, brownish green gray; sandstone, fine to medium sand, angular and subrounded mafic grains
720	760	siltstone, soft, green gray; sandstone, fine to medium sand, angular and subrounded mafic grains
760	770	fanglomerate, soft, fine to coarse sand and trace fine gravel, brownish green gray
770	780	siltstone, hard, green gray; sandstone, medium, feldspathic light brown yellow
780	790	sandstone, fine to medium subrounded sand, feldspathic, multicolor, green-gray fine fraction
790	800	siltstone, hard, blue-green
800	830	sandstone, hard, fine sand, green-gray
830	910	sandstone, hard, fine sand, green-gray, trace angular mafic medium sand
910	930	sandstone, very hard, fine to coarse subrounded sand, green-gray, feldspathic, light gray, red, black sand
930	940	sandstone, hard, fine subrounded sand, green gray, feldspathic sand, mica
940	960	sandstone, soft, fine to medium subrounded sand, green gray, feldspathic, red, green, black sand, mica
960	990	sandstone, soft, fine to medium subrounded sand, light green gray, feldspathic, red, green, black sand, mica; soft brown silt
990	1010	sandstone, soft, fine to medium subrounded sand, light green gray, feldspathic, red, yellow, black sand, mica, wood
1010	1030	sandstone, soft, fine to medium subrounded sand, light brown, feldspathic, red, yellow, black sand, mica,
1030	1040	basalt, broken, black, light gray silt
1040	1050	basalt, broken, black, secondary mineralization iron oxides

Moxee, Washington Well 4

From	To	Material
1050	1060	basalt, broken, black, with hard dark green-gray claystone seams
		basalt, broken, black, with hard dark green-gray claystone seams - drilled
1060	1070	soft at 1060 and hard at 1065
		basalt, broken, black, secondary iron oxide mineralization, med plagioclase
1070	1080	and aphanitic
		basalt, vesicular, black, secondary iron oxide mineralization, med
1080	1090	plagioclase and aphanitic
1090	1100	basalt, vesicular, black, med plagioclase and aphanitic
1100	1105	basalt, fine to med vesicles, black, manganese secondary mineralization
1105	1110	basalt, trace vesicles, black

Moxee Well 4 Hole Penetration Logs

Hole Name Well 4

Hole Diameter 19

Duration in Minutes		Drilling Description i.e. exact depth of breaks, Rough, fractured, smooth soft, hard etc)	Upper Hoist Guage PSI	Duration in Minutes		Drilling Description i.e. exact depth of breaks, Rough, fractured, smooth soft, hard etc)	Upper Hoist Guage PSI
Depth	Minutes		PSI	Depth	Minutes		PSI
5	25	compact gravel	—	155	12	Softer, High clay content	650
10	60	sands gravel		160	15	clay, silt, rough	700
15	60	clays, silts		165	25	Dense clay	700
20	45	clays, silts		170	25	Harder, Rock/silts	650 700
25	60	some cobble, clay		175	20	clay, silt	650 700
30	70	clay, silt		180	20	1.8' Broken UP/rough 12.9-15 High Torque (CLAY)	600 700
35	70	clay, silt		185	30	Broken-up 18.5 High Torque, rough / smooth 18.5-18.7 clay/sandstone MED-197-190	650-700
40	60	clays/silt		190	10		650
45	60	clays/silt		195	15	MEDIUM-HARD sandstone	650
50	70	gravel/cobbles		200	15	sandstone, MED	650
55	70	gravel/cobbles		205	20	HARD	650
60	60	solid dense clay		210	15	206-207 - green clay, high torque CLAY	650 950
65	60	solid dense clay		215	15	Added to BENT clay, silt	900
70	60	solid dense clay		220	12	Brown & Grey clay	900
75	60	solid dense clay		225	20	Harder, some rock	950
80	25	solid, hard silt		230	20	Grey clay, pebbles	950
85	45	solid, hard silt		235	20	Grey clay, Rusty <sup>some rock</sup> pebbles	950-1100
90	20	smooth, HARD	650	240	30	Grey clay, Brown/Blk Mts	950-1100
95	16	MEDIUM, HARD, smooth	650	245	60	Grey clay/brown/blk pebble	1200
100	11	MEDIUM, smooth	650	250	120	Grey clay, Brown/Blk rock	1000-1100
105	14	MEDIUM, SOFT, some CLAY 104' CLAY 105-106	550	255	60	Grey into Brown/green clay	1150
110	12	SOFT, MEDIUM	550-650	260	75	Brown into Blue/grey clay	1150
115	10	SOFT, MEDIUM, smooth	550-650	265	50	Grey/blue clay,	1150
120	16	116' Broken UP, HARD	550-650	270	60	Grey/blue clay, 267 transition	1150
125	17	Broken UP 125-123' HARD 125' clay Broken-up	600-650	275	20	lighter grey, w/ black pebbles some sand	1000
130	15	smooth, soft	650	280	12	grey & 1.25' Boulder some greenish brown clay	1050
135	12	smooth, soft	650	285	15	Grey clay w/ coarse sand	1050
140	15	High clay content, soft	650	290	16	" " " "	1050 1000
145	20	High clay content, soft	650	295	15	Brown clay, sandstone mix smooth	1100-1000
150	40	Harder, solid smooth		300	45	Dense, sticky clay	1200-1000

206' formation change greenish clay

Moxee Well 4 Hole Penetration Logs

Hole Name

Well 4

Hole Diameter

19

Duration in Minutes	Drilling Description i.e. exact depth of breaks, Rough, fractured, smooth soft, hard etc)	Upper Hoist Gauge PSI	Duration in Minutes	Drilling Description i.e. exact depth of breaks, Rough, fractured, smooth soft, hard etc)	Upper Hoist Gauge PSI
305 60	gray clay, traces of sandstone Smooth SOFT	1100-1200	460 18	SOFT, CLAY, SILT	1100
310 30	SOFT Green clay, some green	1100	465 35	SOFT, Broken UP, Roughly 464-465	1200
315 30	SOFT Green clay, some green	1100	470 50	BLUE clay / gray clay	1300
320 30	Gray clay, traces of sandstone	1100	475 1:00	CLAY, soft smooth	1400
325 30	Blue clay, cemented	1100	480 45	H26 - FINE SILT	1000
330 30	Gray clay, pebbles	1100	485 35	SILT, sand, soft	1200
335 35	Gray clay, coarse sand	1100	490 40	H28 HARD, Broken UP, 488-490 SILT, clay	1000-1200
340 30	Gray clay, black coarse sand	1950	495 45	SILT, sand, clay	900
345 60	Black pebbles / coarse sand	1100	500 35	SILT, sand, clay	1300
350 130	SOFT Green clay, very sticky	1250	505 60	BLUE clay, very clay, coarse sand	1400
355 180	SILT, CLAY, SILT, HARD	1150	510 50	CLAY, cemented, fines	1000
360 90	356 Brown clay, black pebbles, coarse sand	1250	515 50	CLAY, sand, coarse sand	1400
365 90	364 Formation change, hard rock, trace of clay	1150	520 45	CLAY, coarse sand	1000
370 80	366 BLUE clay, trace of SILT	1250	525 40	CLAY	1300
375 80	377 some water, soft clay, some SILT	1100	530 45	CLAY	1400
380 47	CLAY, sandstone mix, soft, medium	1250	535 40	CLAY, SILT	1700
385 30	382 WATER	950	540 45	338 SILT, some HARD	1200
390 30	CLAY / sandstone - soft / medium	900	545 40	CLAY, SILT, sand, SILTS, coarse sand	1100
395 190	BLUE clay / gravel mix, some HARD	1000	550 40	546 coarse sand / SILT	1000
400 130	SOFT MEDIUM, some clay	1200	555 30	547 coarse sand, SILT	1000
405 50	SO small gravel with coarse sand	1200	560 25	548 coarse sand, SILT	900
410 40	HARD, Broken UP, slate stone	800-900	565 25	549 SILT, grey clay	950
415 43	403 Broken UP, rough, HARD	800-1000	570 25	569 soft clay, cobble	1100
420 46	HARD, gravel, mix with slate	800-1000	575 25	Very soft, Harder @ 575	1250
425 45	HARD, Broken UP	800-1000	580 35	Dense sand / SILT, hard	1050
430 40	HARD, Black Rock	800-900	585 35	SOFT, rough, Dense clay	1150
435 45	BROKEN UP / gravel / coarse sand	800-900	590 30	SOFT, coarse sand / pebbles	1300
440 40	BROKEN UP / HARD	800-1000	595 30	SOFT, Pea gravel, green clay	1250
445 30	TRAIL OF grey CLAY	800-1000	600 25	SOFT, Green clay w/ sand	10250
450 30	335 BROKEN UP / rough / HARD	800-1000			
455 20	440-441 - HARD - rough	800-1100			
	442-445 SOFT, SILTS some clay	800-1100			
	SOFT, CLAY, SILT	1200-1000			
	SOFT, CLAY, SILT	1200-1000			

Moxee Well 4 Hole Penetration Logs

Hole Name Well 4

Hole Diameter 19

Duration		Drilling Description i.e. exact depth of breaks, Rough, fractured, smooth soft, hard etc)	Upper		Duration		Drilling Description i.e. exact depth of breaks, Rough, fractured, smooth soft, hard etc)	Upper	
In	Minutes		Hoist	Guage	In	Minutes		Hoist	Guage
Depth	Minutes		PSI	Depth	Minutes		PSI		
600	25	Soft Green clay w/ sand	1250	755	15	Soft Green/Grey clay	1350		
605	20	Soft clay w/ rock	1300	760	20	Harder Green/Grey sandy clay	1300		
610	45	60% Very sticky green clay	1350	765	75	Soft grey 765 grey clay	1350		
615	90	Very sticky green clay w/ sand	1350	770	25	765 Harder sand/blue clay	1150		
620	90	Very sticky green clay w/ sand	1300	775	45	Hard soft spot 775	1150		
625	25	Coarse sand & pebbles	1000	780	40	Grout shoe @ 775	490		06/02
630	20	Coarse sand, green clay	1100	785	15	hard to very soft	1050		
635	20	Green clay traces of sand	1000	790	12	Very soft smooth	1050		
640	25	Grey clay, sand silt	1050	795	12	Very soft smooth green/brown	1050		
645	35	Sticky clay, very soft	1150	800	12	Very soft	1050		
650	45	Green clay, sticky	1300	805	12	some rough spots	1050		
655	90	65% Very hard until 65% very soft clay	1350	810	15	65% with black speck	1050		
660	90	Very soft sticky green clay	1300	815	12	soft very rough w/ black speck	1050		
665	60	Getting into some red sand green clay	1250	820	17	soft blue/grey clay	1050		
670	75	Very sticky green clay	1350	825	12	soft blue/grey clay	1050		
675	25	Coarse sand, pebbles blue/grey clay	1100	830	15	soft blue/grey clay	1050		
680	35	Siltstone, blue grey clay	1100	835	12	soft blue/grey clay	1050		
685	90	Very hard mostly rock	1000	840	22	soft blue/grey clay	1050		
690	90	66% soft clay	1000	845	25	Very dense blue clay	1050		
695	75	Very rough/hard	1000	850	18	Dense blue clay	1050		
700	30	Very soft at 695	1300	855	28	Black rock bits	1000		
705	30	Very soft sticky grey/blue clay	1400	860	25	Hard smooth mostly rock chips, blue clay	1050		
710	25	Soft grey/blue clay	1300	865	14	soft @ 64 blue clay	1100		06/03
715	25	Soft grey/blue clay	1300	870	15	soft blue clay	1000		
720	30	Soft green/grey/blue clay	1300	875	20	harder more rock chips, some fine sand	1100		
725	30	Soft green/grey/blue clay	1250	880	22	Very hard until	1100		
730	35	Traces of pebbles	1500	885	12	soft, smooth blue/grey	1150		
735	20	Soft green/blue clay	1300	890	10	Very soft, smooth blue/grey	1150		
740	15	Very soft green/blue clay	1350	895	10	Very soft smooth blue/grey	1150		
745	15	Very soft green/blue clay	1350	900	10	Very soft blue/grey clay	1150		
750	15	Very soft green/blue clay	1350						



Moxee Well 4 Hole Penetration Logs

Hole Name Well 4

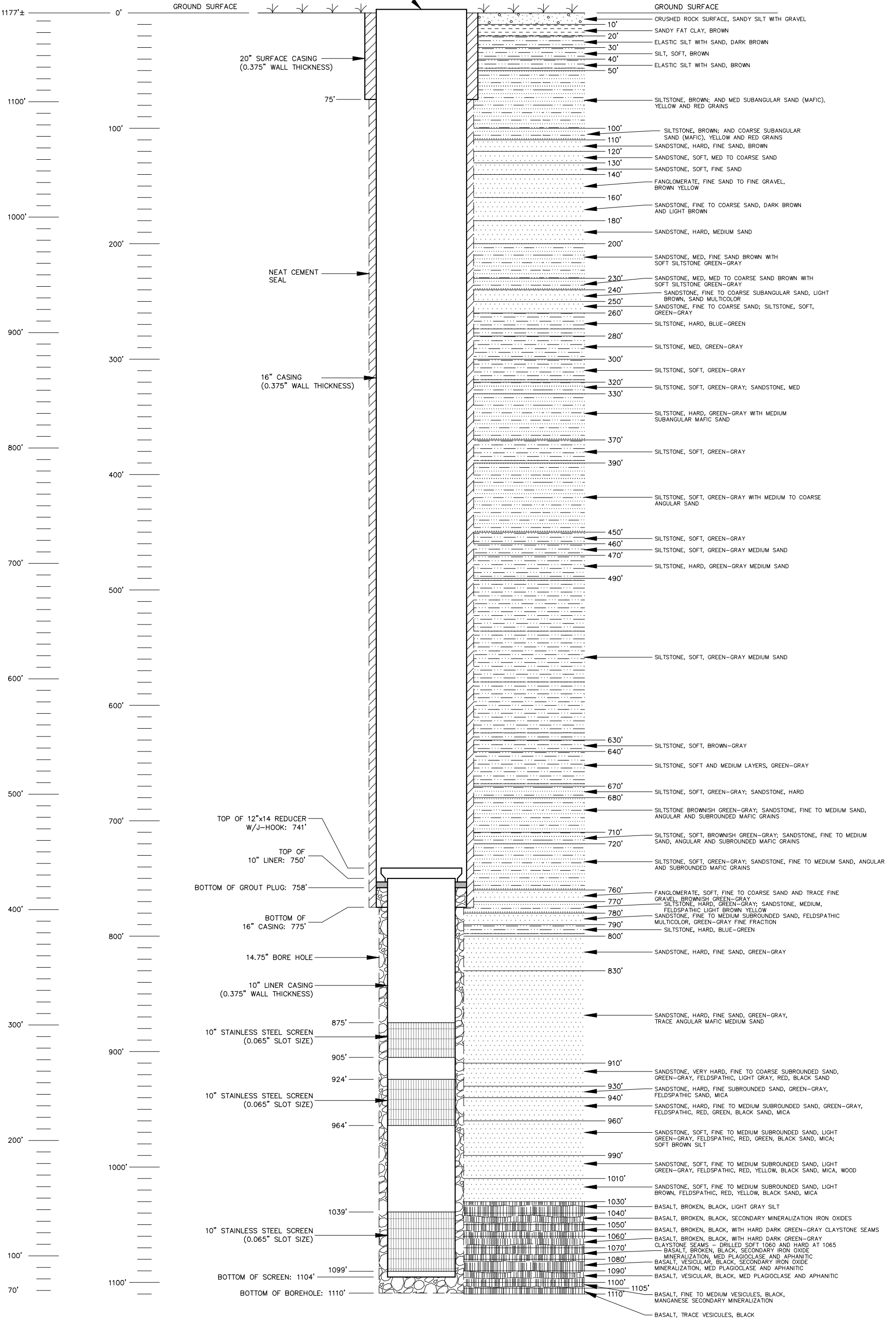
Hole Diameter 19"

Duration in Minutes		Drilling Description i.e. exact depth of breaks, Rough, fractured, smooth soft, hard etc)	Upper Hoist Guage PSI	Duration in Minutes		Drilling Description i.e. exact depth of breaks, Rough, fractured, smooth soft, hard etc)	Upper Hoist Guage PSI
905	10	Very soft Blue smooth grey clay	1150	1060	20	Smooth, Solid Black face	1100
910	12	Some chert here but still soft smooth	1150	1065	24	Slightly harder smooth	1100
915	25	hard Blue/Grey Black face	1100	1070	30	Very Rough some green black	1200
920	28	Solid/Dense Blue/Grey all over chert	1000	1075	75	Very hard/Rough some very rough	1250
925	17	Softer still smooth grey Blue with black spots	1200	1080	90	Very Rough/Block	1200
930	18	Dense hard clay with blue streaks of chert	1100	1085	120	Very hard consistent	1200
935	18	Same very soft sticky sections	1100	1090	120	Very hard consistent	1200
940	10	Very soft Blue Grey with green	1150	1095	90	Very hard smooth	1200
945	10	Very soft Blue Grey with green	1150	1100	90	Solid/Very hard	1200
950	10	Very soft Blue/Grey	1150	1105	90	Solid/Very hard	1200
955	10	Very soft Blue/Grey	1150	1110	130	Very hard/Rough	1200
960	12	Very soft Blue/Grey	1150	1115			
965	10	Very soft Blue/Grey	1150	1120			
970	10	Very soft Blue/Grey	1200	1125			
975	10	Very soft Blue/Grey Brown clump	1200	1130			
980	10	Blue/Grey & Brown	1200	1135			
985	10	Very soft Blue/Grey & Brown	1200	1140			
990	10	Very soft Grey Brown	1200	1145			
995	10	lost circulation little rough soft	1200	1150			
1000	10	wood, black wood	1200	1155			
1005	10	Very soft sand/wood	1200	1160			
1010	8	Very soft sand/wood	1250	1165			
1015	10	Very soft sand	1250	1170			
1020	10	Very soft sand	1250	1175			
1025	43	Slightly harder rough	1150	1180			
1030	28	Rough here but	1150	1185			
1035	65	Very hard/Rough Dark face	1000	1190			
1040	90	Very rough	1200	1195			
1045	90	Very rough Black face	1250	1200			
1050	75	Getting softer / smoother	1200				
1055	30	Softer still hard	1100				

5-14-12 3:30  
U.S.

ELEVATION

DEPTH



NOTE: WELL CONSTRUCTION DETAILS AND FORMATION INFORMATION FROM WELL LOG DATED AUGUST 25, 2020.



2803 River Road  
Yakima, WA 98902  
509.966.7000  
Fax 509.965.3800  
www.hlacivil.com

19218	10-20-20
FILE NAMES:	
DRAWING: 19218.dwg	
DESIGNED BY:	JLB
ENTERED BY:	AJH

**CITY OF MOXEE**  
WELL NO. 4  
WELL CONSTRUCTION SCHEMATIC

## **APPENDIX C**

### **Aspect Field Data Sheets**





# DAILY REPORT

350 Madison Avenue North  
Bainbridge Island, Washington 98110  
(206) 780-9370

710 Second Avenue, Suite 550  
Seattle, Washington 98104  
(206) 328-7443

<b>DATE:</b>	<b>PROJECT NO.</b>	<b>WEATHER:</b>
<b>PROJECT NAME:</b>		<b>CLIENT:</b>
<b>EQUIPMENT USED:</b>		<b>PROJECT LOCATION:</b>

**THE FOLLOWING WAS NOTED:**

<b>COPIES TO:</b>	<b>Aspect Consulting PROJECT MANAGER:</b>
<b>Page 1 of 1      FIELD REP.:</b>	

