

# Commencement Bay Sediment Trap Monitoring Program

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## Commencement Bay Sediment Trap Monitoring Program

Results of monitoring conducted in Hylebos, Sitcum, and Thea Foss Waterways between June 1991 and June 1995

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## Abstract

In November 1988 the Washington State Department of Ecology began a Sediment Trap Monitoring Program in the Nearshore/Tideflats area of Commencement Bay. The program was aimed at evaluating the effectiveness of source control efforts designed to reduce the input of problem chemicals to Thea Foss Waterway. The primary component of this program consisted of a quarterly collection of settling particulate matter (SPM) with the use of moored sediment traps. In-place bottom sediments, current velocity measurements and water column profile data have also been collected periodically as part of the program. Based on the results from Thea Foss Waterway, sediment trap monitoring was expanded in July 1990 to include Hylebos and Sitcum Waterways.

This report presents an update of SPM and bottom sediment monitoring data collected from Hylebos, Sitcum and Thea Foss Waterways between June 1991 and June 1995. Information on the distribution of contaminants, trends in contaminant levels, estimated sedimentation and bottom sediment resuspension rates, and comparisons with the Commencement Bay "Sediment Quality Objectives" is discussed. Recommendations for follow-up work are also presented.

### Summary

The spatial distribution of contaminants in settling particulate matter (SPM) from Hylebos, Sitcum, and Thea Foss Waterways was in relatively good agreement with previous monitoring results collected between November 1988 and November 1992. In all three waterways metals concentrations were typically higher at the head of the waterways and decreased moving toward the mouth. A similar pattern was also present for most of the organic compounds; however, concentrations tended to be more variable.

Overall, SPM data collected from Hylebos and Thea Foss Waterways suggest that concentrations of most problem chemicals have not decreased since sediment trap monitoring was started in July 1990 and November 1988, respectively. In contrast, dramatic reductions in contaminant levels have been observed in Sitcum Waterway comparing pre- and post-dredging conditions. It should be noted that metals data collected after May 1994, for all elements except mercury, were not used to evaluate contaminant trends. A shift in analytical methods after May 1994 is the primary reason for excluding these data. The reader should consult the methods section for a complete discussion of data quality.

Gross sedimentation rates (net + resuspension) in each of the waterways tended to be highest during the summer and fall and lowest during the winter and early spring. The highest average sedimentation and resuspension rates were measured in Sitcum followed by Hylebos, and Thea Foss Waterway. The data collected indicate that 60% to 85% of the material collected by the traps could be bottom sediments which have been resuspended. While the exact proportion of "new" versus "old" material collected by the traps cannot be reliably determined with the available data, it is fair to assume that the trapped material is representative of sediments which are mobile in the waterways.

Specific findings for Hylebos, Sitcum, and Thea Foss Waterways are listed in the following sections:

### **Hylebos Waterway**

• In contrast to the general distribution of contaminants associated with SPM mentioned above, lead, tetrachloroethene, hexachlorobenzene, hexachlorobutadiene, and 1,2,4-trichlorobenzene were elevated in the outer portion of Hylebos Waterway adjacent to Occidental Chemical. Occidental Chemical was identified as a source of hexachlorobenzene and hexachlorobutadiene during the Commencement Bay Remedial Investigation. In addition, between 1947 and 1973 tetrachloroethene was produced in commercial quantities at Occidental's Solvent Plant (Tetra Tech, 1986b).

• Fourteen individual chemicals (3 metals and 11 organics) listed below were measured at concentrations exceeding the Commencement Bay Sediment Quality Objectives (SQOs) in SPM. The greatest number of exceedances were present at the head of the waterway in the upper turning basin.

Metals - arsenic, mercury, and zinc

<u>Organics</u> - low molecular weight polynuclear aromatic hydrocarbons (LPAH), high molecular weight PAH (HPAH), dibenzofuran, 4-methyl phenol, benzoic acid, di-n-butyl phthalate, dimethyl phthalate, bis(2-ethyl hexyl)phthalate, hexachlorobenzene, hexachlorobutadiene, and total polychlorinated biphenyls (PCB)

• Tributyltin (TBT), which has been used extensively in the past as an active ingredient in antifouling paints, was present at relatively high levels (55-1300 ug/kg, dry) in SPM throughout Hylebos. TBT levels in all of the samples analyzed from the waterway exceeded the Puget Sound Dredge Disposal Analysis (PSDDA) interim screening level (ISL) of 30 ug/kg, dry for TBT.

The U.S. Environmental Protection Agency (EPA) Region 10 has also recently established a site specific sediment cleanup level of 19,250 ug TBT/kg organic carbon (OC) for Hylebos Waterway. On an OC normalized basis, TBT levels in Hylebos Waterway ranged from 2,800 to 26,000, with a mean of 11,000 ug TBT/kg OC. Only one value exceeded the Hylebos specific cleanup level in SPM.

- A comparison of PCB levels in bottom sediments collected in 1984, 1991, and 1994 suggests that concentrations have not declined since the Remedial Investigation was conducted in 1984. On average, PCB levels in Hylebos Waterway were approximately five times higher then those measured in Thea Foss Waterway.
- A significant decrease in arsenic levels associated with SPM was observed between July 1990 and May 1994 at the mouth of the waterway.
- The average gross sedimentation rate for Hylebos Waterway between December 1991 and June 1995 was 2.4±1.0 g/cm<sup>2</sup>/yr (see page 26 for explanation of gross sedimentation).
- Resuspension estimates for bottom sediments ranged from 81% (1.3±0.5 g/cm<sup>2</sup>/yr) at the head of Hylebos to 63% (1.9±1.2 g/cm<sup>2</sup>/yr) at the mouth of the waterway (see page 55 for explanation of resuspension).

### Sitcum Waterway

• Prior to dredging in Sitcum Waterway, 7 individual chemicals (1 metal and 6 organics) listed below, exceeded the SQOs in SPM. After the completion of dredging <u>no</u> exceedances of the SQOs were observed for contaminant levels in SPM. Dimethyl phthalate in bottom sediment, near the north corner drain (SI-172), was the only chemical that exceeded the SQOs after dredging was completed.

Metals - arsenic

<u>Organics</u> - LPAH, benzoic acid, benzyl alcohol, pentachlorophenol, dimethyl phthalate, and bis(2-ethyl hexyl)phthalate

- Significant decreases in lead (near north corner drain and Terminal 7 @ Berth B) and zinc (Terminal 7 @ Berth B) levels were noted in SPM between July 1990 and December 1993. This finding is consistent with the fact that as of July 1991, off-loading of black ore (lead and zinc) ceased at Terminal 7.
- In contrast to lead and zinc a significant increase in arsenic levels between July 1990 and December 1993 was noted in SPM near the north corner drain. Recent source control work conducted by the City of Tacoma in the summer of 1995 (storm drain cleaning) may have reduced present arsenic loadings to this area of the waterway.
- The average gross sedimentation rate in Sitcum Waterway between June 1991 and June 1995 was 4.8±1.8 g/cm<sup>2</sup>/yr.
- The highest bottom sediment resuspension rate (85%) for all three waterways was measured at the mouth of Sitcum Waterway.
- Resuspension estimates for Sitcum Waterway ranged from 71% (3.9±1.2 g/cm<sup>2</sup>/yr) at the mouth to 85% (3.5±1.6 g/cm<sup>2</sup>/yr) at the head of the waterway. The resuspension rate at the head of Sitcum was the highest measured in the three waterways being monitored.

### **Thea Foss Waterway**

 Mercury and copper concentrations tended to peak in the central portion of Thea Foss Waterway at two locations: near the mouth of Wheeler-Osgood Waterway (copper) and north of the 15th Street drain (mercury). J.M. Martinac Shipbuilding, located near the mouth of Wheeler-Osgood Waterway, has been identified as a copper source near the mouth of Wheeler-Osgood Waterway in past studies. The 15th Street drain, shoreline banks adjoining the Investco Site, and the inactive Surplus Steam Plant #1 have been identified as potential sources of mercury in this area of the waterway.

- Previous monitoring data (November 1988 to November 1992) indicated a significant increase in mercury levels was occurring north of the 15th Street drain and at the head of the waterway. The lack of a significant regression at these two locations when the most recent data set is included might suggest that mercury concentrations are stabilizing in these areas of the waterway.
- A comparison of bottom sediment samples collected between 1984 and 1994 suggests that mercury, HPAH, and TBT (1989 to 1994) levels in much of the waterway have not decreased during this period.
- Fourteen individual chemicals (3 metals and 11 organics) listed below, exceeded the SQOs in SPM. The greatest number of exceedances (11) were present at the head of the waterway. Mercury, HPAH, PCBs, and benzoic acid were at problem levels throughout the waterway.

Metals - mercury, nickel, and zinc

<u>Organics</u> - LPAH, HPAH, 2-methyl naphthalene, benzoic acid, benzyl alcohol, dimethyl phthalate, di-n-butyl phthalate, di-n-octyl phthalate, butyl benzyl phthalate, bis(2-ethyl hexyl)phthalate, and total PCBs

- TBT levels were elevated (100% of samples at or above the PSDDA ISL of 30 ug/kg) in SPM throughout Thea Foss Waterway, ranging from 30-790 ug/kg. The highest concentrations were measured in the upper portion of the waterway adjacent to the City Marina (station TF-1A). Recent sampling by inspectors at Ecology's Southwest Regional Office has identified high concentrations of TBT (up to 1,400,000 ug/kg) in sediments from catch basins which drain the Picks Cove Marina boat yard in the vicinity of station TF-1A.
- PCB concentrations in SPM ranged from 280 to 610 ug/kg, with a mean of 500 ug/kg. The highest levels were measured north of the 15th Street drain.
- Based on means, the highest concentrations of total organic carbon, mercury, lead, zinc, LPAH, and HPAH were measured in Thea Foss SPM compared to Hylebos and Sitcum Waterways.
- Gross sedimentation in Thea Foss Waterway between November 1992 and June 1995 averaged 1.5±0.9 g/cm<sup>2</sup>/yr.
- During the March to June 1995 collection period a three-fold increase in gross sedimentation was noted at the head of the waterway. The observed increase in this area is probably related to higher bottom sediment resuspension rates caused from bridge construction activities. In March 1995 pile driving was started for the new SR-509 bridge which crosses Thea Foss Waterway at the head.

• Resuspension estimates for bottom sediments in Thea Foss Waterway ranged from 60% (0.6±0.4 g/cm<sup>2</sup>/yr) at the mouth to 82% (1.4±0.9 g/cm<sup>2</sup>/yr) at the head of the waterway.

## Recommendations

Based on the results of information collected during this monitoring program the following recommendations are made:

- Remedial designs and natural recovery evaluations for Hylebos and Thea Foss Waterways should focus on chemicals which exceeded the SQOs in SPM and bottom sediments.
- TBT should be added to the list of chemicals of concern in Hylebos and Thea Foss Waterways based on a comparison of concentrations in SPM and bottom sediments with the Puget Sound Dredge Disposal Analysis (PSDDA) Interim Screening Level (ISL) of 30 ug/kg, dry. Under PSDDA, sediments with TBT concentrations above the ISL would need to undergo biological testing. EPA Region 10 has recently established a site specific sediment cleanup level of 19,250 ug TBT/kg OC for Hylebos Waterway based on a review of the available information concerning the toxicity of TBT to aquatic life. For consistency, EPA should consider adoption of a site wide criteria for TBT in all Commencement Bay problem areas where TBT is a concern.
- Re-evaluate potential sources of PCBs and TBTs in Hylebos and Thea Foss Waterways prior to selecting the final remedial designs for these waterways.
- Continue to include PCBs as a target analyte for SPM analysis in Thea Foss Waterway.
- Continue to monitor arsenic levels at the head of Sitcum Waterway near the north corner drain to see if source control work performed in the summer of 1995 has reduced arsenic levels in SPM in this area of the waterway.
- In-water construction activities, which are planned for heavily contaminated areas, should be designed to minimize disturbance of bottom sediments. This will reduce the potential to redistribute contaminated sediments to other areas of the waterways.
- Determine the comparability of X-Ray Fluorescence and Inductively Coupled Plasma methods for metals by analyzing split samples of SPM during the next routine collection of sediment trap samples. This information will allow trend evaluations for metals other than mercury to be performed in the waterways over a longer time series.
- Conduct seasonal analysis of <sup>210</sup>Pb levels in SPM and adjacent bottom sediments at selected stations in each waterway. This information would be used to help identify the relative importance of the various factors (plankton blooms, vessel traffic, and input of sediment from surface runoff) contributing to resuspension of bottom sediments.

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## **Project Overview**

Hylebos, Sitcum, and Thea Foss Waterways, shown in Figure 1, are three of seven waterways, which along with the Puyallup River mouth make up the tideflats portion of the Commencement Bay Nearshore/Tideflats Superfund Site. Contamination of bottom sediments in the nearshore area of Commencement Bay with a variety of chemicals was documented (Tetra Tech, 1985) during the Commencement Bay Nearshore/Tideflats Remedial Investigation (CBRI).

As a result of the remedial investigation and subsequent studies, source control programs aimed at reducing contaminant loadings to several problem areas in Commencement Bay were implemented by the Department of Ecology, City of Tacoma, and the Tacoma-Pierce County Health Department. The schedule for implementing source identification and control at the Commencement Bay site is described in Appendix C of the Commencement Bay Nearshore/Tideflats Record of Decision (EPA, 1989a).

In 1988 Ecology's Commencement Bay Urban Bay Action Team (UBAT) requested that the Toxics Investigations Section conduct a sediment trap monitoring study to track the progress of source control activities in Thea Foss Waterway. The primary objectives of this monitoring program are:

- Determine the concentrations of problem chemicals associated with settling particulate matter (SPM) entering the waterway
- Estimate present sedimentation rates in the waterway
- Periodically determine problem chemical concentrations in bottom sediments at each of the sediment trap monitoring stations

Based on results obtained in Thea Foss Waterway between November 1988 and May 1989, sediment trap monitoring was expanded to include Hylebos and Sitcum Waterways in 1990.

Problem chemicals for Hylebos, Sitcum, and Thea Foss Waterways were first identified during the CBRI. The list of problem chemicals has subsequently been clarified by EPA Region 10 after re-evaluating the data collected during the CBRI (EPA, 1992). Chemical analysis for the Sediment Trap Monitoring Program focused on the revised list of problem chemicals shown in Table 1. The reader should note that EPA currently requires that each problem chemical identified for the Commencement Bay site meets the associated sediment quality objective regardless of whether the chemical is listed for that waterway (Ngo, 1996).

The results from the Sediment Trap Monitoring Program are being used in conjunction with other available information about Commencement Bay to evaluate the effectiveness of source control





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Waterway	Hy	lebos	Sitcum	r	hea Foss	:
Location	Head	Mouth	,	Head	Mouth	Wheeler-Osgood
Conventionals						
TOC				1		2
Oil and Grease				2		
Metals						
Antimony	2					
Arsenic		1.	. 2			
Cadmium				2		2
Copper	2	3	2	2		2
Lead	2	2	2	1		2
Mercury	2	3		1	3	
Nickel	2			2		
Zinc	1	3	2		3	2
Organics						
Tetrachloroethene	2	2				· · · · · · ·
Trichloroethene		2				
Ethylbenzene	3	3	1			
Xylene	3	3				
Chlorinated Benzenes	3	3				
1,2-Dichlorobenzene		2				2
1,3-Dichlorobenzene		2		·		
1,4-Dichlorobenzene				3		
Hexachlorobenzene		2				· .
LPAH		3	3	2	2	2
НРАН	1	3	3	2	2	2
Dibenzofuran			3			
Methylpyrene	3	3				
Methylphenanthrene	3	3				
Hexachlorobutadiene		2			•	
Biphenyl		3			3	2
Aniline				3		
Phenol	2	3		3	3	2
2-Methylphenol				2		
4-Methylphenol				2	• •	2
Dibenzothiophene	3	3			3	
N-Nitrosodiphenylamine			3	3		2
Benzyl Alcohol	3	3		3		
Butyl benzyl phthalate		(		2		·
Bis(2-ethyl hexyl)phthalate	3	3		2		
An Alkylated benzene isomer	3	3	3			
A Diterpenoid hydrocarbon			3.			
A Pentachlorcyclopentane ison	ner	2				
PCBs		1			3	

Table 1: Summary of problem chemicals in Hylebos, Sitcum, and Thea Foss Waterways.

(1)= Problem chemicals identified during the Commencement Bay

Remedial Investigation (clarified in EPA, 1992)

1,2,or 3= Priority

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efforts in reducing the input of problem chemicals to the waterways, determine when source control has been achieved, model natural recovery potential for bottom sediments and aid in selecting remediation options for sediments in these waterways.

Results from sediment trap monitoring conducted by Ecology in Hylebos (July 1990 to November 1991); Sitcum (July 1990 to June 1991); and Thea Foss (November 1988 to November 1992) Waterways have been previously reported (Norton, 1990 and 1993; Norton and Barnard, 1992a,b). The present document updates SPM and bottom sediment monitoring data collected subsequent to these earlier reports. Data from the following time periods are presented in the "Results" section of this document:

Hylebos Waterway	December 1991	to	June 1995
Sitcum Waterway	June 1991	to	June 1995
Thea Foss Waterway	November 1992	to	June 1995

Monitoring data is continuous for the periods listed above with the exception of Sitcum Waterway. Dredging of bottom sediments pursuant to the Sitcum Waterway Consent Decree occurred between December 1993 and July 1994 (Gilmur, 1995). To avoid conflicts with dredging operations no sediment traps were deployed in the waterway between December 1993 and November 1994. Monitoring was resumed in Sitcum Waterway beginning in November 1994. Between November 1988 and June 1995 a total of 299 sediment traps have been deployed in Hylebos, Sitcum, and Thea Foss Waterways. Of the 299 traps deployed 232 have been successfully recovered (78%). The monitoring network, as presently configured, consists of five stations in Hylebos, three in Sitcum, and four in Thea Foss Waterway.

### Methods

#### Site Selection

Sampling locations in Hylebos, Sitcum, and Thea Foss Waterways are shown in Figures 2-4. Sampling points were selected to provide broad spatial coverage of each waterway and, to the extent possible, correspond to major contaminant sources identified during the CBRI and other source studies. Detailed descriptions of each sampling location are provided in Appendix A, Table A1. Station positions were located using a Magellan<sup>™</sup> Model 11001 Nav 1000 Plus GPS receiver, in conjunction with depth readings. In addition, station positions were noted with reference to fixed structures in the vicinity. It should be noted that some facility names are shown on tables and figures in the report to aid the reader in locating stations. This is not intended to infer that the facilities listed are the primary source of contaminants at that location. Specific linkages between sources and contaminants in SPM are noted where appropriate.

Based on data collected prior to June 1993 and the desire to reallocate resources, several modifications to the monitoring network were made beginning in July 1993. Stations H-4 (near Lincoln Avenue Drain) and H-5 (near the Cenex Plant) in Hylebos Waterway were discontinued due to relatively few chemicals being detected at problem levels. Similarly, station S-2 (north end of Sealand Terminal) in Sitcum Waterway was discontinued. Between December 1993 and November 1994 stations H-4 and H-5 in Hylebos Waterway were temporarily re-started, and a new station (TF-1A) located in the upper portion of Thea Foss Waterway adjacent to the City Marina was added. These changes were made because dredging operations were underway in Sitcum Waterway and, as a result, no monitoring was occurring in the waterway. Finally, in November 1994 the network was returned to its July 1993 configuration since dredging operations had been completed in Sitcum Waterway. The deployment schedule for sediment traps in Commencement Bay between June 1990 and June 1995 is included in Appendix A, Table A2.

Besides the changes made in spatial coverage of the waterways, the number of target analytes in SPM was also reduced beginning in July 1993. The reader is referred to the data tables in Appendix C to determine target analytes during the various monitoring periods.

### **Sample Collection**

#### **Settling Particulate Matter**

SPM was collected at each of the monitoring stations with the use of moored sediment traps, positioned 3 feet above the bottom. The sediment traps were sampled at a three month frequency. Total depths at mean lower low water (MLLW), in the areas sampled ranged as follows: Hylebos 24 to 31 ft., Sitcum 31 to 54 ft., and Thea Foss 18 to 36 ft.



Figure 2: Monitoring Stations in Hylebos Waterway, December 1991 to June 1995.

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Figure 3: Monitoring Stations in Sitcum Waterway, June 1991 to June 1995.



Figure 4: Monitoring Stations in Thea Foss Waterway, November 1992 to June 1995.

The traps used in the present study are straight-sided glass cylinders with a collection area of  $78.5 \text{ cm}^2$  and a height-to-width ratio of five. A schematic of the construction details of the traps and their moorings is presented in Figure 5.

At deployment, the traps were filled with two liters of high salinity water (4% NaCl), which contained sodium azide (2%Na<sub>3</sub>N) as a preservative to reduce microbial degradation of the samples. Prior to deployment, the collection cylinders were cleaned with sequential washes of hot water/Liquinox detergent, 10% nitric acid, distilled/deionized water, and pesticide-grade acetone. They were then air-dried and wrapped in aluminum foil until used in the field.

Upon retrieval of the traps, overlying water in the collection cylinders was removed with a peristaltic pump. The remaining contents were then transferred to half-gallon priority pollutant-cleaned glass jars with teflon-lined lids (supplied by I-Chem, Hayward, CA) and stored at 4EC for transport to the laboratory. At the laboratory all samples were frozen within 12 hours of collection pending processing. Processing consisted of first thawing the samples and then centrifuging to isolate the particulate fraction. All nekton >2 cm in size were removed from the samples prior to determining sample weights and conducting physical/chemical analyses. To obtain sufficient volume for organics analyses, samples from two consecutive collection periods (*i.e.*, semi-annually) were composited. All other analyses were conducted on a quarterly basis. Manipulation of all SPM samples in the laboratory was accomplished with stainless steel spoonulas, cleaned as previously described for the collection cylinders.

#### **Bottom Sediment**

Sampling procedures followed Puget Sound Protocols where applicable (Tetra Tech, 1986a). Three individual grabs of surface sediments (top 2 cm) were collected and composited at each of the sediment trap locations in November 1994 using a 0.1 m<sup>2</sup> stainless steel van Veen grab. For comparability, sampling locations corresponded to previous sites occupied in December 1989 and January 1991.

After retrieving the grab, the top 2 cm layer not in contact with the sidewalls of the sampler was transferred to a stainless steel beaker and homogenized by stirring with a stainless steel spoon. Aliquots for individual analyses were taken from this homogenate and placed in priority-pollutant cleaned glass jars with teflon-lined lids supplied by I-Chem Hayward, CA, wrapped in polyethylene bags, and stored at 4°C for transport to the laboratory. An exception to this procedure was volatiles samples which were collected directly from the grab into appropriate sample containers. Spoons and beakers were pre-cleaned as previously described for the sediment trap collection cylinders.



Figure 5: Schematic of Commencement Bay Sediment Traps and Moorings.

#### Sample Analysis and Quality Assurance

All physical/chemical analyses of samples for this project were conducted using procedures specified in the Puget Sound Protocols (Tetra Tech, 1986a) as amended and updated. In addition, the type and frequency of laboratory quality assurance (QA) samples at a minimum followed those specified in the Ecology/EPA Manchester Laboratory QA manual (Ecology, 1988). A summary of the analytical laboratories and methods used for this project is shown in Table 2.

Quality of the data sets were evaluated with the use of the following sample types: duplicates, matrix spikes, internal standards, surrogate spikes, standard reference materials and method blanks. A summary of QA samples and their frequency of analysis is included in Appendix B, Table B1. Staff at the Ecology/EPA Manchester Laboratory performed detailed QA reviews of each data package. Copies of the case narratives for each data set are included in Appendix B. The reader is referred to the case narratives for specific details concerning data quality.

Overall, few major analytical problems were encountered in the analysis of samples for this project. Notable exceptions to this statement are discussed below. Consequently, the data generated are considered acceptable for use as qualified in the data tables and noted in the case narratives.

Several results for copper, cadmium, and mercury are qualified as estimates due to high results (+20%) obtained on the analysis of certified reference materials. Results qualified for this reason would be expected to slightly overestimate actual environmental levels.

Between November 1988 and May 1994 arsenic, copper, nickel, lead, and zinc determinations were made using X-Ray Fluorescence (XRF) techniques. Beginning in May 1994 the above mentioned metals were analyzed using Inductively Coupled Plasma (ICP) methods. No data are available to evaluate how these two methods compare; however, it is anticipated that XRF would yield higher results since it is a more rigorous analysis. Consequently, data generated using XRF and ICP methods have been separated for temporal analyses in this report. The reader is cautioned about drawing strong conclusions based on concentration differences for elements which were generated using different analytical methods.

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The principal analytical problem encountered in the organics analyses was directly related to chromatographic interferences. When encountered, these interferences were typically caused by the presence of high levels of fats and lipids in the samples. In addition, high native concentrations of PCBs and butyltins in several instances reduced the usefulness of some of the matrix spike information. Both these situations resulted in several compounds being qualified as estimates and produced variable detection limits between sampling periods.

Unless otherwise noted all concentrations in this document are reported on a <u>dry weight</u> basis. Butyltins were determined as butyltin chloride. A summary of the results of physical/chemical

Table 2: Summary of	Table 2: Summary of Analytical Methods for Commer	ncement Bay Sediment Trap	Commencement Bay Sediment Trap Monitoring Project, June 1991 to June 1995.
Analysis	Method	Reference	Laboratory
Conventionals Percent Solids	Dry @ 104°C	Tetra Tech, 1986a	Ecology/EPA-Manchester, WA.
Total Organic Carbon	Combustion/CO2 Measurement	Tetra Tech, 1986a	Analytical Resources, IncSeattle, WA. Laucks Laboratories- Seattle, WA. AMTEST-Redmond, WA. Sound Analytical Services, IncTacoma, WA. Weyerhaeuser Technology Center, Tacoma, WA.
Grain Size	Seive and Pipet	Tetra Tech, 1986a	Soil Technology, IncWinslow, WA.
<u>Metals (6/91-5/94)</u> Arsenic,Copper, Nickel.Lead,Zinc	<u>-5/94)</u> pper, XRF A.Zinc	Nielson and Sanders, 1983	Nielson and Sanders, 1983 Battelle Northwest-Richland, WA.
Antimony, Cadmium Mercury	GFAA CVAA	Tetra Tech, 1986a Tetra Tech, 1986a	Battelle Northwest-Sequim, WA. Battelle Northwest-Sequim, WA.
<u>Metals (5/94–6/95)</u> Arsenic,Copper, I and Tinc	ICP	Tetra Tech, 1986a	Ecology/EPA-Manchester, WA.
Mercury	CVAA	Tetra Tech, 1986a	Leau, Luic Mercury CVAA Tetra Tech, 1986a Ecology/EPA-Manchester, WA.
<u>Organics</u> Volatiles	GC/MS #8240	EPA, 1986	Ecology/EPA-Manchester, WA.
Semivolatiles	GC/MS #8270	EPA, 1986	Ecology/EPA-Manchester, WA.
PCBs	GC/ECD #8080	EPA, 1986	Ecology/EPA-Manchester, WA.
Butyltins	GC/MS-SIM	Tetra Tech, 1986a	Ecology/EPA-Manchester, WA.
<u>Metals</u> XRF= X-Ray Flourescence GFAA= Graphite Furnace Atomic Absorption ICP= Inductively Coupled Plasma CVAA= Cold Vapor Atomic Absorption	ce e Atomic Absorption i Plasma nic Absorption	Organics GC/MS= Gas Chromatography/Mass Spectroscopy GC/ECD= Gas Chromatography/Electron Capture Detector GC/MS-SIM= Gas Chromatography/Mass Spectroscopy-Se	<u>Drganics</u> GC/MS= Gas Chromatography/Mass Spectroscopy GC/ECD= Gas Chromatography/Electron Capture Detector GC/MS-SIM= Gas Chromatography/Mass Spectroscopy-Selective Ion Monitoring

analyses of SPM and bottom sediments from each of the waterways monitored is included in Appendix C and D, respectively.

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## Results

### Hylebos Waterway

#### Chemicals in SPM

Between December 1991 and June 1995 89 sediment traps were deployed in Hylebos Waterway. Of these traps 69 (78%) were successfully recovered.

The results of conventionals and metals analysis of SPM from Hylebos Waterway are tabulated in Appendix C, Table C1. Total organic carbon (TOC) levels ranged from 1.9% to 8.5% in Hylebos SPM with a mean of 4.1%. Based on means, TOC levels at the head of the waterway were elevated by a factor of 3 compared to concentrations at the mouth.

Concentrations (mg/kg) of selected metals associated with SPM from Hylebos Waterway between December 1991 and June 1995 are:

				Detection		
<u>Metal</u>	Range	Mean	<u>C.V.</u>	Frequency	<u>N</u>	
Arsenic	9.6-130	48	57%	100%	77	
Copper	62-180	130	23%	100%	77	
Lead	28-250	81	40%	100%	77	
Mercury	0.16-0.93	0.41	39%	100%	77	
Zinc	66-520	200	38%	100%	77	
C.V.= Coefficient of Variation (SD/Mean) * 100						

N= Number of Samples

Metals concentrations were fairly low in Hylebos SPM over the current monitoring period with the exception of arsenic and mercury in the upper portion of the waterway. As shown in Figure 6, arsenic and mercury levels within one mile from the head were somewhat elevated compared to the remainder of the waterway. The highest concentrations of zinc and copper were also measured in this section of the waterway. Longitudinally, concentrations of most metals in Hylebos SPM tended to peak within one mile of the head of the waterway and decrease moving towards the mouth.

In contrast, lead concentrations were fairly consistent throughout the waterway with a primary peak occurring at station H-6 approximately one-half mile from the mouth of the waterway. The lowest concentrations for all metals were typically present at the mouth of the waterway. Temporally, intra-station metals concentrations tended to be similar, being within a factor of 2-3 in most cases.



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Several volatile organic compounds (VOCs) were identified as problem chemicals in the outer portion of Hylebos Waterway (Tetra Tech, 1985). Consequently, VOC analysis was only conducted at stations H-5, H-6, and H-7, which are located in the outer portion of the waterway. The results of volatile organics analyses of Hylebos Waterway SPM are tabulated in Appendix C, Table C2. Concentrations (ug/kg) of selected volatiles are:

Detection

				Detection	
Compound	Range	<u>Mean</u>	<u>C.V.</u>	Frequency	N
Trichloroethene	0.8-23	4.5	150%	71%	12
Tetrachloroethene	0.53-33	7.7	150%	94%	16
Ethylbenzene	0.4-4.5	0.75	65%	53%	9
Total Xylenes	1.2-16	5.5	96%	65%	11
2-Butanone	22-230	100	63%	65%	11
Isopropyltoluene	4.9-300	98	85%	94%	16

C.V.= Coefficient of Variation (SD/Mean) \* 100

N= Number of Samples with detected concentrations

Thirty VOCs were detected in SPM. Acetone was detected in the highest concentrations, which ranged up to 400 ug/kg. The presence of acetone is in all probability related to the fact that it was used to clean the sediment trap collection cylinders prior to deployment. Levels of most VOCs in SPM were typically under 30 ug/kg. Two solvents, isopropyltoluene and 2-butanone (methyl ethyl ketone), were measured at levels exceeding 100 ug/kg. Isopropyltoluene was consistently detected at all three stations at concentrations ranging from 4.9 to 300 ug/kg. 2-Butanone was detected in 65% of the samples analyzed. The highest concentrations of isopropyltoluene were usually present at station H-5, while the highest levels of 2-butanone occurred at H-6.

Tetrachloroethene was consistently detected (3.4-35 ug/kg) at station H-6 located adjacent to Occidental Chemical. Between 1947 and 1973 tetrachloroethene was produced in commercial quantities at Occidental's solvent plant (Tetra Tech, 1986b).

In addition to the VOCs, 37 target semivolatile organics and PCBs were detected in Hylebos SPM. A complete list of detected compounds is included in Appendix C, Table C3. A summary of detected concentrations of selected organics (ug/kg) is shown below:

		12		Detection		
Compound	<u>Range</u>	<u>Mean</u>	<u>C.V.</u>	Frequency	$\underline{\mathbf{N}}$	
HPAH	830-22000	10000	49%	100%	41	
Hexachlorobenzene	40-130	72	46%	20%	8	
Hexachlorobutadiene	18-200	76	73%	44%	18	
Bis(2EH)phthalate	810-44000	8400	190%	17%	7	
Total PCBs	230-73000	3000	400%	90%	37	
Benzoic Acid	780-5400	2000	59%	29%	12	
C.V.= Coefficient of Variation (SD/Mean) * 100						

N= Number of Samples with detected concentrations

Concentrations of most organics in Hylebos SPM tended to be quite variable (c.v.= 46-400%) over the monitoring period. Polynuclear aromatic hydrocarbons (PAH) were the predominate group of organic compounds quantified in Hylebos SPM. The distribution of high molecular weight PAH (HPAH) is shown in Figure 7. The highest concentrations of HPAH were measured at station H-3 in the upper portion of the waterway. The lowest concentrations were typically measured at the mouth of the waterway.

At all stations, the sum of HPAH exceeded the sum of low molecular weight PAH (LPAH). This enrichment of HPAH in SPM is expected since weathering processes such as evaporation, photochemical oxidation, dissolution, and microbial degradation can preferentially remove PAHs with molecular weights less than that of fluoranthene (Merill and Wade, 1985). The apparent enrichment of HPAHs relative to LPAHs would suggest that historical sources of PAHs have played an important role in the PAH contamination observed in Hylebos SPM.

Hexachlorobenzene, hexachlorobutadiene, and 1,2,4-trichlorobenzene -- which have all been identified as problem chemicals in Hylebos -- were primarily detected at station H-6, located adjacent to Occidental Chemical. Occidental Chemical was identified as a source of hexachlorobenzene and hexachlorobutadiene during the CBRI (Tetra Tech, 1985). Maximum concentrations of Bis(2-ethyl hexyl)phthalate (44,000 ug/kg) occurred in the central portion of the waterway at station H-5.

Several additional compounds, not listed as problem chemicals were also quantified in Hylebos SPM. Benzoic acid was identified throughout the waterway at concentrations ranging from 780-5,400 ug/kg. 4-Methyl phenol concentrations ranged from 49-11,000 ug/kg. The remaining semivolatile organics were typically present at <1,000 ug/kg.

The distribution of total PCBs in Hylebos SPM is also shown in Figure 7. PCBs peaked at stations H-2 and H-3 in the upper portion of the waterway and decreased moving toward either the head or mouth. Total PCB concentrations at stations H-2 and H-3 were significantly higher than levels measured at both the head and mouth of the waterway (Tukey Post Hoc, P<0.001). A secondary peak for PCBs was also observed near station H-6 in the outer portion of the waterway. The current pattern of PCB contamination in Hylebos Waterway is similar to that observed during earlier monitoring, July 1990 to December 1991 (Norton and Barnard, 1992b).

One extremely high value for PCB-1242 of 73,000 ug/kg was measured between December 1991 and May 1992, at station H-2, which is located adjacent to General Metals. Aside from the high concentration, the unusual aspect of this result is that PCB-1242 has been infrequently detected in Hylebos Waterway. One noteworthy exception was a screening study of storm drains which discharge to the waterway (Stinson et al., 1987). In this study PCB-1242 was detected in several catch basin sediments at General Metals. Since this sampling was performed in 1986 a stormwater collection system has been installed at General Metals. Given the unusual type of PCB detected and the fact that it has not been detected at this location (H-2) in three years of subsequent monitoring, this result does not appear to indicate an ongoing problem.



Figure 7: Total PCB and HPAH concentrations in Hylebos Waterway settling particulate matter, December 1991 to June 1995.

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Tributyltin (TBT) has been used extensively in the past as an active ingredient in antifouling paints for private and commercial vessels. Due to concerns over the lack of information on TBT levels in Hylebos Waterway, butyltins were added to the list of target chemicals for the sediment trap program in May 1994. The results of butyltin analysis of SPM are summarized in Appendix C, Table C4.

TBT was detected in 100% of the samples analyzed at concentrations ranging from 55-1,300 ug/kg (mean= 450 ug/kg). Two breakdown products of TBT, monobutyltin (MBT) and dibutyltin (DBT), were also detected extensively throughout the waterway with the following exceptions. MBT and DBT were not detected at the mouth of the waterway. MBT was also not detected in the upper turning basin.

#### Chemicals in Bottom Sediment

The results of conventionals and metals analysis of bottom sediments collected in January 1991 and November 1994 at each of the sediment trap monitoring stations in Hylebos Waterway are listed in Appendix D, Table D1. TOC levels ranged from 1.7 to 12%, with the highest concentrations occurring at H-5. The lowest values were present at the mouth of the waterway. Grain size analysis indicated that most of the sediments sampled consistent of primarily silt and clay size particles. Sediments from H-4 (near Lincoln drain) and the mouth of the waterway tended to have a higher sand content.

Metals concentration were fairly low and consistent throughout the waterway. As was the case for SPM, the lowest concentrations of all metals were typically measured at the mouth. Unlike SPM, strong longitudinal gradients were not observed in bottom sediments for most metals, with the possible exception of arsenic and zinc. These two metals tended to be higher at the head and decreased moving toward the mouth.

Forty-six organic compounds were detected in bottom sediments from Hylebos Waterway. A complete list of organics detected in bottom sediments is included in Appendix D, Table D2. Volatiles were infrequently detected in bottom sediments. When present, concentrations were typically  $\leq 2 \text{ ug/kg}$ , dry. An exception was isopropyltoluene which was measured at 58 ug/kg in the central part of the waterway (station H-5).

PAHs were the predominant group of semivolatile organics detected in SPM. HPAH concentrations varied by an order of magnitude in Hylebos Waterway ranging from 1,800 to 18,000 ug/kg. The lowest HPAH levels were consistently measured at the mouth of the waterway. Hexachlorobenzene and hexachlorobutadiene were consistently detected adjacent to Occidental Chemical (station H-6). Most semivolatile organics were present at <1,000 ug/kg with a few exceptions. Benzoic acid was detected sporadically in the waterway at levels ranging from 200 to 1,700 ug/kg. Bis(2-ethyl hexyl)phthalate (1,500 ug/kg) peaked at H-2 in the upper portion of the waterway, while a single high value for retene (8,900 ug/kg) was measured in the central portion of the Hylebos. Retene is a naturally occurring resin-acid derived compound (Prahl and

Carpenter, 1984), that is often found in sediments in association with wood waste debris. One non-target compound, coprostanol, was present throughout the waterway during the November 1994 survey at concentrations between 690-2,300 ug/kg. Coprostanol is produced in the intestine of mammals by the microbial reduction of cholesterol which is the main steroid in the tissues of vertebrates (Verschueren, 1983).

For perspective, concentrations of total PCBs in Hylebos bottom sediments collected in 1984, 1991, and 1994 are compared in Figure 8. Total PCB levels in the 1994 survey ranged from 510 to 3,000 ug/kg. Peak concentrations were present near ELF-ATO Chem (station H-3) in the upper portion of the waterway. Examination of Figure 8 indicates that total PCB levels are higher in both 1991 and 1994 than those measured in 1984 at approximately similar areas in the waterway. In addition, in five of seven cases concentrations increase with each successive year of sampling. These data suggest that PCB concentrations have not gone down in bottom sediments despite the passage of ten years and in fact may be increasing. It should be noted that factors such as spatial and analytical differences between the present study and the work conducted for the remedial investigation in 1984 may be contributing to the apparent trend in PCBs. However, the samples collected in 1991 and 1994 were from similar locations and employed identical analytical techniques.

Butyltins were detected in bottom sediments throughout Hylebos Waterway. The highest concentrations of TBT (820 ug/kg), MBT (35 ug/kg), and DBT (380 ug/kg) were detected in the central portion of the waterway (H-4). As was the case for most other contaminants the lowest butyltin levels were measured at the mouth of the waterway. A summary of butyltin concentrations in Hylebos bottom sediments can be found in Appendix D, Table D3.

# Comparison to Commencement Bay Sediment Quality Objectives (SQOs)

Of particular concern to source control and sediment cleanup activities in the Commencement Bay Nearshore/Tideflats site are chemicals which exceeded the SQOs. SQOs based on environmental risk (EPA, 1989) are contaminant concentrations above which deleterious effects would always be observed in marine benthic communities. EPA, the lead agency for sediment remediation activities in Commencement Bay, is using the SQOs based on environmental risk (bioassay and benthic community data) as cleanup objectives for contaminated sediments. For PCBs, the cleanup objective is based on the SQO for the protection of human health. A list of SQOs for chemicals detected in SPM during the Sediment Trap Monitoring Program is presented in Table 3. For completeness, SQOs for individual PAHs are also shown in Table 3. However, to simplify presentation of the data only comparisons with the SQOs for LPAH and HPAH are discussed in the report.

A summary of chemicals exceeding the SQOs in Hylebos SPM and bottom sediments are shown in Table 4. A total of 14 individual chemicals (3 metals, 11 organics) were measured at levels above the SQOs in Hylebos SPM. The greatest number of exceedances (9) were measured at the



Figure 8: Comparison of Total PCB concentrations in Bottom Sediments from Hylebos Waterway in 1984, 1991, and 1994.

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	Sediment
Chemical	Cleanup Objective+
Metals (mg/kg, dry weight)	
Antimony	150
Arsenic	57
Cadmium	5.1
Copper	390
Lead	450
Mercury	0.59
Nickel	>140
Zinc	410
Organic Compounds (ug/kg, dry weight) <u>Polynuclear Aromatic Hydrocarbons</u>	
LPAH	5200
Napthalene	2100
Acenapthalene	1300
Acenapthene	500
Fluorene	540
Phenanthrene	1500
Anthrene	960
2-Methylnapthalene	670
HPAH	17000
Fluoranthene	2500
Pyrene	3300
Benzo(a)anthracene	1600
Chrysene	2800
Benzofluoranthenes	3600
Benzo(a)pyrene	1600
Indeno(1,2,3-cd)pyrene	690
Indeno(1,2,3-cd)pyrene Dibenzo(a,h)anthracene	230 720

Table 3: Commencement Bay Sediment Quality Values Based on Environmental Risk for Compounds Detected in Hylebos, Sitcum, and Thea Foss Waterway Settling Particulate Matter.

+= Commencement Bay Nearshore/Tideflats, Record of Decision (EPA, 1989)

\*=Sediment Quality Objective based on human health

	Sediment
Chemical	Cleanup Objective+
Organic Compounds (ug/kg, dry)	
Chlorinated Organics	
1,4 Dichlorobenzene	110
Hexachlorobenzene	22
Total PCBs	1000
Total PCBs	150*
Phthalates	
Dimethylphthalate	160
Diethylphthalate	200
Di-n-butyl phthalate	1400
Butyl benzyl phthalate	900
Bis(2EH)phthalate	1300
Di-n-octyl phthalate	6200
Phenols	、
Phenol	420
4-Methyl phenol	670
Pentachlorophenol	360
Misc. Extractables	
Benzyl alcohol	73
Benzoic acid	650
Dibenzofuran	540
Hexachlorobutadiene	11
N-nitrosodiphenylamine	28
Volatile Organics	
Tetrachloroethene	57
Ethylbenzene	10
Total xylenes	40

Table 3 (cont.): Commencement Bay Sediment Quality Values Based on Environmental Risk for Compounds Detected in Hylebos, Sitcum, and Thea Foss Waterway Settling Particulate Matter.

+= Commencement Bay Nearshore/Tideflats, Record of Decision (EPA, 1989)

\*=Sediment Quality Objective based on human health

Table 4: Summary of metals and organic compounds exceeding the Commencement Bay sediment quality objectives (SQO) in settling particulate matter and bottom sediments from Hylebos Waterway, December 1991 to June 1995.	nd organic co nd bottom sed	mpounds exceed liments from Hy	ling the Comme lebos Waterway	sncement Bay sedimory, December 1991 tc	ent quality obj June 1995.	lectives (SQO)	
Station	H-1	H-2	H-3	H-4	. H-5	H-6	H-7
I	-	@ General	@ Elf	Near	Near	@ Occidental	
Location	Head	Metals	Atochem	Lincoln Drain	Cenex	Chemical <sup>*</sup>	Mouth
Metals							r
Arsenic	$64^{*}_{11}$	62* <sub>13</sub>	62 <sub>13</sub>	28 8			
Mercury	و 11	18 <sub>11</sub>	56 9				
Nickel	bs					bs	
Zinc	9 п	bs					
Organics							
LPAH						33 6	
HPAH	, 17 6	. 29 7	57 7				
Dibenzofuran						17. 6	
4-Methyl phenol	17 6	14 7	. 14 7	50 4	50 4	33 <sub>6</sub>	
Benzoic acid	33* <sub>6</sub>	60 5	33 3	50 2		100 2	60 5
Diethyl phthalate	bs						
Di-n-butyl phthalate		14 7	bs				
Dimethyl phthalate	20 5						
Bis(2-ethyl hexyl)phthalate	$100_{-1}$	$100^{*}$ 1			33 3		33 <sub>6</sub>
N-Nitrosophenylamine		bs					
Hexachlorobenzene			$100^{*}_{1}$		100 2	$100^{*}_{4}$	$100^{*}_{3}$
Hexachlorobutadiene			$100^{*}_{2}$		bs	$100^{*} _{6}$	$100^{*}$ 6
PCBs+	$100^{*}_{6}$	$100^*$ $_7$	$100^{*}$ 7	100* 4	$100^{*}_{4}$	100* 5	$100^{*}$ 7
Values shown are percent of total SPM samples	otal SPM sam		lat exceeded the	collected that exceeded the sediment quality objectives	jectives		
Subscript= Number of samples							
	:						

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\*=SQO also exceeded in bottom sediment

+= SQO for PCBs based on protection of human health bs= Chemical only exceeded SQO in bottom sediments

head of the waterway in the upper turning basin. Two stations (H-4 and H-5) in the central portion of the waterway had the fewest number of exceedances. PCBs were the only chemical that exceeded the SQO in 100% of the samples tested.

TBT is the most toxic of the butyltins. While no SQO has been established for TBT, the Puget Sound Dredge Disposal Analysis (PSDDA) has adopted a Interim Screening Level (ISL) of 30 ug/kg, dry (PSDDA, 1989). Sediments with TBT concentrations above the ISL are required to undergo biological toxicity testing. TBT concentrations in Hylebos SPM ranged from 55-660 ug/kg, with a mean of 450 ug/kg.

At the request of the Hylebos Cleanup Committee, EPA Region 10 has evaluated an appropriate sediment cleanup levels for TBT in Hylebos Waterway. Based on a review of available literature concerning the toxicity of TBT to aquatic organisms, EPA has selected a concentration of 19,250 ug TBT/kg organic carbon (OC) as a site specific sediment cleanup level for Hylebos Waterway. It is anticipated that this level would protect many aquatic organisms from most acute and some chronic effects (EPA, 1996). On an OC normalized basis, TBT levels in Hylebos Waterway ranged from 2,800 to 26,000, with a mean of 11,000 ug TBT/kg OC.

Compared to SPM, fewer chemicals (12) exceeded the SQOs in bottom sediments. Again, PCBs were above the SQO established for the protection of human health at all stations. The highest number of exceedances were present in the upper portion of the waterway, within one mile from the head. As was the case for SPM, 100% of the bottom sediments analyzed (150-820 ug/kg) exceeded the PSDDA ISL of 30 ug/kg for TBT. Only one of seven bottom sediment concentrations exceeded the Hylebos Waterway cleanup level for TBT.

#### **Gross Sedimentation Rates**

A complete list of sediment accumulation rates for Hylebos Waterway determined from sediment trap data is included in Appendix C, Table C5. Two types of accumulation rates have been calculated: gross accumulation (g/cm<sup>2</sup>/yr) which is the measured sediment flux into the traps, and accumulation rate (cm/yr) which is calculated to represent the actual thickness of new sediments once the particulates have consolidated on the bottom. Both these values should be viewed as estimates of gross sedimentation (i.e., net sedimentation + resuspension) in the waterway. Calculations used to generate the reported sedimentation rates are shown below:

• Gross Accumulation  $(g/cm^2/yr) = [(P/A)/D] \times Y$ 

P= Amount of material collected (dry grams)

A= Collection area of cylinder  $(cm^2)$ 

D= Number of days sediment trap was deployed

Y= Number of days in a year (365)

• Accumulation Rate (cm/yr) = Mass accumulation  $(g/cm^2/yr)/Dry$  density  $(g/cm^3)$ 

Dry density= [Wet density x (Bottom Sediment % solids/100)] Wet density= Estimated from Puget Sound Density Model using % solids data from in-situ bottom sediments (Crecelius, 1989)

Gross accumulation rates (dry weight) for Hylebos Waterway between December 1991 and June 1995 are shown in Figure 9. Gross rates ranged from 0.5-5.5 g/cm<sup>2</sup>/yr, with a mean of  $2.4\pm1.0 \text{ g/cm}^2$ /yr. The highest sedimentation rates in the waterway typically occurred in the central portion of the waterway at station H-5 (mean=  $3.4\pm1.2 \text{ g/cm}^2$ /yr), and at the mouth (mean=  $3.1\pm1.2 \text{ g/cm}^2$ /yr). Significantly lower rates (ANOVA, P<0.05) were measured at stations H-1, H-2, and H-6, compared to the two locations (H-5 and H-7) with the highest rates. Temporally, the data collected suggest that sedimentation in most of the waterway tends to be highest during the late summer and fall, and lowest during the winter and spring. Predicted accumulation rates ranged from 0.6-7.4 cm/yr with a mean of  $3.4\pm1.3 \text{ cm/yr}$ .



Figure 9: Gross Sedimentation Rates for Hylebos Waterway, December 1991 to June 1995.

## Sitcum Waterway

#### Chemicals in SPM

Between June 1991 and June 1995 44 sediment traps were deployed in Sitcum Waterway. Of these traps 39 (89%) were successfully recovered. Sediment trap monitoring in Sitcum was temporarily interrupted for a period of 11 months between December 1993 and November 1994 while dredging operations were underway.

The results of conventionals and metals analysis of Sitcum Waterway SPM are summarized in Appendix C, Table C6. TOC levels were fairly low and consistent in Sitcum ranging from 1.2 to 3.0% (mean= 2.0%).

Concentrations of selected metals (mg/kg) associated with SPM from Sitcum Waterway between June 1991 and June 1995 are summarized below. Data collected before and after May 1994 have been separated in this summary due to a shift in analytical methods (see sample analysis and quality assurance section). In addition the two data sets represent pre- and post-dredging conditions in the waterway.

June 1991 to December 1993 (Pre-dredging)

				Detection	
<u>Metal</u>	Range	Mean	<u>C.V.</u>	<b>Frequency</b>	$\underline{\mathbf{N}}$
Arsenic	11-83	37	51%	100%	33
Copper	64-180	120	30%	100%	33
Lead	44-320	170	49%	100%	33
Zinc	91-450	220	41%	100%	33

November 1994 to June 1995 (Post-dredging)

				Detection	
<u>Metal</u>	<u>Range</u>	<u>Mean</u>	<u>C.V.</u>	<b>Frequency</b>	$\underline{\mathbf{N}}$
Arsenic	8.1-24	17	34%	100%	4
Copper	41-99	.75	29%	100%	4
Lead	21-160	110	49%	100%	4
Zinc	50-190	120	41%	100%	4
C.V.=Coe	fficient of Varia	ation (SD/Mea	n)		

N= Number of Samples

Arsenic concentrations, shown in Figure 10, illustrate the characteristic distribution of all metals in Sitcum SPM. Peak concentrations for all metals were measured at the head of the waterway near the north corner drain. A secondary peak was also present along Terminal 7, at Berth B. The lowest concentrations for all metals were measured at the mouth of the waterway.

Datastian





Substantially lower metals concentrations were noted at all monitoring stations after dredging was completed in December 1993. A portion of the observed decrease in metals concentrations is undoubtedly the result of sediment cleanup activities in the waterway. Unfortunately, due to a change in analytical methods for metals (see sample analysis and quality assurance section), which occurred in December 1993, it is difficult to evaluate the actual decrease in metals concentrations in the waterway. Sample splits planned for XRF and ICP analysis during the next routine collection (December 1995) should help in evaluating the comparability of results from these two methods.

Thirty-two organic compounds, primarily PAHs were quantified in SPM. A complete list of semivolatile organics in Sitcum SPM is included in Appendix C, Table C7. Detected concentration (ug/kg) of selected organics in Sitcum SPM are shown below. For comparison pre- and post-dredging data have been separated.

June 1991 to December 1993 (Pre-dredging)

	· •			Detection	
Compound	Range	Mean	<u>C.V.</u>	Frequency	$\underline{\mathbf{N}}$
LPAH	480-5400	1900	76%	100%	19
HPAH	1800-12000	5400	56%	100%	19
Dibenzofuran	28-460	180	73%	100%	19
Bis(2EH)phthalate	34000-170000	110000	52%	16%	3
Benzoic Acid	1400-5500	3200	43%	32%	6

November 1994 to June 1995 (Post-dredging)

			Detection	
Range	Mean	<u>C.V.</u>	Frequency	N
77-2300	1300	71%	100%	3
160-4100	2700	67%	100%	3
120-240	180	33%	67%	2
ND	-	-	2 % #	-
970			33%	1
Variation (SD/Mean)	)			
les				
	77-2300 160-4100 120-240 ND 970	77-2300   1300     160-4100   2700     120-240   180     ND   -     970   -     Variation (SD/Mean)   -	77-2300 1300 71%   160-4100 2700 67%   120-240 180 33%   ND - -   970 - -   Variation (SD/Mean) - -	RangeMeanC.V.Frequency77-2300130071%100%160-4100270067%100%120-24018033%67%ND97033%Variation (SD/Mean)

ND= Not detected

Concentrations of most organics were more variable than metals in Sitcum SPM. PAHs were present in 100% of the samples analyzed from Sitcum Waterway. The distribution of LPAH and HPAH in Sitcum is shown in Figure 11. Based on means, LPAH concentrations were highest at Berth B along terminal 7, while maximum HPAH concentrations occurred near the north corner drain at the head of the waterway. For both LPAH and HPAH the lowest levels were present at the mouth of the waterway. Prior to dredging, Dibenzofuran was detected in 100% of the samples analyzed from Sitcum Waterway at concentrations ranging from 28-460 ug/kg (mean= 180 ug/kg). Chemically, dibenzofuran is closely related to the PAHs. Phthalate acid esters were



Figure 11: LPAH and HPAH Concentrations in Sitcum Settling Particulate Matter, June 1991 and June 1995.

primarily detected near the north corner drain. Retene was present at low levels (150-670 ug/kg) throughout the waterway.

Substantial reductions were noted for most organic compounds associated with SPM after dredging was completed in Sitcum. This was especially true at the mouth of the waterway.

#### Chemicals in Bottom Sediment

Results of conventionals and metals analysis of bottom sediments from Sitcum Waterway, collected in January 1991 and November 1994, are listed in Appendix D, Table D4. TOC values in bottom sediments were quite low and consistent at all stations in the waterway ranging from 1.1 to 2.0%. Grain size analysis indicated that the bottom sediments collected consisted primarily of silt and clay size particles.

Metals concentrations (mg/kg) in bottom sediments were as follows; arsenic= 7-28; copper= 55-160; lead= 19-290; and zinc= 69-470. The highest concentrations for lead and zinc were measured at Berth B, along Terminal 7. The lowest levels for all metals were consistently measured at the mouth of the waterway. As was noted earlier, due to a change in analytical methodologies for metals it is difficult to evaluate actual changes in metals concentrations resulting from dredging in the waterway.

Concentrations of semivolatile organics in Sitcum bottom sediments (Appendix D, Table D5) were <1000 ug/kg with the exception of PAHs. LPAH concentrations in the waterway ranged from 120-5,500 ug/kg. HPAH levels ranged from 950-10,000 ug/kg.

Reductions in contaminant levels were not as apparent in bottom sediments as those observed in SPM. Very little change was noted in contaminant levels at the head and mouth of the waterway between 1991 and 1994. A possible explanation for this finding could be that the areas sampled at the head and mouth of the waterway are just outside the limit of dredging. In contrast, a substantial decrease in PAH levels was noted at Berth B, along Terminal 7. Comparing pre- and post-dredge values, LPAH and HPAH concentrations were lower by 98% and 83%, respectively. In addition, dibenzofuran was not detected at this location in 1994. While this conclusion is based on a limited amount of data it does seem to strongly suggest that contaminant levels at Berth B, along Terminal 7, are lower then those measured prior to dredging. Post-dredging monitoring which is being conducted by the Port of Tacoma will be useful in evaluating changes in bottom sediment conditions.

#### Comparison to Commencement Bay SQOs

In Sitcum Waterway a total of 7 chemicals (1 metal and 6 organics) exceeded the SQOs in SPM (Table 5). The greatest number of exceedances (4) were measured at the head of the waterway near the north corner drain. Arsenic was above the SQO in 55% of the samples tested at this location. Benzoic acid was present above the SQOs at three of the four sites in Sitcum. After

Station	S-1	S-2	S-3	S-4
	Head near	North end	Terminal 7 @	Terminal 7 @
Location	North corner Drain	Sealand Terminal	Berth B	Mouth
Metals				
Arsenic	55 11			
Zinc			bs	
Organics				
LPAH			20* 6	
Dibenzofuran			ps	
Benzoic acid	50 4	33 3		25 4
Benzyl alcohol				100 1
Pentachlorophenol			. 33 3	
Dimethyl phthalate	50* 6			
Bis(2-ethyl hexyl)phthalate	75 4		20 5	
Values shown are percent of total SPM samples collected that exceeded the sediment quality objectives	f total SPM samples collect	ed that exceeded the sedin	nent quality objectives	
Subscript= Number of samples	oles			
*=SQO also exceeded in bottom sediment	ttom sediment			

bs= Chemical only exceeded SQO in bottom sediments

"

Table 5: Summary of metals and organic compounds exceeding the Commencement Bay sediment quality objectives (SQO) in settling narriculate matter and bottom sediments from Sitcum Waterway. June 1991 to June 1995.

completion of dredging in the waterway no chemicals were detected above the SQOs in SPM from Sitcum Waterway.

Four chemicals (zinc, LPAH, dibenzofuran, and dimethyl phthalate) were above the SQOs in Sitcum bottom sediments. Dimethyl phthalate near the north corner drain was the only chemical present above the SQO after dredging was completed in the waterway.

#### **Gross Sedimentation Rates**

Gross sedimentation rates for Sitcum Waterway are shown in Figure 12. Gross accumulation rates in the waterway were quite variable, ranging from 1.3 to 8.3 g/cm<sup>2</sup>/yr. The mean rate for the waterway was  $4.8 \pm 1.8$  g/cm<sup>2</sup>/yr. The highest mean rate in the waterway ( $6.4 \pm 2.2$  g/cm<sup>2</sup>/yr) was measured at Berth B, along Terminal 7. Mean rates at the remaining stations were similar, ranging from  $4.1 \pm 1.6$  g/cm<sup>2</sup>/yr at the mouth to  $4.5 \pm 1.2$  g/cm<sup>2</sup>/yr at the head of the waterway. Predicted accumulation rates for the waterway ranged from 1.3 to 13 cm/yr, with a mean of  $5.1 \pm 2.1$  cm/yr.

Temporally, the lowest sedimentation rates in the waterway occurred during the late winter and spring months (February to May). The highest rates were typically measured between July and November. A complete list of sedimentation rates for Sitcum Waterway between June 1991 and June 1995 is included in Appendix C, Table C8.

## **Thea Foss Waterway**

#### Chemicals in SPM

Between November 1992 and June 1995 44 sediment traps were deployed in Thea Foss Waterway. Of these traps, 41 were retrieved for a recovery rate of 93%. Monitoring results for conventionals and metals in Thea Foss SPM are included in Appendix C, Table C9.

Relatively high TOC concentrations were measured in Thea Foss Waterway, especially at the head. TOC values ranged from 3.8 to 12%. The mean value at the head (9.2%), was elevated by a factor of two compared to the mean value at the mouth (4.6%) of the waterway. TOC is listed as a problem chemical at the head of the waterway.



June 1991 to June 1995.

Concentrations of selected metals (mg/kg) in Thea Foss SPM between November 1992 and June 1995 are:

				Detection	
<u>Metal</u>	Range	<u>Mean</u>	<u>C.V.</u>	Frequency	$\underline{N}$
Copper	97-310	180	24%	100%	41
Lead	90-420	250	36%	100%	41
Mercury	0.4-1.5	0.70	31%	100%	41
Zinc	120-510	300	35%	100%	41

C.V.= Coefficient of Variation (SD/Mean)

The distribution of metals in Thea Foss Waterway during the current monitoring period is consistent with the results from previous monitoring (Norton, 1993). Peak concentrations of cadmium, lead, nickel, and zinc were measured at the head of the waterway near the twin 96" drains. The highest copper concentrations were present near the mouth of Wheeler-Osgood Waterway. On average, mercury levels were at a maximum in the central portion of the waterway near the 15th Street drain and the mouth of Wheeler-Osgood Waterway. The 15th Street drain, shoreline banks adjoining the Investco Site, and the inactive Surplus Steam Plant #1 have been identified as potential sources of mercury in this portion of the waterway (Smith, 1995). The lowest metals concentrations were consistently measured at the mouth of the waterway.

As illustrated in Figure 13, two longitudinal patterns were evident for metals concentrations in Thea Foss Waterway. Lead, nickel, and zinc decreased moving out the waterway, with the highest concentrations occurring at the head and the lowest at the mouth. The distribution of cadmium and nickel, which are not shown in Figure 13, also followed this pattern. In contrast, copper and mercury increased moving away from the head, peaking in the middle, and then decreasing to a minimum at the mouth of the waterway. Consistently higher (8/8) copper values were measured at station TF-3 (mouth of Wheeler-Osgood) compared to TF-2 (15th Street) which is located at the opposite side of the waterway. This result suggests a source of copper is located in or near the mouth of Wheeler-Osgood Waterway. Previous studies have documented high concentrations of copper in sediments at J.M. Martinac Shipbuilding (Norton and Johnson, 1984; Norton, 1995). Sources of copper at Martinac Shipbuilding include the use of granulated slag as sandblasting material, and removal of antifouling paints during vessel renovations (Tetra Tech, 1985).

N= Number of Samples



Figure 13: Selected Metals in Thea Foss Settling Particulate Matter, November 1992 to June 1995.

The results of semivolatile organics and butyltin analysis of Thea Foss Waterway SPM are listed in Appendix C, Table C10. Thirty-two target semivolatile compounds were detected in SPM. Detected concentrations of selected organics in Thea Foss SPM are:

Compound	Range	Mean	<u>C.V.</u>	Detection Frequency	
					N
LPAH	2100-17000	5300	66%	100%	22
HPAH	5200-84000	25000	81%	100%	22
Butyl benzyl phthalate	160-2500	890	75%	73%	16
Bis(2EH)phthalate	590-96000	28000	130%	46%	10
Benzoic Acid	1100-9600	4600	57%	55%	12
Tributyltin	30-790	470	44%	100%	20
C.V.= Coefficient of Varia	ation (SD/Mean)				
N= Number of Samples					

On average, concentrations of most problem chemicals tended to be highest at the head of the waterway. In particular, HPAH concentrations, shown in Figure 14, were substantially higher at the head of the waterway. An exception to this pattern was Bis(2-ethyl hexyl)phthalate. The highest concentration of Bis(2-ethyl hexyl)phthalate (96,000 ug/kg) was measured north of the 15th Street drain. The lowest concentrations of most problem chemicals were typically measured at the mouth of the waterway.

In contrast to metals, few strong longitudinal gradients were evident for semivolatile organics in Thea Foss Waterway, with the exception of HPAH. HPAH concentrations were highest at the head and decreased moving toward the mouth of the waterway. Major PAH sources previously identified at the head of Thea Foss Waterway included discharge from storm drains -- primarily the twin 96" -- and operation of a historic coal gasification plant (Tetra Tech, 1985).

Temporally, <u>intra</u>-station concentrations of most "problem" chemicals tended to be somewhat variable. However, differences in quantitation limits between several of the monitoring periods hinder interpretations of temporal trends (see quality assurance section).

The distribution of TBT in Thea Foss SPM is displayed in Figure 15. TBT and its breakdown products were detected throughout the waterway. TBT concentrations ranged from 30-790 ug/kg with a mean of 470 ug/kg. The highest concentrations were measured in the upper portion of the waterway adjacent to the City Marina (station TF-1A). Recent sampling by inspectors at Ecology's Southwest Regional Office has identified high concentrations of TBT (up to 1,400,000 ug/kg) in sediments from catch basins which drain the Picks Cove Marina boat yard into this section of the waterway (Mercuri, 1995). Longitudinally, TBT levels tended to increase slightly moving away from the head, peaked approximately 0.4 miles from the head









(station TF-1A), and then decreased moving toward the mouth of the waterway. The lowest TBT levels were measured at the mouth of the waterway.

At the request of the City of Tacoma, PCBs were added as a target analyte in Thea Foss Waterway beginning with the November 1994 to June 1994 sample set. During this period PCBs were detected at all monitoring stations in the waterway (Appendix C, Table C11). Concentrations of total PCBs ranged from 280 to 610 ug/kg, with a mean of 500 ug/kg. The highest value was present north of the 15th Street drain (station TF-2). As was the case for most contaminants in the waterway the lowest PCB levels were measured at the mouth.

#### Chemicals in Bottom Sediment

The results of conventionals and metals analyses of bottom sediments collected from each of the sediment trap stations on three occasions -- December 1989, January 1991, and November 1994 -- are summarized in Appendix D, Table D6. TOC concentrations in the waterway ranged from 1.4 to 8.8% with a mean of 5.6%. TOC tended to be highest at the head and decreased moving toward the mouth of the waterway. Grain size analysis indicated that sediments collected from the head and near the mouth of Wheeler-Osgood consisted primarily of silt and clay size particles. Sediments from the area north of 15th Street drain and the mouth tended to have a higher percentage of sand size material.

Metals concentrations were somewhat variable in Thea Foss bottom sediments. The highest concentrations of nickel (190 mg/kg) and zinc (520 mg/kg) were present at the head. Lead was at a maximum north of the 15th Street drain. Peak concentrations for cadmium, copper, and mercury were measured near the mouth of Wheeler-Osgood Waterway. The lowest concentrations for all metals were present at the mouth of Thea Foss Waterway.

Compared to SPM, fewer semivolatile organics (Appendix D, Table D7) were detected in bottom sediments. LPAH and HPAH concentrations tended to be highest at the head of the waterway and decreased moving towards the mouth. At all stations, the sum of HPAH exceeded the sum of LPAH. As previously mentioned the apparent enrichment of HPAHs relative to LPAHs would suggest that historical sources of PAHs have played an important role in the pattern of PAH contamination observed in bottom sediments.

Peak concentrations for the majority of phthalate esters were present north of the 15th Street drain. The lowest concentrations for all organics were measured at the mouth of the waterway.

TBT and its breakdown products MBT and DBT were detected throughout Thea Foss Waterway. Butyltin concentrations in bottom sediments (Appendix D, Table D8) were quite variable, ranging from 10 to 410 ug/kg with a mean of 150 ug/kg. In general, bottom sediment TBT concentrations were typically lower than those measured in SPM. Based on means the highest TBT levels were present near the mouth of Wheeler-Osgood Waterway. As was the case for many of the contaminants in the waterway the lowest TBT levels were present at the mouth of Thea Foss Waterway.

Mercury, TBT, and HPAH levels in bottom sediments collected between 1984 and 1994 are compared in Figure 16. At all locations except the mouth, mercury levels in bottom sediments at each of the sediment trap stations were higher than levels reported in 1984 as part of the CBRI (Tetra Tech, 1985). A similar pattern is evident for HPAH levels at the head of the waterway and near the mouth of Wheeler-Osgood Waterway. For TBT, 1994 levels at three of the four stations were the highest measured between 1989 and 1994. These data suggest that mercury, TBT, and HPAH levels in bottom sediments from much of the waterway have not decreased during the period of monitoring. It should be noted that factors such as spatial and analytical variability between surveys may be contributing to the apparent trends in contaminant levels.

#### Comparison to Commencement Bay SQOs

Chemicals exceeding the SQOs in Thea Foss SPM and bottom sediments are shown in Table 6. A total of 14 compounds in SPM (3 metals and 11 organics) are listed as exceeding the SQOs. The greatest exceedances (11) were present at the head of the waterway near the twin 96" drains. Four chemicals (mercury, HPAH, PCBs, and benzoic acid) were present at concentrations above the SQOs throughout the waterway. Bis(2-ethyl hexyl)phthalate was also measured at problem levels at all monitoring stations except the mouth. PCB levels at all stations except TF-1A were above the SQO for the protection of human health. The fewest number of exceedances (5) were measured in the center of the waterway adjacent to the City Marina.

Comparing TBT levels in Thea Foss SPM to the PSDDA ISL of 30 ug/kg indicates that 100% of the samples (20/20) tested during the monitoring period were above the ISL. Biological testing planned as part of the Thea Foss Pre-Remedial Design Evaluation being conducted by the City of Tacoma should help evaluate the toxicity associated with the observed TBT levels in the waterway.

Thirteen individual chemicals (4 metals and 9 organics) were present above the SQOs in bottom sediments. As was the case for SPM, the greatest number of exceedances (10) were measured at the head of the waterway near the twin 96" drains. The fewest exceedances (3) were present at the mouth of the waterway. Phthalate esters were the predominant group of organic chemicals exceeding the SQOs.

Lower TBT concentrations were present in Thea Foss bottom sediments compared to levels in SPM. However, the majority of bottom sediments still contained TBT levels above the PSDDA ISL.





Table 6: Summary of metals and organic compounds exceeding the Commencement Bay sediment quality obje in settling particulate matter and bottom sediments from Thea Foss Waterway, November 1992 to June 1995.	s and organic compounds t and bottom sediments fi	exceeding the Commenc om Thea Foss Waterway	ement Bay sediment , November 1992 to	exceeding the Commencement Bay sediment quality objectives (SQO) om Thea Foss Waterway, November 1992 to June 1995.	
Station	TF-1	TF-1A	TF-2	TF-3	TF-4
	Head near	Mid-Channel near	North of 15th	Near Mouth of	
Location	Twin 96" Drains	City Marina	Street Drain	Wheeler-Osgood	Mouth
Metals					
Lead	bs		þs		
Mercury	70* 10	100 4	100 8	100* 9	10 10
Nickel	bs	50 2		bs	
Zinc	60* <sub>10</sub>			11 9	
<u>Organics</u>					
LPAH	100* 5		bs	20 5	bs
HPAH	100* 5	100 2	100* 5	40* 5	20* <sub>5</sub>
2-Methyl napthalene	20 5				
1,4-Dichlorobenzene			ps		
Benzoic acid	100 3	100 1	67 3	75 4	100 3'
Benzyl alcohol	bs		100* 1	100 1	
Diethyl phthalate			sq	-	
Dimethyl phthalate	ps		40 5		
Di-n-butyl phthalate	25 4		20* 5		20 5
Di-n-octyl phthalate	25 4				
Butyl benzyl phthalate	100* 4		ps		20 4
Bis(2-ethyl hexyl)phthalate	100 2	100 2	100* 4	50* 2	ps
PCBs+	100 1		100 1	100 2	100 1
Values shown are percent of total SPM samples collected that exceeded the sediment quality objectives	f total SPM samples coll	ected that exceeded the se	ediment quality object	tives	

Subscript= Number of samples Page 45

bs= Chemical only exceeded SQO in bottom sediments \*=SQO also exceeded in bottom sediment

+= SQO for PCBs based on protection of human health

#### **Gross Sedimentation Rates**

Gross sedimentation rates for Thea Foss Waterway are listed in Appendix C, Table C12. Compared to rates measured in the other waterways, mass accumulation rates for Thea Foss Waterway, shown in Figure 17, were fairly low and consistent, ranging from 0.5-4.4 g/cm<sup>2</sup>/yr, with a mean of  $1.5\pm0.9$  g/cm<sup>2</sup>/yr. Significantly higher accumulation rates (ANOVA, P<0.01) were measured approximately 0.4 miles from the head (TF-1A). Sedimentation rates at the remaining stations were not significantly different.

During the March to June 1995 collection period a three-fold increase in sedimentation was noted at the head of the waterway. The observed increase in this area is probably related to higher bottom sediment resuspension rates caused from bridge construction activities. In March 1995 pile driving was started for the new SR-509 bridge which crosses Thea Foss Waterway at the head (Coleman, 1995). Increased bottom sediment resuspension resulting from this type of in-water construction activity has the potential to redistribute contaminants in the waterway. Redistribution of contaminants is a real concern in a heavily contaminated area like the head of Thea Foss Waterway.

In general, the data collected indicate that sedimentation in most of the waterway tends to be highest during the summer and lowest during winter months. Factors such as seasonal variations in plankton populations, increases in vessel traffic during the summer months which could resuspend bottom material, and variations in runoff from the Puyallup River could all be contributing to the seasonal patterns observed. Seasonal analysis of <sup>210</sup>Pb levels in SPM and adjacent bottom sediments could help identify the relative importance of the various factors contributing to resuspension of bottom sediments. This work would involve analyzing <sup>210</sup>Pb levels in SPM on a quarterly basis. <sup>210</sup>Pb activities in SPM could then be compared to levels in adjacent bottom sediments (top 2cm). Predicted accumulation rates on the bottom during the monitoring period ranged from 0.5-8.7 cm/yr with a mean of 2.5±1.8 cm/yr.







# Discussion

## **Contaminant Trends Over Time**

Simple linear regression was used to determine whether changes in contaminant levels associated with SPM have occurred over time. All SPM data collected during the monitoring program were used in this evaluation to provide the longest time-series record possible for each waterway. An exception was metals data collected after May 1994 for all parameters except mercury which was not used in the analysis of temporal trends. The reason for this omission is that metals data collected after 1994 were generated using different analytical methods then were employed prior to May 1994 (see sample analysis and quality assurance section).

In comparison to metals (c.v. = 23-57%), organics concentrations in SPM tended to have a higher degree of temporal variability (c.v. = 33-400%) in SPM. This fact, combined with differences in quantitation limits between monitoring periods, hinders a detailed statistical analysis of trends for organics. Consequently, most of the statistically significant trends noted were for metals.

In most instances little change was evident in contaminant levels associated with SPM in Hylebos and Thea Foss Waterways during the monitoring period. Several exceptions to this general statement are discussed below.

Arsenic in Hylebos Waterway SPM between July 1990 and May 1994 is shown in Figure 18. A significant reduction ( $r^2 = 0.45$ , P<0.01) in arsenic levels was noted at the mouth of the waterway. The reason for the apparent reduction in arsenic at this location is not clear.

In contrast to Hylebos and Thea Foss Waterways, substantial reductions for most metals and organics levels were seen in Sitcum Waterway SPM. Arsenic, lead, and zinc concentrations in Sitcum Waterway SPM are shown in Figure 19. Significant decreases in lead and zinc levels were noted between July 1990 and December 1993 at the following locations:

Lead Head at north corner ( $r^2=0.83$ , P<0.001) Terminal 7 at Berth B ( $r^2=0.49$ , P<0.02)

Zinc Head at north corner drain ( $r^2 = 0.53$ , P<0.005)

This finding is consistent with the fact that, as of July 1991, off-loading and handling of black ore (lead and zinc) ceased at Terminal 7 (Norton and Barnard, 1992).

In contrast to lead and zinc, a significant increase ( $r^2=0.83$ , P<0.001) in arsenic levels was measured near the north corner drain between July 1990 and December 1993. The north corner drain (SI-172) has been documented as a major source of contaminants in past studies







Figure 19: Arsenic, Lead and Zinc Trends in Sitcum Waterway Settling Particulate Matter, July 1990 to December 1993.

(EPA, 1992). Recent work conducted by the City of Tacoma in the summer of 1995 to clean this drain may have already reduced arsenic loadings to the waterway (Mercuri, 1995). The results of this work would not have been evident in the monitoring data shown in Figure 18. Ongoing monitoring in the waterway should prove to be useful in evaluating arsenic trends in this portion of the waterway.

The statement that substantial reductions have occurred for most organic compounds in Sitcum Waterway is primarily based on the fact that, prior to dredging, concentrations of seven individual compounds exceeded the SQOs in SPM. After completion of dredging in the waterway <u>no</u> chemicals were detected above the SQOs in SPM.

Monitoring data collected in Thea Foss Waterway between November 1988 and November 1992 indicated a significant increase in mercury levels was occurring at the head of the waterway and north of the 15th Street drain (Norton, 1993). The lack of a significant regression at these locations when the most recent data set is included might suggest that mercury concentrations are stabilizing in these areas of the waterway.

# Comparison of Problem Chemicals in SPM Between Waterways

A comparison of selected metals and organics concentrations between November 1988 and June 1995 in Hylebos, Sitcum and Thea Foss SPM is presented in Table 7. Based on means, the highest concentrations of TOC, mercury, lead, zinc, LPAH, and HPAH were present in Thea Foss Waterway. Mean arsenic levels in Hylebos Waterway were approximately twice as high as those measured in Thea Foss Waterway. In addition, mean PCB levels in Hylebos were approximate five times higher than levels in Thea Foss Waterway. The highest bis(2-ethyl hexyl)phthalate concentrations were typically present in Sitcum Waterway. Similar concentrations of copper were measured in all three waterways. TBT concentrations in Hylebos and Thea Foss Waterways are also similar.

## **Comparison of Gross and Net Sedimentation Rates**

Sedimentation rate data for Hylebos, Sitcum, and Thea Foss Waterways are summarized in Table 8. Two types of rates are listed. The first is gross sedimentation (net + resuspension) which was determined from sediment trap data collected between September 1991 and June 1995. Also shown are net accumulation rates which have been estimated from several <sup>210</sup>Pb dated cores collected during the remedial investigation in 1984 (EPA, 1992), and a subsequent study in Sitcum Waterway (Hart Crowser, 1992).

Gross accumulation rates reported for the three waterways agreed within a factor of 2. The highest mean rates were measured in Sitcum Waterway ( $4.8\pm1.8 \text{ g/cm}^2/\text{yr}$ ), followed by Hylebos ( $2.4\pm1.0 \text{ g/cm}^2/\text{yr}$ ), and then Thea Foss Waterway ( $1.5\pm0.9 \text{ g/cm}^2/\text{yr}$ ).

Table 7: Comparison of Selected Metals\* and Organics Associated with Settling Particulate Matter from Hylebos, Sitcum, and Thea Foss Waterways, November 1988 to June 1995.

ILYICOUS, SILCUILI, ALLA IL	ica i uss waici ways, inuv	inficous, succuri, and thea ross watchways, inventuel 1200 to Julie 1223.	
Location	Hylebos	Sitcum	Thea Foss
Collection Period	7/90-6/95	7/90-6/95	11/88-6/95
Number of Samples	7-97	26-47	5-92
Total Organic Carbon (%)	3.9 (1.7-8.5)	2.0 (1.2-3.7)	6.1 (1.9–12)
<u>Metals* (mg/kg, dry)</u>			
Arsenic	53 (8.7-130)	34 (6-83)	22 (21-30)
Copper	140 (71-410)	120 (64–190)	190 (120-340)
Lead	9386 (41-250)	180 (44–370)	260 (120-460)
Mercury	0.38 (0.16-0.93)	Nee	0.65 (0.35-1.5)
Zinc	210 (110-520)	240 (91–580)	320 (160-570)
Organics (ug/kg, dry)			
LPAH	3400j (190j-75000j)	2200j (77-8600j)	6000j (250j-32000j)
НРАН	10000j (4100j–49000j)	5800j (160–21000j)	26000j (980j-120000j)
Bis(2EH)phthalate	5100j (730-44000)	41000j (110u-170000)	18000j (590-96000)
Total PCBs	2400j (130-73000)	I	500 (280-610)
Tributyltin	460 (210j-660)	1	360 (11-790)
Mean of detected values (range of all values)	ge of all values)		
u= Not detected at detection limit shown	limit shown		2011.
j= Estimated concentration			

-= No data

\*-- Metals data collected after May 1994 was not included due to different method of analysis

Table 8: Comparison of Sediment Accumulation Rates for Commencement Bay Waterways Obtained from Sediment Trap and Bottom Core Data.

	Collection		No.	Sedimentation Rate(1)	ion Rate(1)
Location	Period	Method	Samples	(g/cm2/yr)	(cm/yr)
		GROSS SEDIMENTATION			
HYLEBOS	HYLEBOS WATERWAY				
Head	12/91-6/95	Sediment Trap	20	1.6±0.5	$2.7\pm0.9$
Mouth	"н		23	$3.1\pm1.2$	3.0±1.2
Overall	Ŧ		145	$2.4\pm1.0$	$3.4\pm1.3$
	CITCIIM WATERWAY				
Head	9/91-6/95	Sediment Trap	19	$4.5\pm 1.2$	5.0±1.3
Mouth	æ	÷	19	<b>4.1</b> ±1.6	4.6±1.9
Overall	-	Ŧ	70	<b>4.8</b> ±1.8	$5.1\pm 2.1$
тие и воес	THEA EOSS WATERWAY				
Uncd FUS	3 WALENWAL	. Sadiment Tran	00	1.7+0.9	3.4+1.7
неаа	C6/0-76/11	deri mammac	27	· · · · · · · · · · · · · · · · · · ·	
Mouth		Ŧ	19	1.0±0.4	0.9+0.4
Overall	<b>2</b> .	-	81	$1.5\pm0.9$	2.5±2.0
		NET SEDIMENTATION(2)			
<b>HYLEBOS</b>	HYLEBOS WATERWAY	to			
Head	5/84	Pb-210 cores (3)	I	0.33	0.77
Mouth	₩.	85	I	1.1	
SITCUM W	SITCUM WATERWAY				
Head	1661	Pb-210 cores (4)	1	0.6 <u>+</u> 0.3	0.6±0.3
THEA FOS	THEA FOSS WATERWAY				
Head	5/84	Pb-210 cores (3)	**	0.3	0.45
Mouth	¥		I	0.45	0,67
(1)=Mean±sd	sd .				
(2)=EPA, 1992	992				
(3)=Estimated values	ed values				
	1001				

(4)= Hart Crowser, 1992

Net accumulation rates for the three waterways ranged from 0.3 to  $1.1 \text{ g/cm}^2/\text{yr}$ . The uncertainty associated with these measurements was not reported.

# Resuspension

Comparison of sedimentation rates from sediment traps (gross sedimentation) and rates from <sup>210</sup>Pb dated cores (net sedimentation) have been used in other investigations to estimate bottom sediment resuspension rates (Baker et.al., 1991). Gross and net sedimentation rates for Hylebos, Sitcum, and Thea Foss Waterways are compared in Table 9 below.

Location	Gross	Net <sup>1,2</sup>	Resuspension	Percent
Hylebos				
Head	1.6±0.5	0.33	1.3±0.5	81%
Mouth	3.0±1.2	1.1	1.9±1.2	63%
Sitcum				
Head	4.5±1.2	0.6±0.3	3.9±1.2	71%
Mouth	4.1±1.6	0.6±0.3	3.5±1.6	85%
Thea Foss		-		
Head	1.7±0.9	0.3	1.4±0.9	82%
Mouth	1.0±0.4	0.45	0.6±0.4	60%

Table 9: Summary of Estimated Resuspension Rates for Hylebos, Sitcum, and Thea Foss Waterways Based on Sediment Trap and Bottom Core Data (g/cm<sup>2</sup>/yr).

Values shown are Mean"S.D.

Resuspension= (gross sedimentation - net sedimentation)

Percent= (resuspension/gross sedimentation) \* 100

 $^{1}$  = EPA, 1992

 $^{2}$ = Net rates for Sitcum reported in Hart Crowser, 1992

Resuspension estimates for the three waterways ranged from 60% at the mouth of Thea Foss Waterway to 85% at the mouth of Sitcum Waterway. The highest resuspension rates were estimated for Sitcum Waterway, followed by Hylebos, and Thea Foss Waterways. The reason that the highest resuspension rates were observed in Sitcum Waterway is probably related to the fact that Sitcum receives the heaviest amount of large ship traffic of the three waterways monitored. While the exact proportion of "new" versus "old" material collected by the traps cannot be reliably determined with the available data, it is fair to assume that the trapped material is representative of sediments which are mobile in the waterways.

Resuspension estimates for Hylebos, Sitcum, and Thea Foss Waterways fall within the range of values  $(0.4-3.6 \text{ g/cm}^2/\text{yr})$  reported by Patmont and Crecelius (1991) for other urban embayments in Puget Sound. In addition, the resuspension estimates for Sitcum Waterway, 71-85%, are similar to values reported along the central Seattle Waterfront (74-92%) in areas that receive a large amount of vessel traffic.
# Conclusions

In general, the spatial distribution of contaminants measured in SPM in Hylebos, Sitcum, and Thea Foss Waterways was in relatively good agreement with previous monitoring results collected between November 1988 and November 1992. In all three waterways metals concentrations were typically higher at the head of the waterways and decreased moving toward the mouth. A similar pattern was also present for most of the organic compounds; however, concentrations tended to be more variable.

In Hylebos Waterway, lead, tetrachloroethene, hexachlorobenzene, hexachlorobutadiene, and 1,2,4-trichlorobenzene were elevated in the outer portion of the waterway adjacent to Occidental Chemical. Occidental Chemical was identified as a source of hexachlorobenzene and hexachlorobutadiene during the Commencement Bay Nearshore/Tideflats Remedial Investigation (Tetra Tech, 1985). In addition, between 1947 and 1973 tetrachloroethene was produced in commercial quantities at Occidental's solvent plant (Tetra Tech, 1986b). Tributyltin (TBT) which has been used extensively in the past as an active ingredient in antifouling paints was present at relatively high levels throughout Hylebos.

Mercury and copper concentrations in Thea Foss Waterway tended to peak in the central portion of the waterway near the mouth of Wheeler-Osgood Waterway (copper) and north of the 15th Street drain (mercury). J.M. Martinac Shipbuilding located near the mouth of Wheeler-Osgood Waterway has been identified as a copper source in past studies (Norton and Johnson, 1984; Norton, 1995). The 15th Street drain, shoreline banks adjoining the Investco Site, and the inactive Surplus Steam Plant #1 have been identified as potential sources of mercury in this area of the waterway (Smith, 1995). As was the case for Hylebos Waterway, TBT was elevated throughout Foss Waterway.

Overall, the SPM data collected from Hylebos and Thea Foss Waterway suggest that concentrations of most problem chemicals have not decreased since sediment trap monitoring was started in July 1990 and November 1988, respectively. In both Hylebos and Thea Foss Waterways, 14 separate chemicals were above the Commencement Bay "Sediment Quality Objectives" (SQOs) in SPM. The greatest number of individual exceedances were present at the head of both waterways.

In contrast, dramatic reductions in contaminant levels have been observed in Sitcum Waterway comparing pre- and post-dredging conditions. Prior to dredging in Sitcum Waterway, seven individual chemicals exceeded the SQOs in SPM. After the completion of dredging <u>no</u> exceedances of the SQOs were observed for contaminant levels in SPM. In all waterways monitored, fewer chemicals exceeded the SQOs in bottom sediments compared to SPM.

The highest gross sedimentation rates (net + resuspension) were measured in Sitcum Waterway  $(4.8\pm1.8 \text{ g/cm}^2/\text{yr})$  followed by Hylebos  $(2.4\pm1.0 \text{ g/cm}^2/\text{yr})$ , and Thea Foss  $(1.5\pm0.9 \text{ g/cm}^2/\text{yr})$  Waterways. Bottom sediment resuspension estimates ranged from 60% at the mouth of Thea Foss Waterway to 85% at the mouth of Sitcum Waterway. The highest resuspension rates were estimated for Sitcum, followed by Hylebos and Thea Foss Waterway. While the exact proportion of "new" versus "old" material collected by the traps cannot be reliably determined with the available data, it is fair to assume that the trapped material is representative of sediments which are mobile in the waterways.

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# Appendices

# Appendix A

Station Location and Deployment Information

Table A1: Station DescriptionsTable A2: Sediment Trap Deployment Schedule

North east corner of Upper Turning Basin @ Mouth of Wheeler-Osgood Waterway Mid-Channel adjacent to City Marina @ Mouth near D-Street Tank Farms North end of Sealand Terminal North end of General Metals Head at North Corner Drain Head near Twin 96" Drains North of 15th Street Drain Near Lincoln Street Drain @ Occidental Chemical Terminal 7 @ Berth B Mouth @ Terminal 7 Station Description Near Cenex Plant @ Elf Atochem @ Mouth (ft @ MLLW) Water Depth 36 20 20 25 54 32 45 18 22 23 25 30 31 4 33 25 52 58 50 59 Longitude 22 50 52 26 4 8 ø ļ 11 2 31 4 24 25 25 52 Position (deg/min/sec) 33 25 53 22 52 24 24 24 25 21 22 122 122 122 122 122 122 22 122 [22] 122 122 122 [22 122 122 122 THEA FOSS WATERWAY HYLEBOS WATERWAY SITCUM WATERWAY 16 35 47 14 35 47 16 20 47 14 50 15 26 47 15 44 8 16 26 47 16 45 47 16 59 47 15 59 47 16 11 r--16 11 Latitude 47 16 16 47 15 47 15 47 47 47 ţ, ÷ TF-1A Station  $\Gamma F - 1$ **FF-2 TF-3 TF-4** H-6 H-7 H-2 H-3 H-5 S-2 S-3 So. H-1 H-4 S-1 S-4

Table A1: Station descriptions for Commencement Bay Sediment Trap Monitoring Program.

# Days	14444000000000000000000000000000000000	I	84	82	1	ł	84	84		83	83	83	83		83	1	83	83	83	# Days		84	84	84	85	1	85	85	*****	84	84	84	84		82	1	82	82	82	
995. Retrieved		NR	5/14/92	5/12/92	NR	NR	5/12/92	5/12/92		5/11/92	5/11/92	5/11/92	5/11/92		5/12/92	I	5/12/92	5/12/92	5/12/92	Retrieved		5/18/93	5/18/93	5/18/93	5/18/93	NR	5/18/93	5/18/93		5/17/93	5/17/93	5/17/93	5/17/93		5/17/93	1	5/17/93	5/17/93	5/17/93	
1991 to June 1 # Days   Deployed		2/20/92	2/20/92	2/20/92	2/20/92	2/19/92	2/18/92	2/18/92		2/18/92	2/18/92	2/18/92	2/18/92		2/19/92	ND	2/19/92	2/19/92	2/19/92	Deployed		2/23/93	2/23/93	2/23/93	2/22/93	2/22/93	2/22/93	2/22/93		2/22/93	2/22/93	2/22/93	2/22/93		2/24/93	Q	2/24/93	2/24/93	2/24/93	
e 1991 t # <sup>Days</sup>		91		91				06					92		93	1			93	# Days		26			96	1		<u>76</u>					86		66				66	·
chedule for Commencement Bay Sediment Traps, June 1991 to June 1995 #Days [Deployed Retrieved # Days [Deployed Retrieved # Days [Deployed Ret		2/20/92	NR	2/20/92	NR	2/19/92	2/18/92	2/18/92		2/18/92	2/18/92	2/18/92	2/18/92		2/19/92	1	2/19/92	2/19/92	2/19/92	Retrieved		2/23/93	2/23/93	2/23/93	2/22/93	NR	2/22/93	2/22/93		2/22/93	2/22/93	2/22/93	2/22/93		2/24/93	1	2/24/93	2/24/93	2/24/93	
ediment <sup>*</sup> Deployed		11/21/91	11/21/91	11/21/91	11/21/91	11/21/91	11/20/91	11/20/91		11/20/91	11/18/91	11/20/91	11/18/91		11/18/91	ND	11/18/91	11/18/91	11/18/91	Deployed		11/18/92	11/18/92	11/18/92	11/18/92	11/18/92	11/18/92	11/17/92		11/16/92	11/16/92	11/17/92	11/16/92		11/17/92	ND	11/17/92	11/17/92	11/17/92	
t Bay S€ # <sup># Days</sup>		56	56	1	. 1	56	2	55		63	63	1	63		61	I	62	63	62	# Days		98	98	98	66	66	66	98		98	98	1	98		86	I	66		66	
nencemen Retrieved	949	11/21/91	11/21/91	NR	NR	11/21/91	11/20/91	11/20/91		11/20/91	11/18/91	NR	11/18/91		11/18/91	ı	11/18/91	11/18/91	11/18/91	Retrieved		11/18/92	11/18/92	11/18/92	11/18/92	11/18/92	11/18/92	11/17/92		11/16/92	11/16/92	NR	11/16/92		11/17/92	I	11/17/92	11/17/92	11/17/92	
edule for Comi		9/26/91	9/26/91	9/17/91	9/26/91	9/26/91	16/11/6	9/26/91		9/18/91	9/16/91	9/18/91	9/16/91		9/18/91	QN	16/11/6	9/16/91	9/11/91	Deployed		8/12/92	8/12/92	8/12/92	8/11/92	8/11/92	8/11/92	8/11/92		8/10/92	8/10/92	8/10/92	8/10/92		8/11/92	DN	8/10/92	8/11/92	8/10/92	
hedule f # Days		- 89	1	89	I	I	I	I		ł	I	92	I		1	I	16	91	91	# Days		1	90	92	89	89	16	16			16	91	91		91	1		ł	60	
Table A2: Deployment Sc Station Deployed Retrieved	-Anno 1997 - Anno 1997	16/11/6	NR	16/11/6	NR	NR	NR	NR		NR	NR	9/18/91	NR		NR	I	9/16/91	9/16/91	9/16/91	Retrieved		NR	8/12/92	8/12/92	8/11/92	8/11/92	8/11/92	8/11/92		8/10/92	8/10/92	8/10/92	8/10/92		8/11/92	1	8/10/92	NR	8/10/92	
A2: Depl Deployed	Vaterway	6/20/91	6/18/91	6/20/91	6/18/91	6/20/91	6/18/91	6/18/91	'aterway	6/19/91	6/20/91	6/18/91	6/20/91	Thea Foss Waterway	6/17/91	DN	16/11/9	6/11/91	6/11/91	Deployed	Hylebos Waterway	5/14/92	5/14/92	5/12/92	5/14/92	5/14/92	5/12/92	5/12/92	<sup>t</sup> aterway	5/11/92	5/11/92	5/11/92	5/11/92	Thea Foss Waterway	5/12/92	Q	5/12/92	5/12/92	5/12/92	NR= Not recovered ND= Not deployed
Table /	Hylebos Waterway	<u>H-1</u>	Н-2	Н-3	H-4	H-5	H-6	H-7	Sitcum Waterway	<u>S-1</u>	S-2	S-3	S-4	Thea Fos	TF-1	TF-1A	TF-2	TF-3	TF-4	Station	Hylebos V	H-1	H-2	H-3	H-4	H-5	H-6	Н-7	Sitcum Waterway	S-1	S-2	S-3	S-4	Thea Fos	TF-1	TF-1A	TF-2	TF-3	TF-4	NR= Not ND= Not

Table	A2(cont):	Table A2(cont): Deployment Schedule for Commencement Bay Sediment Traps, June 1991 to June 1995	ant Sch	edule for	Commenc	ement E	3ay Sedii	ment Trap	s, June	1991 to J	lune 1995.	
Station	Deployed	Ketrieved	# Days	# Days Deployed	Retrieved	# Days	# Days Deployed	Retrieved	# Days	# Days Deployed	Retrieved	# Days
Hylebos	Hylebos Waterway	*******										
H-1	7/20/93	9/28/93	70		NR	~~~~	12/7/93	3/28/94	111	3/28/94	5/24/94	15
H-2	7/20/93	9/28/93	70	9/28/93	12/7/93		12/7/93	3/28/94	111	3/28/94	5/24/94	57
H-3	7/20/93	NR	I	9/28/93	12/7/93	70	12/7/93	3/28/94	111	3/28/94	5/24/94	57
H-4	ND	1	1	DN.		ţ.	12/7/93	3/28/94	111	3/28/94	5/24/94	57
H-5	ND	1	*	DN	1	1	12/7/93	3/28/94	111	3/28/94	5/24/94	57
H-6	7/20/93	9/28/93	02	9/28/93	12/7/93	70	12/7/93	3/28/94	111	3/28/94	5/24/94	57
Н-7	7/20/93	9/28/93	70	9/28/93	12/7/93	02	12/7/93	3/28/94	111	3/28/94	5/23/94	56
Sitcum 1	Waterway											
S-1	S-1 7/19/93	9/27/93	70	9/27/93	11/9/93	43	ND	1	i	QN	1	ł
S2	ND	I	I	DN	I	1	DN	1	1	QN	ŧ	I
S-3	7/19/93	10/01/93	74		11/9/93		QN	***	1 <sup>.</sup>	QN	ľ	I
S-4	7/19/93	9/27/93	70	9/27/93	11/9/93	43	QN	1	1	QN	1	I
Thea Fo	Thea Foss Waterway											
TF-1	7/19/93	9/27/93	70	9/27/93	12/13/93	11	12/13/93	3/29/94		3/29/94	5/23/94	55
TF-1A	QN	ł	1	DN	I	I	12/13/93	3/29/94		3/29/94	5/23/94	55
<b>TF-2</b>	7/19/93	9/27/93	22	9/27/93	12/13/93	17	12/13/93	3/29/94	106	3/29/94	5/23/94	55
TF-3	7/19/93	9/27/93	10	9/27/93	12/13/93	17	12/13/93	3/29/94	106	3/29/94	5/23/94	55
TF-4	7/20/93	9/27/93	69		12/13/93.	11	12/13/93	3/29/94	106	3/29/94	5/23/94	55
Station	Deployed	Retrieved	# Days	Deployed	Retrieved	# Days	Deployed	Retrieved	# Days	Deployed	Retrieved	# Days
Hulehos	Hylehoe Waterway							·				
H_1	2/74/94	8/27/94	06	8/22/94	11/30/94	100	11/30/94	3/7/95	67	3/7/95	6/13/95	98
H-2	5/24/94	8/22/94	8 06		11/30/94	100	11/30/94	3/7/95	79 7	3/7/95	6/13/95	86
H-3	5/24/94	8/22/94	6		11/30/94	100	11/30/94	3/7/95	1:6	3/7/95	6/13/95	. 98
H-4	5/24/94	8/22/94	96		11/30/94	100	Q	ł	1	ŊD	I	I
H-5	5/24/94	8/23/94	91	8/23/94	11/30/94	66	QN	I	I	DN	ł	ł
9H	5/24/94	8/23/94	16	8/23/94	11/29/94	98	11/29/94	NR	1	3/7/95	NR	I
H-7	5/24/94	8/24/94	92	8/24/94	11/29/94	67	11/29/94	3/7/95	98	3/7/95	NR	i
Sitcum	Sitcum Waterway											
S-1	QN	*	l	ND	1	1	11/30/94	3/6/95	96	3/6/95	6/17/95	86
S-2	QN	I	I		I	I	QN	I	1	Ð	ł	I
S-3	Q	·	I		I.	1	11/30/94	3/6/95	96	3/6/95	NR	1
S-4	QN	I	I	QN	I	1	11/30/94	NR	ł	3/6/95	6/12/95	98
Thea Fc	Thea Foss Waterway										1000	
TF-1	5/23/94	8/23/94	92		11/29/94	98	11/29/94	3/6/95	<i>L</i> 6	3/6/95	6/12/95	98
TF-1A	5/23/94	8/23/94	92		11/29/94	98	Ð	QN	1			I
TF-2	5/23/94	NR	i		11/29/94	86	11/29/94	3/6/95	<i>L</i> 6	3/6/95	NR	1 0
TF-3	5/23/94	8/23/94	92		NR	1	11/29/94	3/6/95	26	3/6/95	6/12/95	8, 8
TF-4	5/23/94	8/23/94	92	8/23/94	11/29/94	98	11/29/94	3/6/95	61	3/6/95	6/12/95	98
NR= N. ND=No	NR= Not recovered ND=Not Denloved											

ND=Not Deployed

# Appendix B

# Quality Assurance Information

# Table B1: Summary of Quality Assurance SamplesCase Narratives for Data Packages

Table B1: Summary of Quality Assurance Samples and Frequency of Analysis for th Commencement Bay Sediment Trap Monitoring Program.

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	Internal	Surrogate	Method	Matrix	Reference	Field
Analyte	Standards	Spikes	Blank	Spike*	Materials* Duplicate*	Duplicate*
Percent Solids		I	***	****	***	1/Batch
Grain Size	ı	ŀ	I	I	ı	1/Batch
TOC	I	I	1/Batch	ł	i	1/Batch
Metals						
Antimony	I	I	1/Batch	ł	1/Batch	1/Batch
Arsenic	I	1	1/Batch	I	1/Batch	1/Batch
Cadmium	I	I	1/Batch	1	1/Batch	1/Batch
Copper	I	1	1/Batch	I	1/Batch	1/Batch
Lead	I	i	1/Batch	**	1/Batch	1/Batch
Mercury	ł	ł	1/Batch	I	1/Batch	1/Batch
Nickel	I	1	1/Batch	i	1/Batch	1/Batch
Zinc	I	I	1/Batch	1	1/Batch	1/Batch
Organics	·					
<b>Volatiles</b>	1/sample	1/sample	1/Batch	1/Batch	I	1/Batch
Semivolatiles	1/sample	1/sample	1/Batch	1/Batch	(1/Batch)	1/Batch
PCBs	1/sample	1/sample	1/Batch	1/Batch	1/Batch	1/Batch
Butyltins	1/sample	1/sample	1/Batch	1/Batch	-	1/Batch
*= Samples analyzed in duplicate	in duplicate	_				
() = PAH only						
<b>Certified Reference Materials</b>	<u>Aaterials</u>					•
Metals- NIST #1646 (Estuarine Sediment)	6 (Estuarine	Sediment)				
PAH- NRCC #HS-6 (Harbour Marine)	6 (Harbour	Marine)		·		

PCBs- NRCC #HS-2 (Harbour Marine)

# Case Narratives

Conventionals Analysis

June 1991 to June 1995

6/91-12/91

# WASHINGTON STATE DEPARTMENT OF ECOLOGY ENVIRONMENTAL INVESTIGATIONS AND LABORATORY SERVICES MANCHESTER ENVIRONMENTAL LABORATORY

January 21, 1991

TO:

Dale Nortor FROM:

Sediment Traps - Commencement Bay amended report SUBJECT:

#### SAMPLE RECEIPT

The samples were received by the Manchester Laboratory on 12/19/91, in good condition.

#### **HOLDING TIMES:**

All analyses were performed within the suggested holding time for TOC and % Solids(6 months-frozen).

#### **INSTRUMENT CALIBRATION:**

Instrument calibration was performed before the run, and checked by initial calibration verification standards and blanks. Continuing calibration standards were analyzed at the frequency of 10% during the run. All initial and continuing verification standards were within the control limits of + /-10%.

#### **PROCEDURAL BLANKS:**

The blank values were within the acceptable range and showed no analytically significant levels.

#### PRECISION DATA

The results of the duplicate samples were used to evaluate analytical precision related to the sample set. The RPD's were within a 5% window of acceptance for duplicate analysis, except sample 518509, 518543, and 518568. Triplicate analysis were run on these samples.

#### **STANDARD REFERENCE MATERIAL:**

The analysis of the external verification standards gave no unusual results, and were within the acceptable established criteria.

#### SUMMARY

The data generated by the analysis of the samples above may be used noting any stated qualifications.

For TOC analysis, the samples were dried to constant weight at 71 degrees C.

If you have any questions concerning the results, please feel free to call me at SCAN 744-4737.

12/91-5/92

# WASHINGTON STATE DEPARTMENT OF ECOLOGY ENVIRONMENTAL INVESTIGATIONS AND LABORATORY SERVICES MANCHESTER LABORATORY

August 20, 1992

TO: Project Officer

FROM: David A. Thomson Not

SUBJECT: Commencement Bay Sediment TOC Results

# SAMPLE RECEIPT:

The samples from Commencement Bay were received by the Manchester Laboratory on 6/25/92 in good condition.

The Total Organic Oxygen analyses were performed by Analytical Resources Incorporated using Puget Sound Estuary Protocol (PSEP) for TOC.

# **PROCEDURAL BLANKS:**

All blanks are within the method guideline.

# **PRECISION DATA:**

The % RSD for all the replicate analyses were all less than 5%.

# STANDARD REFERENCE MATERIAL:

Standard reference material (KHP) and external verification standards were all within 10% of the stated value.

# SUMMARY:

The data generated by the analysis of the Commencement Bay samples can be used without qualification.

If you have any questions about the results or the methods used to obtain these results please call me at (206) 871-8823.

5/92 - 11/92



#### STATE OF WASHINGTON

# DEPARTMENT OF ECOLOGY

MANCHESTER ENVIRONMENTAL LABORATORY

7411 Beach Drive East • Port Orchard, Washington 98366-8204 • (206) 895-4737 • SCAN 744-4737

February 9, 1993

TO: Project Officer

FROM: David A Thomson

SUBJECT: Quality Assurance memo for the Commencement Bay Sediment Traps TOC Results

# SAMPLE RECEIPT

The samples from the Commencement Bay Sediment Traps were received by the Manchester Laboratory on December 9, 1992 in good condition. The analyses for these samples were subsequently contracted to Laucks Testing Laboratories. The samples were run using the Puget Sound Estuary Program (PSEP1986) for TOC.

#### HOLDING TIMES

All analyses were performed within 30 days of sample collection.

#### PROCEDURAL BLANKS

The procedural blanks associated with these samples showed no analytically significant levels of analytes.

# **PRECISION DATA**

Five samples were run in duplicate to evaluate precision on this sample set. The Relative Percent Difference (RPD) for all analytes was within the +/-10% window for duplicate analysis.

# LABORATORY CONTROL SAMPLE (LCS) ANALYSES

LCS analyses were within the windows established for TOC.

# SUMMARY

The data generated by the analysis of these samples can be used without qualifications.

Please call David A Thomson at SCAN 744-4737 to further discuss this project.

11/92 - 5/93

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#### STATE OF WASHINGTON

# DEPARTMENT OF ECOLOGY

MANCHESTER ENVIRONMENTAL LABORATORY

7411 Beach Drive East • Port Orchard, Washington 98366-8204 • (206) 871-8860 • SCAN 871-8860

July 7, 1993

TO: Project Officer

FROM: David A Thomson

SUBJECT: Quality Assurance memo for the Bremerton/Bellingham Storm Drains TOC Results

# SAMPLE RECEIPT Commencement Bue

The samples from the Bremerton/Bellingham Storm Drains were received by the Manchester Laboratory on May 28, 1993 in good condition. The analyses for these samples were subsequently contracted to Analytical Resources Inc. The samples were run using the Puget Sound Estuary Program (PSEP1986) for TOC.

#### HOLDING TIMES

All analyses were performed within 30 days of sample collection.

# PROCEDURAL BLANKS

The procedural blanks associated with these samples showed no analytically significant levels of analytes.

# PRECISION DATA

Two samples were run in duplicate to evaluate precision on this sample set. The Relative Percent Difference (RPD) for all analytes was within the +/-10% window for duplicate analysis.

# SPIKED SAMPLE ANALYSIS

Spike and duplicate spike analysis were performed on two samples. All spike recoveries were within limits of 100 + 1/- 30%.

# LABORATORY CONTROL SAMPLE (LCS) ANALYSES

LCS analyses were within the windows established for TOC.

#### SUMMARY

The data generated by the analysis of these samples can be used noting the data qualifications discussed in this memo.

Please call David A Thomson at SCAN 871-8822 to further discuss this project.

Toc 6/93-12/93

# State of Washington Department of Ecology Manchester Environmental Laboratory 7411 Beach Dr. East Port Orchard WA. 98366

February 16, 1994

Project:	Commencement Bay Sediment Traps
Samples:	028502 through 028582

Laboratory:

Weyerhaeuser Analytical and Testing Services

13807

Karin Feddersen K By:

These samples were received at the Manchester Laboratory on January 10, 1994, and were sent to Weyerhaeuser Analytical and Testing Services on January 12, 1994 for TOC analysis using PSEP.

# HOLDING TIMES

The holding time for frozen sediments is six (6) months. There have been no studies performed to indicate the effect of holding time after the samples have been thawed for analysis. Therefore an evaluation of the results with regard to holding time is not feasible. All samples were stored in the proper containers at 4 degrees C after being thawed. All analyses were performed within twenty-nine (29) days of thawing.

# PROCEDURAL BLANKS

The procedural blanks associated with these samples demonstrated that the processes were free from contamination.

# INITIAL CALIBRATION

The % Relative Standard Deviation (%RSD) was within QC limits of  $\leq 20\%$ .

# **CHECK STANDARDS**

All Check Standard recoveries are reasonable, acceptable, and within QC limits of 90% to 100%.

# **DUPLICATES**

The Relative Percent Difference (RPD) of the duplicate analyses to the original analyses are within QC limits of  $\leq 10\%$ .

## SUMMARY

This data is acceptable for use as amended.

Page 1 of 1

TOC 12/93-5/94

State of Washington Department of Ecology Manchester Environmental Laboratory 7411 Beach Dr. East Port Orchard WA. 98366 June 22, 1994

#### Project: Commencement Bay

Samples: 228503, 228504, 228507, 228510, 228513, 228516, 228519, 228522, 228525, 228528, 228531, 228540, 228544, 228545, 228548, 228551, 228554, 228557, 228560, 228560, 228566, 228569, 228572, 228575, 228578, 228581

Laboratory: Sound Analytical Services, Inc. 40700

By: Karin Feddersen KF

These samples were received at the Manchester Laboratory on May 31, 1994, and were sent to Sound Analytical Services, Inc. on June 1, 1994 for TOC analysis using the following method: Puget Sound Estuary Program

# DATA QUALIFIER DEFINITIONS

U - The analyte was not detected at or above the reported result.

# **HOLDING TIMES**

All analyses were performed within the method holding times of six (6) months from the date of collection.

#### PROCEDURAL BLANKS

The procedural blanks associated with these samples have demonstrated that the process is free from contamination.

# CHECK STANDARDS

All recoveries were within QC limits of +/- 20%.

#### TRIPLICATE

Samples 228503 and 228560 were run in triplicate. The triplicate analyses are in acceptable agreement with the original analyses. The Relative Percent Differences (RPD) were within the +/-20% window for triplicate analyses.

## SUMMARY

For consistency with Manchester Environmental Laboratory reporting requirements, all non-detect values have been qualified with a "U". This data is acceptable for use as amended.

Patientates 5/au-11/acr

State of Washington Department of Ecology Manchester Environmental Laboratory 7411 Beach Dr. East Port Orchard WA. 98366 February 3, 1995

Project:	Commencement Bay	
Samples:	518132, 35, 40, 43, 47, 50, 53, 56, 60, 63, 68, 69, 72, 76, 79, 84, 87, 91, 94, 98 518201, 518206, 518207	3,

Laboratory: Sound Analytical Services, Inc. 45364

By: Karin Feddersen KF

These samples were received at the Manchester Environmental Laboratory (MEL) on December 23, 1994, and were sent to Sound Analytical Services, Inc. on December 23, 1994 for TOC analysis using the following method: Puget Sound Estuary Program.

# **HOLDING TIMES**

The PSEP TOC holding time for frozen sediments is six (6) months. It is unknown how holding time affects samples that have not been stored frozen prior to analysis. No studies to my knowledge have been performed to indicate whether there is any effect. Therefore, an evaluation of the results with regard to holding time is not feasible. The samples were stored in the proper containers at 4 degrees C until analysis.

#### PROCEDURAL BLANKS

The procedural blanks associated with these samples have demonstrated that the process is free from contamination.

# CHECK STANDARDS

All recoveries were within QC limits of +/- 20% of the expected values.

## TRIPLICATE

Samples 518132 and 518187 were analyzed in triplicate. The Relative Standard Deviation (RSD) was within 20% for all samples.

#### SUMMARY

The result for sample 518135 exceeded the concentration of the highest standard analyzed with these samples. The result for this sample has therefore been qualified with a "J" (the reported result is an estimated quantity).

For consistency with MEL reporting protocol, all non-detect values have been qualified with a "U" (the analyte was not detected at or above the reported result). This data is acceptable for use as amended.

Bottom Secturety 11/94 Grain Size

State of Washington Department of Ecology Manchester Environmental Laboratory 7411 Beach Dr. East Port Orchard WA. 98366

December 28, 1994

Project: Commencement Bay

Samples: 49-8180-81,84-88, 90-98

Pam Covey

Laboratory: Soil Technology

By:

# Case Summary

The Commencement Bay samples required sixteen (16) Grain Size analyses on sediment using Puget Sound Esturary Program Portocol with wet preparation.

These samples were received at the Manchester Environmental Laboratory on December 5, 1994 and transported to Soil Technology on December 8, 1994 for Grain Size analyses. These analyses were reviewed for qualitative and quantitative accuracy, validity and usefulness.

The results are acceptable for use as reported.

Bottom Sectured

State of Washington Department of Ecology Manchester Environmental Laboratory 7411 Beach Dr. East Port Orchard WA. 98366 January 11, 1995

# Project: Commencement Bay

Samples: 498180, 498181, 498184 through 498188, and 498190 through 498198

Laboratory: Sound Analytical Services, Inc. 45009

By: Karin Feddersen KF

These samples were received at the Manchester Laboratory on December 5, 1994, and were sent to Sound Analytical Services, Inc. on December 8, 1994 for TOC analysis using the following method: Puget Sound Estuary Program

# DATA QUALIFIER DEFINITIONS

U - The analyte was not detected at or above the reported result.

## HOLDING TIMES

The PSEP TOC holding time for frozen sediments is six (6) months. No studies to my knowledge have been performed to indicate the effect of holding time on samples that have not been stored frozen prior to analysis. Therefore an evaluation of the results with regard to holding time is not feasible. The samples were stored in the proper containers at 4 degrees C until analysis.

# PROCEDURAL BLANKS

The procedural blanks associated with these samples have demonstrated that the process is free from contamination.

#### CHECK STANDARDS

All recoveries were within QC limits of +/- 20% of the expected values.

## TRIPLICATE

Sample 498180 was run in triplicate. The triplicate analyses are in acceptable agreement with the original analysis. The Relative Standard Deviation (RSD) was within 20%.

# SUMMARY

For consistency with Manchester Environmental Laboratory reporting requirements, all non-detect values have been qualified with a "U". This data is acceptable for use as amended.

11/94-6/95

State of Washington Department of Ecology Manchester Environmental Laboratory 7411 Beach Dr. East Port Orchard WA. 98366 August 8, 1995

# Project: Commencement Bay

Samples: 248702 through 2478086

Laboratory: Sound Analytical Services, Inc. 49648

By: Karin Feddersen KF

These samples were received at the Manchester Environmental Laboratory (MEL) on June 16, 1995, and were sent to Sound Analytical Services, Inc. on June 22, 1995, for TOC analysis using the following: Puget Sound Estuary Program.

# HOLDING TIMES

These samples were analyzed within the specified PSEP TOC holding time for frozen sediments of six (6) months.

# PROCEDURAL BLANKS

The procedural blanks associated with these samples have demonstrated that the process is free from contamination.

# CHECK STANDARDS

All recoveries were within QC limits of +/- 20% of the expected values.

# TRIPLICATE

Sample 248702 was analyzed in triplicate. The Relative Standard Deviation (RSD) was within 20%.

# SUMMARY

For consistency with MEL reporting protocol, all non-detect values have been qualified with a "U" (the analyte was not detected at or above the reported result). This data is acceptable for use as amended.

# Case Narratives

Metals Analysis

June 1991 to June 1995

Particulates June - Dec, 1991

# WASHINGTON STATE DEPARTMENT OF ECOLOGY ENVIRONMENTAL INVESTIGATIONS AND LABORATORY SERVICES MANCHESTER ENVIRONMENTAL LABORATORY

January 28, 1992

TO:

FROM:

Dale Norton Craig Smith

SUBJECT: Commencement Bay - Sediment Traps

#### SAMPLE RECEIPT

The samples were received by the Manchester Laboratory on 12/19/91, in good condition.

#### HOLDING TIMES

All analyses were performed within the specified holding time for Metals analysis (28 days for Hg, 180 days for all other metals).

#### INSTRUMENT CALIBRATION

Instrument calibration was performed before the run, and checked by initial calibration verification standards and blanks. Continuing calibration standards were analyzed at the frequency of 10% during the run, and again at the end of the analytical run.

#### **PROCEDURAL BLANKS**

The blank values were within the acceptable range and showed no analytically significant levels except for Sb. The blank value for Sb is significant, and a "B" qualifier will be attached to the data.

#### PRECISION DATA

The results of the duplicate samples were used to evaluate analytical precision related to the sample set. The RPD's for all parameters were within the +/-20% window of acceptance for duplicate analysis.

#### STANDARD REFERENCE MATERIAL

The analysis of the external verification standards for GFAA gave no unusual results, and were within the acceptable established criteria. The external reference materials for XRF analyses were within acceptable limits.

#### SUMMARY

During the XRF run, an additional external verification standard was run, USGSAGV-1. All of the values obtained were within acceptable limits except for Ni. This is the only reported SRM which bracketed the analytical run. PACS-1 and 1646 apparently were not analyzed at the beginning of the run. All of the PACS-1 and 1646 values were within acceptable limits. Caution might be called for with the Ni data.

The data generated by the analysis of the samples above may be used noting any stated qualifications.

If you have any questions concerning the results, please feel free to call me at SCAN 744-4737.



12/91-5/92

# WASHINGTON STATE DEPARTMENT OF ECOLOGY ENVIRONMENTAL INVESTIGATIONS AND LABORATORY SERVICES MANCHESTER ENVIRONMENTAL LABORATORY

August 27, 1992

TO: Dale Norton

FROM: Bill Kammin, Environmental\_Lab\_Director

SUBJECT: June 1992 Commencement Bay Metals Results

# SAMPLE RECEIPT

The samples from the June 1992 Commencement Bay Metals project were received by the Manchester Laboratory on 6/25/92 in good condition. The analytical work was subsequently contracted to Battelle/Sequim. Analytical methods were specified by the project officer and included: X-ray fluorescence for arsenic, copper, nickel, lead, and zinc; mercury by cold vapor generation, and cadmium and antimony by graphite furnace atomic absorption.

# **HOLDING TIMES**

All analyses were performed within the specified holding times for metals analysis (28 days for mercury, 180 days for all other metals).

# **INSTRUMENT CALIBRATION**

Instrument calibration was performed before each analytical run and checked by initial calibration verification standards and blanks. Continuing calibration standards and blanks were analyzed at a frequency approximately 10% during atomic absorption analytical runs. AA calibration gave correlation coefficients greater than the criteria of 0.995. A correlation coefficient of 0.995 or higher means that the calibration is acceptable.

# **PROCEDURAL BLANKS**

The procedural blanks associated with these samples showed detectable levels of mercury and antimony. Reporting limits were adjusted up in these instances, and for mercury the levels in the blanks were less than 10% of sample values, so no blank qualifiers were required. For antimony, six samples were between the adjusted reporting limit and the estimated reliable quantitation limit. These results were flagged P, B -- denoting possible blank influence on results, and results in an area of less-certain quantitation. Results from atomic absorption analyses were not background corrected.



5/92 - 11/92



STATE OF WASHINGTON

# DEPARTMENT OF ECOLOGY

MANCHESTER ENVIRONMENTAL LABORATORY

7411 Beach Drive East • Port Orchard, Washington 98366-8204 • (206) 895-4737 • SCAN 744-4737

March 22, 1993

TO: Dale Norton

FROM: Bill Kammin, Environmental\_Lab\_Director

SUBJECT: December 1992 Commencement Bay Metals Results

# SAMPLE RECEIPT

The samples from the December 1992 Commencement Bay Metals project were received by the Manchester Laboratory on 12/4/92 in good condition. The analytical work was subsequently contracted to Battelle/Sequim. Analytical methods were specified by the project officer and included: X-ray fluorescence for arsenic, copper, nickel, lead, and zinc; mercury by cold vapor generation, and cadmium and antimony by graphite furnace atomic absorption. The review of this data consisted of evaluation of QA/QC/reference material results, spot checks of calculations, and examination of data and final reports for transcription errors.

# HOLDING TIMES

All analyses were performed within the specified holding times for metals analysis (28 days for mercury, 180 days for all other metals).

# **INSTRUMENT CALIBRATION**

Instrument calibration was performed before each analytical run and checked by initial calibration verification standards and blanks. Continuing calibration standards and blanks were analyzed at a frequency approximately 10% during atomic absorption analytical runs. AA calibration gave correlation coefficients greater than the criteria of 0.995. A correlation coefficient of 0.995 or higher means that the calibration is acceptable. With regard to XRF analyses for copper, please find as an attachment information from Battelle Richland on problems with the XRF instrument. Because copper results in the reference materials were over 20% high, copper data is qualified as estimated (J).

# **PROCEDURAL BLANKS**

The procedural blanks associated with these samples showed no analytically significant blank levels. Results from atomic absorption analyses were not background corrected.

# **REFERENCE SAMPLE ANALYSIS**

Results from the analysis of two different reference samples were used to evaluate the accuracy of the results submitted by the contract lab. The reference materials used were NIST 1646, Estuarine Sediment, and the Canadian material, PACS-1. For all anlytes except copper and mercury, the mean results for the analysis of all reference materials were within the +/- 20% of the certified analyte value given by the reference material manufacturers. For copper and mercury, the mean results from the analysis of the above mentioned reference materials were at or outside the +/- 20% limit. For this reason, all mercury and cadmium results are qualified with J, indicating estimated values.

With the exception of cadmium results under 1 mg/kg, all results are reported to 3 significant figures. When using these numbers in a final report (particularly for mercury), it may be more appropriate to round to two significant figures.

#### SUMMARY

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The data generated by the analysis of the above referenced samples can be used with the above-mentioned qualifications.

If you have any questions about the results or the methods used to obtain these results, please call me at SCAN 744-4737.

11/92-5/93



STATE OF WASHINGTON

# DEPARTMENT OF ECOLOGY

MANCHESTER ENVIRONMENTAL LABORATORY

7411 Beach Drive East • Port Orchard, Washington 98366-8204 • (206) 895-4737 • SCAN 744-4737

July 28, 1993

TO: Dale Norton

FROM: Bill Kammin, Environmental\_Lab\_Director

SUBJECT: May 1993 Commencement Bay Metals Results

## SAMPLE RECEIPT

The samples from the May 1993 Commencement Bay Metals project were received by the Manchester Laboratory on 05/28/93 in good condition. The analytical work was subsequently contracted to Battelle/Sequim. Analytical methods were specified by the project officer and included: X-ray fluorescence for arsenic, copper, nickel, lead, and zinc; mercury by cold vapor generation, and cadmium and antimony by graphite furnace atomic absorption. The review of this data consisted of evaluation of QA/QC/reference material results, spot checks of calculations, and examination of data and final reports for transcription errors.

# HOLDING TIMES

All analyses were performed within the specified holding times for metals analysis (28 days for mercury, 180 days for all other metals).

# **INSTRUMENT CALIBRATION**

Instrument calibration was performed before each analytical run and checked by initial calibration verification standards and blanks. Continuing calibration standards and blanks were analyzed at a frequency approximately 10% during atomic absorption analytical runs. AA calibration gave correlation coefficients greater than the criteria of 0.995. A correlation coefficient of 0.995 or higher means that the calibration is acceptable.

# PROCEDURAL BLANKS

The procedural blanks associated with these samples showed no analytically significant blank levels. Results from atomic absorption analyses were not blank corrected.

# **REFERENCE SAMPLE ANALYSIS**

Results from the analysis of two different reference samples were used to evaluate the accuracy of the results submitted by the contract lab. The reference materials used were NIST 1646, Estuarine Sediment, and the Canadian material, PACS-1. Generally, results for these reference materials indicated acceptable data quality.

With the exception of cadmium results under 1 mg/kg, all results are reported to 3 significant figures. When using these numbers in a final report (particularly for mercury), it may be more appropriate to round to two significant figures.

# SUMMARY

The data generated by the analysis of the above referenced samples can be used with the above-mentioned qualifications.

If you have any questions about the results or the methods used to obtain these results, please call me at SCAN 744-4737.



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STATE OF WASHINGTON

# DEPARTMENT OF ECOLOGY

MANCHESTER ENVIRONMENTAL LABORATORY

7411 Beach Drive East • Port Orchard, Washington 98366-8204 • (206) 871-8860 • SCAN 871-8860

April 13, 1994

TO: Dale Norton

FROM: Bill Kammin, Environmental\_Lab\_Director

SUBJECT: January 1994 Commencement Bay Metals Results

# SAMPLE RECEIPT

The samples from the January 1994 Commencement Bay Metals project were received by the Manchester Laboratory on 1/10/94 in good condition. The analytical work was subsequently contracted to Battelle/Sequim. Analytical methods were specified by the project officer and included: X-ray fluorescence (XRF) for arsenic, copper, nickel, lead, and zinc; mercury by cold vapor generation followed by atomic fluorescence (CVAF), and cadmium and antimony by graphite furnace atomic absorption (GFAA). The review of this data consisted of evaluation of QA/QC/reference material results, spot checks of calculations, and examination of data and final reports for transcription errors.

# HOLDING TIMES

All analyses were performed within the specified holding times for metals analysis (28 days for mercury, 180 days for all other metals).

# INSTRUMENT CALIBRATION

XRF calibration was evaluated by the analysis of the following reference materials:

1. PACS-1 2. NIST 1646 3. MESS-1

All XRF recoveries for these reference materials were within +/- 20% of the certified values, with the following exception: arsenic in PACS-1. These results ranged from 22% to 28% high. No qualification of the XRF data was necessary.

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The GFAA data was of typical interest. The calibration regime used by Battelle is highly unusual. It consists of the following steps:

- Analysis of NIST 1646 by the Method of Standard Additions (MSA).
- Then, samples are analyzed and analyte response is equated to the analyte response for 1646 on the MSA curve.

This technique might be acceptable if 1646 was very similar to the sediment trap samples. However, NIST reference material 1646 is a highly processed, very homogeneous material with very low organic content. The sediment trap samples generally have very high organic content. In the absence of any spike recovery data for the sediment trap samples, it is impossible to know how well analytes are being recovered by Battelle's method. The recoveries of these analytes do seem reasonable, but how accurate these GFAA results are is really unknown.

These GFAA calibration techniques do not appear in any USEPA methodology, and carry no regulatory approval.

Continuing calibration standards and blanks were analyzed at a frequency approximately 10% during atomic absorption analytical runs. GFAA reference material results were within +/-20% of the certified values.

For mercury, reference material recoveries were within +/-20% of the certified values. A sample independent calibration was performed.

## PROCEDURAL BLANKS

The procedural blanks associated with these samples showed no analytically significant blank levels. Results from atomic absorption analyses were not blank corrected.

#### SUMMARY

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The data generated by the analysis of the above referenced samples can be used with the above-mentioned qualifications.

If you have any questions about the results or the methods used to obtain these results, please call me at SCAN 744-4737.

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#### STATE OF WASHINGTON

# DEPARTMENT OF ECOLOGY

MANCHESTER ENVIRONMENTAL LABORATORY

7411 Beach Drive East • Port Orchard, Washington 98366-8204 • (206) 871-8860 • SCAN 871-8860

August 29, 1994

TO: Dale Norton

FROM: Bill Kammin, Environmental\_Lab\_Director

SUBJECT: June 1994 Commencement Bay Metals Results

# SAMPLE RECEIPT

The samples from the June 1994 Commencement Bay Metals project were received by the Manchester Laboratory on 1/10/94 in good condition. The analytical work was subsequently contracted to Battelle/Sequim. Analytical methods were specified by the project officer and included: X-ray fluorescence for arsenic, copper, nickel, lead, and zinc, and mercury by cold vapor generation. The review of this data consisted of evaluation of QA/QC/reference material results, spot checks of calculations, and examination of data and final reports for transcription errors.

### HOLDING TIMES

All analyses were performed within the specified holding times for metals analysis (28 days for mercury, 180 days for all other metals), with the following exception: mercury. Mercury was analyzed 29 days after the requested analysis date of 6/20/94. Mercury data is qualified with J, denoting estimated values.

#### **INSTRUMENT CALIBRATION**

Atomic fluorescence calibration gave correlation coefficients greater than the criteria of 0.995. A correlation coefficient of 0.995 or higher means that the calibration is acceptable. Continuing calibration standards and blanks were analyzed at a frequency approximately 10% during atomic fluorescence analytical runs. Results for the analysis of these standards were acceptable.

#### PROCEDURAL BLANKS

The procedural blanks associated with these samples showed no analytically significant blank levels. Results from atomic fluorescence analyses were not blank corrected.

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# **REFERENCE SAMPLE ANALYSIS**

Results from the analysis of two different reference samples were used to evaluate the accuracy of the results submitted by the contract lab. The reference materials used were NIST 1646, Estuarine Sediment, and the Canadian material, PACS-1. Generally, results for these reference materials indicated acceptable data quality. For mercury, the analysis of 1646 showed high recoveries (135% and 143%). However, analysis of PACS-1 and BEST-1 showed acceptable recoveries

# SUMMARY

The data generated by the analysis of the above referenced samples can be used with the above-mentioned qualifications.

If you have any questions about the results or the methods used to obtain these results, please call me at SCAN 744-4737.



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#### STATE OF WASHINGTON

# DEPARTMENT OF ECOLOGY

MANCHESTER ENVIRONMENTAL LABORATORY

7411 Beach Drive East • Port Orchard, Washington 98366-8204 • (206) 871-8860 • SCAN 871-8860

March 17, 1995

TO: Dale Norton, Project Officer

FROM: Myrna McIntosh, Metals Chemist

SUBJECT: Metals Quality Assurance memo for the Commencement Bay Project

# QUALITY ASSURANCE SUMMARY

Data quality for this project is good. No significant quality assurance issues are noted for mercury analysis, ICP analysis, or graphite furnace (GFAA) analysis.

## SAMPLE INFORMATION

The samples from the Week 51 Commencement Bay Sediment project were received by the Manchester Laboratory on 12/23/94 in good condition.

# HOLDING TIMES

All analyses were performed within the USEPA Contract Laboratory Program (CLP) holding times for metals analysis (28 days for mercury, 180 days for all other metals).

## **INSTRUMENT CALIBRATION**

Instrument calibration was performed before each analytical run and checked by initial calibration verification standards and blanks. Continuing calibration standards and blanks were analyzed at a frequency of 10% during the run and again at the end of the analytical run. All initial and continuing calibration verification standards were within the relevant USEPA (CLP) control limits. AA calibration gave a correlation coefficient (r) of 0.995 or greater, also meeting CLP calibration requirements.

# PROCEDURAL BLANKS

The procedural blanks associated with these samples show no analytically significant levels of analytes.

# SPIKED SAMPLE ANALYSES

Spiked and duplicate spiked sample analyses were performed on this data set. All spike recoveries are within the CLP acceptance limits of +/-25%.

# PRECISION DATA

The results of the spiked and duplicate spiked samples are used to evaluate precision on this sample set. The Relative Percent Difference (RPD) for all analytes is within the 20% CLP acceptance window for duplicate analysis.

# LABORATORY CONTROL SAMPLE (LCS) ANALYSES

LCS analyses are within the windows established for each parameter. Our normal LCS is a sediment from ERA (Environmental Resource Associates) and this was run with the samples and recovered well. NIST 1646 was analyzed in duplicate for GFAA and ICP and in triplicate for mercury. It gave approximately 45% recovery of arsenic. Because our normal LCS gave good recovery, the sample results are not qualified.

Please call Bill Kammin at SCAN 206-871-8801 to further discuss this project.

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Washington State Department of Ecology Manchester Environment Laboratory

February 24, 1995

TO: Dale Norton

FROM: Bill Kammin, Environmental\_Lab\_Director

SUBJECT: Metals Quality Assurance memo for the Week 49 Commencement Bay Project

# QUALITY ASSURANCE SUMMARY

Both the contract data and the in-house data showed few quality assurance issues. The contract mercury data showed high recoveries for SRM 1646, and this data may be biased high. Split sample results showed generally good agreement, with the following exception: nickel. Nickel results were significantly higher (150 mg/Kg vs. 40 mg/Kg) by digestion and analysis by ICP compared to X Ray Fluorescence (XRF). However, quality assurance results for both techniques were in specification. It is possible that this sample has an interference affecting one of the analytical techniques.

# SAMPLE INFORMATION

These samples from the Week 49 Commencement Bay project were received by the Manchester Laboratory on 12/5/94 in good condition. All ICP and arsenic results are reported on a dry weight basis. Mercury results are reported on a wet weight basis.

# HOLDING TIMES

All analyses were performed within the USEPA Contract Laboratory Program (CLP) holding times for metals analysis (28 days for mercury, 180 days for all other metals).

# INSTRUMENT CALIBRATION

Instrument calibration was performed before each analytical run and checked by initial calibration verification standards and blanks. Continuing calibration standards and blanks were analyzed at a frequency of 10% during the run. All initial and continuing calibration verification standards were within the relevant USEPA (CLP) control limits. AA calibration gave a correlation coefficient (r) of 0.995 or greater, also meeting CLP calibration requirements.

### PROCEDURAL BLANKS

The procedural blanks associated with these samples showed no analytically significant levels of analytes. These samples were digested by our normal sediment digestion technique with one modification: 200 microliters of hydrofluoric acid was added to the digestion mix to help solubilize mineralized analytes.

### SPIKED SAMPLE ANALYSES

### Manchester Results

Spike and duplicate spike sample analyses were performed on this data set. All spike recoveries were within the CLP acceptance limits of +/- 25%.

### Battelle Results

No spikes were performed for XRF, mercury or cadmium analyses. It is not possible to assess matrix interference for this data.

# PRECISION DATA

#### Manchester Results

The results of the spike and duplicate spike samples were used to evaluate precision on this sample set. The Relative Percent Difference (RPD) for all analytes was within the 20% CLP acceptance window for duplicate analysis.

#### Battelle Results

Based on recoveries of Standard Reference Materials, precision met CLP specification for all results except mercury. Mercury precision results did not meet CLP criteria. Battelle mercury results are qualified J, denoting estimated values.

# LABORATORY CONTROL SAMPLE (LCS) AND STANDARD REFERENCE MATERIAL (SRM) ANALYSES

#### Manchester Results

LCS (Environmental Resource Associates soil sample) analyses were within the windows established for each parameter. NIST SRM 1646 was analyzed for ICP and arsenic. For ICP, NIST 1646 recoveries ranged from 72-99%. For arsenic, NIST 1646 recoveries were both 78%. For mercury, recoveries for multiple analyses of NIST 2704 ranged from 93-95%. Precision for all LCS and SRM analyses was within CLP acceptance criteria.

#### Battelle Results

Analyses of all SRM's was acceptable, with one exception: mercury. NIST 1646 showed very high recoveries and poor precision. Battelle mercury results are qualified J, denoting estimates.

Please call Bill Kammin at SCAN 206-871-8801 to further discuss this project.

### WRK:wrk

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Mussel Tissue 3.5/95

STATE OF WASHINGTON

# DEPARTMENT OF ECOLOGY

MANCHESTER ENVIRONMENTAL LABORATORY

7411 Beach Drive East • Port Orchard, Washington 98366-8204 • (360) 871-8860 • FAX (360) 871-8850

July 31, 1995

TO: Dale Norton, Project Officer

FROM: Myrna McIntosh, Metals Chemist

SUBJECT: Metals Quality Assurance memo for the Commencement Bay Project Sediment Sample Numbers: 95248730 - 95248786 Tissue Sample Numbers: 95248788 - 95248791

# QUALITY ASSURANCE SUMMARY

Data quality for this project is generally good. None of the sediment samples have any QA issues. There is a small amount of copper in the tissue digestion blank. Because the amount of copper was low in the samples it was necessary to qualify these results as estimates, "J".

The arsenic recoveries for the tissue spikes and LCS were low and so these arsenic results are qualified with "J" as estimates.

# SAMPLE INFORMATION

The samples from the Commencement Bay project were received by the Manchester Laboratory on 6/16/95 in good condition.

# HOLDING TIMES

All analyses were performed within the USEPA Contract Laboratory Program (CLP) holding times for metals analysis (28 days for mercury, 180 days for all other metals).

# INSTRUMENT CALIBRATION

Instrument calibration was performed before each analytical run and checked by initial calibration verification standards and blanks. Continuing calibration standards and blanks were analyzed at a frequency of 10% during the run and again at the end of the analytical run. All initial and continuing calibration verification standards were within the relevant USEPA (CLP) control limits. AA calibration gave a correlation coefficient (r) of 0.995 or greater, also meeting CLP calibration requirements.

# PROCEDURAL BLANKS

The procedural blanks from the tissue digestion show a trace of copper contamination. The copper level of the samples is low (within ten times the detection limit) so the copper results are qualified with "J" as estimates.

# SPIKED SAMPLE ANALYSES

Spiked and duplicate spiked sample analyses were performed on this data set. All spike recoveries from the sediment digestion are within the CLP acceptance limits of +/-25%. Arsenic recovery in the tissue samples is low at approximately 35%.

# PRECISION DATA

The results of the spiked and duplicate spiked samples are used to evaluate precision on this sample set. The Relative Percent Difference (RPD) for all analytes is within the 20% CLP acceptance window for duplicate analysis.

# LABORATORY CONTROL SAMPLE (LCS) ANALYSES

LCS analyses are within the windows established for each parameter, with the exception of arsenic in the tissue LCS. The tissue arsenic results are qualified "J", as estimates. The certified value for lead in the tissue LCS is too low to be useful as a control.

Please call Bill Kammin at SCAN 360-871-8801 to further discuss this project.

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# Case Narratives

Organics Analysis

June 1991 to June 1995

7411 Beach Drive E, Port Orchard Washington 98366

# CASE NARRATIVE

# March 6, 1992

Subject: Sediment Traps - Commencement Bay

Samples: 91 - 518524 to 518527, - 518533, - 518539, - 518543, - 518546, - 518550 to 518552 - 518555, - 518556, -518560, -518561, -518567, -518569 and -518572.

Case No. DOE-020Q

Officer: Dale Norton

By: Dickey D. Huntamer Organics Analysis Unit

### SEMIVOLATILE ORGANICS

#### ANALYTICAL METHODS:

The semivolatile soil samples were Soxhlet extracted with acetone following the Manchester modification of the EPA CLP and SW 846 8270 procedure with capillary GC/MS analysis of the sample extracts. Normal QA/QC procedures were performed with the analyses.

### HOLDING TIMES:

The sediment samples were composited samples acquired over a period of time. The samples were stored frozen between collections. After compositing and prior to analysis the samples were kept frozen following Puget Sound Estuary Program guidelines. The samples were thawed, extracted and analyzed within the recommended holding times. No data qualifiers were added due to holding times.

### **BLANKS:**

Low levels of some target compounds were detected in the laboratory blanks. The EPA five times rule was applied to all target compounds which were found in the blank. Compounds that were found in the sample and in the blank were considered real and not the result of contamination if the levels in the sample are greater than or equal to five times the amount of compounds in the associated method blank.

### SURROGATES:

The normal CLP surrogates were added to the sample prior to extraction. Most surrogate spike recoveries were within acceptable QC limits except for six samples which each had one out of the recommended limits. All surrogates were out in sample, 518524 due to interferences and problems with spiking. Consequently all the data is reported as "REJ", rejected except for the tentatively identified compounds. Sample data was salvaged by using one of the matrix spikes which serendipitously was not spiked with the target compounds but had the surrogates added. This is reported as a duplicate analysis for 518524.

### MATRIX SPIKE AND MATRIX SPIKE DUPLICATE:

Three sets of matrix spikes were analyzed with these samples. These were on samples 518524, 518533 and 518555. One of the matrix spike samples for 518524 was not available due to a problem with spiking. All of the spikes had problems with matrix interferences and the high native concentrations of some target compounds complicated calculating the corrected recoveries. This was particularly true for sample 518533.

### SPECIAL ANALYTICAL PROBLEMS:

The requirement for low level analysis resulted in analysis of the smallest possible extract volume. This caused rapid deterioration of the capillary column resulting in frequent column reconditioning and some column changes. Most of the samples exhibited large "humps" of eluting material loosely referred to as "biogenic yuck", which appeared to consist of mixtures of fatty acid, steroids, assorted biological molecules along with the usual petroleum hydrocarbons. The tentatively identified compounds reported consisted primarily of unidentifiable hydrocarbons or compounds along with sterols and fatty acids. In sample 518543 some possible resin acids were also detected.

A Canadian reference sample was analyzed in duplicate with this sample set. 518572 and 518572D. These were analyzed as blind samples without having the "true" values and we would appreciate receiving a copy of the actual values for comparison.

The data is acceptable for use as qualified.

# DATA QUALIFIER CODES:

U	-	The analyte was not detected at or above the reported value.
l	-	The analyte was positively identified. The associated numerical value is an estimate.
UJ	-	The analyte was not detected at or above the reported estimated result.
REJ	-	The data are <u>unusable</u> for all purposes.
EXP		The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .
NAF	-	Not analyzed for.
Ň	<b></b>	For organic analytes there is evidence the analyte is present in this sample.
NJ	-	There is evidence that the analyte is present. The associated numerical result is an estimate.
E	-	This qualifier is used when the concentration of the associated value exceeds the known calibration range.
*	-	The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)

7411 Beach Drive SE, Port Orchard Washington 98366

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# CASE NARRATIVE

### February 4, 1992

 Subject:
 Sediment Traps - Commencement Bay

 Samples:
 91 - 518533, -518539, -518543, -518546, -518550, -518551, and -518571

 Case No.
 DOE-020Q

Officer: Dale Norton

By:

Dickey D. Huntamer Organics Analysis Unit

# POLYCHLORINATED BIPHENYLS-PCB

# ANALYTICAL METHODS:

The sediment trap samples were Soxhlet extracted using acetone as the solvent. Analysis was done by EPA Method 8080 using dual column capillary GC analysis with Electron Capture Detectors (ECD).

### HOLDING TIMES:

The sediment samples were composited samples acquired over a period of time. The samples were stored frozen between collections. After compositing and prior to analysis the samples were kept frozen following Puget Sound Estuary Program guidelines. The samples were thawed, then analyzed within the recommended 14 day holding time. No data qualifiers were added due to holding times.

### BLANKS:

No target compounds were detected in the laboratory blanks.

#### SURROGATES:

Soil surrogate recoveries ranged from 79% to 189% for decachlorobiphenyl (DCB) and 77% to 120% for 4,4'dibromooctafluorobiphenyl (DBFB). All surrogate spike recoveries for DBFB were within acceptable QC limits. Three surrogate recoveries for DCB exceeded 150%. These high values were most likely caused by interference from coeluting peaks. No additional data qualifiers were added based on surrogate recoveries.

Page 2 Sediment Traps - Commencement Bay - PCB

# MATRIX SPIKE AND MATRIX SPIKE :

The soil matrix spikes recoveries ranged from 84% to 118%. The Relative Percent Differences (RPD) ranged from 1% to 5%. All recoveries and RPD were within acceptable QC limits.

# SPECIAL ANALYTICAL PROBLEMS:

There were no significant problems with the PCB analysis. All results are reported on a dry weight basis.

Sample Number	% Solids
91- 518533	43.8
91 - 518539	39.5
91 - 518543	43.5
91 - 518546	45.5
91 - 518550	51.7
91 - 518551	37.0
91 - 518571	100

# DATA QUALIFIER CODES:

U	-	The analyte was not detected at or above the reported value.
1	-	The analyte was positively identified. The associated numerical value is an estimate.
IJ	-	The analyte was not detected at or above the reported estimated result.
REJ	-	The data are <u>unusable</u> for all purposes.
EXP	•	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .
NAF	-	Not analyzed for.
N	-	For organic analytes there is evidence the analyte is present in this sample.
NJ	-	There is evidence that the analyte is present. The associated numerical result is an estimate.
E	-	This qualifier is used when the concentration of the associated value exceeds the known calibration range.
*	-	The analyte was present in the sample. (Visual Aid to locate detected

compound on report sheet.)

7411 Beach Drive E, Port Orchard Washington 98366

# CASE NARRATIVE

### March 6, 1992

Subject: Sediment Traps - Commencement Bay

Samples: 91 - 518540 and 518543

Case No. DOE-020Q

Officer: Dale Norton

By:

Dickey D. Huntamer

# POLYCHLORINATED BIPHENYLS (PCB)

### ANALYTICAL METHODS:

Both samples were Soxhlet extracted using acetone as the solvent. Sample 518540 was extracted for PCB and 518543 was extracted for semivolatile organics. Consequently no PCB surrogate compounds were added to 518543. Analysis was done by EPA Method 8080 using dual column capillary GC analysis with Electron Capture Detectors (ECD).

### HOLDING TIMES:

Both samples were extracted within the recommended holding times. Sample 518540 was analyzed for PCB within the extract holding time but sample 518543 was analyzed seven days past the recommended 40 day holding time. Given the long term stability of the PCB this should have no discernible effect upon the results.

#### BLANKS:

No target compounds were detected in the laboratory blanks.

### SURROGATES:

All surrogate spike recoveries were within acceptable QC limits for 518540. No PCB surrogates were present in 518543 since the extract for the semivolatile analysis was used.

# SPECIAL ANALYTICAL PROBLEMS:

Confusion on the desired tests requested resulted in a failure to analyzed sample 518543 for PCB. It was analyzed for semivolatiles but was also intended to be the duplicate sample for 518540 (PCB only). Consequently the semivolatile extract was analyzed for PCB since it involves the same extraction procedure. No PCB surrogate recoveries are available for the semivolatile extract, but the semivolatile surrogate recoveries are available in the semivolatile data. The results are listed in the table below along with the Relative Percent Difference (RPD). The data is acceptable for use as qualified.

PCB	1254	1260	RPD
518540	540 ug/Kg	670 ug/Kg	43%
518543	350 ug/Kg	450 ug/Kg	39%

# DATA QUALIFIER CODES:

U	••	The analyte was not detected at or above the reported value.
J	-	The analyte was positively identified. The associated numerical value is an estimate.
UJ	-	The analyte was not detected at or above the reported estimated result.
REJ		The data are <u>unusable</u> for all purposes.
EXP	-	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .
NAF	-	Not analyzed for.
N	-	For organic analytes there is evidence the analyte is present in this sample.
NJ	-	There is evidence that the analyte is present. The associated numerical result is an estimate.
Е	-	This qualifier is used when the concentration of the associated value exceeds the known calibration range.
*	-	The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)

Particulates 12/91-5/92

7411 Beach Drive E, Port Orchard Washington 98366

# CASE NARRATIVE

#### September 23, 1992

Subject: Commencement Bay Sediment Traps

Samples: 92 - 268433, -268444, -268465 to -268467

Case No. DOE-370Y

Officer: Dale Norton

By:

Dickey D. Huntamer (C) Organics Analysis Unit

# VOLATILE ORGANIC ANALYSIS

### ANALYTICAL METHODS:

Volatile organic compounds were analyzed using Manchester modification of the EPA SW 846 Method 8240 purge-trap procedure with capillary GC/MS analysis. Normal QA/QC procedures were performed on the samples.

#### BLANKS:

Low levels of the common laboratory solvents acetone and methylene chloride were detected in the laboratory blanks The EPA five times rule was applied to all target compounds which were found in the blank. Compounds that were found in the sample and in the blank were considered real and not the result of contamination if the levels in the sample are greater than or equal to five times the amount of compounds in the associated method blank.

### SURROGATES:

Surrogate recoveries were within acceptable limits for the soil samples.

### HOLDING TIMES:

The soil samples were analyzed within the recommended 14 day holding time.

### MATRIX SPIKE AND MATRIX SPIKE DUPLICATE:

Soil matrix spikes were within acceptable QC limits for both percent recovery and Relative Percent Difference (RPD) for most compounds. Four compounds, trans-1,3-dichloropropene, cis-1,3-dichloropropene, 4-methyl-2-pentanone and 1,2,3-trichlorobenzene had spike recoveries outside acceptable limits and the results for these compounds in sample 268465 were qualified with the "J" flag.

Page 2 Commencement Bay Sediment Traps - VOA

### SPECIAL ANALYTICAL PROBLEMS:

No analytical problems were encountered in the analysis. The data is acceptable for use as qualified.

# DATA QUALIFIER CODES:

The analyte was not detected at or above the reported value. U The analyte was positively identified. The associated numerical value is an J estimate. The analyte was not detected at or above the reported estimated result. UJ The data are unusable for all purposes. REJ The result is equal to the number before EXP times 10 to the power of the EXP number after EXP. As an example 3EXP6 equals  $3 \times 10^6$ . NAF Not analyzed for. For organic analytes there is evidence the analyte is present in this sample. Ν There is evidence that the analyte is present. The associated numerical result NJ is an estimate. This qualifier is used when the concentration of the associated value exceeds Ε the known calibration range. The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)

7411 Beach Drive E, Port Orchard Washington 98366

# CASE NARRATIVE

#### October 8, 1992

Subject: Com

Commencement Bay Sediment Traps

Samples: 92 - 268426 to -268430, -268433, -268436, -268444, -268452, -268463 to 268467, -268494 to -268498.

Case No. DOE-370Y

Officer: Dale Norton

By: Dickey D. Huntamer OCC Organics Analysis Unit

# SEMIVOLATILE ORGANICS

### ANALYTICAL METHODS:

The semivolatile soil samples were Soxhlet extracted with acetone following the Manchester modification of the EPA SW 846 8270 procedure with capillary GC/MS analysis of the sample extracts. Normal QA/QC procedures were performed with the analyses.

# HOLDING TIMES:

All sample and extraction holding times were within the recommended limits.

### **BLANKS:**

Low levels of some target compounds were detected in the laboratory blanks. The EPA five times rule was applied to all target compounds which were found in the blank. Compounds that were found in the sample and in the blank were considered real and not the result of contamination if the levels in the sample are greater than or equal to five times the amount of compounds in the associated method blank.

### SURROGATES:

The normal surrogates compounds were added to the sample prior to extraction. Only three surrogate spike recoveries were outside acceptable QC limits all of the others were acceptable. Four samples, blanks BS2189 and BS2189D along with samples 92-268396 and -268397 (Canadian reference material HR-6), under went the silica gel polynuclear aromatic hydrocarbon (PAH) clean-up procedure. Consequently only surrogate recoveries for 2-fluorobiphenyl, d10-pyrene and d14-terphenyl are valid.

# MATRIX SPIKE AND MATRIX SPIKE DUPLICATE:

Matrix spike recoveries and Relative Percent Differences (RPD) were acceptable for most of the Contract Laboratory Program (CLP) check compounds except for phenanthrene and pyrene. Both of these compounds were found in the sample at relatively high levels and correction of the matrix spikes could not be made. Spike recoveries for most of the remaining non-CLP compounds were also affected by the high native concentrations and could not be calculated.

### SPECIAL ANALYTICAL PROBLEMS:

No special analytical problems were encountered in the semivolatile analyses. The HR-6 reference material samples, -268396 and -97 were mistakenly given the silica gel clean-up procedure. Since HR-6 is primarily certified for the PAH compounds this should not be a major problem and the results should still be valid.

A smaller sample size than normal, five to 10 grams, was used in the extraction. The small sample size was compensated for by concentrating to a lower volume for injection. This resulted in similar sample quantitation limits compared to previous analyses. Analysis and review at these low levels was complicated by the large amounts of interfering biogenic material which affected the chromatography.

In all samples except, 268427, 268433, 268463 and 268465, results for benzo(b)fluoranthene and benzo(k)fluoranthene could not resolved. Consequently the results were reported as benzo(b) fluoranthene but includes benzo(k)fluoranthene as well and the results for benzo(k)fluoranthene were flagged as not detected, "U".

Due to interferences in the samples several internal standard area counts were outside the acceptable limits. Those samples and compounds affected by this were qualified by adding the "J" qualifier.

# DATA QUALIFIER CODES:

U	•	The analyte was not detected at or above the reported value.
J	-	The analyte was positively identified. The associated numerical value is an estimate.
យ	-	The analyte was not detected at or above the reported estimated result.
REJ	-	The data are <u>unusable</u> for all purposes.
EXP	•	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .
NAF		Not analyzed for.
N	-	For organic analytes there is evidence the analyte is present in this sample.
NJ	-	There is evidence that the analyte is present. The associated numerical result is an estimate.
Е	-	This qualifier is used when the concentration of the associated value exceeds the known calibration range.
*	-	The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)

7411 Beach Drive E, Port Orchard Washington 98366

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# CASE NARRATIVE

### September 22, 1992

Subject:

Commencement Bay Sediment Traps

Samples:

92 - 268433, -268436, -268444, -268452, -268463 to -268465, -268467.

Case No. DOE-370Y

Officer: Dale Norton

By:

Dickey D. Huntamer

# POLYCHLORINATED BIPHENYLS

### ANALYTICAL METHODS:

The sediment trap samples were Soxhlet extracted using acetone as the solvent. Analysis was done by EPA Method 8080 using dual column capillary GC analysis with Electron Capture Detectors (ECD).

# HOLDING TIMES:

All sample extraction and analysis holding times were met.

# **BLANKS**:

No target compounds were detected in the laboratory blanks.

#### SURROGATES:

All surrogate spike recoveries were within acceptable QC limits. Surrogates ranged from 60% to 99% recovery.

Surrogate recovery for 4,4-Dibromooctafluorobiphenyl (DBOB) is not reported in the matrix spikes due to interference from the spiking compounds.

# MATRIX SPIKE AND MATRIX SPIKE :

The matrix spikes recoveries ranged from 84% to 89%. The Relative Percent Differences (RPD) ranged from 2.3% to 5.8%. All recoveries and RPD were within acceptable QC limits.

### SPECIAL ANALYTICAL PROBLEMS:

There were no significant problems with the Pesticide/PCB analysis. Several PCB's were detected in the samples. These included PCB-1242, PCB-1254 and PCB-1260. The station identification for one sample, -268433, was entered incorrectly. The correct station description is PUY-1C not DUY-1C.

Samples 92-268398 and -268399, labeled PCBREF and PCBREFD (duplicate), were EPA PCB in soil reference materials. The sample was a natural sediment characterized for two aroclors by the National Institute of Standards and Technology (NIST) and obtained from EPA-Cincinnati.

Table 1. Comparison of PCB concentrations in NIST sediment sample with results from EPA Region X inter-laboratory study and results from sediment trap analyses. All results are reported as ug/Kg.

Aroclor	NIST	EPA - Interlab *	92 -268398	92-268399
1242	8160 +/- 280	11,300 +/- 368	8000	. 7500
1260	6810 +/- 140	14,000 +/- 705 **	15,000	15,000

\* +/- one standard deviation

\*\* data from 33 laboratories.

The PCB-1242 sediment trap results were comparable to the NIST value and lower than the EPA-Interlab results. Results for PCB-1260 in the inter laboratory study were 2 to 2.5 times higher than the NIST value but were comparable to the inter-laboratory results. A conversation between Bob Rieck, EPA Manchester, Laboratory and Jim Longbottom, EPA, Cincinnati confirmed the disparity with the NIST reference values.

Page 3 Commencement Bay Sediment Traps - PCB

# DATA QUALIFIER CODES:

U	-	The analyte was not detected at or above the reported value.
J	-	The analyte was positively identified. The associated numerical value is an estimate.
UJ	-	The analyte was not detected at or above the reported estimated result.
REJ	-	The data are <u>unusable</u> for all purposes.
EXP	-	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .
NAF	-	Not analyzed for.
N	-	For organic analytes there is evidence the analyte is present in this sample.
NJ	<b>-</b>	There is evidence that the analyte is present. The associated numerical result is an estimate.
E	-	This qualifier is used when the concentration of the associated value exceeds the known calibration range.
*	-	The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)

Patriculates 5/92-11/92

7411 Beach Drive E, Port Orchard Washington 98366

# CASE NARRATIVE

### May 24, 1993

Subject: Commencement Bay Sediment Traps

Samples: 92 - 498276 to -498279

Case No. DOE-555Y

Officer: Dale Norton

By:

Dickey D. Huntamer O's Organics Analysis Unit

# VOLATILE ORGANIC ANALYSIS

# ANALYTICAL METHODS:

Volatile organic compounds were analyzed using Manchester modification of the EPA SW 846 Method 8240 purge-trap procedure with capillary GC/MS analysis. Normal QA/QC procedures were performed on the samples.

# **BLANKS:**

Low levels of the common laboratory solvents acetone and methylene chloride were detected in the laboratory blanks The EPA five times rule was applied to all target compounds which were found in the blank. Compounds that were found in the sample and in the blank were considered real and not the result of contamination if the levels in the sample are greater than or equal to five times the amount of compounds in the associated method blank.

### SURROGATES:

All surrogate recoveries were within acceptable limits for soil samples.

### **HOLDING TIMES:**

The soil samples were stored frozen following Puget Sound Estuarine Protocols and were analyzed within the recommended 14 day holding time after thawing out..

# MATRIX SPIKE AND MATRIX SPIKE DUPLICATE:

Matrix spike recoveries and Relative Percent Differences (RPD) were within acceptable limits.

Page 2 Commencement Bay Sediment Traps - VOA

# SPECIAL ANALYTICAL PROBLEMS:

There were no analytical problems with these analyses. The data is acceptable for use as qualified.

# DATA QUALIFIER CODES:

U	-	The analyte was not detected at or above the reported value.
J	a_	The analyte was positively identified. The associated numerical value is an estimate.
UJ	-	The analyte was not detected at or above the reported estimated result.
REJ	-	The data are <u>unusable</u> for all purposes.
EXP	**	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .
NAF	-	Not analyzed for.
N	-	For organic analytes there is evidence the analyte is present in this sample.
NJ	-	There is evidence that the analyte is present. The associated numerical result is an estimate.
Е	-	This qualifier is used when the concentration of the associated value exceeds the known calibration range.
*	•	The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)

# CN\_CBAY2.DOC-3

7411 Beach Drive E, Port Orchard Washington 98366

# CASE NARRATIVE

### March 31, 1993

Subject: Commencement Bay

Samples: 92 - 498272 to -498277, -498279, -498288 -498303 to -498305, -498307, -498331 to -498335

Case No. DOE-555Y

Officer: Dale Norton

By: Dickey D. Huntamer Duranics Analysis Unit

# SEMIVOLATILE ORGANICS

### ANALYTICAL METHODS:

The semivolatile soil samples were Soxhlet extracted with acetone following the Manchester modification of the EPA SW 846 8270 procedure with capillary GC/MS analysis of the sample extracts. The sample extracts were cleaned up prior to analysis using silica gel. By eluting with various mixtures of solvents the semivolatile target compounds could be recovered. Normal QA/QC procedures were performed with the analyses. A pair of Canadian reference materials (HS-6) were also analyzed with the samples.

# HOLDING TIMES:

The samples were stored frozen until sample preparation following Puget Sound Estuary Program guidelines. All sample and extraction holding times were within the recommended limits.

### **BLANKS:**

Low levels of some target compounds were detected in the laboratory blanks. The EPA five times rule was applied to all target compounds which were found in the blank. Compounds that were found in the sample and in the blank were considered real and not the result of contamination if the levels in the sample are greater than or equal to five times the amount of compounds in the associated method blank.

### SURROGATES:

The normal surrogates compounds were added to the sample prior to extraction. All surrogate spike recoveries were within acceptable QC limits.

Page 2 Commencement Bay - Semivolatiles

# MATRIX SPIKE AND MATRIX SPIKE DUPLICATE:

Matrix spike recoveries for the EPA Construct Laboratory Program (CLP) compounds were within acceptable limits. Eight other non-CLP compounds had less than 50% recovery and the results for those compounds in the matrix sample, 92-498303 were qualified by adding "J". The base compounds were not part of the spiking mixture and therefore are reported as no analyte found (NAF). One compound, Hexachloroethane had low recoveries < 4%, and also was reported as NAF.

# SPECIAL ANALYTICAL PROBLEMS:

No special analytical problems were encountered in the semivolatile analyses. The silica gel clean-up for semivolatiles worked well, eliminating much of the interferences caused by the presence of fats and lipids. This allowed the samples to be analyzed at a smaller volume and improved chromatography which resulted in better quantitation limits.

The Canadian reference material (HS-6) results are reported as sample 93498335 SRM1 and SRM2.

#### DATA QUALIFIER CODES:

The analyte was not detected it or above the reported value. U The analyte was positively iterrified. The associated numerical value is an J estimate. The analyte was not detected it or above the reported estimated result. UJ The data are unusable for all purposes. REJ The result is equal to the number before EXP times 10 to the power of the EXP number after EXP. As an example 3EXP6 equals 3 X 106. Not analyzed for. NAF For organic analytes there is evidence the analyte is present in this sample. Ν There is evidence that the analyte is present. The associated numerical result NJ is an estimate. This qualifier is used when the concentration of the associated value exceeds Έ the known calibration range. The analyte was present in the sample. Wisual Aid to locate detected bold compound on report sheet.)

CN\_CBAY6.DOC

7411 Beach Drive E, Port Orchard Washington 98366

# CASE NARRATIVE

#### April 2, 1993

Subject: Commencement Bay Sediment Traps

Samples: 92 - 498272 to -498277 and -498279

Case No. DOE-555Y

Officer: Dale Norton

By:

Dickey D. Huntamer Organics Analysis Unit

### POLYCHLORINATED BIPHENYLS

### ANALYTICAL METHODS:

The sediment samples were Soxhlet extracted using acetone as the solvent. Analysis was done by EPA Method 8080 using dual column capillary GC analysis with Electron Capture Detectors (ECD). Interferences were removed by acid treatment.

#### HOLDING TIMES:

The samples were stored frozen following Puget Sound Estuary Program guidelines until time for extraction. The extracts were analyzed within the recommended holding times.

### BLANKS:

No target compounds were detected in the laboratory blanks.

### SURROGATES:

Surrogates ranged from 77% to 129% recovery for most of the samples. Decachlorobiphenyl had 187% recovery in sample -498277. This could be due to chromatographic interference since the other surrogate recovery was acceptable. The "J" data qualifier was added to all results for sample -498277 because of the high surrogate recovery. Surrogate recoveries for the first blank were low, less than 15%, however the duplicate blank recoveries were acceptable. Surrogate recovery for Dibutylchlorendate (DBC) was not reported in some of the samples because it was removed by the acid treatment.

Page 2

Commencement Bay Sediment Traps - PCB

# MATRIX SPIKE AND MATRIX SPIKE :

The matrix spikes recoveries for PCB-1242 ranged from 125% to 130%. The Relative Percent Differences (RPD) was 3.9%. All recoveries and RPD were within acceptable QC limits.

# ANALYTICAL COMMENTS:

There were no significant problems with the Pesticide/PCB analysis. PCB-1254 was detected in all of the samples.

A Canadian PCB reference material was analyzed in duplicate with the samples. These samples are identified in the report as 93-498279 SRM1 and 93-498279 SRM2. Significant interferences from sulfur were encountered in the analysis. One Arochlor PCB-1260 was quantitated and reported, a smaller amount of PCB-1254 also appears to be present, however it could not be quantitated separately due to the interferences and was incorporated into the PCB-1260 value.

### DATA QUALIFIER CODES:

U	-	The analyte was not detected at or above the reported value.
J	- ·	The analyte was positively identified. The associated numerical value is an estimate.
UJ	-	The analyte was not detected at or above the reported estimated result.
REJ	-	The data are unusable for all purposes.
EXP	-	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .
NAF	-	Not analyzed for.
N	-	For organic analytes there is evidence the analyte is present in this sample.
NJ	-	There is evidence that the analyte is present. The associated numerical result is an estimate.
Е	-	This qualifier is used when the concentration of the associated value exceeds the known calibration range.
*		The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)

CN\_CBAY5.DOC

Particulates 11/92-5/93

7411 Beach Drive E , Port Orchard Washington 98366

# CASE NARRATIVE

### August 4, 1993

Subject: Commencement Bay Sediment Traps

Samples: 93 - 228219, -228249 and -228250

Case No. DOE-761Y

Officer: Dale Norton

By:

Dickey D. Huntamer Corganics Analysis Unit

# VOLATILE ORGANIC ANALYSIS

### ANALYTICAL METHODS:

Volatile organic compounds were analyzed using Manchester modification of the EPA SW 846 Method 8240 purge-trap procedure with capillary GC/MS analysis. Normal QA/QC procedures were performed on the samples.

### BLANKS:

Low levels of the common laboratory solvents acetone and methylene chloride were detected in the laboratory blanks The EPA five times rule was applied to all target compounds which were found in the blank. Compounds that were found in the sample and in the blank were considered real and not the result of contamination if the levels in the sample are greater than or equal to five times the amount of compounds in the associated method blank.

### SURROGATES:

Surrogate recoveries were within acceptable limits for soil samples.

# HOLDING TIMES:

The samples were analyzed twelve days past the recommended holding time of fourteen days due to instrument problems. All of the sample results were qualified with "J to indicate the increased uncertainty in their quantitation.

# MATRIX SPIKE AND MATRIX SPIKE DUPLICATE:

Soil matrix spikes were within acceptable QC limits for both percent recovery and Relative Percent Differences (RPD) for all but seven compounds. These were chloromethane, chloroform, trans-1,3-dichloropropene, p-isopropyltoluene, butylbenzene, naphthalene and hexachlorobutadiene. All of the results for these compounds in the matrix spike sample were given the "J" qualifier. Acetone was also outside the limits but that was due to the high background in the sample matrix and the "REJ" qualifier was applied to the acetone results.

### ANALYTICAL COMMENTS:

Other than the holding time no analytical problems were encountered in the analysis. The data is acceptable for use as qualified. A number of chlorinated butadienes were detected in the samples.

### DATA QUALIFIER CODES:

- U The analyte was not detected at or above the reported value.
   J The analyte was positively identified. The associated numerical value is an estimate.
- UJ The analyte was not detected at or above the reported estimated result.
- REJ The data are <u>unusable</u> for all purposes.
- EXP The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals  $3 \times 10^6$ .
- NAF Not analyzed for.
- N For organic analytes there is evidence the analyte is present in this sample.
- NJ There is evidence that the analyte is present. The associated numerical result is an estimate.
- E This qualifier is used when the concentration of the associated value exceeds the known calibration range.
  - The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)
- CN\_CBAY3.DOC 3

7411 Beach Drive E, Port Orchard Washington 98366

# CASE NARRATIVE

# August 30, 1993

Subject: Commencement Bay

Samples: 93 - 228219, -228243 to -228247, -228250, -228277 to -228281, -228308 to -238312

Case No. DOE-761Y

Officer: Dale Norton

By: Dickey D. Huntamer

## SEMIVOLATILE ORGANICS

### ANALYTICAL METHODS:

The semivolatile soil samples were Soxhlet extracted with acetone following the Manchester modification of the EPA SW 846 8270 procedure with capillary GC/MS analysis of the sample extracts. The sample extracts were cleaned up prior to analysis using silica gel. By eluting with various mixtures of solvents the semivolatile target compounds could be recovered. Normal QA/QC procedures were performed with the analyses. A pair of Canadian reference materials (HS-6) sample numbers 93-228400 and -228401, were also analyzed with the samples.

### HOLDING TIMES:

The samples were stored frozen until sample preparation following Puget Sound Estuary Program guidelines. All sample and extraction holding times were within the recommended limits.

### BLANKS:

Low levels of some target compounds were detected in the laboratory blanks. The EPA five times rule was applied to all target compounds which were found in the blank. Compounds that were found in the sample and in the blank were considered real and not the result of contamination if the levels in the sample are greater than or equal to five times the amount of compounds in the associated method blank.

### SURROGATES:

The normal surrogates compounds were added to the sample prior to extraction. All surrogate spike recoveries were within acceptable QC limits except for pyrene-d10 in sample 93-228312 which was 6.6%. Since the other surrogates and terphenyl-d14 had acceptable recoveries no additional qualifiers were added.

# MATRIX SPIKE AND MATRIX SPIKE DUPLICATE:

Matrix spike recoveries for eight of the EPA Contract Laboratory Program (CLP) compounds were within acceptable limits. Three compounds were outside the CLP limits. Thirteen other non-CLP compounds had less than 50% recovery and the results for those compounds, along with the three CLP compounds, were qualified by adding "J" to the results for the matrix sample, 93-228280. The base compounds were not part of the spiking mixture and therefore are reported as no analyte found (NAF).

# SPECIAL ANALYTICAL PROBLEMS:

No special analytical problems were encountered in the semivolatile analyses. The silica gel clean-up as with the previous set of semivolatiles samples worked well, eliminating much of the interferences caused by the presence of fats and lipids. This allowed the samples to be analyzed at a smaller volume and improved chromatography which resulted in better quantitation limits.

The Canadian reference material (HS-6) results are reported as sample 93-228400 SRM1 and 93-228401 SRM2.

The samples reported as LDP1 are dilutions to bring the sample into the quantitation range, not sample duplicates.

# DATA QUALIFIER CODES:

U	-	The analyte was not detected at or above the reported value.
1.	-	The analyte was positively identified. The associated numerical value is an estimate.
UJ	-	The analyte was not detected at or above the reported estimated result.
REJ	-	The data are <u>unusable</u> for all purposes.
EXP	-	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .
NAF	-	Not analyzed for.
N	-	For organic analytes there is evidence the analyte is present in this sample.
NJ	-	There is evidence that the analyte is present. The associated numerical result is an estimate.
Е	•	This qualifier is used when the concentration of the associated value exceeds the known calibration range.
*	-	The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)

# CN\_CBAY8.DOC

7411 Beach Drive E, Port Orchard Washington 98366

# CASE NARRATIVE

### August 24, 1993

Subject: Commencement Bay Sediment Trap

Samples: 93 - 228219, -228243 to -228247 and -228247

Case No. DOE-761Y

Officer: Dale Norton

By:

Dickey D. Huntamer

Organics Analysis Unit

# POLYCHLORINATED BIPHENYLS

### ANALYTICAL METHODS:

The soil sample was Soxhlet extracted using acetone as the solvent. Analysis was done by EPA Method 8080 using dual column capillary GC analysis with Electron Capture Detectors (ECD).

# HOLDING TIMES:

The samples were stored frozen until extraction. The samples were analyzed within forty days of extraction.

### **BLANKS:**

No target compounds were detected in the laboratory blanks.

### SURROGATES:

All surrogate spike recoveries were within acceptable QC limits. Surrogates ranged from 50% to 142% recovery.

### MATRIX SPIKE AND MATRIX SPIKE :

The matrix spikes recoveries ranged from 91% to 106%. The Relative Percent Differences (RPD) ranged from 10% to 11%. All recoveries and RPD were within acceptable QC limits.

# SPECIAL ANALYTICAL PROBLEMS:

There were no significant problems with the Pesticide/PCB analysis. Two reference samples were analyzed with the sample set. These are identified as 93-228314 and -228315. Canadian reference material PACS-1 was used in the analysis. Both PCB-1254 and PCB-1260 were detected in the reference samples. The RPD was 27% for PCB-1254 and 42% for PCB-1260.

# DATA QUALIFIER CODES:

U	+	The analyte was not detected at or above the reported value.
J	•	The analyte was positively identified. The associated numerical value is an estimate.
UJ	-	The analyte was not detected at or above the reported estimated result.
REJ	-	The data are <u>unusable</u> for all purposes.
EXP	-	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .
NAF	-	Not analyzed for.
N	-	For organic analytes there is evidence the analyte is present in this sample.
NJ	. •	There is evidence that the analyte is present. The associated numerical result is an estimate.
E	-	This qualifier is used when the concentration of the associated value exceeds the known calibration range.
*	-	The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)

CN\_CBAY5.DOC - 3

7411 Beach Drive E , Port Orchard Washington 98366

# CASE NARRATIVE

### Anguest 5, 1993

Commencement Bay Sediment Trans Subject:

93 - 228308, -228309 and 238311 Samples:

DOE-761Y Case No.

Dale Norton Officer:

Dickey D. Huntamer By:

Organics Analysis Unit

# TRIBUTYL TINS

# ANALYTICAL METHODS:

The samples were extracted following the methods given in Puget Sound Estuary Program (PSEP) "Recommended Guidelines for Measuring Organic Compounds in Puget Sound Sediment and Tissue Samples" Recommended Methods for Organotin Compounds. The samples were Soxhlet extracted using acetone and tropolone, 0.2% by weight, solvent exchanged to hexane and dried using sodium sulfate. The organotin compounds were hexylated using the Grignard reaction given in Krone et al (1989) including the silica gel/alumina cleanup. Analysis was done by capillary Gas Chromatography using a Ion Trap Detector (FPD). All results are on a dry weight basis.

### **HOLDING TIMES:**

The samples were stored frozen following PSEP Guidefines until extraction. After extraction all samples were analyzed within the recommended 40 day extract time.

### **BLANKS:**

No target compounds were detected in the laboratory blanks.

#### SURROGATES:

The surrogate, Tripropyltin had consistently poer recoveries for all of the analyses. Recoveries ranged from not found to 5.2%. The reason for the poor recoveries is not readily apparent. The possibility that the surrogates were not added is unlikely since all of the samples were the same and a second set of tin analyses prepared and extracted at a different time also showed a similar problem. This would suggest a possible problem with the spiking solution but it was used in a previous sample set and recoveries were acceptable. Another potential problem area is the derivitization reagent, but it appeared to behave normally. If it was not working properly then an zibutyltin (TBT) would be detected and TBT was detected in all samples except the laboratory manks. A possible scenario is that the Tripropyltin surrogate failed to elute off the silica gel/alumina commun used to clean-up the extracts. Unfortunately the real reason can not be determined at this time. All of the sample data was qualified with the "J" flag because of the low surrogate recoveries.

### MATRIX SPIKE AND MATRIX SPIKE DUPLICATE:

One of the matrix spike samples went dry during the Soxhlet extraction. No tin compounds were recovered in that sample. The other matrix spike had 79.5% recovery of tributyltin and 117% recovery of dibutyltin. The tetrabutyl and mono butyl tin spikes were not detected. No recovery or RPD limits have been established for this method.

### ANALYTICAL COMMENTS:

Sample -228309 also went dry during the Soxhlet process although apparently not as long as the one matrix spike because tributyltin was detected in that sample. All of sample -228309 was used up in the initial extractions and re-extractions could not be done.

The "J" data qualifier was added to all of the results due to the low surrogate recoveries and is used to indicate that the quantitation values and quantitation limits are estimates. The reason for the low surrogate recovery is not apparent at this time and additional testing will be required to ascertain the problem. Tributyltin was detected in all three samples and in the Sequim Bay reference material sample, -228313 at 54.1 ug/Kg dry or 31.2 ug/Kg wet weight.

The Sequim Bay Reference Sediment is presumably spiked with 100 ng/gm (100 ug/Kg) wet weight of tributyltin. No value for tributyltin has been established for the Sequim Bay Reference Sediment so the accuracy of the analysis cannot be determined.

#### DATA QUALIFIER CODES:

U	-	The analyte was not detected at or above the reported value.
J	-	The analyte was positively identified. The associated numerical value is an estimate.
UJ	-	The analyte was not detected at or above the reported estimated result.
REJ	-	The data are <u>unusable</u> for all purposes.
EXP	-	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .
NAF	-	No analyte found.
N	-	For organic analytes there is evidence the analyte is present in this sample.
NJ	-	There is evidence that the analyte is present. The associated numerical result is an estimate.
E	-	This qualifier is used when the concentration of the associated value exceeds the known calibration range.
*	-	The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)

CN\_CBAY4.DOC - 3

Particulates 7/93-12/43

7411 Beach Drive E, Port Orchard Washington 98366

# CASE NARRATIVE

February 17, 1994

Subject: Commencement Bay Sediment Traps

Samples: 94 - 028587 to -028589

Case No. DOE-957Y

Officer: Dale Norton

By:

Dickey D. Huntamer 60 Organics Analysis Unit

# VOLATILE ORGANIC ANALYSIS

#### ANALYTICAL METHODS:

Volatile organic compounds were analyzed using Manchester modification of the EPA SW 846 Method 8260 purge-trap procedure with capillary GC/MS analysis. Normal QA/QC procedures were performed on the samples.

### **BLANKS:**

Low levels of the common laboratory solvents acetone and methylene chloride were detected in the laboratory blanks The EPA five times rule was applied to all target compounds which were found in the blank. Compounds that were found in the sample and in the blank were considered real and not the result of contamination if the levels in the sample are greater than or equal to five times the amount of compounds in the associated method blank.

#### SURROGATES:

Surrogate recoveries were within acceptable limits for the soil samples except for 1,2-dichloroethane-d4 in matrix spike samples -028588 LMX1 and LMX2 which were close to 400% recovery. This appears to be interference from the matrix spiking since the recovery in the source sample -028588 itself was acceptable.

#### HOLDING TIMES:

The soil samples were analyzed within the recommended 14 day holding time.

### MATRIX SPIKE AND MATRIX SPIKE DUPLICATE:

Soil matrix spikes were within acceptable QC limits for both percent recovery and Relative Percent Differences (RPD) for most of the compounds. The "J" qualifiers were added to results in the matrix source sample -028588 for acetone, 2-butanone, p-isopropyl toluene and naphthalene the "R" reject qualifier was added to results for 4-methyl-2-pentanone, 2-hexanone.

Commencement Bay Sediment Trap - VOA

# **ANALYTICAL COMMENTS:**

No analytical problems were encountered in the analysis. The data is acceptable for use as qualified.

# **DATA QUALIFIER CODES:**

U	-	The analyte was not detected at or above the reported value.
1	<b>-</b> ·	The analyte was positively identified. The associated numerical value is an estimate.
UJ		The analyte was not detected at or above the reported estimated result.
REJ	<b></b>	The data are <u>unusable</u> for all purposes.
EXP	-	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .
NAF	-	Not analyzed for.
N	-	For organic analytes there is evidence the analyte is present in this sample.
NJ	-	There is evidence that the analyte is present. The associated numerical result is an estimate.
E	<u></u>	This qualifier is used when the concentration of the associated value exceeds the known calibration range.
*	-	The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)
3A02.D0	DC - 4	All Data Whatball

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7411 Beach Drive E, Port Orchard Washington 98366

# CASE NARRATIVE

#### April 10, 1994

Subject:

Commencement Bay Sediment Traps

Samples: 94 -

94 - 028526 to -028530, -028533, -028584 to -028587, -028589 to -028595

Case No. DOE-957Y

Officer: Dale Norton

By:

Dickey D. Huntamer

# SEMIVOLATILE ORGANICS

#### ANALYTICAL METHODS:

The semivolatile soil samples were Soxhlet extracted with acetone following the Manchester modification of the EPA SW 846 8270 procedure with capillary GC/MS analysis of the sample extracts. The sample extracts were cleaned up using silica gel prior to analysis. Normal QA/QC procedures were performed with the analyses.

### HOLDING TIMES:

The samples were composites from two collections and were stored frozen following Puget Sound Estuary Program guidelines until analysis. Upon release of the samples from the project officer to the laboratory all sample and extraction holding times were kept within the recommended limits of fourteen days for extraction and forty days for analyses.

#### **BLANKS:**

Low levels of some target compounds were detected in the laboratory blanks. The EPA five times rule was applied to all target compounds which were found in the blank. Compounds that were found in the sample and in the blank were considered real and not the result of contamination if the levels in the sample are greater than or equal to five times the amount of compounds in the associated method blank.

#### SURROGATES:

The normal surrogates compounds were added to the sample prior to extraction. All surrogate spike recoveries were within acceptable QC limits.

Page 2 Commencement Bay Sediment Traps - Semivolatiles

# MATRIX SPIKE AND MATRIX SPIKE DUPLICATE:

Matrix spike recoveries were acceptable for most of the compounds however the Relative Percent Differences (RPD) for the majority of the earlier eluting compounds were low and consequently the "J" qualifier was added to the results for those compounds in sample -028592. An additional problem was the high native concentrations in the source sample, -028592. This interfered with the recovery calculations and resulted in some of the RPD's being outside of the 25% limit.

### ANALYTICAL COMMENTS:

During the extraction two samples were labeled as -028528 and no sample -028526. Both extracts were shot as -028528a and -028528b with the rest of the samples. Since there was no way to distinguish which sample was which both were re-extracted, but after the holding time, and analyzed. The results of the second analysis are reported for these samples. Normally all results would be qualified with "J" as estimates since holding times were exceeded. This action was not felt to be necessary since a compound by compound comparison of the re-analyses with the original analyses showed that the results in the second analysis were higher and no loss of analytes due to holding times is apparent.

Other than those items discussed above no special analytical problems were encountered in the semivolatile analyses and the data is acceptable for use as qualified.

### DATA QUALIFIER CODES:

U	-	The analyte was not detected at or above the reported value.
J	-	The analyte was positively identified. The associated numerical value is an estimate.
UJ	-	The analyte was not detected at or above the reported estimated result.
REJ	-	The data are <u>unusable</u> for all purposes.
ЕХР	-	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .
NAF	*	Not analyzed for.
N	-	For organic analytes there is evidence the analyte is present in this sample.
NJ	-	There is evidence that the analyte is present. The associated numerical result is an estimate.
E	*	This qualifier is used when the concentration of the associated value exceeds the known calibration range.
*	-	The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)

CN CBA03.DOC - 4
7411 Beach Drive E, Port Orchard Washington 98366

#### **CASE NARRATIVE**

#### February 11, 1994

Subject:

Commencement Bay Sediment Trap

Samples: 94 - 028533, -028584 to -028587, -028589, -028596 and -028597

Case No. DOE-957Y

Officer: Dale Norton

By:

Dickey D. Huntamer (D) Organics Analysis Unit

#### POLYCHLORINATED BIPHENYLS

#### ANALYTICAL METHODS:

The soil sample was Soxhlet extracted using acetone as the solvent. Analysis was done by EPA Method 8080 using dual column capillary GC analysis with Electron Capture Detectors (ECD).

#### HOLDING TIMES:

The samples were stored frozen until extraction. The samples were analyzed within forty days of extraction.

#### BLANKS:

No target compounds were detected in the laboratory blanks.

#### SURROGATES:

Four surrogate compounds were analyzed with the PCB's. Surrogates ranged from 47% to 206% recovery. Surrogate spike recoveries were within the acceptable QC limits of 50% to 150% for most of the samples. In the few cases where surrogate recoveries exceeded the limits only one of the four surrogates was outside the range. Since the remaining surrogates were acceptable no data was qualified because of surrogate recoveries.

#### MATRIX SPIKE AND MATRIX SPIKE :

The matrix spikes recoveries ranged from 90% to 111%. The Relative Percent Differences (RPD) ranged from 5.1% to 10.4%. All recoveries and RPD were within acceptable QC limits.

#### SPECIAL ANALYTICAL PROBLEMS:

There were no significant problems with the Pesticide/PCB analysis. Two reference samples were analyzed with the sample set. These are identified as 94-028596 and -028597. Canadian reference material HS-2 was used in the analysis. Both PCB-1254 and PCB-1260 were detected in the reference samples. The RPD was 1.6% for PCB-1254 and 2.0% for PCB-1260.

### DATA QUALIFIER CODES:

U	-	The analyte was not detected at or above the reported value.
J	-	The analyte was positively identified. The associated numerical value is an estimate.
UJ	-	The analyte was not detected at or above the reported estimated result.
REJ	-	The data are <u>unusable</u> for all purposes.
EXP	<b>-</b>	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .
NAF	-	Not analyzed for.
N	*	For organic analytes there is evidence the analyte is present in this sample.
IJ	-	There is evidence that the analyte is present. The associated numerical result is an estimate.
Е		This qualifier is used when the concentration of the associated value exceeds the known calibration range.
*	-	The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)

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7411 Beach Drive E, Port Orchard Washington 98366

#### CASE NARRATIVE

#### July 8, 1994

Subject: Commencement Bay Sediment Traps

Samples: 94 - 028526, -028528 to -028530

Case No. DOE-957Y

Officer: Dale Norton

By:

Dickey D. Huntamer

#### ORGANO TINS

#### ANALYTICAL METHODS:

The samples were extracted following the methods given in Puget Sound Estuary Program (PSEP) "Recommended Guidelines for Measuring Organic Compounds in Puget Sound Sediment and Tissue Samples" Recommended Methods for Organotin Compounds. The samples were Soxhlet extracted using acetone and tropolone, 0.2% by weight, solvent exchanged to hexane and dried using sodium sulfate. The organotin compounds were hexylated using the Grignard reaction given in Krone et al (1989) including the silica gel/alumina cleanup. Analysis was done by capillary Gas Chromatography/Mass Spectrometry using the selected ion monitoring (SIM) mode to decrease quantitation limits. Percent solids were available so all results are on a dry weight basis.

#### HOLDING TIMES:

The samples were stored frozen following PSEP Guidelines until extraction. After extraction all samples were analyzed within the recommended 40 day extract time.

#### BLANKS:

No organotin compounds were detected in the laboratory blanks.

#### SURROGATES:

The surrogate, tripropyltin chloride was inadvertently spiked in a a concentration ten times higher than planned due to a dilution error. Recovery of the surrogate spike, Tripropyltin, ranged from 46% to 106%. No limits have been established for this method.

#### MATRIX SPIKE AND MATRIX SPIKE :

The matrix spikes were also added at a concentration ten times higher than desired and were outside the linear calibration range. The spiked samples were diluted, reanalyzed and the results from the dilution are reported. Spike recoveries ranged from 41.6% to 160.6%. The Relative Percent Differences (RPD)

Page 2 Sediment Bay Sediment Traps - Organozius

were high, greater than 50%, for all except dibutyltin chloride, at 26%. The recoveries for the tributyltin chloride were compromised by the high ributyltin found in the matrix spike source sample.

#### ANALYTICAL COMMENTS:

Despite the use of SIM mode GC/MS analysis their were still chromatographic interferences which contributed interfering ions. This resulted in some hits for the other non-tributyl tin species being marked as undetected due to poor spectra. The minutyltin had relatively high concentrations and presented minimal problems. Tetrabutyl tin was not detected in any of the samples.

Two additional samples were analyzed with the sediment samples. This was a Sequim Bay Reference Sediment which presumably was spiked with 100 ng/gm (100 ug/Kg) wet weight of tributyltin. No value for tributyltin has been established for the Sequim Bay Reference Sediment so the accuracy of the analysis cannot be determined.

SQ4139	110	1g/Kg	(wet weight)	Tributyltin
-	33.3		• •	Monobutyltin
SQ4139D	83.3	•		Tributyltin
-	12.3	•	100 · 10	Dibutyltin

#### ANALYTICAL COMMENTS:

No analytical problems were encountered in the analysis, the data is acceptable for use as qualified.

#### DATA QUALIFIER CODES:

U	-	The analyse was not detected at or above the reported value.
J	-	The analyse was positively identified. The associated numerical value is an estimate.
ហ	-	The analyte was not detected at or above the reported estimated result.
REJ	-	The data are <u>unusable</u> for all purposes.
EXP	-	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .
NAF	-	Not analyzed for.
N	-	For organic analyzes there is evidence the analyte is present in this sample.
NJ	-	There is evidence that the analyte is present. The associated numerical result is an estimate.
Е	-	This qualifier is used when the concentration of the associated value exceeds the known calibration range.
*	-	The analyte was present in the sample. (Visual Aid to locate detected compound as report sheet.)

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Particulates 12/93-5/94

# Manchester Environmental Laboratory

7411 Beach Dr E, Port Orchard Washington 98366 July 20, 1994

Project:	Commencement Bay
Samples:	94228587, 94228588, 94228589, 94228590
Project Officer:	Dale Norton
By:	Karin Feddersen KF

These samples were analyzed by EPA Method 624 for Volatile Organic Compounds.

#### **Holding Times:**

The samples were analyzed within method holding times.

## Method Blanks:

Low levels of several analytes were detected in both method blanks. When these analytes are detected in the samples at a concentration greater than five (5) times that detected in the corresponding method blank, the contribution to the results due to laboratory contamination is considered negligible. No qualification of the results is necessary.

When these analytes are detected in the samples at a concentration less than five (5) times that detected in the corresponding method blank, they are most likely present due to laboratory contamination and not native to the samples. Results for these analytes in the corresponding samples have been changed (qualifier "U") to indicate that these analytes were not detected at a level above the suspected contamination.

# GC/MS Tuning and Calibration:

Calibration against Bromofluorobenzene (BFB) is acceptable for the initial calibration, continuing calibration and all associated sample analyses.

#### Initial Calibration:

The initial calibration met the minimum response criteria for the average relative responses. The % Relative Standard Deviations were within the maximum of 30% with the exception of Acetone. All results for Acetone have been qualified with a "J".

#### **Continuing Calibration:**

The average relative response factors for all target analytes were above the minimums, and the percent deviations between the initial and continuing calibration standards were within the maximum of 25% with several exceptions. In the corresponding samples: positive results for these analytes have been qualified with a "J"; non-detect results for these analytes have been qualified with a "UJ".

### Surrogates:

All surrogate recoveries for these samples, the matrix spikes, and the associated method blanks are reasonable, acceptable, and within QC limits.

#### Matrix Spikes (MS/MSD):

Matrix spike recovery and precision data are reasonable, acceptable, and within advisory QC limits of 50% to 150% with several exceptions. The matrix spike duplicate recovery for Isopropyltoluene, and both matrix spike recoveries for Acetone, 2-Butanone, and Napthalene are outside QC limits. Due to the high concentration of these analytes present in the original sample compared to the amount spiked, the matrix spike recovery data is unreliable. Also, all surrogates for these samples are within QC limits. Therefore, sample results have not been qualified on the basis of the spike data. All affected spike recoveries for these analytes have been qualified with "NAR". Isopropylbenzene co-eluted with an unknown compound which resulted in artificially elevated recoveries for this analyte. The spike recoveries for Isopropylbenzene have been for 1,2,4-Trichlorobenzene, recoveries "NAR". Matrix spike qualified with Hexachlorobutadiene, and 1,2,3-Trichlorobenzene are slightly below QC limits. All positive results for these analytes have been qualified with a "J"; the instrument sensitivity is sufficient that the non-detect results are unaffected.

#### Sample Results:

This data is acceptable for use as amended.

#### DATA QUALIFIER CODES:

- U The analyte was not detected at or above the reported value.
- J The analyte was positively identified. The associated numerical value is an estimate.
- UJ The analyte was not detected at or above the reported estimated result.
- NAR No viable analytical result obtained for this analyte.
- \* The analyte was present in the sample. (Visual Aid to locate detected analyte on report sheet.)

7411 Beach Drive E, Port Orchard Washington 98366

#### CASE NARRATIVE

#### August'3, 1994

Subject: Commencement Bay

Samples: 94 - 228582 to -228588 and -228590 to -228592

Case No. DOE-118X

Officer: Dale Norton

By:

Dickey D. Huntamer

# POLYCHLORINATED BIPHENYLS

#### ANALYTICAL METHODS:

The soil sample was Soxhlet extracted using acetone as the solvent. Analysis was done by EPA Method 8080 using dual column capillary GC analysis with Electron Capture Detectors (ECD).

#### HOLDING TIMES:

All extraction and analysis holding times were met.

#### BLANKS:

No target compounds were detected in the laboratory blanks.

#### SURROGATES:

Surrogates recoveries for the samples ranged from 25% to 117%. In several samples, surrogate recovery for Dibutylchlorendate (DBC) is low, the other surrogates are okay, consequently no qualifiers were added to the results. Surrogate recoveries in the blanks tended to be lower than in the samples and possibly reflects losses during sample concentration.

#### MATRIX SPIKE AND MATRIX SPIKE :

The matrix spike compounds were apparently not added to the samples. Since the sample, (-228582), used for the matrix spikes contained some native PCB's, the native amount was also calculated in the two "spiked" samples to provide a triplicate analysis.

Page 2 Commencement Bay - PCB

# ANALYTICAL COMMENTS:

There were no significant problems with the PCB analysis other than the problem with the matrix spikes. Samples -228591 and -228592 are duplicates of Canadian Reference Material HS-2.

# DATA QUALIFIER CODES:

U	-	The analyte was not detected at or above the reported value.
J ·	-	The analyte was positively identified. The associated numerical value is an estimate.
UJ	-	The analyte was not detected at or above the reported estimated result.
REJ	- ·	The data are <u>unusable</u> for all purposes.
EXP	<b>-</b>	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .
NAF	-	Not analyzed for.
Ν	•	For organic analytes there is evidence the analyte is present in this sample.
NJ	-	There is evidence that the analyte is present. The associated numerical result is an estimate.
Е	-	This qualifier is used when the concentration of the associated value exceeds the known calibration range.
*	-	The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)

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7411 Beach Drive E, Port Orchard Washington 98366

#### CASE NARRATIVE

#### August 30, 1994

Subject: Commencement Bay

Samples: 94 - 228585 Matrix Spikes

Case No. DOE-118X

Officer: Dale Norton

By:

Dickey D. Huntamer

#### POLYCHLORINATED BIPHENYLS

#### ANALYTICAL METHODS:

The soil samples were Soxhlet extracted using acetone as the solvent. Analysis was done by EPA Method 8080 using dual column capillary GC analysis with Electron Capture Detectors (ECD).

#### HOLDING TIMES:

These samples were extracted after the recommended 14 day holding time. However given the persistence and stability of Arochlor (PCB) mixtures, exceeding the holding times will have no detrimental effect on the concentrations.

#### BLANKS:

No target compounds were detected in the laboratory blanks.

#### SURROGATES:

Surrogates recoveries for the samples ranged from 61% to 99%.

#### MATRIX SPIKE AND MATRIX SPIKE :

Matrix spike recoveries ranged from 71% to 104% and the Relative Percent Differences (RPD) ranged from 2.0% to 4.1%.

Page 2 Commencement Bay - PCB

# ANALYTICAL COMMENTS:

There were no significant problems with the PCB analysis. Sample -228585 was used since it had the most material remaining after the earlier analysis.

# DATA QUALIFIER CODES:

U	-	The analyte was not detected at or above the reported value.
J	*	The analyte was positively identified. The associated numerical value is an estimate.
IJ	-	The analyte was not detected at or above the reported estimated result.
REJ	-	The data are <u>unusable</u> for all purposes.
EXP	<b>.</b>	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .
NAF	-	Not analyzed for.
N	-	For organic analytes there is evidence the analyte is present in this sample.
NJ	•	There is evidence that the analyte is present. The associated numerical result is an estimate.
E	*	This qualifier is used when the concentration of the associated value exceeds the known calibration range.
*	-	The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)

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12/93 - 5/94 Partrenleto

# **Manchester Environmental Laboratory**

7411 Beach Dr E, Port Orchard Washington 98366

# CASE NARRATIVE

September 23, 1994

Subject: Commencement Bay

Samples: 942285582 - 228588

Case No. DOE-118X

Officer: Dale Norton

By: **Greg Perez** Organics Analysis Unit

# SEMIVOLATILE ANALYSIS

#### ANALYTICAL METHODS:

The semivolatile water samples were extracted with methylene chloride following the Manchester Laboratory modification of the EPA CLP and SW 88846 8270 procedure with capillary GC/MS analysis of the sample extracts. Normal QA/QC procedures were performed with the analyses.

#### **BLANKS:**

Low levels of some target compounds were detected in the laboratory blanks. The EPA five times rule was applied to all target compounds which were found in the blank. Compounds that were found in the sample and in the blank were considered real and not the result of contamination if the levels in the sample are greater than or equal to five times the amount of compounds in the associated method blank.

#### SURROGATES:

Surrogate recoveries were within acceptable limits for the soil samples. Surrogate recoveries for both the blanks were unusually low and below the recommended limits. Why this occurred has not been fully determined. The problem seems to have been restricted to the blanks, as the recoveries for the samples were reasonable. No samples were qualified because of the problems with the blanks.

#### HOLDING TIMES:

All sample and extraction holding times were within the recommended limits.

#### MATRIX SPIKE AND MATRIX SPIKE DUPLICATE:

Spiked compounds which were recovered but had recoveries below the recommended limits were given a J qualifier in the native sample only. Several compound were not recovered at all in either of the matrix spikes. This an effect of column degradation or injection port contamination which often occurs with sediment samples. Data for these compounds was rejected (qualifier REJ) in all samples

#### ANALYTICAL COMMENTS:

This data is acceptable for use as qualified.

# DATA QUALIFIER CODES:

U	-	The analyte was not detected at or above the reported value.
J	-	The analyte was positively identified. The associated numerical value is an estimate.
UJ	-	The analyte was not detected at or above the reported estimated result.
REJ	-	The data are <u>unusable</u> for all purposes.
EXP	••	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .
NAF	-	Not analyzed for.
N	-	For organic analytes there is evidence the analyte is present in this sample.
NJ	•••	There is evidence that the analyte is present. The associated numerical result is an estimate.
E		This qualifier is used when the concentration of the associated value exceeds the known calibration range.
*	-	The analyte was present in the sample. (Visual Aid to locate detected

compound on report sheet.)

7411 Beach Drive E, Port Orchard Washington 98366

#### CASE NARRATIVE

October 24, 1994

Subject: Commencement Bay

Samples: 94 - 228532 to -228537

Case No. DOE-118X

Officer: Dale Norton

By: Dickey D. Huntamer Organics Analysis Unit

#### TRIBUTYL TINS

#### ANALYTICAL METHODS:

The samples were extracted following the methods given in Puget Sound Estuary Program (PSEP) "Recommended Guidelines for Measuring Organic Compounds in Puget Sound Sediment and Tissue Samples" Recommended Methods for Organotin Compounds. The samples were Soxhlet extracted using acetone and tropolone, 0.2% by weight, solvent exchanged to hexane and dried using sodium sulfate. The organotin compounds were hexylated using the Grignard reaction given in Krone et al (1989) including the silica gel/alumina cleanup. Analysis was done by capillary Gas Chromatography using Single Ion Monitoring (SIM) mode GC/MS. All samples are reported on a dry weight basis.

#### HOLDING TIMES:

The samples were stored frozen following PSEP Guidelines until extraction. After extraction all samples were analyzed within the recommended 40 day extract time.

#### **BLANKS:**

No target analytes were detected in the laboratory blanks.

#### SURROGATES:

Recovery of the surrogate spike, Tripropyltin, ranged from 28% to 156% except for sample -228534 LMX2 which was 235%. Most of the recoveries were greater than 100% due to chromatographic interference with the quantitation ion. The "J" qualifier was added to sample results in those samples where the surrogate recovery was over 150%. No surrogate recovery QC limits have been established for this method.

#### MATRIX SPIKE AND MATRIX SPIKE :

Both the matrix spike and spike duplicate had significant chromatographic interference. Consequently recoveries for only tetrabutyltin and monobutyltin are reported. Recoveries of the other analytes in the matrix spikes could not be determined due to chromatographic interference which gave high (>200%) recoveries. No spike recovery QC limits have been established for Organotins at this time. The Relative Percent Differences (RPD) for tetrabutyltin was 1.1% and monobutyltin, 9.4%.

#### ANALYTICAL COMMENTS:

The samples were analyzed at a higher concentration, which is reflected in the lower quantitation limits. Consequently most of the samples had to be diluted and reanalyzed to bring the organotin concentrations within the linear calibration range. The results for the diluted samples were generally within about 10% of the result from the undiluted sample. One sample -228536 was not diluted and consequently an "E" flag was added to indicate that it was outside the calibration curve. Based on the dilution results from the other samples it is probably still within 10% of the diluted "true" value.

The principle analytical problems encountered were directly related to the chromatographic interferences in some of the samples. This was a problem with tributyltin and dibutyltin in the matrix spikes. Analysis of diluted sample extracts also had interference problems.

Two additional samples were analyzed with the sediment samples. This was a Sequim Bay Reference Sediment which presumably was spiked with 100 ng/gm (100ug/Kg) wet weight of tributyltin. No value for tributyltin has been established for the Sequim Bay Reference Sediment so the accuracy of the analysis cannot be determined. These samples are identified as -228598 (SRM) and -228599 (SRM2).

SRM	85.8	ug/Kg (wet weight)	Tributyltin
SRM2	120	ug/Kg (wet weight)	Tributyltin

Note that the data sheets report these values as dry weight. The percent solids is 58% for these samples.

Page 3 Commencement Bay - Organotins

# DATA QUALIFIER CODES:

U	-	The analyte was not detected at or above the reported value.
l	-	The analyte was positively identified. The associated numerical value is an estimate.
UJ	-	The analyte was not detected at or above the reported estimated result.
REJ	-	The data are unusable for all purposes.
EXP	•	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example $3EXP6$ equals $3 \times 10^6$ .
NAF	- '	Not analyzed for.
N	-	For organic analytes there is evidence the analyte is present in this sample.
NJ	•	There is evidence that the analyte is present. The associated numerical result is an estimate.
Е	-	This qualifier is used when the concentration of the associated value exceeds the known calibration range.
*	•	The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)

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# **Manchester Environmental Laboratory**

7411 Beach Dr E, Port Orchard Washington 98366

# CASE NARRATIVE

#### March 6, 1995

Subject:	Commencement Bay sediments-51
Samples:	94518195 - 518209

94518195 - 518209

Case No.

Officer:

By:

170494 **Dale Norton** 

Greg Perez

Organics Analysis Unit

# **VOLATILE ORGANIC ANALYSIS**

#### SUMMARY:

Carryover from a previous sample affected the results for sample 518203. The sample was reanalyzed. Both results have been reported, however the duplicate more accurately represents the character of the sample.

#### **ANALYTICAL METHODS:**

Volatile organic compounds were analyzed using Manchester modification of the EPA SW 846 Method 8260 purge-trap procedure with capillary GC/MS analysis. Normal QA/QC procedures were performed on the samples.

#### BLANKS:

Low levels of the common laboratory solvents acetone and methylene chloride were detected in the laboratory blanks. If the concentrations of the compounds in the sample are greater than or equal to five times the concentrations of the compounds in the associated method blank, they are considered native to the sample.

#### SURROGATES:

Surrogate recoveries were within acceptable limits for the sediment samples.

#### **HOLDING TIMES:**

The sediment samples were analyzed within the recommended 14 day holding time.

# MATRIX SPIKE AND MATRIX SPIKE DUPLICATE:

Sediment matrix spikes were within acceptable QC limits for both percent recovery and Relative Percent Differences (RPD).

# DATA QUALIFIER CODES:

U	-	The analyte was not detected at or above the reported value.
J	-	The analyte was positively identified. The associated numerical value is an estimate.
UJ	-	The analyte was not detected at or above the reported estimated result.
REJ	-	The data are <u>unusable</u> for all purposes.
EXP	-	The result is equal to the number before EXP times 10 to the power of the
		number after EXP. As an example 3EXP6 equals 3 X 10 <sup>6</sup> .
NAF	-	Not analyzed for.
N	-	For organic analytes there is evidence the analyte is present in this sample.
NJ	-	There is evidence that the analyte is present. The associated numerical
	res	ult is an estimate.
E	-	This qualifier is used when the concentration of the associated value exceeds
		the known calibration range.
bold	-	The analyte was present in the sample. (Visual Aid to locate detected
		compound on report sheet.)

7411 Beach Drive E, Port Orchard Washington 98366

#### CASE NARRATIVE

#### March 24, 1995

Subject: Commencement Bay Sediment Traps - 51

Samples: 94 - 518136, -518137, -518144, -518147, -518150, -518157, -518164, -518173, -518180, -518188, -518195, -518202 and -518209.

Case No. 1704 - 94

Officer: Dale Norton

By: Dickey D. Huntamer

#### SEMIVOLATILE ORGANICS

#### ANALYTICAL METHODS:

The semivolatile soil samples were extracted with acetone following the Manchester modification of the EPA CLP and SW 846 8270 procedure. The sample extracts were cleaned up with Gel Permeation Chromatography followed by silica gel chromatography. Analysis was by capillary GC/MS. Normal QA/QC procedures were performed with the analyses.

#### HOLDING TIMES:

All sample and extraction holding times were within the recommended limits except for sample -518202. No surrogates were added in the original extraction of sample -518202 and the sample was re-extracted after the recommended fourteen day holding time. All results for sample -518202 were given the "J" qualifier.

#### BLANKS:

Low levels of some target compounds were detected in the laboratory blanks. The EPA five times rule was applied to all target compounds which were found in the blank. Compounds that were found in the sample and in the blank were considered real and not the result of contamination if the levels in the sample are greater than or equal to five times the amount of compounds in the associated method blank.

#### SURROGATES:

The normal Manchester Laboratory surrogates were added to the sample prior to extraction. All surrogate recoveries were within acceptable limits except for blank, BLNK5652 (BS5039) associated with the re-extraction of sample -518202. BLNK5652 apparently was not spiked with surrogates. No recoveries are available and the "J" data qualifier was added to all results for this sample.

### MATRIX SPIKE AND MATRIX SPIKE DUPLICATE:

Matrix spike recoveries were low for hexchloroethane, 1,3 and 1,2-dichlorobenzene, 2,4-dinitrophenol, 4mirroganiline 4,6-dinitro-2-methylphenol. The "J" qualifier was added to the results for sample -518188 these compounds. The following compounds, hexachlorocylcopentadiene, 3 and, 4-chloroaniline, and 4nitrophenol had zero percent recovery in the matrix spikes and were qualified as rejected, "REJ" in the matrix spike source sample, -518188.

#### ANALYTICAL COMMENTS:

No other analytical problems were encountered in the semivolatile analyses. The data is acceptable for use as qualified.

#### DATA QUALIFIER CODES:

U	-	The analyte was not detected at or above the reported value.
1	-	The analyte was positively identified. The associated numerical value is an estimate.
w	-	The analyte was not detected at or above the reported estimated result.
REJ	-	The data are <u>unusable</u> for all purposes.
EXP	-	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .
NAF	-	Not analyzed for.
N	-	For organic analytes there is evidence the analyte is present in this sample.
NJ	-	There is evidence that the analyte is present. The associated numerical result is an estimate.
E	-	This qualifier is used when the concentration of the associated value exceeds the known calibration range.
bold	<b></b>	The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)

#### CN\_CBA51.DOC

7411 Beach Drive E, Port Orchard Washington 98366

#### CASE NARRATIVE

#### January 28, 1995

Subject: Commencement Bay Seriments -51

Samples: 94 - 518164, -518173, -518180, -518181, -518188, -518195, and -518202

Case No: 1704-94

Officer: Dale Norton

By:

Dickey D. Huntamer

#### **POLYCHLORINATED BIPHENYLS**

#### ANALYTICAL METHODS:

The soil samples were Soxhlet extracted using acetone as the solvent. Analysis was done by EPA Method 8080 using dual column capillary GC analysis with Electron Capture Detectors (ECD).

#### HOLDING TIMES:

All extraction and analysis holding times were met.

#### **BLANKS:**

No target compounds were detected in the laboratory blanks.

#### SURROGATES:

Surrogates recoveries for the samples ranged from 57% to 145% and were within the acceptable limits.

#### MATRIX SPIKE AND MATRIX SPIKE :

Matrix spike recoveries ranged from 11% to 121%. The Relative Percent Differences (RPD) for the PCB-1242 was 3.4%. Recovery of PCB-1260 was good, 86% in one sample (LMX1) but only 11% in the other sample (LMX2). This could be due to the high native PCB-1260 present in the sample interfering. No qualifiers were added because of matrix spike recoveries. Page 2 Commencement Bay - PCB

### ANALYTICAL COMMENTS:

There were no significant problems with the PCB analysis and the data is acceptable as qualified. samples labeled HS-24828 and HS-24829 are Canadian Reference Material samples.

# DATA QUALIFIER CODES:

U	-	The analyte was not detected at or above the reported value.
1	<b></b>	The analyte was positively identified. The associated numerical value is an estimate.
UJ	-	The analyte was not detected at or above the reported estimated result.
REJ		The data are <u>unusable</u> for all purposes.
EXP	••	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .
NAF	-	Not analyzed for.
N	-	For organic analytes there is evidence the analyte is present in this sample.
NJ	-	There is evidence that the analyte is present. The associated numerical result is an estimate.
Е	-	This qualifier is used when the concentration of the associated value exceeds the known calibration range.
*	-	The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)

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7411 Beach Drive E, Port Orchard Washington 98366

#### CASE NARRATIVE

#### April 6, 1995

Subject: Commencement Bay Sediments Traps - 51

Samples: 94 - 518136, -518137, -518144, -518147, -518150, -518157, -518164, -518173, -518180, -518188, -518195, -518202, and -518209.

Case No. 1704 - 94

Officer: Dale Norton

By:

Dickey D. Huntamer

#### TRIBUTYL TINS

#### ANALYTICAL METHODS:

The samples were extracted following the methods given in Puget Sound Estuary Program (PSEP) "Recommended Guidelines for Measuring Organic Compounds in Puget Sound Sediment and Tissue Samples" Recommended Methods for Organotin Compounds. The samples were Soxhlet extracted using acetone and tropolone, 0.2% by weight, solvent exchanged to hexane and dried using sodium sulfate. The organotin compounds were hexylated using the Grignard reaction given in Krone et al (1989) including the silica gel/alumina cleanup. Analysis was done by capillary Gas Chromatography using Single Ion Monitoring (SIM) mode GC/MS. All samples are reported on a dry weight basis.

#### HOLDING TIMES:

The samples were extracted within the recommended fourteen day holding time. After extraction all samples were analyzed within the recommended 40 day extract holding time.

#### BLANKS:

No tributyltin was detected in the laboratory blanks.

#### SURROGATES:

Recovery of the surrogate spike, Tripropyltin, ranged from 55% to 166% except for two samples, -518209 and SQ5012 which had 259% and 199% respectively. Data for both these samples was "J" qualified due to the high surrogate recoveries. No surrogate recovery QC limits have been established for this method.

#### MATRIX SPIKE AND MATRIX SPIKE :

Both the matrix spike and spike duplicate had significant native compound interference. Recoveries ranged from 81% to 156% except for a high recovery, 310%, of dibutyhin chloride in LMX1. The compound interferences for tributyhin chloride in LMX2 resulted in very high recoveries and consequently there is No Analytical Result (NAR) reported for this sample. The Relative Percent Differences (RPD) ranged from 13.7% to 31.7% for tetrabutyl and monobutykin. No percent recovery or RPD limits have been established for this method.

#### ANALYTICAL COMMENTS:

The samples were analyzed at a higher concentration, which is reflected in the lower quantitation limits. Consequently most of the samples had to be diluted and reanalyzed to bring the organotin concentrations within the linear calibration range. The "E" flag was given to tributyltin in two samples. -518150 and -518180, since even after dilution the concentration was just above the calibration curve.

The principle analytical problems encountered were directly related to the chromanographic interference's in some of the samples. This was a problem with tributyltin and dibutyltin in the matrix spikes

Two additional samples were analyzed with the sediment samples. These were Sequim Bay Reference Sediments which presumably were spiked with 100 ng/gm (100 ug/Kg) wer weight of the reference in No value for tributyltin has been established for the Sequim Bay Reference Sediment so the accuracy of the analysis cannot be determined. These samples are identified as SQ5012 and SQ5012D

SQ5012	108	ug/Kg (wet weight)	Tributyltin
SQ5012D	95.4	ug/Kg (wet weight)	Tributyltin

Note that the data sheets report these values as dry weight. The percent solids is 60.4% for these samples.

# DATA QUALIFIER CODES:

U	-	The analyte was not detected at or above the reported value.
1	-	The analyte was positively identified. The associated numerical value is an estimate.
ហ	-	The analyte was not detected at or above the reported estimated result.
REJ	-	The data are <u>unusable</u> for all purposes.
EXP	-	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .
NAF	-	Not analyzed for.
N	-	For organic analytes there is evidence the analyte is present in this sample.
NJ .	-	There is evidence that the malyte is present. The associated numerical result is an estimate.
E	-	This qualifier is used when the concentration of the associated value exceeds the known calibration range.
bold	-	The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)

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Hybbos Bottom Sections 12/94

# Manchester Environmental Laboratory

7411 Beach Dr E, Port Oschard Washington 98366

# CASE NARRATIVE

#### February 15, 1995

Subject: Commencement Bay sediments 49

Samples: 94498187 - 190

Case No. 160894

Officer:

By:

160894 Dale Norton Greg Perez

Organics Analysis Unit

# **VOLATILE ORGANIC ANALYSIS**

#### SUMMARY:

No difficulties were encountered in the analysis of these samples. The data is usable as qualified.

#### ANALYTICAL METHODS:

Volatile organic compounds were analyzed using Manchester manification of the EPA SW 846 Method 8260 purge-trap procedure with capillary GC/MS analysis. Normal QAQC procedures were performed on the samples.

#### BLANKS:

Low levels of the common laboratory solvents acetome and methylene chloride were detected in the laboratory blanks. If the concentrations of the compounds in the sample are greater than or equal to five times the concentrations of the compounds in the associated method blank, they are considered native to the sample.

#### SURROGATES:

Surrogate recoveries were within acceptable limits for the sediment samples.

#### HOLDING TIMES:

The sediment samples were analyzed within the recommended 14 day holding time.

# MATRIX SPIKE AND MATRIX SPIKE DUPLICATE:

Sediment matrix spikes were within acceptable QC limits for both percent recovery and Relative Percent Differences (RPD).

### DATA QUALIFIER CODES:

1

U - The analyte was not detected at or above the reported value.

- The analyte was positively identified. The associated numerical value is an estimate.

UJ - The analyte was not detected at or above the reported estimated result.

REJ - The data are unusable for all purposes.

EXP  $\frac{1}{5}$  The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals  $3 \ge 10^6$ .

NAF - Not analyzed for.

N - For organic analytes there is evidence the analyte is present in this sample.

NJ - There is evidence that the analyte is present. The issociated numerical result is an estimate.

E - This qualifier is used when the concentration of the associated value exceeds the known calibration range.

bold - The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)

Bottom Scelimit 11/94

7411 Beach Drive E, Port Orchard Washington 98366

#### CASE NARRATIVE

#### January 27, 1995

Subject: Commencement Bay Sediments

Samples: 94 - 498180, -498181, -498184 to -498188, and -498191 to-498198

Case No. 1608-94

Officer: Dale Norton

By:

Dickey D. Huntamer (00," Organics Analysis Unit

#### SEMIVOLATILE ORGANICS

#### ANALYTICAL METHODS:

The semivolatile soil samples were Soxhlet extracted with acetone following the Manchester modification of the EPA SW 846 8270 procedure with capillary GC/MS analysis of the sample extracts. Normal QA/QC procedures were performed with the analyses.

#### HOLDING TIMES:

All sample and extraction holding times were within the recommended limits.

#### **BLANKS:**

A number of target compounds were detected in the laboratory blanks. The EPA five times rule was applied to all target compounds which were found in the blank. Compounds that were found in the sample and in the blank were considered real and not the result of contamination if the levels in the sample are greater than or equal to five times the amount of compounds in the associated method blank.

The target compounds detected in the lab blanks may be due to carryover in the automated Gel Permeation Chromatography (GPC) unit. Procedures have been implemented to reduce this problem in the future.

#### SURROGATES:

The normal surrogates compounds were added to the sample prior to extraction. All surrogate recoveries were within acceptable limits, except for the two laboratory blanks which had double the expected recovery. This probably due to these samples being double spiked with surrogate compounds. No qualifiers were added due to the high recoveries.

# MATRIX SPIKE AND MATRIX SPIKE DUPLICATE:

Matrix spikes were extracted with these samples Recoveries were acceptable with the exception of the following compounds 1,3-dichlorobenzene, hexachloroethane, 4-chloroaniline, hexachlorocyclopentadiene, 3 and 4-nitroaniline, and di-n-butylphthalate. The "J" qualifier was added to the results for these compounds in the matrix spike source sample, -498192.

#### ANALYTICAL COMMENTS:

There were no other problems with the analysis and the data is acceptable for use as qualified.

#### DATA QUALIFIER CODES:

U	÷	The analyte was not detected at or above the reported value.		
1		The analyte was positively identified. The associated numerical value is an estimate.		
UJ	-	The analyte was not detected at or above the reported estimated result.		
REJ	•	The data are <u>unusable</u> for all purposes.		
EXP	<b>-</b> .	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .		
NAF	-	Not analyzed for.		
N	•	For organic analytes there is evidence the analyte is present in this sample.		
NJ	-	There is evidence that the analyte is present. The associated numerical result is an estimate.		
E	<b>-</b> 1	This qualifier is used when the concentration of the associated value exceeds the known calibration range.		
bold	-	The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)		

CN\_CBAY4.DOC - 5

# MANCHESTER ENVIRONMENTAL LABORATORY 7411 Beach Drive E, Port Orchard Washington 98366 CASE NARRATIVE March 3, 1995

Subject: **Commencement Bay Sediments** 94 - 498190 Samples: Case No. 1608-94 Officer: Dale Norton

Dickey D. Huntamer

Organics Analysis Unit

By:

# SEMIVOLATILE ORGANICS

#### **ANALYTICAL METHODS:**

This sample was inadvertently left off the request for analysis. Consequently it was not analyzed with the other semivolatile samples. The project officer requested that it be analyzed if possible, despite the fact that the fourteen day holding time was exceeded. The semivolatile soil samples were Soxhlet extracted with acetone following the Manchester modification of the EPA SW 846 8270 procedure with capillary GC/MS analysis of the sample extracts. Normal QA/QC procedures were performed with the analyses except for matrix spikes.

#### **HOLDING TIMES:**

The sample was extracted after the recommended fourteen day holding time. All sample results were qualified by adding "J". Extract holding times were within recommended limits.

#### BLANKS:

One blank was analyzed with the sample. Several target compounds were detected in the laboratory blank. The EPA five times rule was applied to all target compounds which were found in the blank. Compounds that were found in the sample and in the blank were considered real and not the result of contamination if the levels in the sample are greater than or equal to five times the amount of compounds in the associated method blank.

#### SURROGATES:

The normal surrogates compounds were added to the sample prior to extraction. All surrogate recoveries were within acceptable limits.

MATRIX SPIKE AND MATRIX SPIKE DUPLICATE:

No matrix spikes were extracted with these samples.

# ANALYTICAL COMMENTS:

There were no problems other than the holding time which resulted in all of the data being "J" qualified.

# DATA QUALIFIER CODES:

U	-	The analyte was not detected at or above the reported value.
1	<b>-</b> .	The analyte was positively identified. The associated numerical value is an estimate.
UJ	-	The analyte was not detected at or above the reported estimated result.
REJ	-	The data are <u>unusable</u> for all purposes.
EXP	-	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .
NAF	-	Not analyzed for.
N		For organic analytes there is evidence the analyte is present in this sample.
NJ	-	There is evidence that the analyte is present. The associated numerical result is an estimate.
E	-	This qualifier is used when the concentration of the associated value exceeds the known calibration range.
bold	-	The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)

CN\_CBAY6.DOC - 5

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Bottom Sectional 12/94

7411 Beach Drive E, Port Orchard Washington 98366

#### CASE NARRATIVE

#### January 28, 1995

Subject: Commencement Bay Sediments -49

Samples: 94 - 498180, -498181, -498184 to -498188, and -498190

Case No: 1608-94

Officer: Dale Norton

By: Dickey D. Huntamer Organics Analysis Unit

#### POLYCHLORINATED BIPHENYLS

#### ANALYTICAL METHODS:

The soil samples were Soxhlet extracted using acetone as the solvent. Analysis was done by EPA Method 8080 using dual column capillary GC analysis with Electron Capture Detectors (ECD).

#### HOLDING TIMES:

All extraction and analysis holding times were met.

#### BLANKS:

No target compounds were detected in the laboratory blanks.

#### SURROGATES:

Surrogates recoveries for the samples ranged from 59% to 151% and were within the acceptable limits.

#### MATRIX SPIKE AND MATRIX SPIKE :

Matrix spike recoveries ranged from 67% to 82% and were within acceptable limits. The Relative Percent Differences (RPD) ranged from 7.7% to 20%.

#### **ANALYTICAL COMMENTS:**

Page 2 Commencement Bay - PCB

### AMALYTICAL COMMENTS:

There were no significant problems with the PCB analysis and the data is acceptable as qualified. No Canadian sediment reference materials were analyzed with this sample set.

### DATA QUALIFIER CODES:

U	•	The analyte was not detected at or above the reported value.
1	-	The analyte was positively identified. The associated numerical value is an estimate.
UJ	-	The analyte was not detected at or above the reported estimated result.
REJ	-	The data are <u>unusable</u> for all purposes.
EXP	•	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .
NAF	<b>-</b> ·	Not analyzed for.
N	-	For organic analytes there is evidence the analyte is present in this sample.
NJ	-	There is evidence that the analyte is present. The associated numerical result is an estimate.
E	-	This qualifier is used when the concentration of the associated value exceeds the known calibration range.
*	-	The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)

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7411 Beach Drive E, Port Orchard Washington 98366

#### CASE NARRATIVE

#### March 31, 1995

Subject: Commencement Bay Sediments - 49

Samples: 94 - 498180, -498181, -498184 to -498188, -498190, -498194 to -498198

Case No. 1608-94

Officer: Dale Norton

By:

Dickey D. Huntamer

#### TRIBUTYL TINS

#### ANALYTICAL METHODS:

The samples were extracted following the methods given in Puget Sound Estuary Program (PSEP) "Recommended Guidelines for Measuring Organic Compounds in Puget Sound Sediment and Tissue Samples" Recommended Methods for Organotin Compounds. The samples were Soxhlet extracted using acetone and tropolone, 0.2% by weight, solvent exchanged to hexane and dried using sodium sulfate. The organotin compounds were hexylated using the Grignard reaction given in Krone et al (1989) including the silica gel/alumina cleanup. Analysis was done by capillary Gas Chromatography using Single Ion Monitoring (SIM) mode GC/MS. All samples are reported on a dry weight basis.

#### HOLDING TIMES:

The samples were extracted within the recommended fourteen day holding time. After extraction all samples were analyzed within the recommended 40 day extract holding time.

#### BLANKS:

A small amount of tributyltin was detected in the duplicate laboratory blank, BLNK5608. No tributyltin was detected in the first blank, BLNK5607. The EPA five times rule was applied to all target compounds which were found in the blank. Compounds that were found in the sample and in the blank were considered real and not the result of contamination if the levels in the sample are greater than or equal to five times the amount of compounds in the associated method blank.

#### SURROGATES:

Recovery of the surrogate spike, Tripropyltin, ranged from 30% to 106%. The "J" qualifier was added to the results for samples -498184, -498197 and SQ4348 because the surrogate recoveries were less than 50%. No surrogate recovery QC limits have been established for this method.

#### MATRIX SPIKE AND MATRIX SPIKE :

Both the matrix spike and spike duplicate had significant native compound interference. Recoveries ranged from 20% to 109%. The Relative Percent Differences (RPD) ranged from 5.4% to 95%. No percent recovery or RPD limits have been established for this method.

#### ANALYTICAL COMMENTS:

The samples were analyzed at a higher concentration, which is reflected in the lower quantitation limits. Consequently most of the samples had to be diluted and manalyzed to bring the organotin concentrations within the linear calibration range.

The principle analytical problems encountered were directly related to the chromatographic interference's in some of the samples. This was a problem with tributyltin and dibutyltin in the matrix spikes

Two additional samples were analyzed with the sectiment samples. These were Sequim Bay Reference Sediments which presumably were spiked with 100 ng/gm (100 ug/Kg) wet weight of tributyltin. No value for tributyltin has been established for the Sequim Bay Reference Sediment so the accuracy of the analysis cannot be determined. These samples are identified as SQ4348 and SQ4348D

SQ4348	75.3	ug/Kg (wei weight)	Tributyltin
SQ4348D	61.9	ug/Kg (wet weight)	Tributyltin

Note that the data sheets report these values as dry weight. The percent solids is 58.4% for these samples.

# DATA QUALIFIER CODES:

U	*	The analyte was not detected at or above the reported value.		
J	-	The analyte was positively identified. The associated numerical value is an estimate.		
UJ	-	The analyte was not detected at or above the reported estimated result.		
REJ	-	The data are <u>unusable</u> for all purposes.		
EXP	-	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .		
NAF	-	Not analyzed for.		
N	-	For organic analytes there is evidence the analyte is present in this sample.		
IJ	-	There is evidence that the analyte is present. The associated numerical result is an estimate.		
E	-	This qualifier is used when the concentration of the associated value exceeds the known calibration range.		
bold	•	The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)		

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Particulates,

11/94-6/95

#### MANCHESTER ENVIRONMENTAL LABORATORY

7411 Beach Drive E, Port Orchard Washington 98366

#### **CASE NARRATIVE**

August 28, 1995

Subject: Commencement Bay

Samples: 95 - 248762

Case No. 1942 - 95

Officer: Dale Norton

By: Dickey D. Huntamer OUM Organics Analysis Unit

#### **VOLATILE ORGANIC ANALYSIS**

#### ANALYTICAL METHODS:

Volatile organic compounds were analyzed using Manchester modification of the EPA SW 846 Method 8260 purge-trap procedure with capillary GC/MS analysis. Normal QA/QC procedures were performed on the samples.

#### **BLANKS:**

Low levels of the common laboratory solvents acetone and methylene chloride were detected in the laboratory blanks The EPA five times rule was applied to all target compounds which were found in the blank. Compounds that were found in the sample and in the blank were considered real and not the result of contamination if the levels in the sample are greater than or equal to five times the amount of compounds in the associated method blank.

#### SURROGATES:

Surrogate recoveries were within acceptable limits for sediment samples.

#### HOLDING TIMES:

The samples were analyzed within the recommended 14 day holding time.

#### MATRIX SPIKE AND MATRIX SPIKE DUPLICATE:

Matrix spikes were within acceptable QC limits for both percent recovery and RPD except for acetone, 2butanone, bromodichloromethane, cis and trans-1,3-dichloropropene, dibromochloromethane, bromoform, and 1,2,4--trichlorobenzene. The "J" qualifier was added to the sample results for these compounds. Hexachlorobutadiene, naphthalene p-isopropyltoluene and 1,2,3-trichlorobenzene were outside acceptable limits due to high native amounts interfering. No qualifiers were added to these compound results.

#### ANALYTICAL COMMENTS:

There were problems with the blanks due to carry over. A continuing check standard was analyzed just prior to the laboratory blank, KBS5178. Numerous compounds were detected in the blank. Normally two blanks are run after a standard and the first one is discarded. Due to holding times and the length of the sample analyses run the second blank was not analyzed. A follow-up blank was done after the samples and it also showed considerable contamination. The EPA five times rule was applied to the sample data in those situations where target compounds were detected in both the sample and blank. This resulted in higher than normal quantitation limits for some compounds and the possibility of not detecting some compounds which may have been present.

### DATA QUALIFIER CODES:

U	-	The analyte was not detected at or above the reported value.
J		The analyte was positively identified. The associated numerical value is an estimate.
យ	-	The analyte was not detected at or above the reported estimated result.
REJ	-	The data are <u>unusable</u> for all purposes.
EXP	-	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .
NAF	-	Not analyzed for.
N	-	For organic analytes there is evidence the analyte is present in this sample.
NJ	•	There is evidence that the analyte is present. The associated numerical result is an estimate.
E	-	This qualifier is used when the concentration of the associated value exceeds the known calibration range.
bold	-	The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)

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Particulates 11/914 - 6/95

#### Manchester Environmental Laboratory

7411 Beach Dr E, Port Orchard Washington 98366 October 2, 1995

Project: Commencement Bay

Samples: 95248706, 248707, 248710, 248717, 248727, 248734, 248743, 248750, 248762, 248773, 248774, 248786

By: Karin Feddersen KF

These samples were analyzed for Semivolatiles (BNA's) by Puget Sound Estuary Program (PSEP) protocols.

#### Holding Times:

The samples were extracted and analyzed within PSEP holding times.

#### Method Blanks:

Many target analytes were detected in the method blanks. When these analytes were also detected in the samples at less than five (5) times the on-column amount detected in the method blanks, the results were qualified with a "U". In these instances, these analytes are most likely present due to laboratory contamination and not native to the sample.

When these analytes were detected in the samples at greater than 5 times the method blank, no corrective action was required. The amount of laboratory contamination contributing to the results is not significant.

#### GC/MS Tuning and Calibration:

Calibration against Decafiuorotriphenylphosphine (DFTPP) is acceptable for the initial calibration, continuing calibrations and all associated sample analyses.

#### Initial Calibration:

The initial calibration met the minimum response criteria for the average relative responses. The % Relative Standard Deviations were within the maximum of 30% with two exceptions which do not affect the results.

#### Continuing Calibration:

The average relative response factors for all target analytes were above the minimums, and the percent deviations between the initial and continuing calibration standards were within the maximum of 25% with two notable exceptions on September 7, Benzidine and 3,3-Dichorobenzidine. These analytes were not detected in any of the samples. All results for these two analytes have been qualified with a "UJ" in the corresponding samples.

#### Matrix Spikes (MS/MSD):

Matrix spike recovery and precision data are reasonable, acceptable, and within QC limits with several exceptions. Results for these analytes in the samples may be biased low. This condition is typical due to the extensive cleanup procedures performed on these samples. The most volatile compounds exhibited 0% recovery.

Therefore, analytes with recoveries below 50%, when detected, have been qualified with a "J", and non-detected analytes been qualified with a "UJ". Analytes with both spike recoveries below 20% have been qualified "REF" when not detected in the samples.

#### Surrogates:

All surrogate recoveries were within QC limits with two exceptions. Recovery of 2-Fluorophenol was slightly below the QC limit of 25% in sample HS65213 (19.2%) and HS65213D (24.5%). Some analytes may have been affected in the same manner as the surrogates. However, the detection limits for these analytes are low enough that they would still be visible to the detector. No qualification of the results is necessary for this condition.

#### Sample Results:

This data is acceptable for use as amended.

#### DATA QUALIFIER CODES:

U - The analyte was not detected at or above the reported value.

- J The analyte was positively identified. The associated numerical value is an estimate.
- UJ The analyte was not detected at or above the reported estimated result.
- REJ The data are unusable for all purposes.

Porticilates 11/94 - 6/95

## Manchester Environmental Laboratory

7411 Beach Dr E Port Orchard Washington 98366 September 25, 1995

Project: Commencement Bay

Samples: 248734, 248743, 248750, 248751, 248762

By: Karin Feddersen KF

These samples were analyzed by EPA Method 8080 for Polychlorinated Biphenyls (PCBs) employing the dual column confirmation technique.

#### Holding Times:

These samples were extracted six days past the method holding time of fourteen days. PCBs are extremely stable. Therefore, extraction beyond the holding time should not affect the results. The samples were analyzed within the method holding time of forty days from extraction.

#### Method Blanks:

No analytes of interest were detected in the method blanks.

Initial Calibration:

The % Relative Standard Deviations were within the maximum of 30% for all target analytes.

#### **Continuing Calibration:**

The Percent Differences between the initial and continuing calibrations were within the maximum of 25% for all target analytes.

#### Surrogates:

Four surrogates were added to each sample. The recommended range for surrogate recovery is between 60% and 150%. Dibutylchlorendate (DBC) recoveries were low in most of the samples. An acid cleanup was performed on these samples. DBC is very susceptible to degradation by acid. Since PCBs are not susceptible to acid degradation, and because the other surrogates demonstrated acceptable recoveries, qualification of the results is not required.

4,4-Dibrooctafluorobiphenyl was low in sample 95248750. Decachlorobiphenyl and Tetrachloro-m-xylene recoveries were acceptable. No qualification of the data was warranted.

# Matrix Spikes (MS/MSD):

All matrix spike recoveries were between 75% and 110%. These recoveries are reasonable and acceptable.

# • Sample Results:

When the RPD between the two columns was greater than 30% for an analyte, the result was qualified with a "J".

This data is acceptable for use with the qualifications mentioned.

# DATA QUALIFIER CODES:

U - The analyte was not detected at or above the reported value.

J - The analyte was positively identified. The associated numerical value is an estimate.

UJ - The analyte was not detected at or above the reported estimated result.

NJ - There is evidence that the analyte is present. The associated numerical result is an estimate.

NAF - Not analyzed for.

REJ - The data are unusable for all purposes.

Particulates 11/14-6/95

## Manchester Environmental Laboratory

7411 Beach Dr. E Port Orchard Washington 98366 October 11, 1995

#### Project: Commencement Bay

Samples: 248706, 248710, 248717, 248720, 248727

By: Karin Feddersen Kr

These samples were analyzed according to Puget Sound Estuary Program (PSEP) protocol for Polychlorinated Biphenyls (PCBs) employing the dual column confirmation technique.

#### **Holding Times:**

These samples were extracted within the PSEP holding time. The samples were analyzed twenty-one days past the method holding time of forty days from extraction. PCBs are extremely stable. Therefore, analysis beyond the holding time should not affect the results.

#### Method Blanks:

No analytes of interest were detected in the method blank.

#### **Initial Calibration:**

The % Relative Standard Deviations were within the maximum of 30% for all target analytes.

#### **Continuing Calibration:**

The Percent Differences between the initial and continuing calibrations were within the maximum of 25% for all target analytes.

#### Surrogates:

There was insufficient sample to perform a separate extraction for PCBs on these samples. Therefore, the BNA extracts were analyzed for PCBs, and only the surrogates normally added for BNA analysis are present in these extracts. The results for the surrogates normally added to extracts for PCB analysis have been qualified "NAF".

One BNA surrogate, 2-Fluorobiphenyl (FBP), is chemically similar to PCBs, and its recovery was therefore evaluated for each sample. The EPA method 8270 (BNA) QC limits for FBP recovery are 30% to 115%. No limits have been established for this surrogate's use in the PCB method. FBP recoveries for these samples ranged from 41% to 53%. These recoveries are acceptable, although lower than those reported for the BNA analyses of these samples. The

1

differences in surrogate recovery can be most likely explained by differences in the two methods used, and are not due to loss of sample material in any way.

These recoveries are acceptable and within expected parameters, and no qualification of the results is necessary.

### Sample Results:

This data is acceptable for use as reported.

# **DATA QUALIFIER CODES:**

- U The analyte was not detected at or above the reported value.
- J The analyte was positively identified. The associated numerical value is an estimate.
- UJ The analyte was not detected at or above the reported estimated result.
- NJ There is evidence that the analyte is present. The associated numerical result is an estimate.

NAF - Not analyzed for.

REJ - The data are unusable for all purposes.

Particulates 11/94-6/95

#### MANCHESTER ENVIRONMENTAL LABORATORY

7411 Beach Drive E, Port Orchard Washington 98366

### CASE NARRATIVE

#### November 3, 1995

Subject: Commencement Bay

Samples:

: 95 - 248706, -248707, -248710, -248717, -248720, -248727, -248734, -248743, -248750 and -248762

Case No. 1942 -95

Officer: Dale Norton

By:

Dickey D. Huntamer

#### TRIBUTYL TINS

#### ANALYTICAL METHODS:

The samples were extracted following the methods given in Puget Sound Estuary Program (PSEP) "Recommended Guidelines for Measuring Organic Compounds in Puget Sound Sediment and Tissue Samples" Recommended Methods for Organotin Compounds. The samples were Soxhlet extracted with acetone and tropolone, 0.2% by weight, solvent exchanged to hexane and dried using sodium sulfate. The organotin compounds were hexylated using the Grignard reaction given in Krone et al (1989) including the silica gel/alumina cleanup. Analysis was done by capillary Gas Chromatography using Single Ion Monitoring (SIM) mode GC/MS. Verification screening to check for interference's was done using Atomic Emission Spectroscopy. All samples are reported on a dry weight basis.

#### HOLDING TIMES:

The samples were stored frozen following PSEP Guidelines until extraction. After extraction all samples were analyzed within the recommended 40 day extract time.

#### **BLANKS:**

No target compounds were detected in the laboratory blank.

#### SURROGATES:

Recovery of the surrogate spike, Tripropyltin, ranged from 54% to 150%. Some of the recoveries were greater than 100% due to chromatographic interference with the quantitation ion. No surrogate recovery QC limits have been established for this method.

#### MATRIX SPIKE AND MATRIX SPIKE :

The matrix caused some interference problems with determining the organotin recoveries. A quick screen was made using the atomic emission detector (AED) and organotin recoveries reported were based on the analyses.

#### ANALYTICAL COMMENTS:

Some samples had chromatographic interference's with the organotin peaks, particularly monobutyl tin. The samples were screened on the AED to confirm whether organotins were present.

Internal standard area counts fell below the +/-50% limits for several samples. The "J" qualifier was added to the data results for those samples. Generally results from the samples with low area counts was comparable to the results from the dilution analyses indicating that there was little effect on the quantitation.

Two additional samples were analyzed with the sediment samples. This was a Sequim Bay Reference Sediment which presumably was spiked with 100 ng/gm (100 ug/Kg) wet weight of tributyltin. No value for tributyltin has been established for the Sequim Bay Reference Sediment so the accuracy of the analysis cannot be determined. These samples are identified as SBR53598 and SBR53599.

SBR53598	67	ug/Kg (wet weight)	Tributyltin
SBR53599	57	ug/Kg (wet weight)	Tributyltin

Note that the data sheets report these values as dry weight. The percent solids is 61.2% for these samples.

#### DATA QUALIFIER CODES:

U	-	The analyte was not detected at or above the reported value.
J	-	The analyte was positively identified. The associated numerical value is an estimate.
UJ	-	The analyte was not detected at or above the reported estimated result.
REJ	-	The data are <u>unusable</u> for all purposes.
EXP	-	The result is equal to the number before EXP times 10 to the power of the number after EXP. As an example 3EXP6 equals $3 \times 10^6$ .
NAF	-	Not analyzed for.
N	**	For organic analytes there is evidence the analyte is present in this sample.
NJ	<b>-</b>	There is evidence that the analyte is present. The associated numerical result is an estimate.
Е	-	This qualifier is used when the concentration of the associated value exceeds the known calibration range.
bold	-	The analyte was present in the sample. (Visual Aid to locate detected compound on report sheet.)

#### CN\_CBYT3.DOC

# Appendix C

# Physical/Chemical Analysis of Settling Particulate Matter (Dry Weight) June 1991 to June 1995

# Hylebos Waterway

Table C1: Conventionals and Metals Analysis
Table C2: Volatiles Analysis
Table C3: Semivolatiles and PCB Analysis
Table C4: Butyltins Analysis
Table C5: Sediment Accumulation Rates

Sitcum Waterway Table C6: Conventionals and Metals Analysis Table C7: Semivolatiles Analysis Table C8: Sediment Accumulation Rates

Thea Foss Waterway Table C9: Conventionals and Metals Analysis Table C10: Semivolatiles and Butyltins Analysis Table C11: PCB Analysis Table C12: Sediment Accumulation Rates

# Hylebos Waterway

Table C1: Results of Conventionals and Metals Analysis of Settling Particulate Matter from Hylebos Waterway,

$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	Location						Ū	Upper Turning Basin	ing Basin			·			
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	ation No.		-					H	1	******	******				
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	ollection	12/91-	2/92-	5/92-	8/92-	11/92-	2/93-		9/93-	12/93-	3/94-	5/94-	8/94-	-11/94-	3/95-
	eriod	2/92	5/92	8/92	11/92	2/93	5/93	9/93	12/93	3/94	5/94	8/94	11/94	3/95	C6/9
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	olids (%)	43		1	30	46	40	39	І	42	30	34	37	45	28
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	oc (%)	. 6.0	I	I	I	8.5	7.5	7.1	ł	5.1	7.1	6.3	5.2	4.4	5.6
	etals														
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	ntimony	16	I	i	14	35	11	yund yund	ł	ла	na	na	na	па	na
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	rsenic	16	ł	ı	130	100	60	87	ŧ	59	74	54	45	34	44
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	admium	16.0	1	*	1.3	0.88	0.93	1.7	I	na	na	ла	na	па	na
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	opper	120	ł	ł	170 j	130	130	170	I	140	160	130	130	80	130
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	lercury	0.61 j	1	I	0.28 j	0.29	0.37	0.38	I	na	na	0.43	0.36	0.32	0.39
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	ead .	75	I	I	87	80	70	78	1	99	69	55	50	35	49
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	ickel	40	I	1	48	45	51	39	I	49.	38	na	มล	na	na
n $H-2$ $H-2$ No. $H-2$ $H-2$ No. $11/9$ $9/92$ $11/92$ $2/92$ $3/92$ $11/92$ $3/94$ $5/94$ $8/94$ $11/94$ $31$ $2/92$ $5/92$ $8/92$ $11/92$ $2/93$ $7/93$ $3/94$ $5/94$ $8/94$ $11/94$ $31$ $7/92$ $5/92$ $8/92$ $11/92$ $2/93$ $7/93$ $3/94$ $5/94$ $8/94$ $11/94$ $31$ $7/92$ $5/92$ $5/93$ $5/93$ $9/93$ $12/93$ $3/94$ $5/94$ $8/94$ $11/94$ $31$ $7/9$ $ 4.2$ $5.3$ $5.5$ $2.8$ $4.2$ $5.9$ $4.5$ $4.7$ $37$	inc	220	1	1	520	270	280	310	1	250	310	250	260	150	220
n $H-2$ $H-2$ No. $11-2$ $11-2$ $11-2$ No. $11-2$ $11-2$ $11-2$ No. $1291 2192 5192 8192 11/92 2193 3794 5794 8794 11/9$ $\pi/2$ $-1$ $41$ $45$ $68$ $45$ $41$ $44$ $43$ $374 374 374 317 \pi/2$ $-1$ $41$ $45$ $68$ $45$ $41$ $44$ $43$ $37 37-$											-				
No. $H-2$ on         12/91-         2/92-         5/92-         11/92-         2/93-         3/94-         5/94-         8/94-         11/9 $2/92$ 5/92-         11/92-         2/93-         7/93-         9/93         12/93-         3/94-         5/94-         8/94-         11/94-         3/ $\%$ -         42         41         45         68         45         41         44         3/         3/7 <td< td=""><td>cation</td><td></td><td></td><td></td><td></td><td></td><td></td><td>@ Genera</td><td>l Metals</td><td></td><td></td><td></td><td></td><td></td><td></td></td<>	cation							@ Genera	l Metals						
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	ation No.							H-	ŗ						
2/92 $5/92$ $8/92$ $11/92$ $2/92$ $8/92$ $11/92$ $2/93$ $3/94$ $5/94$ $8/94$ $11/94$ $31$ $8/9$ $ 42$ $41$ $45$ $68$ $45$ $41$ $44$ $43$ $34$ $37$ $37$ $8/9$ $ 4.6$ $5.5$ $5.5$ $2.8$ $4.2$ $5.0$ $4.6$ $4.5$ $5.1$ $4.8$ $5.0$ $4$ $8/9$ $ 11$ $11$ $12$ $7.1$ $11$ $1.8$ $5.0$ $4.5$ $5.1$ $4.8$ $5.0$ $4$ $8/9$ $8/9$ $69$ $68$ $61$ $67$ $42$ $41$ $8/9$ $170$ $180$ $71$ $160$ $170$ $180$ $140$ $11$ $8/9$ $170$ $170$ $180$ $71$ $116$ $120$ $120$ $120$ $120$ $120$ $120$ $120$	llection		2/92-	5/92-	8/92-	11/92-	2/93-	7/93-	9/93	12/93-	3/94-	5/94-	8/94-	-+	3/95-
$\pi$	riod	2/92	5/92	8/92	11/92	2/93	5/93	9/93	12/93	3/94	5/94	8/94	11/94	3/95	6/95
b) $-4.6$ 5.5 5.5 2.8 4.2 5.0 4.6 4.5 5.1 4.8 5.0 4 -11 11 11 12 7.4 30 7.1 11 1a na	lids (%)		42	41	45	68	45	41	44	43	34	37	37	37	33
uy       -       11       11       12       7.4       30       7.1       11       na       na <t< td=""><td>C (%)</td><td>ł</td><td>4.6</td><td>5.5</td><td>5.5</td><td>2.8</td><td>4.2</td><td>5.0</td><td>4.6</td><td>4.5</td><td>5.1</td><td>4.8</td><td>5.0</td><td>4.7</td><td>4.9</td></t<>	C (%)	ł	4.6	5.5	5.5	2.8	4.2	5.0	4.6	4.5	5.1	4.8	5.0	4.7	4.9
-       11       11       12       7.4       30       7.1       11       na       na <t< td=""><td>etals</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td></t<>	etals														
-     81     81     85     30     98     69     68     61     67     42     41       -     1.3     1.6     1.6     0.51     1.2     1.4     1.5     na     na     na     na       -     160     170     180     71     160     170     180     170     170     130     140     1       -     0.6     j     0.54     j     0.27     0.7     0.48     0.5     na     na     0.53     0.55     0       -     110     100     110     49     100     100     100     93     91     73     71       -     47     46     45     42     52     41     45     47     46     na	ntimony	1	11	11	12	7.4	30	7.1	11	na	ла	na	na	na	na
-     1.3     1.6     1.6     0.51     1.2     1.4     1.5     na     na     na     na       -     160     170     180     71     160     170     180     130     140     1       -     0.6     0.54     0.54     0.27     0.7     0.48     0.5     na     na     0.53     0.55     0       -     110     100     110     49     100     100     100     93     91     73     71       -     47     46     45     52     41     45     47     46     na     na	rsenic	I	81	81	85	30	98	69	68	61	67	42	41	43	45
-       160 j       170 j       180 j       71       160       170       130       140       1         -       0.6 j       0.54 j       0.27       0.7       0.48       0.5       na       na       0.53       0.55       0         -       110       100       110       49       100       100       100       93       91       73       71         -       47       46       45       42       52       41       45       47       46       na       na       0.53       0.55       0	admium	I	1.3	1.6	1.6	0.51	1.2	1.4	1.5	na	па	na	na	រាន	na
-     0.6 j     0.54 j     0.57 j     0.7 0.48 0.5     0.5 na     na     0.53 0.55 0.55 0.       -     110     100     110     49     100     100     100     93     91     73     71       -     47     46     45     42     52     41     45     46     na     na	opper	I	160 j	170 j	180 j	11	160	170	170	160	170	130	140	130	120
- 110 100 110 49 100 100 100 93 91 73 71 - 47 46 45 42 52 41 45 47 46 na na	fercury		0.6 j	0.54 j	0.54 j	0.27	0.7	0.48	0.5	na	na	0.53	0.55	0.50	0.48
1 - 47 46 45 42 52 41 45 47 46 na na	cad	*	110	100	110	49	100	100	100	93	91	73	11	75	68
	lickel	ł	47	46	45	42	52	41	45	47	46	na	na	na	na
	Zinc	*	300	300	300	190	310	270	290	270	300	240	240	230	210
							•								

Table C1(cont): Results of Conventionals and Metals Analysis of Settling Particulate Matter from Hylebos Waterway,

H - 3           Station No.           Partical 12/91- 2/92- 5/92- 8/92- 11/92- 2/93- 7/93- 9/93- 12/93- 3/94           Period         2/92         5/92- 8/92- 8/92- 11/92- 2/93- 7/93- 9/93- 12/93- 3/94           Solids (%)         49         41         47         49         51         48         -         503- 12/93- 3/93           Solids (%)         49         71         13         5.3         4.8         4.0         3.6         -         4.2         4.4           Antimony         15         12         097         13         1.2         0.93         12/93         3/94           Antimony         15         10         13         1.3         0.79         0.97         1.3         1.2         0.93         2         4.4         4.0         5.5         9/93         12/93         3/94           Antimony         15         0.79         0.97         1.0         110         110         110         110         120         120         120         120           Antimory         0.79         0.97         1.30         1.70         1.20         2.93         2.93         2.93         2.93         2.94         3.94 <t< th=""><th></th><th></th><th></th><th></th><th>@ Elf Atochem</th><th>ochem</th><th></th><th></th><th></th><th></th><th></th><th></th><th></th></t<>					@ Elf Atochem	ochem							
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$				*********	H-3				5				
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	2/92-	5/92-	8/92	11/92-	2/93-	-£6/1	9/93-	12/93-	3/94-	5/94-	8/94-	11/94- <sup>-</sup>	3/95-
	5/92	8/92	11/92	2/93	5/93	9/93	12/93	3/94	5/94	8/94	11/94	CK/S	CK/0
	44	47	49	51	48	ł	50	46	38	36	4	42	37
		5.3	4.8	4.0	3.6	ł	4.2	4,4	5.2	5.0	4.2	4.6	5.4
		12	11	17	18	I	10	na	na	na	na	na	na
		80	61	70	71	ł	49	65	. 89	45	40	46	47
	· • • •	1.3	1.2	0.95	0.93	ł	1.2	na	na	na	na.	រាន	na
		160 j	170 j	150	130	I	160	150	150	130	130	130	120
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	•	0.58 j	0.61 j	0.55	0.93	I	0.48	na	na	na	na	0.64	0.47
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$		110	110	100	93	I	95	<u>8</u>	88	72	68	71	61
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$		45	47	53	50	ł	45	45	36	na	na	па	na
No.       Near Lincoln Drain         No. $H-4$ No. $H-4$ No. $H-4$ No. $12/91 2/92 8/92 11/92 2/93 9/93 12/93$ $2/92$ $5/92$ $8/92 11/92 2/93 7/93 9/93 12/93 2/92$ $5/92$ $8/92 11/92 2/93 7/93 9/93 12/93 2/92$ $5/92$ $8/92 11/92 2/93 7/93 9/93 12/93 3/1-1/9 2/92$ $5/92 8/92 11/92 2/93 7/93 9/93 12/93 3/1-1/9 8/9   4.3$ $4.0$ $3.6$ $na$ $na$ $mu$ $   4.3$ $4.0$ $3.6$ $na$ $na$ $mu$ $               -$		260	250	220	230	1	230	220.	230	190	180	170	170
No. $H-4$ an       12/91-       2/92-       5/92-       8/92-       11/92-       2/93-       9/93-       12/93 $2/92$ 5/92       5/92       8/92-       11/92-       2/93-       9/93-       12/93       3/ $8/9$ -       -       48       51       44       47       na       na       na $8/9$ -       -       4.5       4.3       4.0       3.6       na       na       3 $8/9$ -       -       4.5       4.3       4.0       3.6       na       na       3 $8/9$ -       -       -       4.5       9.7       na       na       3 $9/9$ -       -       -       4.0       3.6       na       na       3 $9/9$ -       -       -       4.3       4.0       3.6       na       na       3 $9/9$ -       -       -       -       4.3       9.0       10.9       12       9 $9/9$ -       -       -       -       1.1       9.7       na       na       10				2	lear Linco	ln Drain							
$\begin{array}{cccccccccccccccccccccccccccccccccccc$					Η̈́	4							
$\begin{array}{rrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrr$	2/92-	5/92-	8/92-	11/92-	2/93-	7/93-	9/93-	12/93-	3/94-	5/94-	8/94-	11/94-	3/95-
(x)     -     -     48     51     44     47     na     na     3       (x)     -     -     -     4.5     4.3     4.0     3.6     na     na     3       ny     -     -     -     8.4     8.2     9.5     9.7     na     na     1a       ny     -     -     8.4     8.2     9.5     9.7     na     na       nu     -     -     1.1     0.99     0.8     0.72     na     na       nu     -     -     1.1     0.99     0.8     0.72     na     na       nu     -     -     160     150     150     150     na     na       y     -     -     9.41     0.46     0.49     na     na	5/92	8/92	11/92	2/93	5/93	9/93	12/93	3/94	5/94	8/94	11/94	3/95	6/95
x)       -       -       4.5       4.3       4.0       3.6       na       na       3         ony       -       -       4.5       4.3       4.0       3.6       na       na       1a         cony       -       -       8.4       8.2       9.5       9.7       na       na       na       na         c       -       -       55       50       60       55       na       na       na       na         nm       -       -       1.1       0.99       0.8       0.72       na       na       na         r       -       -       160       150       150       150       150       na       1         r       -       -       0.41       0.46       0.49       na       na <t< td=""><td></td><td>48</td><td>51</td><td>44</td><td>47</td><td>na</td><td>na</td><td>51</td><td>40</td><td>39</td><td>43</td><td>na</td><td>па</td></t<>		48	51	44	47	na	na	51	40	39	43	na	па
ony     -     -     8.4     8.2     9.5     9.7     na     na       lc     -     -     55     50     60     55     na     na       lum     -     -     1.1     0.99     0.8     0.72     na     na       r     -     -     111     0.99     0.8     0.72     na     na       r     -     -     160     j     150     150     150     na     na       ry     -     -     9.41     0.46     0.49     na     na       ry     -     -     95     93     96     88     na     na	1	4.5	4.3	4.0	3.6	na	na	3.7	3.7	4.0	3.6	па	na
yy     -     8.4     8.2     9.5     9.7     na     na       m     -     -     55     50     60     55     na     na       m     -     -     1.1     0.99     0.8     0.72     na     na       m     -     -     1.1     0.99     0.8     0.72     na     na       m     -     -     160     150     150     150     na     na     1       r     -     -     0.41     0.46     0.49     na     na     na       r     -     -     95     93     96     88     na     na													
m     -     55     50     60     55     na     na       m     -     -     1.1     0.99     0.8     0.72     na     na       -     -     -     160     j     150     j     150     na     na     1       r     -     -     -     160     j     150     j     0.49     na     na     1       r     -     -     -     0.41     j     0.46     0.49     na     na       r     -     -     95     93     96     88     na     na	*	8.4	8.2	9.5	6.7	na	na	na	113	na	na	na	na
m 1.1 0.99 0.8 0.72 na na 160 j 150 j 150 150 na na 1 / 0.43 j 0.41 j 0.46 0.49 na na na 95 93 96 88 na na	• 1	55	50	60	55	na	na	38	130	30	27	na	na
/ – – – 160 j 150 j 150 150 na na 1 / – – – 0.43 j 0.41 j 0.46 0.49 na na – – 95 93 96 88 na na	;	1.1	0.99	0.8	0.72	na	na	na	na	na	na	na	na
/ – – 0.43 ј 0.41 ј 0.46 0.49 па па – – 95 93 96 88 па па	I	160 j	150 j	150	150	na	na	150	130	120	120	na	na
– – – 95 93 96 88 na na	I	0.43 j	0.41 j	0.46	0.49	na	па	ua	na	na	na	na	na
	I	95	93	96	88	па	Пâ	76	. 71	61	58	па	na
1 40 40 48 44 na na	I	40	40	48	44	na	na	34	33	ua	na	na	na
		230	200	200	200	na	na	170	180	150	130	na	na

Table C1(cont): Results of Conventionals and Metals Analysis of Settling Particulate Matter from Hylebos Waterway,

Heise         Heise         Heise         Heise         Sign Sign Sign Sign Sign Sign Sign Sign				ı			Near Cenex	Shex							
$ \  \  \  \  \  \  \  \  \  \  \  \  \ $	ation No.						Η						- 100 TWO		
n $n$ $n$ $n$ $n$ $q$ <th>ollection</th> <th>12/91-</th> <th>2/92-</th> <th>5/92- 8/92</th> <th>8/92- 11/92</th> <th>11/92 2/93</th> <th>2/93 5/93</th> <th>7/93 9/93</th> <th>9/93- 12/93</th> <th>12/93 3/94</th> <th>3/94 5/94</th> <th>5/94 8/94</th> <th>8/94-</th> <th>11/94- 3/95</th> <th>3/95- 6/95</th>	ollection	12/91-	2/92-	5/92- 8/92	8/92- 11/92	11/92 2/93	2/93 5/93	7/93 9/93	9/93- 12/93	12/93 3/94	3/94 5/94	5/94 8/94	8/94-	11/94- 3/95	3/95- 6/95
	POT 7	a (1			54				50	40	41	42	43	na	na
(b)         3.4         -         4.5         4.5         -         -         na         na         7.4         -         7.4         -         1.4         0.7         9.4         -         9.4         9.4         9.5         9.4         -         -         1.4         0.3         4.4         9.4         0.3         4.4         1.4         0.3         4.4         1.4         0.3         4.4         1.4	lids (%)	50	1	00	55	ł	***	113	BII	÷ .	F.		2		
	oc (%)	3.4	I	4.5	4.5	I	I	na	па	ৰ ৰ	4.0	3.9	4.2	119	na
	letals														
$ \  \  \  \  \  \  \  \  \  \  \  \  \ $	untimony	11	. 1	10	8.8	I	ł	na	na	na	na	na	na	па	na
	vrsenic	52	I	46	50	1	ł	na	na	39	35	26	26	na	na
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	admium	0.7 j	1	0.95	1.0	ŧ	ł	па	na	ั้ยน	na	na	па	na	na
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	opper	140	t	150 j	150 j	1	ŧ	na	na	140	120	110	110	na	na
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	fercury	0.51	I	0.38 j	0.41 j	I	I	na	na	па	na	na	na	na	na
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	ead	92	. I	76	100	I	I	na	na	81	74	61	60	na	na
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	lickel	39	1	40	38	I	t	na	na	34	31	na	na	na	na
n         H-6           No         H-6           No         1291-         2/92-         5/92-         8/92-         11/92-         2/93-         3/94-         5/94-         8/94-         11/94-           %         49         49         5/92         8/92-         11/92-         2/93-         7/93-         9/93-         12/93-         8/94-         11/94-         3/95-           %         49         49         5/2         8/92         11/92-         2/93-         3/94-         5/94-         8/94-         11/94-         3/95           %         49         49         5/2         3/2         12/93-         3/94-         5/94-         8/94-         11/94-         3/95           %         49         49         5/2         3/2         2/93         3/94         5/94-         8/94-         11/94-           %         2/2         2/2         2/2         2/2         2/2         2/2         2/2         2/2         3/2         2/6         3/0         7/9         3/9           %         1/10         1/10         1/10         1/10         1/10         1/10         1/10         1/10         1/10         1/10	inc	170	I	200	200	I	1	na	na	170	160	120	120	na	na
No. $H-6$ on         12/91-         2/92-         5/92-         5/92-         11/92-         2/93-         7/93-         9/93-         12/93-         8/94-         8/94-         8/94-         11/94-         3/95-         3/94-         3/94-         3/94-         3/94-         3/95-         3/95-         3/94-         3/94-         3/94-         3/94-         3/94-         3/95-         3/95-         3/95-         3/95-         3/95-         3/94-         3/95-         3/9	cation		2			0	Occidental	Chemica	1						
$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$	ation No.						Η̈́	6					********		
2/92         5/92         8/92         11/92         2/93         5/93         9/93         12/93         3/94         5/94         8/94         11/94           %)         49         49         48         52         53         49         48         52         33         41         45           %)         2.5         2.7         3.6         3.3         2.9         49         48         52         38         41         45           %)         2.5         2.7         4.0         7         2.6         3.0         3.0           m         0.6 j         0.66         0.77         0.67         0.55         0.38         0.72         0.5         3.0         50         3.0           m         0.6 j         0.66         0.77         0.67         0.55         0.38         0.72         0.5         7         2         2         16         16         16           m         0.6 j         0.56         0.77         0.67         0.55         0.38         0.72         0.5         16         16         16         16           m         110         120         120         120         110         120 </td <td>ollection</td> <td>12/91-</td> <td>2/92-</td> <td>5/92-</td> <td>8/92-</td> <td>11/92-</td> <td>2/93-</td> <td>-£6//</td> <td>9/93-</td> <td>12/93</td> <td>3/94</td> <td>5/94-</td> <td>8/94</td> <td>11/94</td> <td>3/95</td>	ollection	12/91-	2/92-	5/92-	8/92-	11/92-	2/93-	-£6//	9/93-	12/93	3/94	5/94-	8/94	11/94	3/95
(5)       49       49       48       52       32       39       49       48       52       38       41       45         (5)       2.5       2.7       3.6       3.3       2.9       -       3.0       3.2       2.3       2.9       -       3.0       3.2       2.6       3.0         (7)       2.5       3.5       3.5       3.5       4.8       3.5       2.7       4.0       na       na       na       na       na         (7)       0.6       0.77       0.67       0.55       0.38       0.72       0.5       na       <	briod	2/92	5/92	8/92	11/92	2/93	5/93	9/93	12/93	3/94	5/94	8/94	11/94	3/95	6/95
2.52.73.63.32.9-3.03.22.32.32.63.0435.13.23.54.83.52.74.0nananana293329294027242722211616101201301201301201301201101201090931101201301201301201301201101201201109093130120130120130120130120110120120110909313012017011025010093110110821006434383447362831313130nana4615017015016015017014010064	olids (%)	49	49	48	52	52	39	49	48	52	38	41	45	1	I
ony $43$ $5.1$ $3.2$ $3.5$ $4.8$ $3.5$ $2.7$ $4.0$ $na$ $na$ $na$ $na$ $c$ $29$ $33$ $29$ $29$ $40$ $27$ $24$ $27$ $22$ $21$ $16$ $16$ $um$ $0.6$ $0.77$ $0.66$ $0.77$ $0.67$ $0.55$ $0.38$ $0.72$ $0.5$ $na$ $na$ $na$ $r$ $110$ $120$ $130$ $120$ $130$ $120$ $110$ $120$ $110$ $90$ $93$ $ry$ $0.277$ $0.35$ $0.35$ $0.31$ $0.36$ $0.41$ $0.26$ $0.26$ $na$ $na$ $na$ $ry$ $0.277$ $0.35$ $0.35$ $0.31$ $0.36$ $0.41$ $0.26$ $120$ $110$ $90$ $93$ $ry$ $0.277$ $0.35$ $0.35$ $0.34$ $0.31$ $0.36$ $0.41$ $0.26$ $0.26$ $na$ $na$ $ry$ $0.277$ $0.35$ $0.35$ $0.34$ $0.36$ $0.41$ $0.26$ $0.26$ $na$ $0.23$ $0.24$ $ry$ $130$ $120$ $170$ $110$ $250$ $100$ $93$ $110$ $100$ $93$ $0.24$ $ry$ $38$ $38$ $34$ $47$ $36$ $28$ $31$ $31$ $30$ $na$ $ry$ $140$ $150$ $170$ $150$ $170$ $140$ $100$ $93$ $ry$ $ry$ $ry$ $ry$ $ry$ $ry$ $ry$ $ry$ $ry$	oc (%)	2.5	2.7	3.6	3.3	2.9	I	3.0	3.2	2.3	3.2	2.6	3.0	I	1
5.1       3.2       3.5       4.8       3.5       2.7       4.0       na	letals														
33       29       29       40       27       24       27       22       21       16 <t< td=""><td>Antimony</td><td>43</td><td>5.1</td><td>3.2</td><td>3.5</td><td>4.8</td><td>3.5</td><td>2.7</td><td>4.0</td><td>na</td><td>na</td><td>па</td><td>na</td><td>na</td><td>na</td></t<>	Antimony	43	5.1	3.2	3.5	4.8	3.5	2.7	4.0	na	na	па	na	na	na
0.66       0.77       0.67       0.55       0.38       0.72       0.5       na       na </td <td>Arsenic</td> <td>29</td> <td>33</td> <td>29</td> <td>29</td> <td>40</td> <td>27</td> <td>24</td> <td>27</td> <td>22</td> <td>21</td> <td>16</td> <td>16</td> <td>I</td> <td></td>	Arsenic	29	33	29	29	40	27	24	27	22	21	16	16	I	
120     130     120     130     120     130     120     110     120     110     90     93       0.35     j     0.35     j     0.36     0.41     0.26     0.26     na     na     0.23     0.24       120     170     110     250     100     93     110     110     82     100     64       38     34     47     36     28     31     31     30     na     na       150     170     150     160     140     150     170     140     100     93	Cadmium	0.6 j	0.66	0.77	0.67	0.55	0.38	0.72	0.5	na	na	na	na	па	пa
0.35     j     0.3     j     0.36     0.41     0.26     0.26     na     na     0.23     0.24       120     170     110     250     100     93     110     110     82     100     64       38     34     47     36     28     31     31     30     na     na       150     170     150     160     140     150     170     190     93	Copper	110	120	130 j	120 j	130	120	110	120	120	110	60	93	l	I
120     170     110     250     100     93     110     110     82     100     64       38     34     47     36     28     31     31     30     ла     ла       150     170     150     160     140     150     170     140     100     93       ла=Not analyzed	Mercury	0.27 j	0.35 j	0.35 j	0.3 j	0.36	0.41	0.26	0.26	na	na	0.23	0.24	I	I
38         38         34         47         36         28         31         31         30         na         na           150         170         150         160         140         150         170         93           na=Not analyzed	ead	130	120	170	110	250	100	93	110	110	82	100	2	ł	I
150 170 150 160 140 150 170 140 140 100 na=Not analyzed	Vickel	34	38	38	34	47	36	28	31	31	30	na	114	na	na
	Zinc	140	150	170	150	160	140	150	170	140	140	100	93	-	-
	=Estimated	concentration					ìa=Not an	alyzed							

Table C1(cont): Results of Conventionals and Metals Analysis of Settling Particulate Matter from Hylebos Waterway, December, 1991 to June, 1995 (mg/kg, dry weight).

Location			-			Mouth	ų							
Station No.			·			Н-7	•							
Collection	12/91-	2/92-	5/92-	8/92-	11/92-	2/93-	-2/63-	9/93-	12/93-	3/94	5/94-	8/94-	11/94-	3/95-
Period	2/92	5/92	8/92	11/92	2/93	5/93	9/93	12/93	3/94	5/94	8/94	11/94	3/95	6/95
Solids (%)	55	53	56	60	60	53	54	55	58	41	53	56	53	ł
TOC (%)	2.0	2.3	2.7	2.5	2.2	2.4	2.2	2.4	2.1	2.3	2.2	1.9	2.0	ł
Metals														
Antimony	3.2	3.3	3.5	7	2.8	2.9.	11	2.5	na	na	na	na	na	na
Arsenic	19	20	18	19	23	24	17	14	19	15	9.6	10	11	ł
Cadmium	0.4 j	0.41	0.64	1.8	0.38	0.38	0.41	0.38	na	na	na	na	na	na
Copper	86	82	93 j	95 j	100	95	92	94	82	87	76	-01	62	1
Mercury	0.27	0.24 j	0.31 j	0.24 j	0.21	0.25	0.18	0.2	na	na	0.22	0.19	0.16	ł
Lead	51	48	47	44	52	52	48	52	42	41	36	31	28	1
Nickel	29	28	26	28	35	38	30	27	30	24	na	na	na	na
Zinc	110	110	130	160	120	140	120	120	110	120	88	71	66	
j=Estimated concentration	concentration					na= Not analzyed	alzyed							
-=No sample														

-=No sample

Table C2: Summary of Volatile Organic Compounds Detected in Settling Particulate Matter from Hylebos Waterway, December 1991 to June 1995 (ug/kg, dry weight).

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Location		******	ž	Near Cenex						@ Occi	@ Occidental Chemical	ical		
Cretion No.			i	H-5							H-6			
Collection	12/91-	5/92	11/92-	1.1	12/93-	5/94-	11/94-	12/91-	5/92	11/92-	7/93	12/93-	5/94-	11/94-
Period	5/92	11/92	5/93	12/93	5/94	11/94	6/95	5/92	11/92	5/93	12/93	5/94	11/94	6/95
TOC (%)	3.4	4.5		na	4.2	4.1	na	2.6	3.5	2.9	3.1	2.8	2.8	ł
Co (Jo)		n 8	1	па	4.1 i	6.8 u	na	2 :	8 u	12 j	12	5.2 j	27 uj	1
Trichlorofluoromethane	f 6 0		1	na	1.6 i	0.53	na	5 uj	8 u	2 j	3.8 uj	0.78 j	2.9 u	I
	l ( ). I≜n ni	400 i	I	na	290 i	69 ui	na	110 uj	290 j	ι	200 uj	400	63 uj	I
Accivite V?1 Cit14	fn 17		1	eu Bu	4.5 u	3.4 u	па	5 n	n 8	8 uj	3.8 u	0.42 j	1.34 uj	ł
VIIIVI CIUOLIUE	יי ג ייי נ	3 ⊑ > ≪	I	na	4.5 u	7.4	na	0.8 j	л 8	9 ju	15 uj	4.1 u	9.2 uj	I
Carout Disultac		 	I	na	5	1.6 j	na	6 j	4 j	11 j	6	8.4 j	2.7 j	1
Trans 1 7-Dichloroethene	- 10 - 11	-, ⊐ • 00	<b>ا</b>	na	4.5 u	3.4 u	na	0.2 j	8 8		0.6 j	į 0.1	2 uj	I
Trichloroethene	,	8 U	I	na	1.3 j	1.1 j	na	3 j	3 j.	6 j	3.6 uj	11	1.1 j	ł
Tetrachloconcurate Tetrachloconethene		8 1	ł	nà	2.5 j	1.6 j	na	16	7 j	35 j	5.3	12	3.4	I
Chloroform	E	n 8	I	na	4	5	na	15	7 j	57 j	4	22	3.2 uj	3
Outotothe O_Heronome	 	- n - 80	I	na	4.5 u	17 uj	na	5 u	8 u	8 uj	37 uj	3.5 j	20 uj	I
2-Ileadium	, 25 1 1	100 i	ł	na	170		na	. 66	71 j	120 uj	130 j	230	30 uj	I
4-Dutation A-Mathref?-Dantanone	2 2 2	г ост С ост	I	na	4.2 i	3.4 u	na	5 uj	8 u	8 uj	74 u	4.6	2.7 u	ţ
Tolyana	s	н ж о		na	44 > -1	1.8 i	na	•••••	8 u	3 uj	1.3 j	1.3 j	2.7 uj	<b>i</b>
L'uturit l'eonronstholmene	140	230	1	na	170	300 j	na	14	43	38 j	74 j	170	11	ŧ
Hthrikenzene.	0.85 1	n 8	I	na	4.5 u	0.69 j	na	0.4 j	n 8	2 j	0.79 j	0.45 j	0.64 j	I
1 0 A_Trimethvilhonzene	, <b>7</b>	: = : ~	I	na	7.5 1	4.2	na	6	8 8	16 j	14	4.7	4.9	ŝ
1, 2, 7-1, 1, methylbenzene	3 = 3 VC	1 = 1 0 00	1	na	4.5 u	2.8	na		n 8	10 j	6.6	2.5 j	3.5 j	1
Total Vulenes	s . vr	. = > 00	ı	na	2.9 j	3.6 j	na		8 8	16 j	9.2 uj	3.2 j	3.4 j	1
Teonronvihenzene	n N	8 n	I	na	4.5 u	3.4 u	па	5 u	n 8	8 uj	11	4.1 u	2.9 u	3
1 1 1-Trichloroethane	2.1	8 С	I	na	4.5 u	3.4 u	na	5 u	8 u	8 uj	3.8 u	0.8 j	2.9 u	I
Styrene	0,4 j	n 8	I	па	4.5 u	0.59 j	na	5 u	n 8	8 uj	1.4 j	0.4 uj	0.32 j	ŧ
Carbon Tetrachloride	S u	8 n	ł	na	4.5 u	3.4 u	na	5 u	n 8	S. j	4 u	0.25 j		I
Methylene Chloride	5 u	8 u	I	na	4.5 u	3.2 j	រាង	5 u	8 8	8 uj	4 u	[]	2.9 uj	1
1 4 Dichlorohenzene	5 u	8 1	1	na	4.5 u	0.53 j	na	5 u	. 8 u	8 uj	4 u	4.1 u	3.7 u	I
1 1 2 2-Tetrachloroethane	5 U	8 8		na	4.5 · u	3.4 u	na	5 u	n 8	8 uj	4 u	4.1 u	2.9 u	I
1 1 Dichloroethane	5 tt	8 U	I	na	4.5 u	3.4 u	na	5 u	8 u	8 uj	4 u	4.1 u	2.9 u	I
4-Chlorotolnene	5 u	п 8	ł	na	4.5 u	1.4 j	na	5 u	8 u	8 uj	4 u	4.1 u	2.9 u	ł
Renzene	5 u	8 8	I	Шâ	4.5 u	0 <i>.77</i> j	na	5 u	8 u	8 uĵ	4 u	4.1 u	1.2 . j	I
Chlorohenzene	- 2 2	n <sub>.</sub> 8	1	na	4.5 u	3.4 u	na	ς α	8 u	4 j	4 u	4,1 u	2:9 u	-
-=No sample						j=Estimate	j=Estimated concentration	ion						
no-Nict analysis				~		uj=Estimat	uj=Estimated detection limit	limit						
lig-14.00 anter 22.00						,								

u=Not detected at detection limit shown

Table C2(cont): Summary of Volatile Organic Compounds Detected in Settling Particulate Matter from Hylebos Waterway, December 1991 to June 1995 (ug/kg, dry weight).

H-7           Station No.           JOJ         JOJ         JOJ         JOJ           Collection         JOJ         JOJ         JOJ         Collection           Period         S/92         JIOJ         JOJ         JOJ         JOJ         JOJ         JOJ         Collection         JOJ         JOJ <th< th=""><th>Location</th><th></th><th></th><th>Mout</th><th>Mouth of Waterway</th><th>ıy</th><th></th><th></th><th>1</th></th<>	Location			Mout	Mouth of Waterway	ıy			1
ion $12/91$ $5/92$ $11/92$ $5/92$ $11/92$ $5/93$ $12/93$ $5/94$ $87$ $2.2$ $2.6$ $2.3$ $2.3$ $2.2$ $2.2$ $2.2$ $2.2$ $2.2$ $2.2$ $2.2$ $2.2$ $2.2$ $2.2$ $2.2$ $2.2$ $2.3$ $2.3$ $2.3$ $2.3$ $2.3$ $2.3$ $2.3$ $2.3$ $2.3$ $2.3$ $2.3$ $2.3$ $2.3$ $2.3$ $2.3$ $2.2$ $2.2$ $2.4$ $3$	Station No.			•	H-7				1
5/92 $1/92$ $5/93$ $12/93$ $5/94$ $87$ $2.2$ $2.6$ $2.3$ $2.3$ $2.2$ methane1j7u $12$ j $3.3$ ufor fluoromethane1j7u $12$ j $3.3$ u $0.66$ je83uj $250$ j $550$ uj $220$ uj $3.3$ u $0.66$ je83uj $250$ j $550$ uj $220$ uj $3.3$ u $0.66$ jDisulfide1j1jjjj $1.1$ j $2.4$ jlor ortheme1j1jjj $2.4$ jjlor ortheme1jjj $2.1$ $3.3$ u $1.4$ jlor ortheme1jjj $2.1$ $3.3$ u $1.4$ jlor ortheme1jjj $2.1$ $2.4$ j $1.1$ $2.4$ jlor ortheme0.8j7u23j $3.3$ uj $1.4$ jlor ortheme1jjjjj $2.4$ j $1.7$ $1.7$ lor ortheme1jjjjjj $1.4$ j $1.7$ lor ortheme2jujjjjjjjlor ortheme3uj<	Collection	12/91-	5/92	11/92-	7/93-	12/93-	5/94-	11/94-	I
2.2         2.6         2.3         2.3         2.2           fluoromethane         1         7         u         12         3.3         u         2.66         j           fluoromethane         1         7         u         12         3.3         u         2.66         j           bisulfide         1         j         1         j         3.3         u         3.66         j           bisulfide         1         j         1         j         3.3         uj         3.7         u         3.3         u         3.3         u         3.3         u         3.3         u         1.4         j         u         2.0         j         3.3         u         4.7         j         j         u         4         j         j         1.1         j	Period	5/92	11/92	5/93	12/93	5/94	11/94	6/95	1
1         7         12         3.3         u         2.8         j           methane         4         1         7         1         15         3.3         u         2.8         j           ib         1         7         u         20         uj         3.3         u         14         j         u         1         j         u         1         j         2.4         j         1.1         j         2.4         j         1.1         j         2.4         j         1.4         j         2.4         j         1.4         j         2.4         j         1.4         j         2.4         j         1.4         j         1.4         j         2.4         j         j         1.4         j         j         1.4         j         j         j         j         j         j         j         j         j         j         j         j         j         j         j         j         j	TOC (%)	2.2	2.6	2.3	2.3	2.2	2.1	2.0	I
4 u       7 u       16 uj       3.3 u       0.66 j         83 uj       250 j       550 uj       200 uj       3.3 u         1 j       7 u       20 uj       3.3 u       3.3 u         1 j       1 j       1 j       1 j       1 j       3.3 u         1 j       1 j       1 j       1 j       200 uj       3.2 u         1 j       0 j       3 j       1.1 j       2.4 j         1 j       0 j       3 j       0.3 j       1.4 j         1 j       7 u       23 j       3.3 uj       1.4 j         2 j       7 u       2 j       2.4 j       1.4 j         2 j       7 u       2 j       3.3 uj       1.4 j         2 j       7 u       2 j       3.3 uj       4.7 j         3 su       5 j       2 uj       3.3 uj       1.4 j         2 4 u       7 u       10 uj       3.3 uj       1.3 j         4 4 u       7 u       10 uj       3.3 uj       3.1 j         6       1 j       1 u       1 u       3.3 u       3.3 u         1 1 2 j       7 u       1 j       1 u       3.3 u       3.3 u         1 1 2 j	Chloromethane	44 •9	7 u	12 j	3.3 u	2.8 j		1.8 u	
83 uj       250 j       550 uj       200 uj       320 j         4 u       7 u       30 uj       17 uj       3.3 u       3.3 u         1 j       1 j       1 j       1 j       1 j       2.4 j         1 j       7 u       30 uj       17 uj       3.3 u       3.3 u         1 j       7 u       30 uj       17 uj       3.3 u       3.3 u         1 j       0.9 j       33 j       0.31 j       1.4 j         2 j       7 u       20 uj       33 uj       1.4 j         2 j       7 u       20 uj       33 uj       4.7 j         2 j       7 u       20 uj       57 uj       57         4 u       7 u       20 uj       57 uj       57         4 u       7 u       20 uj       57 uj       57         4 u       7 u       10 uj       33 uj       4.7 j         5 4 j       7 u       10 uj       33 uj       4.7 j         3 8 u       51 j       29 uj       33 uj       4.7 j         4 u       7 u       10 uj       33 u       3.3 u       3.3 u         6       1 j       7 u       10 uj       3.3 u       3.3 u	Trichlorofluoromethane	4 u	7 u		3.3 u	0.66 j	3.9 u	1.8 u	
4 u       7 u       20 uj       3.3 u       3.3 u         1 j       7 u       30 uj       17 uj       3.3 u         1 j       1 j       1 j       1 j       24 j         1 j       1 j       1 j       1 j       24 j         1 j       1 j       1 j       1 j       2.4 j         1 j       0.9 j       33 j       0.31 j       1 j         2 j       7 u       23 j       3.3 uj       1.4 j         2 j       7 u       20 uj       33 uj       4.7 j         2 j       7 u       20 uj       67 u       3.6         4 u       7 u       20 uj       33 uj       4.7 j         38 u       51 j       290 uj       67 u       3.6         4 u       7 u       10 uj       3.3 u       1.3 j         4 u       7 u       15 uj       3.3 u       3.3 j         4 u       7 u       16 uj       3.3 u       3.3 j         1 1 2 j       7 u       16 uj       3.3 u       3.3 j         4 u       7 u       19 uj       3.3 u       3.3 u         1 1 2 j       7 u       19 uj       3.3 u       3.3 u	Acetone	83 uj	250 j			320 j	150 j	30 n	
I       J       T       30       U $17$ $3.3$ U $1.1$ $3.3$ U $1.1$ $3.3$ U $1.1$ $3.3$ U $1.4$ J         0.8 $7$ $0.9$ $1.1$ $1.1$ $2.4$ $1.10$ $2.4$ $1.10$ $2.4$ $1.10$ $2.4$ $1.10$ $2.4$ $1.10$ $2.4$ $1.10$ $2.4$ $1.10$ $2.4$ $1.10$ $2.4$ $1.10$ $2.4$ $1.10$ $2.4$ $1.10$ $2.4$ $1.11$ $2.4$ $1.10$ $2.4$ $1.10$ $2.4$ $1.10$ $2.4$ <td>Vinyl Chloride</td> <td>4 u</td> <td>n 7 u</td> <td></td> <td></td> <td>3.3 u</td> <td>3.9 u</td> <td>1.8 u</td> <td></td>	Vinyl Chloride	4 u	n 7 u			3.3 u	3.9 u	1.8 u	
I       I	Carbon Disulfide	i i	7 u			3.3 u	18	3.8 u	
4 u       7 u       96 j       0.31 j       1 j         0.8 j       7 u       23 j       3.3 uj       1.4 j         2 j       7 u       23 j       3.3 uj       1.4 j         2 j       7 u       10 j       0.86 j       2.4 j         2 j       7 u       10 j       0.86 j       2.4 j         2 j       7 u       10 j       0.86 j       2.4 j         38 u       51 j       290 uj       67 u       3.6         4 u       7 u       10 uj       3.3 uj       4.7 j         45       7 u       15 uj       3.3 uj       3.1 j         1 j       7 u       10 uj       3.3 uj       3.1 j         1 j       7 u       10 uj       3.3 uj       3.1 j         1 j       7 u       10 uj       3.3 uj       3.1 j         1 j       7 u       10 uj       3.3 uj       3.3 uj         1 1 j       7 u       19 uj       3.3 uj       3.3 uj         1 1.2 j       7 u       19 uj       3.3 u       3.3 u         1 1.2 j       7 u       19 uj       3.3 u       3.3 u         1 4 u       7 u       19 uj       3.3 u	Cis 1,2-Dichloroethene	مسز ، دستا	i i	15 j	1.1 j	2.4. j	1.4 j	0.41 j	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	Trans 1,2-Dichloroethene	4 u	7 u	· [ 96	0.31 j		3.9 u	1.8 j	
110.9j33j0.93j1.8j2j7uj0.86j2.4j4u7u20uj57u17038u51j200uj67u3.6457u20uj67u3.6457u15uj3.3u1.3457u7u15uj3.3u467u7u10uj3.3u417u10uj3.3u0.5j447u7u19uj3.3u3.3u447u7u19uj3.3u3.3u447u7u19uj3.3u3.3u447u7u19uj3.3u3.3uane4u7u19uj3.3u3.3u4u7u19uj3.3u3.3u1.44u7u19uj3.3u3.3u1.44u7u19uj3.3u3.3u1.44u7u19uj3.3u	Trichloroethene	0.8 j	7 u	23 j		1.4 j	0.89 j		
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Tetrachloroethene		0.9 j	33 j	0.93 j	1.8 j	0.88 j	0.53 j	
4 u       7 u       20 uj       33 uj $4.7$ j         38 u       51 j       290 uj $67$ u $3.6$ 4 u       7 u $15$ uj $67$ u $3.6$ 45       7 u $15$ uj $67$ u $3.6$ $45$ 7 u $15$ uj $3.3$ u $1.3$ j $45$ 7 u $15$ uj $3.3$ u $1.3$ j $41$ 7 u $15$ j $3.3$ uj $3.1$ j $44$ 7 u $16$ j $10$ uj $3.3$ uj $3.1$ j $e$ $11j$ 7 u $19$ uj $3.3$ u $0.5$ j $u$ $4$ u       7 u $19$ uj $3.3$ u $3.3$ u $3.3$ u $3.3$ u $4$ u       7 u $19$ uj $3.3$ u $3.3$ u $3.3$ u $4$ u       7 u $19$ uj $3.3$ u $3.3$ u $3.3$ u $4$ u       7 u $19$ uj $3.3$ u $3.3$ u $3.3$ u $1.2$ j       7 u $19$ uj $3.3$ u $3.3$ u $3.3$ u $4$ u       7 u $19$ u	Chloroform	5	7 u	10 j	0.86 j	2.4 j	3.9 u		
38 u51 j290 uj67 u1704 u7 u12 uj67 u3.6457 u15 uj3.3 u1.3 j457 u10 uj3.3 uj3.1 j467 u15 j3.3 uj3.1 j47 u7 u15 j3.3 uj3.1 j4 u7 u15 j3.3 uj3.3 u12 j7 u16 j10 u3.3 u4 u7 u19 uj3.3 u3.3 uane4 u7 u19 uj3.3 u3.3 u $4 u$ 7 u19 uj3.3 u<	2-Hexanone	4 u	7 u			4.7 j			
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2-Butanone	38 u	Ś1 j		67	170	47 j		
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	4-methyl-2-Pentanone	4 u	7 n			3.6	3.9 u		
45       7 u       44 j       4.9       57         4 u       7 u       10 uj       3.3 uj       3.1 j         ane       1 j       7 u       25 j       3.3 uj       3.1 j         ane       1 j       7 u       25 j       3.3 uj       3.1 j         ane       1 j       7 u       15 j       3.3 uj       3.1 j         ane       1.2 j       7 u       16 j       10 u       3.9 j         4 u       7 u       19 uj       3.3 u       3.3 u       3.3 u         4 u       7 u       19 uj       3.3 u       3.3 u       3.3 u         4 u       7 u       19 uj       3.3 u       3.3 u       3.3 u         hame       4 u       7 u       19 uj       3.3 u       3.3 u       3.3 u         hame       4 u       7 u       19 uj       3.3 u       3.3 u       3.3 u         hame       4 u       7 u       19 uj       3.3 u       3.3 u       4.0 $t_1$ 7 u       19 uj       3.3 u       3.3 u       3.3 u       4.0 $t_2$ 19 uj       3.3 u       3.3 u       3.3 u       3.3 u       4.0	Toluene	0.6 j	7 n			1.3 j	1.8 j		
4 u $7$ u $10$ uj $3.3$ uj $3.1$ j         ane $1$ j $7$ u $15$ j $3.3$ uj $3.1$ j         ane $1$ j $7$ u $15$ j $3.3$ uj $3.1$ j $1.2$ j $7$ u $15$ j $3.3$ uj $3.3$ u $3.3$ u $4$ u $7$ u $19$ uj $3.3$ u $3.3$ u $3.3$ u $4$ u $7$ u $19$ uj $3.3$ u $3.3$ u $3.3$ u $4$ u $7$ u $19$ uj $3.3$ u $3.3$ u $3.3$ u $4$ u $7$ u $19$ uj $3.3$ u $3.3$ u $3.3$ u $4$ u $7$ u $19$ uj $3.3$ u $3.3$ u $3.3$ u $4$ u $7$ u $19$ uj $3.3$ u $3.3$ u $3.3$ u $4$ u $7$ u $19$ uj $3.3$ u $3.3$ u $3.3$ u $4$ u $7$ u $19$ uj $3.3$ u $3.3$ u $3.3$ u $4$ u $7$ u $19$ uj $3.3$ u $3.3$ u $3.3$ u $4$ u $7$ u $19$ uj <td>Isopropyltoluene</td> <td>45</td> <td>7 u</td> <td>44 j</td> <td>4.9</td> <td>57</td> <td>53</td> <td>110</td> <td></td>	Isopropyltoluene	45	7 u	44 j	4.9	57	53	110	
ane       4 u       7 u       25 j       3.3 uj       3.1 j         ane       1 j       7 u       15 j       3.3 uj       3.3 u $1.2$ j       7 u       16 j       10 u       3.9 j $4$ u       7 u       19 uj       3.3 u       3.3 u $4$ u       7 u       19 uj       3.3 u       3.3 u $4$ u       7 u       19 uj       3.3 u       3.3 u $4$ u       7 u       19 uj       3.3 u       3.3 u $4$ u       7 u       19 uj       3.3 u       3.3 u $4$ u       7 u       19 uj       3.3 u       3.3 u $4$ u       7 u       19 uj       3.3 u       3.3 u $4$ u       7 u       19 uj       3.3 u       3.3 u $4$ u       7 u       19 uj       3.3 u       3.3 u $4$ u       7 u       19 uj       3.3 u       3.3 u $4$ u       7 u       19 uj       3.3 u       3.3 u $4$ u       7 u       19 uj       3.3 u       3.3 u $4$ u       7 u       19 uj       3.3 u       3.3 u $4$ u       7 u <td>Ethylbenzene</td> <td>4 u</td> <td>7 u</td> <td></td> <td>3.3 u</td> <td>0.5 j</td> <td>0.47 j</td> <td></td> <td></td>	Ethylbenzene	4 u	7 u		3.3 u	0.5 j	0.47 j		
ane       1 $7$ $15$ $3.3$ $yi$ $3.3$ $yi$ $3.3$ $yi$ $1.2$ $1$ $1.2$ $7$ $16$ $10$ $0$ $3.9$ $j$ $4$ $1$ $7$ $19$ $19$ $yi$ $3.3$ $u$ $3.3$ $u$ $4$ $1$ $7$ $19$ $yi$ $3.3$ $u$ $4$ $u$ $7$ $u$ $3.3$ $u$ $7$ $u$ $10$ $u$ $3.3$ $u$ $10$ $u$ $10$ $u$ $10$	1,2,4-Trimethylbenzene	4 u	7 u	25 j	3.3 uj	3.1 j	2.4 j	1.9	
1.2 j       7 u       16 j       10 u       3.9 j         4 u       7 u       19 uj       3.3 u       3.3 u         4 u       7 u       19 uj       3.3 u       3.3 u         4 u       7 u       19 uj       3.3 u       3.3 u         4 u       7 u       19 uj       3.3 u       3.3 u         4 u       7 u       19 uj       3.3 u       3.3 u         4 u       7 u       19 uj       3.3 u       3.3 u         10 uj       3.3 u       3.3 u       3.3 u       3.3 u         10 uj       3.3 u       3.3 u       3.3 u       3.3 u         10 uj       3.1 u       19 uj       3.3 u       7.9         10 uj       3.3 u       7.0       3.3 u       7.9         10 uj       3.3 u       7.0       3.3 u       7.9         10 uj       3.3 u       7.0       3.3 u       3.3 u         11 u       7 u       19 uj       3.3 u       3.3 u         4 u       7 u       19 uj       3.3 u       3.3 u         4 u       7 u       19 uj       3.3 u       3.3 u         20 uj       3.3 u       3.3 u       3.3 u       <	1,3,5-Trimethylbenzene	1	7 u	15 j		3.3 u	1.1 j	1.8 j	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	Total Xylenes	1.2 j	7 u	16 j	10 u	3.9 j	2.4 j		
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	Isopropylbenzene	4 u	7 u		3.3 u	3.3 u	3,9 . u		
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	1,1,1-Trichloroethane	4 u	7 u		3.3 u	3.3 u	3.9 u		
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	Styrene	4 u	7 u		3.3 u	0.08 j	0.33 j	1.8 u	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	Carbon Tetrachloride	4 u	7 n		3.3 u	3.3 u	3.9 u	1.8 u	
zene $0.5 j$ $7 u$ $19 uj$ $3.3 u$ $3.3 u$ oroethane $4 u$ $3 j$ $21 uj$ $3.3 u$ $3.3 u$ ane $4 u$ $7 u$ $95 uj$ $3.3 u$ $7.9$ 4 u $7 u$ $19 uj$ $3.3 u$ $3.3 u4 u$ $7 u$ $19 uj$ $3.3 u$ $3.3 u4 u$ $7 u$ $19 uj$ $3.3 u$ $3.3 u4 u$ $7 u$ $19 uj$ $3.3 u$ $3.3 u$	Methylene Chloride	4 u	7 u		3.3 u	3.3 u	3.9 uj	1.8 u	•—
orocethane4 u3 j21 uj3.3 u3.3 uane4 u7 u95 uj3.3 u7.9 $4$ u7 u19 uj3.3 u3.3 u3.3 u $4$ u7 u19 uj3.3 u3.3 u14 u7 u19 uj3.3 u14 detection limit shown	1,4-Dichlorobenzene	0.5 j	7 u		3.3 u	3.3 u	0.57	1.8	
and $4 u 7 u 95 uj 3.3 u 7.9$ 4 u 7 u 19 uj 3.3 u 3.3 u 3.3 u 4 u 7 u 19 uj 3.3 u 3.3 u 4 u 7 u 19 uj 3.3 u 3.3 u 10 uj 3.3 u 3.3 u 10 uj 4 u 7 u 19 uj 3.3 u 3.3 u 10 uj 4 u 4 u 7 u 19 uj 3.3 u 3.3 u 10 uj 4 u 4 u 7 u 19 uj 3.3 u 3.3 u 10 uj 4 u 4 u 7 u 19 uj 3.3 u 3.3 u 10 uj 4 u 4 u 7 u 19 uj 3.3 u 3.3 u 10 uj 4 u 4 u 7 u 19 uj 3.3 u 3.3 u 10 uj 4 u 4 u 7 u 19 uj 3.3 u 3.3 u 10 uj 4 u 4 u 7 u 19 uj 3.3 u 3.3 u 10 uj 4 uj 4 u 10 uj 4 uj 4 u 10 uj 4	1,1,2,2-Tetrachloroethane	4 u	3. j.		3.3 u	3.3 u	3.9 u	1.8 u	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	1,1-Dichloroethane	4 u	7 u		3.3 u	7.9	3,9 u	1.8 u	
4 u         7 u         19 uj         3.3 u         3.3 u           e         4 u         7 u         19 uj         3.3 u         3.3 u           zed         at detection limit shown	4-Chlorotoluene	4 u	7 u		3.3 п	3.3 u	3.9 u	1.8 u	
e 4 u 7 u 19 uj 3.3 u 3.3 u zed zed at detection limit shown	Benzene	4 u	1 n		3,3 u	3.3 u	2.2 j	1.8 u	
zed ad at detection limit shown	Chlorobenzene	4 u	n /		3.3 u	3.3 u	3.9 u	1.8 u	. 1
	-=No sample						j=Estimated	i concentra	ion
n=Not detected at detection limit shown	na=Not analyzed						uj=Estimato	ed detection	limit
	u=Not detected at detection 1	imit shown							

Table C3: Summary of Semivolatile Organics and Polychlorinated Biphenyls Detected in Settling Particulate Matter from Hylebos Waterway. December 1991 to June 1995 (ug/kg. drv weight).

from Hylebos Waterway, December 1991 to June 1995 (ug/kg, dry weight)	/aterw	ay, D	ecemt	ber 19.	91 to J	une 19.	130) CV	kg, ar	y weigi	at).								4149-C			****
Location Station No.		H	lead @ Uf	Head @ Upper Turning Basin H-1	ng Basin					@ Gen	@ General Metals H-2	10					@ 11	@ Elf Atochem H-3		1	
Period	12/91- 5/07	5/92-	1/92- 5/93		12/93- 5/94	5/94- 1 11/94	11/94-1	12/91- 5/92	<u>5/92- 11</u> 11/92	11/92-7 5/93 1	7/93- 12 12/93	12/93- 5 5/94 1	5/94- 1) 11/94	11/94- 1 6/95	12/91- 5/92	5/92- II 11/92	11/92-7 5/93 1	7/93- 12 12/93	12/93- 5 5/94 1	5/94- 11 11/94 11	11/94- 6/95
TOC (%)			8.0	7.1	6.1	5.8		.			4.8		4.9	4.8	3.9	5.1	3.8	4.2	4.8	4.6	5.0
Acenaphthene	190	\$	0/1	140 j	110	220 j	88 j	48 j	100 j	110	120 j	160	82 j	110 j	320 j	340	80 j	40 j	230	360	380
Acenaphthylene	230 u	ı	<i>51</i> j	73 j	56 j	96 j	59 j	410 u	62 j	48 uj	68 j	76 j	54 j	66 j	110 j	94 j	63 j	20 j	99 190	140 j 150 :	120 j 730 r
Naphthalene	53 j	ı	49 j	170 u	68	160 j	190 u	85 j	130 u	2	150 j	8	100	230 u	130 j	<u>8</u>	n n	9 E	120	l vci	n 052
Fluorene	240	ı	290	160 j	170	220 j	110 j	410 u	120 j	120	150	220	110 j	120 120	420	350	120	[ ]],	2/0	065	01/ 2500
Anthracene	280	f	540	360	310	420	240 u	380 j	380	270		490	530	430 u	000	07/	0/6	040		7200	2000
Phenanthrene	1200	1	1300	840	770	1200	670 u	750	720	670		1800	640	n 066	2300	1800	10001		.,	1 0017	
Sum LPAH	2000 j		2400 j	1600 j	1500 j	2300 j	260 j	1300 j	1400 j	1300		2800 j	1300 j	350 J	3900 J	1 0055	1,000	-		4100 ]	f 00/0
Fluoranthene	4700	1	2000	2200	1400	3200		2200	1970	2000	2000	3500		1 010	2800 1650	00/0	00077		1300	1600	1600 u
Benzo(a)anthracene	4000 u	ł	89	088	Qç Ş	0011	n 000,	3/00 II	0026	0031		2300	000	3600	1600	3400	2100			3900	3500
Chrysene	4600	ŧ	1400	0007	110	00/7	1900	1800	1040	1800		3400	1600	1900	4100	3600	2100				3800
Pyrene	2400 j	I	0011	1600	016	0007		7000	04-5T	1800			2600	2700	940	3900	2800				4100
Benzofluoranthenes	fn 0035	1	1400	onte		00001	n 02.5	00.77	800	400			740	066	06/	1200	640			1400	240
Benzo(a)pyrene	f oner	•	110 n		n 1020	5000 i		1000 u	190	100	250	260	160	240 u	300 uj	390	180	100 j	j 99	280	240
DIOCHZO(A, II) MILITACELIE V-4/1 2 2 40	n 0/c	. 1	10 tr		050	002		410 u	630	300	850	870	530	690 u	460	800	440	370	690	870	830 u
mucno(1,2,3-cu)pyrate Benrafa h ihnerviene		I	130	580	120	680	1	410 u	410	<i>TT</i> u	950	600	570	680 u	p	510	370		620	800	820
Citra HPAH		*	7100	12000	5100	16000 i	3400	12000	12000	8700 1	17000 1	19000 1					12000 1				17000
Total PAH		1	9500	14000 j	6600 i	18000	.,,	13000 j	••••	10000 j 1	19000 j 2	22000 j 1	11000 j	10000 j	•	23000 j	13000 j 1	11000 j 2	21000 j 2	26000 j 2	24000 j
1-Methylnapthalene	50	1	93 i		na	-	1	410 u	<u>57</u> j	40 j	na	na	I	1	96 j	150	40 j	na	na	ı	1
2-Methylnaphthalene	51 j	ı	8	170 u	120	130 j	190 u	44 j	58 j	36 j	180	80 j	19 j	280 u	92 j	160	41 j	140 u	120	170	290 u
Dihenzofuran	140	I	160	110 j	110	160 j	190 u	71 J	110 j	170 j	110 j	150	85 j	230 u	240	270	82 j	48 j	190	280	330 u
Carbazole	1200 uì	I	170	170 u	100	240 u	190 u	2100 uj	110 j	110 j	160 u	200	150 u	230 u	1100 uj	180	120	50 j	120	150 j	750
Isonhrone		I	110 u	170 u	64 u	86 j	190 u	410 u	130 u	76 u	160 u	120 u	25 j	230 u	200 u	110 u	100 u	140 u	66 u	52 j	230 u
Retene	230 u	ı		па	240	1	i 061	410 u	290	130	na	330	ı	240	180 u	310	210	na	250	ı	330
Conrostanol	na	I	Da	na	ца	na	7600 u	Пâ	na	ПÂ	na	na	na	9300 u	na	na	na	na	11.8	0.8	n 0006
Phenol	370 u	I	110 u	170 u	64 u	150 j	190 u	410 u	130 u	17 u	330	52 j	210	230 u	270	390	100 u	140 u	.n 99	240 u	230 u
Pentachlorophenol	ŧ	١	55 uj	1700 u	640 u	2400 u	950 u	ŧ	130 u	38 uj	1600 u	1200 u	1500 u	1200 u	I	110 u	500 uj	1400 u	660 u	2400 u	1100 u
4-Methylphenol	230 u	1	110 u	170 u	64 u	1500	190 u	410 u	420	<i>77</i> u	570	49 j	830	230 u	180 u	0//	100 u	140 u	390	200	230 u
Benzoic Acid	3000 uj	I	ju 000	5400 ]	1700 j	3500 uj	380 uj	5300 uj	1500	1100	2500 uj	1400 j	2500 uj	470 uj	2600 uj	770 u	320 uj	(in 006	1400 j	2600 uj	450 uj
Hexachloroethane	230 u	1	110 uj	170 u	t	1	1	410 u	130 u	76 u	160 u	120 u	\$	1	200 u	110 n	100 n)	140 u	гş	י. ני <u>י</u>	: : : :
Hexachlorobenzene	230 u	I	110 u	170 u	64 и	ı	· 190 u	410 u	130 u	10 n	160 u	120 u	I	230 u	n 007	n orr	100 u	140 u	9 (	f (†	n
1,2,4-Trichlorobenzene	230 u	ı	110 u	170 u	64 u	1	190 u	410 u	130 u	76 u	160 u	120 u	II J	230 u	n 007	n ort		140 ti	n : 00	( )	1 077 n
Hexachlorobutadiene	570 u	1	110 u	. 170 u	64 u	ı	190 u	1000 u	26	10 n	160 u	120 u	1	fn 057	[ 66,	41 ]	n 81	140 H	n :: 00	- 090	n 077
Dimethylphthalate	230 u	I	110 u	120 j	(2 1 1	210 j	[ j 011	410 u	130 u	11 n	160 u	120 u	n OCI	n 067	n 027	n off		n 041	220 H	n	330 n
Butylbenzylphthalate	<i>57</i> 0 uj	۱. 	110 u	860 u			190 u	1000 и	130 u		200 uj	580 u	150 j	400	45U u	017		140 ml	ມ 000 ຳ າຕ	1 021	340 1
Di-n-Butyiphthalate	230 u	;	110 u	170 u			330 u	410 u	130 u	60 uj	120 ]	2400		34U U	n 00/	n nit	n oor	n 041	(m T)		
Bis(2EH)phthalate	4000 uj	1	4000 uj	i 2500 uj	j 3200 uj	j 2300	1800 u	2400 u	2500	12000 uj	2200 uj	3300 uj	2500 uj	2700 u	ZZ00 u	n 0007	în mot	În ANCO	Town m	fn nner	n McT
PCBs														 5	+ 000	: 011	1001	140	83 n	120	81 11
1242	46 u	I	110 u				95 u	73000	130 u	a 28	1001	140 u	n 001	orc	n 0000	1100	1 000 C	* 041	3 64,6	1100	5 50
1254	160 u	ı	230	390	240	330	230	4200 u	1100	510	850	016	610	078	2000	noct	1000	800	00 <del>1</del>	0017	20
1260	250	ı	110 u	190	120	190	130 j	1300 u	130 u	190	460	360	410	200	8/0	n orr	/30	AN	490	0007	0021
Total PCB'S	250	ı	230	580	360	520	460 j	73000	1100	700	1300	1300	1000	1600	2900	1300	2500	1000	1900	TANG	MCT
u=Not detected at detection limit shown	on limit sh	UMD						1	=No sample												
j=Estimated concentration	0								na=Not analyzed	/zed											
uj=Estimated detection limit	mit	•																			

Table C3(cont.): Summary of Semivolatile Organics and Polychlorinated Biphenyls Detected in Settling Particulate Matter from Hylebos Waterway December 1991 to June 1995 (ug/kg. drv weight).

$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	Location Near Lincoln Drain		Near	Near Lincoln Drain	Drain	*				Nea	Near Cenex						@ Occid	@ Occidental Chemical	mical		1
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	Station No.			H-4							1						00.1	1	00/04		101
	ion		11/92- 5/93	7/93- 12/93	2/93- 5/94						~	· ++			2/91- 5/92		5/93		5/94		6/95
			3.8	na	3.7		na	3.4	4.5		na	4.2	4.1	па	2.6	3.5	2.9	3.1	2.8	2.8	¥
$ \begin{bmatrix} 7 & 7 & 1 \\ 7 & 1$	Accounthene	- 75	i 68 i	na	86	1.02	Па	55 ]	120 j	ł	Пâ	230	210	na	98 j	320	141	580	940	310 j	1
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Acenaphthylene	- 69	107	шa	51	57 j	ца	28 j	44 j	I	113	76	95 j	na	37 ]	22	66 j	93 j	130	6 I 6	I
$ \left[ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Nanhthalene	- 130	, 10 11	<b>D</b> .a	120	130 j	na	86 j	146 u	ı	na	310	380	ра	110 j	250	130	370	200	240 ]	ł
$ \begin{bmatrix} 50 & n & 50 & 100 &$	Fluorenc	- 97	i 85 j	ПА	130	88 j	na	200 j	146 j	1	na	320	290	па	180 j	400	181	640	770	370 j	ŧ
450         a         100	Anthracene	- 270	250	<u>n</u> a	250 -	190 j	na	200 j	240	I	na	370	400	11.2	300	470	410	<b>06</b>	730	520 j	i
$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$	Phenanthrene	- 570	450	na	610	490	Da	1200	680	ı	na	1200	1200	na	920	1700	086	2800	2700	1700 j	1
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Sum I PAH	- 1200	i 1000 i	na	1300	1000	na	1800	1200 j		na	2500	2600 j	DA	1600 j	3200 j	1900 j	5300 j	5800	3200	1
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Threathene	- 1200	1100	Ц	1100	1000	na	3200	1000	۱,	na	1500	1800	na	1400	2200	1600	3600	2900	2100 j	ı
$ \begin{bmatrix} 1200 & m & 1200 & 1000 & m \\ 880 & m & 1200 & 1000 & m \\ 880 & 1200 & 1000 & 100 & 1800 & 1400 & 1800 & 1400 & 1800 & 1400 & 110 \\ 880 & 1800 & 1400 & 1800 & 1400 & 1200 & 1800 & 1400 & 110 & 180 & 1400 & 110 \\ 1 & 171 & m & 200 & 230 & 10 & 201 & 110 & 230 & 140 & 110 & 180 & 1400 & 100 & 130 & 1400 & 110 \\ 1 & 171 & m & 300 & 1300 & 1 & 200 & 201 & 1 & 10 & 1300 & 1400 & 100 & 130 & 100 & 130 & 100 & 130 & 100 & 130 & 100$	Turolaunto Dama Carlination	- 550	6	ELL	480	420	na	2000	390	ı	ពង	550	620	IIA	1300	800	620	1200	906	770 j	ı
		- 105		i u	1200	1000	03	1600 i	870	I	na	1100	1400	na	1300	. 098	1200	1800	1600	1200 j	ŧ
$ \begin{bmatrix} 1000 & 111 & 100 & 400 & 113 & 500 & 1120 & - 114 & 500 & 1100 & 200 & 1400 & 200 & 140 & 200 & 110 & 200 & 120 & 200 & 20 & 2$	Curysene	1300	•	1	1200	1000	u a	2100 i	1000	ł	Па	1100	1600	Da	1100	1800	970	2600	2100	1600 j	ı
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	ryrene r		-	1 2	1700	4800	μ	5400 mi	1200	\$	Ta	1400	1900	na	2100	1800	1400	2500	1400	1500 j	ι.
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Benzol(uofantucues	0027 -	-		490	480	1	670 i	120 i	I	Цâ	380	620	Da	520	009	340	950	580	610 j	t
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	benzo(a)pyrene	110				230 ni	, e	590 n	1 22	ı	113	96		113	590 u	110 j	89 j	160	110	<i>97</i> j	ŧ
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $		110	-,	ă ș	002	6 092	Ê	- 730 n	230 i	I	na	280		ца	320	390	230	570	370	380 j	ŧ
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	Indeno(1,2,5-cd)pyrene	1			2 6	330	12	230 n	150	ı	Цâ	180	400	na	290	210	160	520	140	360 j	ł
$ \begin{bmatrix} 3 & 201 & 1 & 1 & 700 & 1 & 1 & 1000 & 1 & 2000 & 1 & 1000 & 1 & 2000 & 1 & 1000 & 1 & 1000 & 1 & 1000 & 1 & 1$	Benzo(g,h,1)perylene	- 1/0			040	0020	H I	15000 1				6600	0100	84	8900	8800 i	6600 ì	14000	10000	8600 j	1
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Sum HPAH	- 6800	· .	a i	/000/	5000 11000				L	193 193	9100	12000 i	E E	11000 i	12000	8500 j	19000 j	16000	12000 j	ŧ
$ \begin{bmatrix} 1 & 71 \\ 71 & 11 \\ 71$	I otal PAH	- www	_	Пă	Donto	f onort		1 27	202					Ē	60	160	. 0L	na.	DA	*	;
$ \begin{bmatrix} 5 & 1 & 1 & 1 & 1 & 2 & 1 & 1 & 1 \\ 1 & 7 & 1 & 1 & 1 & 0 & 7 & 1 & 1 & 1 & 1 & 1 & 1 & 1 & 1 & 1$	1-Methylnapthalene	- 62		na	an K	. 93		6 6	۲ ۲ ۲ ۲	1	2011 113	200	210	us Us	51 1	160	16 ]	420	440	230 j	1
$ \begin{bmatrix} 1 & 0 & 1 \\ 1 & 1 & 1 \\ 1 & 0 & 1 \\ 1 & 1 & 1 \\ 1 & 1 & 1 \\ 2 & 1 & 1 \\ 2 & 1 & 1 \\ 2 & 1 & 1 \\ 2 & 1 & 1 \\ 2 & 1 & 2 \\ 2 & 1 & 1 & 1 \\ 2 & 1 & 1 & 1 \\ 2 & 1 & 1 & 1 \\ 2 & 1 & 1 & 1 \\ 2 & 1 & 1 & 1 \\ 2 & 1 & 1 & 1 \\ 2 & 1 & 1 & 1 \\ 2 & 1 & 1 & 1 \\ 2 & 1 & 1 & 1 \\ 2 & 1 & 1 & 2 \\ 2 & 1 & 1 & 1 \\ 2 & 1 & 1 & 2 \\ 2 & 1 & 1 & 1 \\ 2 & 1 & 1 & 2 \\ 2 & 1 & 1 & 1 \\ 2 & 1 & 1 & 2 \\ 2 & 1 & 1 & 1 \\ 2 & 1 & 1 & 2 \\ 2 & 1 & 1 & 1 \\ 2 & 1 & 1 & 2 \\ 2 & 1 & 1 & 1 \\ 2 & 1 & 1 & 2 \\ 2 & 1 & 1 & 1 \\ 2 & 1 & 1 & 2 \\ 2 & 1 & 1 & 1 \\ 2 & 1 & 1 & 2 \\ 2 & 1 & 1 & 1$	2-Methylnaphthalene	80 1		Ħ	* 5				f 0.1	ı	ļ	250	200	na	, 66	290	130	460	700	270 j	1
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Dibenzofuran	5 E	<b></b> .	en 1	100	n 020		f oct	1 95 1 95	1		\$98	110	na	280 i	1001	87 ]	220	190	80 uj	ı
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Carbazole		-, :					730 mi	150 "	1	en en	67 n	1	na	280 u	130 u	94 u	140 u	71 n	4	ı
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Isophrone	- 120	3	и 1					250	۱	60	370	1	па	620	400	300	na	410	ı	ı
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Retene				nkc K	I ç			2 2 2 4	I		500	113	ца	113	Da	na	Da	nas	Пâ	ı
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Coprostanol	8 i	EL 22			130 C	a c	13 U II	11 YE	l t		37 :	190 1	U9	240 u	130 u	94 u	140 u	n 11	80 uj	1
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Phenol	- 151	8	Ξ.		230	133	n 0c7	00 n 991	<b>i</b> 1	al c	f /c	1900 n	an eu	, 2 1	130 u	470 ui	i 1400 u	i 710 u	800 uj	1
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Pentachlorophenol	- 120	n 200	i na		11 00.07 1	ă ș	- USC	1400	I		n 19	2009	EL EL	240 u	2500	94 u	140 u	71 u	11000 j	t
u       100       u       330       u       150       u       500       u       150       1       1       1       1       u       1       1       1       1       u       1<	4-Methylphenol			1	 ; ;		an c		- UUS	I		1000	2000 mi	2	1700	n 066	2400 i	4100 u	i 1400 uj	1400 uj	ι
u       100       u       44       u       100       u       100       u       130       j       100       u       130       j       17       j       75       j       71       u         0       u       100       u       ma       44       u       230       u       150       u       21       u       100       j       97       j       94       u       19       j       71       u         0       u       100       u       ma       44       230       u       150       u       130       u       100       u       130       j       94       u       140       u       71       u         0       u       130       u       150       u       150       u       130       u       130       u       130       u       140       u       120       u       130       u       130       u       130       u       130       u       140       u       120       u       130       u       130       u       140       u       11       u       140       u       10       u       130       u	Benzoic Acid	007I -	0075 n		1 040			1300 m	150 4	ł			ר י י	Da	280 u	130 u	94 ui	i 140 u	•	;	<b>.</b>
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Hexachloroethane	120	n n			:		330 "	150 1	I	1 E	67 n	ł	na	280 u	130	1 16	75. ]	n 11 n	98 j	1
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Hexachlorobenzene	- 120	3		; ;	220	4	230 a	150 1	ı	Ĩ	e7 n	ı	Цâ	280 u	26	94 u	19	n 1/	21 J	ī
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	1,2,4-1 richlorobenzene	- 120	a.	n i	‡ 3		4		18 :	ı	1	67 n	21 1	na	110 i	97 j	190	<b>1</b> 28	200	120 j	ŧ
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Hexachlorobutadiene	- 00	n :		‡ 3	220	1	330 n	150 1	1	en a	67 n	n 061	na	240 u	130	94 u	140 u	1 11 n	27 j	ŧ
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Dimetayipnnalate	-	⇒;		- 90c	+	1	5 (M) 11	1 20 n	1	U3	330 u	950 ui	Пâ	590 u	230	94 u	n 069	i 360 u	400 uj	ł
0       u       420       u       1300       uj       1400       uj       1200       uj       100       uj       100       uj       100       uj       100       uj       100       uj       120       uj       100       uj       120	Butylbenzylphinalate	- 120	n 10		10		1	5 u 025	1007	I	U3	84 ui	1300	na	380 L	130 u	04 n	210 u	ij 71 uj	80 uj	I
u 100 u na 59 u 120 u na 160 u 150 u - na 83 u 95 u na 160 u 130 uj 470 u 140 u 100 u 760 na 420 450 na 650 830 - na 330 320 na 350 130 uj 470 u 140 u 100 u 0 u 410 na 320 590 na 940 150 u - na 480 640 na 330 130 uj 430 j 450 250 1 -=No sample na=Not analyzed	Di-n-Butylphthalate		1 40 1 40		10061	1400			1 028	I		14000	810	na	4500 u	i 1200 u	1800 ul	i 5400 u	ij 1200 uj	940 j	ı
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Bis(2EH)phthalate	2100	19091 n		MCT	247	म्	10 AN / 7	20							5	-		•		
u 400 u 140 u 14 350 u 450 u 650 830 - na 330 320 na 350 1800 j 670 270 240 0 760 na 420 450 na 940 150 u - na 480 640 na 380 130 uj 430 j 450 250 1 0 1200 na 740 1000 na 1600 830 - na 810 960 na 730 1900 uj 1100 j 720 490 2 -=No sample na=Not analyzed	PCBs	005			03	120	ŝ	160 n	150 n	ł	Ug	83 u	95 u	Цâ	160 L	i 130 uj	470 u	. 140 u		600	I
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	1242	140	n 100		ŝ	150	¥ 4	- 057 720		1	66	330	320	Пâ	350	1800	670	270	240	640 j	ł
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	1254	- 820		ла		004		0.00	150	1		Vor	240		380	130	430 i	450	250	1600	I
) 1200 na /40 1000 na / 1000 oo oo - ua oro oo - ua oro oo ua oro oo	1260	- 120	Ħ	па		040		046	n Oct		917 G	810	090	e c	054			720	490	2800	ł
	Total PCB'S	- 820		Da	/40	non	ца	1000	- L.		PI	010	8		22						
	u=Not detected at detection	umit spown						na-Not an	livred												
	j=Estimated concentration							**** 1AL T BIT													

Table C3(cont.): Summary of Semivolatile Organics and Polychlorinated Biphenyls Detected in Settling Particulate Matter from Hylebos Waterway, December 1991 to June 1995 (ug/kg, dry weight).

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Irom Hylebos waterway, Decentuel	erway,	necelli			•		(ug/ng,	m	
Location			Mouth	Mouth of Waterway	way				
Station No.				H-7					1
Collection Period	12/91- 5/92	5/92- 11/92	11/92- 5/93	7/93- 12/93	12/93- 5/94	<b>.</b>	5/94- 11/94	11/94 6/95	
TOC (%)	2.2	2.6	2.3	2.3	2.2		2.1	2.0	
Acenaphthene	49 j	92 j	16	290	170	•	64 j	$\mathcal{II}$	· ••••
Acenaphthylene	38 j	49 j	50 j	59 j	52	•••••	61 j	58	•••••
Naphthalene	120 j	110	120	170	130		150	170	p
Fluorenc	120 j	110	140	350	320		100 j	100	- <del>•</del> ,
Anthracene	260	190	250	390	560		220	230	n
Phenanthrene	540	560	770	1500	1100		450	550	¤
Sum LPAH	1100 j	1100 j	1400 j	2800 j	2300	j	1000 ĵ	230	
Fluoranthene	710	850	1200	1700	1300		970	950	p
Benzo(a)anthracene	1300 u	340	530	530	480		400	170	n
Chrysene	1400	620	810	810	820		690	830	
Pyrene	630	840	820	1400	1100		830	840	p.
Benzofluoranthenes	2100 uj	840	1200	1100	1100		970	470	n
Benzo(a)pyrenc	270	280	330	410	350		370	360	n
Dibenzo(a,h)anthracene	480	57 j	į 69	16	56	·,	280	170	n
Indeno(1,2,3-cd)pyrene	190 u	180 j	230 -	260	210		220	230	n
Benzo(g,h,i)perylene	190 u	85 j	150	280	140		220	230	=
Sum HPAH	7300	4100 j	5300 j		j 5600	•	5000	830	
Total PAH	8400 j	5200 j	5700 j	9400	7900	, j	6000 j	118	
1-Methylnapthalene	52 j	54 j	n 06	na	na		, ,	I	
2-Methylnaphthalene	47 j	52 j	<u>51</u> j	180	130	_	81 j	170	n
Dibenzofuran	77 j	66	100	260	190	_	89 j	170	n
Carbazole	350 j	41 j	70 j	180	150	_	140 u	170	n
Isophrone	190 u	97 u	90 u	130	u 68	n	ı	170	n
Retene	350	360	290	ца	350	_	5	<del>6</del>	:
Coprostanol	na	na.	na	na	Пâ		na	5100	<b>ب</b> ر
Phenol	190 u	07 u	90 u	130	u .59	·,	140 j	170	n
Pentachlorophenol	ŗ	n 16	450 uj		uj 680	8	1400 u	840	Þ
4-Methylphenol	190 u	580	90 u			<b>n</b> -		0/1	<b>"</b>
Benzoic Acid	1500 j.	170 uj	20		uj 780	,	1500 uj	340	6
Hexachloroethane	190 u	n 16	90 U	130	י : ס.		ŀ	I G	
Hexachlorobenzene	40 j	54 j	n 06	<del>\$</del>	j 68	<b>p</b>	\$	51	3
1,2,4-Trichlorobenzene	190 u	12 j	90 u	130	uj 68	3	:	0/1	p.
Hexachlorobutadiene	39 j	51 j	110	20	j (88	n	43 1	4 7	-
Dimethylphthalate	190 u	97 u	90 u	130	u 68	, p		0/1	n
Butylbenzylphthalate	480 u	120	и 06	130		n (	1300 uj	170	a
Di-n-Butylphthalate	440 u	110 u	180 uj			5	170	170	
Bis(2EH)phthalate	7000	790 u	1000 uj	90	uj 750	(n)	1300	49000	'n
PCBs								ļ	
1242	100 u	97 u	90 n		u 110	n	n Li	\$	p,
1254	330	320	. 230	180	100	r.	120	011	•~~ ·
1260	100 u	<b>190</b>	190	290	170	_	270	170	
Total PCB'S	330	1100	420	470	270	i	390	280	·
u=Not detected at detection limit shown	imit shown				-=No sample	aple			
j=Estimated concentration					na=Not analyzed	nalyzec	_		
uj=Estimated detection limit									

Table C4: Results of Butyltin\* Analysis of Settling Particulate Matter from Hylebos Waterway, May 1994 to June 1995 (ug/kg, dry weight).

weig
dry
(ug/kg,
\$ 1995
June
5
1994
May

, ,

ľ ocation	Unner Turning	Irning	@ General	neral	@ Elf	11	Near Lincoln	ncoln			@ Occidenta	idental		
TOTATA	Dosin	0	Matals	ale	Atochem	em	Ē	Drain	Near Cenex	enex	Chemical	nical	Mout	h
Station	II-H	E	H	H-2	H-3	100	H-4	+	H-5	Ś	Щ	H-6	H-7	
Collection	5/04-	11/94-	5/94-	11/94-	5/94-	11/94-		11/94-	5/94-	11/94-	5/94-	11/94-	5/94-	11/94-
Date		6/95	11/94	6/95		6/95	11/94	6/95	11/94	6/95	11/94	6/95	I	6/95
Monobutvltin	13 u	24 u	50	20 ui	140	24 uj	58	na	76	na	55		32 u	20 uj
Dibutvltin	230	110	210	7.3 i		28		na	150	па	64	t	16 u	9.4 u
Tributvltin	620	280	450	160 ,	530 j	1300	660	na	480	па	240	I	210 j	55
Tetrabutyltin		12 u	13 u	10 u	13 u	12 u	10 j	па	11 u	na	12 u	1	8.3 u	9.8 u
* Domentod or oblowido	ablanido													

\*=Reported as chloride u=Not detected at detection limit shown

j=Estimated concentration

uj=Estimated detection limit

-=No sample na= Not analyzed

Table C5: Sediment Accumulation Rates for Hylebos Waterway fr	om Sediment
Trap Data, December 1991 to June 1995.	

		Number	Collection	SPM	SPM	BS	SPM	Dry	Gross	Accumulation
Station	Deployment Period	Days Deployed	Area cm2	Wet g Collected	Percent Solids	Percent Solids	Dry g Collected	Density* g/cm3	Accumulation g/cm2/year	Rate cm/year
I-1A	12/91-2/92	91	78.5	65.1	43	43	28	0.58	1.4	2.4
-1B		91	78.5	68.9	42	43	29	0.58	1.5	2.5
-1A	11/92-2/93	97	78.5	79.0	45	43	36	0.58	1.7	2.9
-1B	6)	97	78.5	77.3	46	43	36	0.58	1.7	
-1A	2-5/93	84	78.5	56.5	. 39	43	22	0.58	1.2	2.1
-1B	"	84	78.5	55.0-	41	43	23	0.58	1.2	2.1
-1A	7-9/93	70	78.5	82.2	39	43	32	0.58	2.1	. 3.0
-1B		70	78.5	83.6	39	43	33	0.58	2.2	. 3.1
-1A	12/93-3/94	111	78.5	74.3	43	43	32	0.58	1.3	2.3
-1B	**	111	78.5	73.9	42	43	31	0.58	1.3	2.:
-1A	35/94	57	78.5	35.7	29	43	10	0.58	0.8	1.4
-1B		57	78.5	36.6	30	43	11	. 0.58	0.9	1.:
-1A	5-8/94	90	78.5	92.0	33	43	30	0.58	1.6	2.1
-1B	` <b>u</b>	90	78.5	90.0	34 .	43	31	0.58	1.6	2.1
-1A	8-11/94	100	78.5	78.0	36	43	28	0.58	1.3	2.:
-1B		100	78.5	79.0	37	43	· 29	0.58	1.4	2.
-1A	11/94-3/95	97	78.5	133.0	45	43	60	0.58	2.9	4.
-1B	0	97	78.5	135.0	45	43	61	0.58	2.9	5.
-1A	3/95-6/95	98	78.5	123.0	28	43	34	0.58	1.6	. 2.
-1B	\$ <b>1</b>	98	78.5	102.0	28	43	29	0.58	1.4	2.
								mean=	1.6	2.
H-2A	2-5/92	84	78.5	54.1	41	43	22	0.58	1.2	2.
2B	ti	84	78.5	54.1	42	43	23	0.58	1,3	2.
-2A	5-8/92	90	78.5	109.5	43	43	47	0.58	2.4	4.
-2B	¥#	90	78.5	109.4	42	43	46	0.58	2.4	4.
-2A	8-11/92	98	78.5	114.3	45	43	51	0.58	2.4	4.
-2B	**	98	78.5	116.2	45	43	52	0.58	2.5	4.
-2A	2-5/93	84	78.5	74.9	44	43		0.58	1.8	3.
-2B	23	84	78.5	76.4	46	43		0.58	1.9	3.
-2A	7-9/93	70	78.5	86.4	41	43	35	0.58	2.4	4.
-2B		70	78.5	85.6	42	43	36	0.58	2.4	4.
-2A	9-12/93	70	78.5	69.4	44	43	31	0.58	2.0	3.
2B	**	70	78.5	77.7	43	43	33	0.58	2.2	
-2A	12/93-3/94	111	78.5	74.7	44	43	33	0.58	1.4	2.
-2B	н .	111	78.5	73.9	43	43	32	0.58	1.3	2
-2A	3-5/94	57	78.5	43.9	34	43	15	0.58		
-2B	11	57	78.5	43.0	34	43	15	0.58		
-2A	5-8/94	90	78.5	108.0	37	43	40	0.58	2.1	3
-2B	13	90	78.5	103.0	38	43	39	0.58		
-2A	8-11/94	100	78.5	103.0	38	43	39	0.58	1.8	3
-2B		100	78.5	102.0	37	43	38	0.58	1.8	3
-2A	11/94-3/95	97			37	43	3 28	0.58	1.4	2
-2B	IJ	97				43	3 29	0.58	1.4	2
-2A	3/95-6/95	98				43	35	0.58	1.7	2
-2B	te	98			33	43	3 35	0.58	1.6	2

BS= Bottom Sediment

\*=Estimated from Puget Sound Sediment Density Model (Crecelius, 1989)

Table C5(cont): Sediment Accumulation Rates for	· Hylebos Waterway from Sediment
Trap Data, December 1991 to June 1995.	

		Number	Collection	SPM	SPM	BS	SPM	Dry	Gross	Accumulation
G4+41++	Deployment	Days Deployed	Area cm2	Wet g Collected	Percent Solids	Percent Solids	Dry g Collected	Density* g/cm3	Accumulation g/cm2/year	Rate cm/year
Station	Period	91	78.5	134.6	49	41	66	0.55	3.4	6.2
H-3A -3B	12/91-2/92	91 91	78.5	134.0	49 49	41	65	0.55	3.3	6.1
-38 -3A	25/92	82	78.5	82.0	44	41	36	0.55	2.0	3.7
-3A	2	92	78.5	152.5	47	• 41	- 72	0.55	3.6	6.6
-3R	-0/ <i>72</i>	92	78.5	155.4	46	41	71	0.55	3.6	6.6
-3A	8-11/92	98	78.5	162.8	49	41	80	0.55	3.8	6.9
-3B	0	98	78.5	161.6	48	41	78	0.55	3.7	6.7
-3A	11/92-2/93	97	78.5	99.1	51	41	51	0.55	2.4	4.4
-3B	14	97	78.5	93.3	50	41	47	0.55	2.2	4.1
-3A	2-5/93	84	78.5	74.9	. 48	41	. 36	0.55	2.0	3.6
-3B	11	84	78.5	75.5	48	41	36	0.55	2.0	3.7
3A	9-12/93	70	78.5	88.5	46	41	41	0.55	2.7	4.9
-3A	12/93-3/94	111	78.5	125.2	46	41	58	0.55	2.4	
-3B	. 11	111	78.5	122.3	45	41	55	0.55	2.3	4.2
-3A	3-5/94	57	78.5	68.8	39	41	27	0.55	2.2	4.0
3B	55	57	78.5	71.9	38	41	27	0.55	2.2	4.1
-3A	5-8/94	90	78.5	136.0	37	41	50	0.55	2.6	4.8
3B	0	· 90	78.5	139.0	36	41	50	0.55	2.6	4.7 5.1
-3A	8-11/94	100	78.5	149.0	40	41	60 50	0.55 0.55	2.8 2.8	5.1
-3B		100	78.5	148.0	40	41	59	0.55	2.8	3.6
-3A	11/943/95 "	97 97	78.5	98.0	42	41	41 41	0.55	2.0	3.6
-3B		97	78.5	96.0	43 38	41 41	52	0.55	2.0	4.6
-3A	3/95-6/95	98	78.5 78.5	138.0 134.0	36	41	32 48	0.55	2.3	4.0
-3B	······	98	18.5	154,0		41	40	mean=	2.6	4.8
TT AA	5-8/92	. 89	78.5	139.8	48	51	67	0.75	3.5	4.7
H-4A -4B	5-0/92	. 89	78.5	141.5	48	51		0.75	3.5	4.7
-4A	8-11/92	99	78.5	160.1	51	51		0.75	3.8	5.1
-4A -4B	0-11172 "	99	78.5	161.4	51	51		0.75	3,9	5.2
-4A	11/92-2/93	96	78:5	90.0	40	51		0.75	1.7	2.3
-4B	11)/2 2///	96	78.5	90.4	52	51		0.75	2.3	3.0
-4A	2-5/93	85	78.5	94.4	46	51		0.75	2.4	3.2
~4B	11	85	78.5	91.6	49	51		0.75	2.5	3.3
-4A	12/93-3/94	111	78.5		51	51		0.75	2.2	2.9
-4B	64 64	. 111	78.5		50	51		0.75	2.2	3.0
-4A	3-5/94	57	78.5	58.4	40	51	. 23	0.75	- 1.9	2.5
-4B		57	78.5	54.6	40	51	22	0.75	1.8	2.4
-4A	5-8/94	90	78.5		39	51	49	0.75	2.5	3.4
-4B	4	90	78.5	132.0	38	51	50	0.75	2.6	3.5
-4A	8-11/94	100	78.5	81.0	43	51	35	0.75	1.6	2.2
-4B	н	100	78.5	79.0	. 43	51	34	0.75	1.6	2.1
	4							mean=	2.5	3.3
H-5A	12/91-2/92	90	78.5			51			2.9	3.9
-5B	n	90	78.5			51			2.8	3.8
-5A	58/92	89	78.5			51			4.9	6.5
-5B	44	89	78.5			51			5.0	
-5A	8-11/92	99	78.5			51			5.5	7.4
-5B		99	78.5			51			5.4	
-5A	12/93-3/94	111	78.5			51			2.2	
-5B	**	111	78.5			51			2.2	
-5A	3-5/94	. 57	78.5			51			1.8	
-5B	H	57	78.5						2.0	
-5A	5-8/94	. 91	78.5			51			2.8	
-5B	"	91	78.5						3.0	
-5A -5B	8-11/94	99	78.5						3.3 3.3	
^ R		99	78.5	169.0	i 42	21	ι /1	0.75	2.3	·••

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BS= Bottom Sediment

\*=Estimated from Puget Sound Sediment Density Model (Crecelius, 1989)

Table C5(cont): Sediment Accumulation Rates for Hylebos Waterway from Sediment Trap Data, December 1991 to June 1995.

		Number	Collection	SPM	SPM	BS	SPM	Dry	Gross	Accumulation
Station	Deployment Period	Days Deployed	Area cm2	Wet g Collected	Percent Solids	Percent Solids	Dry g Collected	Density* g/cm3	Accumulation g/cm2/year	Rate cm/year
H-6A	12/91-2/92	90	78.5	66.1	49	51	32	0.75	1.7	2.2
6B	*1	90	78.5	65.9	49	51	32	0.75	1.7	2.2
-6A	2-5/92	84	78.5	56.7	49	51	28	0.75	1.5	2.1
-6B	12	84	. 78.5	56.1	48	51	27	0.75	1.5	2.0
-6A	5-8/92	91	78.5	119.7	48	51	57	0.75	2.9	3.9
-6B	11	91	78.5	122.5	48	51	59	0.75	3.0	4.0
-6A	8-11/92	99	78.5	120.4	52	51	63	0.75	2.9	3.9
-6B		99	78.5	120.5	52	51	63	0.75	2.9	3.9
-6A	11/92-2/93	96	78.5	61.9	52	51	32	0.75	1.6	2.1
-6B	H	96	78,5	61.8	51	51	32	0.75	1.5	2.0
-6A	2-5/93	85	78.5	22.5	39	51	9	0.75	0.5	0.6
-6B	*1	85	78.5	21.5	39	51	. 8	0.75	0.5	0.6
-6A	7-9/93	70	78.5	102.0	48	51	49	0.75	3.3	4.3
-6B	<b>4</b>	70	78.5	102.3	49	51	50	0.75	3.3	4.5
-6A	9-12/93	70	78.5	74.4	47	51	35	0.75	2.3	. 3.1
6B	**	70	78.5	72.8	. 48	51	35	0.75	2.3	3.1
-6A	12/93-3/94	111	78.5	87.7	53	51	46	0.75	1.9	2.6
-6B	60 60	111	78.5	89.1	52	. 51	46	0.75	1.9	2.6
-6A	3-5/94	57	78.5	46.7	37	51	17	0.75	1.4	1.9
6B	н н	57	78.5	45.8	39	51	18	0.75	1.5	1.9
-6A	5-8/94	92	78.5	109.0	42	51	46	0.75	2.3	3.1
-6B	"	92	78.5	109.0	40	51	44	0.75	2.2	2.9
-6A	8-11/94	97	78.5		45	51	62	0.75	3.0	4.0
-6B	B	97	78.5	129.0	45	51	58	0.75	2.8	3.7
								mean=	2.1	2.8
H-7A	12/91-2/92	90	78.5	81.7	55	62	45	1.01	2.3	2.3
-7B		90	78.5	80.1	- 55	62		1.01	2.3	2.2
-7A	2-5/92	84	78.5	75.0	54	62	41	1.01	2.2	2.2
7B		84	78.5	73.7	52	. 62		1.01	2.1	2.1
-7A	5-8/92	91	78.5	177.0	56	62		1.01	5.1	5.0
-7B		91	78.5		56	62	103	1.01	5.3	5.2
-7A	8-11/92	98	78.5		59	62		1.01	5.0	4.9
-7B		98	78.5		60	62		1.01	5.1	5.0
-7A	11/922/93	97	78.5		62	62		1.01	3.1	3.0
-7B	fa	97	78.5		59	62		1.01	3.0	2.9
-7A	2-5/93	85	78.5		56	62		1.01	2.3	2.2
-7B	"	85	78.5		50	62		1.01	2.1	2.0
-7A	7-9/93	70	. 78.5		54				4.0	3.9
-7A	9-12/93	70							3.3	3.2
-7B	"	70			55					
-7A	12/93-3/94	111	78.5							
-7B	,∠195" 319 <del>"</del> "	111	78.5							
-7B -7A	3-5/94	56				62			1.8	
-7A -7B	<i>үшү үч</i> н	56								
-7в -7в	5-8/94	92								
-7в 7А	3-8/94 8-11/94	92 97					•			
-7A -7A	8-11/94 11/94-3/95	98								
-7A -7B	11/94-5/95	98 98								
	Settling Particu		/0.5		54			mean=		

BS= Bottom Sediment

\*=Estimated from Puget Sound Sediment Density Model (Crecelius, 1989)

# Sitcum Waterway

Table C6: Results of Conventionals and Metals Analysis of Settling Particulate Matter from Sitcum Waterway, June 1991 to June 1995 (mg/kg, dry weight).

Location			5 5 7 7 7 7 7 7 7 7 7 7 7 7 7 7 7 7 7 7			Terminal 7 @ North Corner Drain	North Corr.	ter Drain					
Station No.							S-1						
Collection	6/91-	9/91-	11/91-	2/92-	5/92-	8/92-	11/92-	2/93-	-1/93-	9/93-	12/93-	11/94-	3/95-
Period	16/6	11/91	2/92	5/92	8/92	11/92	2/93	5/93	9/93	12/93	11/94	3/95	6/95
Total Solids (%)		50	62	57	59	58	60	57	58	58	na	56	41
TOC %	I	2.1	1.9	2.3	2.8	2.6	2.1	2.3	2.4	2.3	ла	2.0	2.4
Metals													
Arsenic	I	53	58	57	70	58	66	83	40	49	na	19	24
Copper	\$	150	150	150	180 j	170 j	150	150	130	150	na	85	66
Lead	I	320	260	260	260	210	230	220	190	200	na	110	160
Zinc	I	360	310	290	330	260	280	310	230	260	na	140	190
I ociation		and the second se	-		19944 P. P.	Scal	Sealand Terminal	1					
Station No.							S-2						
Collection	6/91-	9/91-	11/91-	2/92-	5/92-	8/92-	11/92-	2/93-	7/93–	9/93-	12/93-	11/94-	3/95-
Period	16/6	11/91	2/92	5/92	8/92	11/92	2/93	5/93	9/93	12/93	11/94	3/95	6/95
Total Solids (%)		57	62	56	59	51	62	56	na	na	na	na	na
TOC %	I	2.0	1.2	1.7	2.6	1.7	1.4	1.7	na	na	na	na	na
Metals													
Arsenic	I	28	22	24	23	22	21	19	na	na	UA	na	na
Copper	I	130	95	86	100 j	j 06	80	20	<b>n</b> a	na	na	na	Пâ
Lead	I	110	94	96	95	80	85	80	na	na	na	na	na
Zinc	1	160	150	140	160	130	150	120	na	na	na	na	na
and the surfaced													

na=Not analyzed j=Estimated concentration

Table C6(cont.): Results of Conventionals and Metals Analysis of Settling Particulate Matter from Sitcum Waterway, June 1991 to June 1995 (mg/kg, dry).

s-3	Location						Termi	Terminal 7 @ Berth B	hВ					
minipolity         691-         919-         1191-         292-         592-         1192-         293-         1293-         1194-         395-         1194-         395-         1194-         395-         1194-         395-         1194-         395-         1194-         395-         1194-         395-         1194-         395-         1194-         395-         1194-         395-         1194-         395-         1194-         395-         1194-         395-         1194-         395-         1194-         395-         1194-         395-         1194-         395-         1194-         395-         1184-         395-         1194-         395-         1194-         395-         1184-         395-         1194-         395-         1184-         113         1	Station No.				·			S-3						
991         1191         292         592         892         1192         293         593         1293         1194         395           % bitks (%)         53         -         65         61         58         -         62         54         10         395           %         2.2         -         1.5         1.6         2.6         -         62         54         10         59         10         50           %         49         -         45         44         48         -         52         43         32         47         10         13           %         140         -         150         120         150         1         10         120         130         10         10         130           %         140         -         200         200         200         100         100         100         100           %         200         200         200         200         200         200         200         100           %         200         200         200         200         200         200         100         100           %         100         200 <th>Collection</th> <th>6/91-</th> <th>9/91-</th> <th>11/91-</th> <th>2/92-</th> <th>5/92-</th> <th>8/92-</th> <th>11/92-</th> <th>2/93-</th> <th>7/93</th> <th>9/93-</th> <th>12/93-</th> <th>11/94-</th> <th>3/95-</th>	Collection	6/91-	9/91-	11/91-	2/92-	5/92-	8/92-	11/92-	2/93-	7/93	9/93-	12/93-	11/94-	3/95-
	Date	16/6	11/91	2/92	5/92	8/92	11/92	2/93	5/93	9/93	12/93	11/94	3/95	6/95
$\pi$ $2.2$ $ 1.5$ $1.6$ $2.6$ $ 1.6$ $2.1$ $1.9$ $2.0$ $1.3$ $1.3$ $\pi$ $4.9$ $ 4.5$ $4.4$ $4.8$ $ 5.2$ $4.7$ $1.3$	Total Solids (%)	53		65	61	58		62	58	60	54	na	59	1
sinc         49         -         45         44         48         -         52         43         32         47         na         17           cr         140         -         130         120         130         130         130         na         75           cr         140         -         250         240         250         -         230         240         na         130           450         -         300         290         260         -         260         230         230         309         na         130           6/91         9/91         11/91         2/92         5/92         8/92         11/92         2/93         7/93         9/93         11/94           6/01         9/91         11/91         2/92         5/92         11/92         2/93         12/93         11/94           6/01         9/91         11/91         2/92         5/92         11/92         2/93         12/93         11/94           6/01         9/91         11/91         2/92         5/92         11/92         2/93         11/94         3/95           6/01         0         1         1         1 <td>FOC %</td> <td>2.2</td> <td>ŧ</td> <td>1.5</td> <td>1.6</td> <td>2.6</td> <td>i</td> <td>1.6</td> <td>2.1</td> <td>1.9</td> <td>2.0</td> <td>na</td> <td>1.3</td> <td>I</td>	FOC %	2.2	ŧ	1.5	1.6	2.6	i	1.6	2.1	1.9	2.0	na	1.3	I
tic 49 - 45 44 48 - 52 43 32 47 na 17 14 140 - 130 120 120 150 na 75 290 - 250 240 250 - 220 170 190 240 na 130 130 145 145 130 110 120 120 130 na 130 130 145 145 130 145 145 145 145 145 145 145 145 145 145	Metals												:	
er         140         -         130         120         150         130	Arsenic	49	ł	45	44	48	I	52	43	32	47	na	17	1
	Conter	140	ł	130	120	150 j	I	130	110	120	150	na	75	ŧ
450         -         300         290         260         -         260         230         210         na         110           No.           6)1         9/91         11/91         2/92         5/92         11/92         2/93         9/93         12/93         11/94 $nNo.         6/91         9/91         11/91         2/92         8/92         11/92         2/93         9/93         12/93         11/94           g/91         11/91         2/92         8/92         11/92         2/93         9/93         12/94         3/95           solids (%)         -         56         56         5/92         8/93         9/93         12/94         3/95           solids (%)         -         2.0         1.7         1.9         3.05         5/93         9/93         12/94         3/95           solids (%)         -         2.0         1.7         1.5         1.7         1.9         1.9         1         1/94         3/95           solids (%)         -         2.0         3.0         5/93         9/93         12/94         3/95           solids (%)         -         1.5         1.5         $	Lead	290	I	250	240	250	ł	220	170	190	240	na	130	I
tori No. $\frac{6/91-}{6/91-} \frac{9/91-}{9/91-} \frac{11/91-}{11/91-} \frac{2/92-}{2/92-} \frac{5/92-}{5/92-} \frac{8/92-}{11/92-} \frac{11/92-}{2/93-} \frac{2/93-}{9/93-} \frac{12/93-}{12/93-} \frac{11/94-}{3/95-} \frac{3/95-}{3/95-} \frac{11/94-}{3/95-} \frac{3/95-}{2/93-} \frac{11/94-}{3/95-} \frac{3/95-}{2/93-} \frac{11/94-}{3/95-} \frac{3/95-}{2/93-} \frac{11/94-}{2/93-} \frac{3/95-}{2/93-} \frac{11/94-}{2/93-} \frac{3/95-}{2/93-} \frac{11/94-}{2/93-} \frac{3/95-}{2/93-} \frac{11/94-}{2/93-} \frac{3/95-}{2/93-} \frac{11/94-}{2/92-} \frac{3/95-}{2/93-} \frac{11/92-}{2/93-} \frac{2/93-}{2/93-} \frac{11/94-}{2/94-} \frac{3/95-}{2/93-} \frac{11/94-}{2/94-} \frac{3/95-}{2/94-} \frac{11/94-}{2/94-} \frac{3/95-}{2/94-} \frac{11/94-}{2/94-} \frac{3/95-}{2/94-} \frac{11/94-}{2/94-} \frac{11/94-}{2/94-} \frac{11/94-}{2/94-} \frac{3/95-}{2/94-} \frac{11/94-}{2/94-} 11/94$	Zinc	450	I	300	290	260	***	260	230	220	310	BI	110	-
S-4           S-4           stion $6/91 9/91 11/91 2/92 5/92 11/92 2/93 12/93 12/93 11/94-$ stion $9/91$ $11/91 2/92 5/92 8/92 11/92 2/93 7/93 9/93 11/94 3/95$ solids (%)         -         56         56         55         55         56 $11/94 3/95$ solids (%)         -         2.0 $1.7$ $1.92$ $1/92 1/93 1/94 3/95$ solids (%)         -         56         56         55         56         56 $11/94 3/95$ solid< (%)	ocation						Mou	th @ Termin <sup>®</sup>	al 7					
tion $6/91 9/91 11/91 2/92 5/92 8/92 11/92 2/93 12/93 11/94 9/91$ $11/91$ $2/92$ $5/92$ $8/92 11/92 2/93 12/93 11/94 8/91$ $11/91$ $2/92$ $5/92$ $8/92$ $11/92 2/93 12/93 11/94-$ Solids (%) $ 56$ $56$ $56$ $56$ $86$ $11/94-$ Solids (%) $ 2.0$ $11/7$ $11/9$ $2/93$ $11/94 3/95$ Solids (%) $ 2.0$ $11/7$ $11/9$ $3/92$ $11/94 3/95$ Solids (%) $ 2.0$ $11/7$ $11/9$ $11/7$ $11/7$ $11/7$ $11/7$ $11/7$ $11/7$ $11/7$ $11/9$ $11/94-$ Solid $ 3/92$ $11/7$ $11/7$ $11/7$ $11/7$ $11/7$ $11/7$ $11/7$ $11/7$ $11/7$ $11/7$ $11/7$ $11/7$ $11/7$ $11/7$ $11/7$ <td>tation No.</td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td>4-2</td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td>	tation No.							4-2						
9/91 $11/91$ $2/92$ $5/92$ $5/92$ $5/92$ $5/92$ $5/92$ $5/92$ $5/92$ $5/92$ $11/94$ $3/94$ $%$ $ 56$ $56$ $55$ $56$ $56$ $11/94$ $3/94$ $%$ $ 2.0$ $1.7$ $1.9$ $3.0$ $1.7$ $1.9$ $56$ $56$ $1.9$ $11/94$ $3/94$ $%$ $ 2.0$ $1.7$ $1.9$ $3.0$ $1.7$ $1.9$ $1.94$ $3/94$ $%$ $ 2.0$ $1.7$ $1.9$ $3.0$ $1.7$ $1.9$ $1.9$ $1.9$ $1.9$ $%$ $ 3.0$ $1.7$ $1.9$ $3.0$ $1.7$ $1.9$ $1.9$ $1.9$ $1.9$ $%$ $ 3.0$ $1.7$ $1.7$ $1.5$ $1.9$ $1.9$ $1.9$ $%$ $ 1.0$ $7.0$ $7.0$ $1.9$ $1.9$	-	6/91-	9/91-	11/91-	2/92	5/92-	8/92	11/92-	2/93	-£6//	9/93-	12/93-	11/94-	3/95-
	Jate	16/6	11/91	2/92	5/92	8/92	11/92	2/93	5/93	9/93	12/93	11/94	3/95	6/95
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	otal Solids (%)	1	56	56	52	57	62	60	55	56	56	па	1	52
-       32       17       19       17       11       25       16       14       11       na         -       -       110       76       76       99       56       73       54       70       71       na         -       250       99       100       85       44       82       72       53       70       na         -       370       130       140       130       91       130       130       130       130       130       na	OC %	I	2.0	1.7	1.9	3.0	1.7	1.5	1.7	1.9	1.9	na	I,	1.8
-       32       17       19       17       11       11       11       11       11a         -       110       76       76       76       99       56       73       64       70       71       1na         -       250       99       100       85       44       82       72       63       70       na         -       370       130       140       130       91       130       130       130       na	Actals			<b>.</b>							:			ć
- 110 76 76 99 ј 66 ј 73 64 70 71 па - 250 99 100 85 44 82 72 63 70 па - 370 130 140 130 91 130 130 110 130 па	Arsenic	I	32	17	19	17	11	25	16	14	11	na	1	0.1
- 250 99 100 85 44 82 72 63 70 na - 370 130 140 130 91 130 130 110 130 na	Copper		110	76	. 76	· [ 66	66 j	73	2	70	71	na	I	4
– 370 130 140 130 91 130 130 110 130 na	Lead	I	250	66	100	85	44	82	72	63	70	na	1	21
-=No sample na=Not anaivzed	Zinc	I	370	130	140	130	91	130	130	110	130	na	-	20
aa⊒Nut anaivred														
	na=Not analvzed													

not allaryzou

j=Estimated concentration

Table C7: Summary of Semivolatile Organics Detected in Settling Particulate Matter from Sitcum Waterway, June 1991 to June 1995 (ug/kg, dry weight).

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Location		5	Head	Head @ North Corner Drain 8-1	orner Drain							Sealand Terminal S-2	rminal			
Station No. Collection Date	<u>6/91–</u> 11/91	11/91- 5/92	5/92- 11/92	11/92- 5/93	7/93- 12/93	12/93- 5/94	5/94- 1 11/94	11/94- 6/95	6/91- 11/91	11/91- 5/92	5/92- 11/92	11/92- 5/93	7/93- 1 12/93	12/93- 5/94	5/94- 11 11/94	11/94- 6/95
TOC (%)	2.1	2.1	2.7	2.2	2.4	na	na	2.2	2.0	1.5	2.2	1.6	na	na	na	na
Acenanhthene	140	200	270	350	200	na	na	120	56 j	35 j	72	72 j	па	na	na	na
Acenanhthylene	19 1	50 i	63 j	81 j	66	na	na	33 j	21 j	18 j	20 j	24 j	na	na	ца	na
Nanhthalene	110	, 061	220	250	180 j	па	na	59 u	240	59 j	120	210	na	na	na	ла
Fluorene	130	270	310	530 j	320	na	na	180	76 j	200 u	71 j	74 j	па	na	na	na
Anthracene	87 ;	320	310	710	510	na	па	360	240 u	66 j	62	100	113	Пa	na	na
Phenanthrene	880	1400	1400	2700	1400	na	na	780	360	330	290	300	na	na	na	na
Sum LPAH	1400 j	2400 j	2600 j	4600 j	2700 j	na	na	1500 j	750 j	500 j	650 j	680 j	na .	na	na	па
Fluoranthene	1400	2400	2000	3600	2200	na	na	1200	1800	670	470	530	na	na	na	na
Benzo(a)anthracene	700	1800	. 650	1100 j	800	na	na	350 u	380	360	160	220	na	na	na	na
Chrysene	430 1	2000	930	1400	1400	na	па	590	340	450	210	280	na	na	na	na
Durene	850 i	2100	1600	1800	1600	na	na	810	1400	630	380	340	na	na	na	na
Renzofluoranthenes	650 ui	2100	1300	1800	1400	na	na	580	610	630	320	450	na	na	na	na
Renzo(a)mrene		510	570	380	560	na	na	320	260 u	310	120	130	na	na	na	na
Dihenzo(a, h)anthracene	270 u	440 u	74	120	100 j	na	na	59 u	600 u	500 u	21 j	n 66	na	na	na	na
Indeno(1, 2-cd)nvrene			380	320	360	na	na	220 u	240 u	130 j	85 j	81 j	na	na	na	na
Renzo(a h i)nervlene	110 u	490	290	180	360	na	na	210	240 u	110 j	50 j	60 j	na	na	na	na
Sum HPAH	4400 i	12000	7800	11000 j	8800 j	na	na	3700	5900	3300 j	1800 j	2100	na	na	na	ца
Total PAH	5800 i	14000	100001	16000 j	12000 j	na	na	5200 j	6700 j	3800 j	2500 j	2900	na	na	na	na
Phenol	110 u	1	80 u	560 u	180 u	na	na	59 u	240 u	200 u	71 u	n 66	ла	na	na	na
Pentachloronhenol		I	80 u	430 u	1400 uj	па	na	300 u	ŧ	ł	71 u	490 uj	Ла	na	пa	na
4-Methylnhenol	110 u	180 u	730	560 u	200	na	na	ł	240 u	200 u	390	n 66	na	па	na	na
1-Methylnaphthalene	68 i	120 j	140	150	I	na	na	1	86 j	42 j	62 j	63 j	na	na	na	na
2-Methylnanhthalene	62 i	100 j	130 j	200	80· j	na	na	86 u	75 j	31 j	53 j	56 j	na	na.	na	na
Carhazole	550 uj	440 j	96 ,	260 j	100 j	na	па	59 u	100 j	1000 uj	25 j	46 j	na	na	na	na
Benzoic Acid		3300 uj	310 u	4200	2500 uj	па	na	120 uj	3400uj	2200 j	n 009	420 uj	na	'na	na	na
Benzv1 Alcohol	550 u	, I	80 u	85 uj	180 u	na	na	59 u	1200 u	1	71 u	n 66	na	Пâ	па	пa
Retene	210 j	460	390	440	l	na	na	250	660	670	370	450	na	na	113	пa
Coprostanol	, na	na	na	na	na	na	na	2600	па	na	na	na	na	na	na	na
Dihenzofuran	110	210	270	340 j	220	na	na	120	83 j	44 j	69		na	na	na	na
Diethvlohthalate	110 u	180 u	80 u	85 u	180 uj	na	na	59 u	240 u	200 u	71 u	99. u	na	na	na	na
Dimethylohthalate	55 i	390	230	110	230	na	na	140	240 u	200 u	71 u	99 u	na	na	na	ца
Rutvlhenzvlnhthalate	270 u	440 u	80 u	120	200 uj	na	na	59 u	600 u	500 u	71 u	n 66	na	па	na	na
Di-n-hutvlnhthalate	110 u	1000 u	600	85 u	270 uj	na	na	340 u	240 u	570 u	71 u	22 uj	na	na	na	na
Di-n-octv/nhfhalate		180 ui	200 ui	85 u	540 u	na	na	59 u	240 uj	200 uj	180 uj	n 66	na	NA	na	na
Ris(7FH)nhthalate				34000	170000	IIA	па	1400 u	1000 u	n 086	530 u	780 uj	na	na	na	па
-=No sample	1					***			j=Estimated concentration	concentrat	uo					
u=Not detected at detection limit shown	m limit show	u							na=Not analyzed	lyzed						
ui=Estimated detection limit	nit															

Table C7(cont.): Summary of Semivolatile Organics Detected in Settling Particulate Matter from Sitcum Waterway, June 1991 to June 1995 (ug/kg, dry weight).

691- $11/91$ $5/92 11/92 5/92 11/92 5/93-$ ne $2.2$ $1.6$ $2.6$ $1.9$ $500$ $500$ $500$ fence $220$ $370$ $350$ $500$ $300$ $200$ $300$ e $130$ $1100$ $220$ $370$ $360$ $500$ $71$ $1$ $1100$ $220$ $370$ $360$ $500$ $1100$ $1300$ $1300$ $1100$ $2200$ $1200$ $2000$ $1100$ $2500$ $1400$ $1200$ $1200$ $570$ $1100$ $2500$ $1400$ $1200$ $360$ $570$ $1100$ $2500$ $1400$ $1200$ $370$ $2400$ $1100$ $2500$ $1400$ $1200$ $570$ $11100$ $2500$ $1400$ $1200$ $2200$ $1400$ $11100$ $1900$ $1100$ $1200$ $370$ $11100$ $1900$ $1100$ $1100$ $1200$ $1200$ $11100$ $1100$ $1100$ $1100$ $1100$ $1100$ $11100$ $1100$ $1100$ $1100$ $1100$ $1100$ $11100$ $1100$ $1100$ $1100$ $1100$ $1100$ $1100$ $230$ $1100$ $1100$ $1100$ $1100$ $1100$ $1100$ $1100$ $1100$ $1100$ $1100$ $1100$ $1100$ $1100$ $1100$ $1100$ $1100$ $1100$ $230$ $1100$ $1200$ $1100$ $11$	Location Station No.			Ļ	Terminal 7 @ Berth B S-3	) Berth B						- <b>-</b>	Mouth @ lermmal / S-4	srmnal /			
	Collection Date	6/91- 11/91	11/91- 5/92			7/93- 12/93	12/93- 5/94	1	11/94- 6/95	6/91– 11/91	11/91- 5/92	5/92 11/92	11/92- 5/93		12/93- 5/94		11/94- 6/95
	TOC (%)	2.2	1.6	2.6	1.9	2.0	na	na	1.3	2.0	1.8	2.4	1.6	1.9	na	na	1.8
	Acenaphthene	250 j	390	290	500	200	na	па	350	36 j	47 j	74 j	75 J	86 j	na	na	27 ;
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Acenaphthylene	22 j	43			30 j	na	na	23 j	S. j	11 j	15 j	95 u	21 j	na	na	15 j
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	Naphthalene	130 ]	190	200	300	150 j	na	na	96 u	45 j	59 j	63 j	86 j	130 j	na	na	n 0/
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	Fluorene	250 ]	780	350	620	500	na	na	340	31 j	64 j	67 j	100	87 ]	па	na	35 ]
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	Anthracene	71 j	1100	220	570	880	na	na	290	35 j	150	62 j	130	90 j	na	< na	10 n
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	Phenanthrene	1100 j	2900	1200	2000	1600	па	na	1300	330.	470	310	530	390	na	na	170 u
1400         200         1430         1800         1430         1800         1430         1800         1430         1800         1430         1800         1430         1800         1400         1800         1400         1800         1400         1800         1400         1400         1800         1400         1800         1400         1800         1400         1800         1400         1800         1400         1800 <th< td=""><td>Sum LPAH</td><td>1900 j</td><td>5400 j</td><td>2300</td><td>4000</td><td>3400 j</td><td>na</td><td>na</td><td>2300 j</td><td>480 j</td><td>800 j</td><td>590 j</td><td>920 j</td><td>800 j</td><td>ла</td><td>na</td><td>77 j</td></th<>	Sum LPAH	1900 j	5400 j	2300	4000	3400 j	na	na	2300 j	480 j	800 j	590 j	920 j	800 j	ла	na	77 j
365         1400         430         140         430         140         330         130         430         140         330         130         430         270         430         740         730 </td <td>Fluoranthene</td> <td>1400 j</td> <td>2900</td> <td>1400</td> <td>1800</td> <td>1480</td> <td>na.</td> <td>na</td> <td>1500</td> <td>069</td> <td>. 061</td> <td>460</td> <td>870</td> <td>630</td> <td>na</td> <td>na</td> <td>290 u</td>	Fluoranthene	1400 j	2900	1400	1800	1480	na.	na	1500	069	. 061	460	870	630	na	na	290 u
	Benzo(a)anthracene	565 j	1400	430	570	430	na	na	350	670	480	140	330	190	na	na	70 u
780 1900 1200 030 40 1200 030 147 1 ma ma 1000 620 750 390 540 500 ma ma 223 130 030 640 120 110 1 ma ma 235 125 130 130 130 130 130 130 130 130 130 130	Chrysene	250 j	1000	610	590	600	па	па	530	310	480	220	450	270	na	na	160
490         [500         830         740         na         450         870         630         640         400         na         na         130           255         330         660         601         601         77         na         na         550         200         210         100         110	Pyrene	780 i	1900	1200	930 j	1200	na	па	1000	620	750	390	540	500	· na	na	200 u
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	Renzoffmoranthenes	490 i	1500	830	840	740	па	na	450	870	630	300	640	400 j	na	ла	120 u
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Renzo(a)nvrene	235 1	390	360	220 i	330	na	na	260	350	240	120	215	150 j	na	na	70 u
	Dihenzo(a, h)anthracene	26 1		60 j	60 1	47 j	па	na	55 u	250 u	430 u	12 j	95 u	180 u	na	na	70 u
98         140         180         69         180         na         140         150	Indeno(1.2.3-cd)pvrene	130 i	160	220 j	148	, 061 190	na	na		260	110 j	81	170	110 j	na	na	n 07
	Benzo(g.h.i)pervlene	1 86		180	69	180	na	na	140 u	150	170	57 j	150	120 j	па	. na	70 <sup>.</sup> u
5900j 5000j 7600j 7600j 800j anan6400j 4900j 2400j 4300j 3200j ananan390u=87u77u170u00035u1800uanma<	Sum HPAH	4000 i	00/6	5300 j	5200	5200 j	na	na	4100	3900	4100 j	1800 j	3400	2400 j	na	na	160
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	Total PAH	5900 j	15000 j	7600 j	9200 j	8600 j	na	na	6400 j	4400 j	4900 j	2400 j	4300 j	3200 j	na	na	240 j
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Phenol	420 j				170 uj	na	na	63 u	<u>97</u> u	170 u	1000	95 u	180 и	na	na	81 u
530 j 230 390 74 u 230 na na 55 u 97 u 170 u 3600 95 u 180 na na 72 120 j 170 140 200 na na 16 120 j 120 j 140 270 u 90 uj 100 uj na na 160 u 22 j 56 j 57 j 97 j na na 9 110 u 1900 u 1200 u 980 uj 100 uj na na 110 uj 360 uj 550 j 57 j 97 j na na 9 1430 u - 21 j 74 uj 170 uj na na 150 u 230 460 230 j 570 uj 180 u na na 22 200 j 540 3300 j 5700 uj na na 22 200 j 540 3300 j 5700 uj na na 22 200 j 440 230 450 j 2700 uj na na 22 200 j 240 230 460 230 460 na na na na na 18 na 2200 j 420 2700 uj na na 22 200 j 220 460 230 460 230 460 na na na na 18 na 22 200 j 420 2700 uj na na 22 20 j 420 2700 uj na na 22 20 j 420 2700 uj na na 22 20 j 220 460 230 460 230 460 na na na na 18 na 22 200 j 420 270 uj na na 22 20 j 230 460 230 460 230 460 130 u na na 24 20 200 j 420 170 u 200 u 95 u 180 uj na na 23 20 210 u 350 u 87 u 74 u 170 u 200 u 95 u 180 uj na na 23 u 140 u 220 uj 74 u 170 u na na 55 u 97 u 170 u 200 u 95 u 180 uj na na 18 83 u 140 u 220 uj 74 u 170 u na na 55 u 97 u 170 u 200 u 95 u 180 uj na na 18 83 u 140 u 220 uj 74 u 170 u na na 55 u 97 u 170 u 200 u 95 u 180 uj na	Pentachlorophenol	390 u	I		370	1700 uj	na	na		500 u	I	6L	470 uj	1800 u	na	na	350 u
	4-Methylphenol	530 j	230	390		230	na	na		97 u	170 u	3600	95 u	180	na	na	n 0/
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	1-Methylnaphthalene	120 j	170	140	200	na	na	na	1	22 j	56 j	53 j	65 j	na	na	na	1
	2-Methylnaphthalene	110_1	120 ]	140	270	170 j	na	па		22 j	44 j	43 j	57 j	97 j	na	na	10 n
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Carbazole	59 j	310 j	54 j	230	220	na	na	83	84 j	860 uj	22 j	81 j	180 u	na	113	10 n
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Benzoic Acid	1100 u	1900 u	1200 u			na	na		360 uj	3300 j	5500	2500 j	2700 uj	na	na	0/6 0/6
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Benzyl Alcohol	430 u	I	21 j		0	na	na		500 u	I ;	81	95 u	180 u	na	na.	n 0/
nadate210u350u370u370u370u370u370u370u370u120unanananahalate83u140u77u370u370u370u370u370u370u370u120unanananaater440u1100<	Retene	200 j	540	330	430	na	na	na	150	230	460	230	460	пa	na	na	007
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Coprostanol	11a	na	na	na	na	na	na	5000	na	na	na	na	ца П	na	113	
	Dibenzofuran	200 j	420	270	460	270	. 113	na	240	28 j	62 j	2	[ 7]	[ <u>.</u>	118	na	n 2 2
	Diethylphthalate	83 u	140 u	87 u	74 u	170 uj	na	па	55 u	97 u	170 u	62	95 u	180 uj	na	រាន	n 0
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	Dimethylphthalate	83 u	140 u	87 u	74 u	33 j	na	na	55 u	97 u	170 u	200 u	95 u	180 u	na	na	n 0/
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Butylbenzylphthalate	210 u	350 u	87 u	74 u		na	na	55 u	250 u	430 u	n 62	95 u	180 uj	пa	na	n 0/
140 u $220$ uj $74$ u $840$ u na na $55$ u $97$ uj $170$ uj $190$ uj $95$ u $180$ u na na $1100$ u $850$ u $870$ uj $470$ uj $470$ uj na na $12j=Estimated concentrationna=Not analyzed$	Di-n-butylphthalate	88 u	140 u	14000 j	74 u	170 u	ЛA	na	82 u	97 u	1300 u	370 u	95 u	200	na	na	n 061
1100 u 850 u 870 uj 470 uj na na 19000 uj 530 u 1200 u 640 u 860 uj 1000 uj na na j=Estimated concentration na=Not analyzed	Di-n-octylphthalate	83 u			74 u	840 u	na	na	55 u	97 uj	170 uj	190 uj	95 u		ла	na	n 0/
	Bis(2EH)phthalate	440 u					na	na		530 u	1200 u	640 u		Ŧ	na	na	1200 u
	-=No sample										j=Estimate	d concentra	tion				
ui=Estimated detection limit	u=Not detected at detectio	n limit show	a								na=Not an	alyzed					
	uj=Estimated detection lin	uit															

		Number	Collection		SPM	BS		Dry	Gross	Accumulation
	Deployment	Days	Area		Percent		SPM		Accumulation	Rate
Station	Period	Deployed	cm2	Wet g	Solids	Solids	Dry g	g/cm3	g/cm2/year	cm/year
S-1A	9-11/91	63	78.5	142.8	44	57	63	0.89	4.6	5.2
-1B	ti-	63	78.5	144.9	56	57	81	0.89	6.0	6.7
-1A	11/91-2/92	90	78.5	176.1	63	57	111	0.89	5.7	6.4
-1B	*1	90	78.5	177.6	60	57	107	0.89	5.5	6.2
-1A	2-5/92	83	78.5	114.9	56	57	64	0.89	3.6	4.1
-1B		83	78.5	108.5	57	57	62	0.89	3.5	3.9
-1A	5-8/92	91	78.5	210.6	59	57	124	0.89	6.3	7.1
-1B	41	91	78.5	209.7	60	57	126	0.89	6.4	7.2
-1A	8-11/92	98	78.5	182.4	59	57	108	0.89	5.1	5.7
-1B	**	98	78.5	178.0	62	57	110	0.89	5.2	5.9
-1A	11/92-2/93	98	78.5	111.7	60	57	67	0.89	3.2	3.6
-1B	**	98	78.5	111.4	58	57	65	0.89	3.1	3.4
-1A	2-5/93	84	78.5	96.6	56	57	54	0.89	3.0	3.4
-1B	н	84	78.5	96.5	58	57	56	0.89	3.1	3.5
-1A	7-9/93	70	78.5	112.4	59	57	66	0.89	4.4	5.0
-1B	**	70	78.5	130.5	57	57	74	0.89	4.9	5.6
-1B	9-12/93	43	78.5	70.8	58	57	41	0.89	4.4	5.0
-1A	11/94-3/95	96	78.5	181.0	47	62	85	1.01	4.1	4.1
-1B	"	96	78.5	175.0	36	62	63	1.01	3.1	3.0
								mean=	4.5	5.0
S-2A	9-11/91	62	78.5	170.0	57	62	97	1.01	7.3	7.2
2A	11/91-2/92	92	78.5	126.3	61	62	77	1.01	3.9	3.8
-2B	н	92	78.5	131.7	62	62	82	1.01	4.1	4.1
-2A	2-5/92	83	78.5	93.8	55	62	52	1.01	2.9	2.9
-2B	<b>1</b> 5	83	78.5	97.8	. 57	62	56	1.01	3.1	3.1
2A	5-8/92	91	78.5	207.1	59	62	122	1.01	6.2	6.2
2B		91	78.5	200.5	59	62	118	1.01	6.0	6.0
-2A	8-11/92	98	78.5	226.6	65	62	147	1.01	7.0	<sub>,</sub> 6.9
-2B		98	78.5	235.2	63	62	148	1.01	7.0	6.9
-2A	11/92-2/93	98	78.5	47.0	60	62	28	1.01	1.3	1.3
-2B	47	98	78.5	49.7	63	62	31	1.01	1.5	1.5
2A	2-5/93	84	78.5	75.1	56	62	42	1.01	2.3	2.3
-2B	**	84	78.5	74.9	56	62	42	1.01		
	Settling Partic	ulate Matte						mean=	4.2	4.2

Table C8: Sediment Accumulation Rates for Sitcum Waterway from Sediment Trap Data, June 1991 to June 1995.

BS= Bottom Sediment

\*=Estimated from Puget Sound Sediment Density Model (Crecelius, 1989)

BS Dry Gross Accumulation Number Collection SPM Area SPM Percent Percent SPM Density\* Accumulation Rate Deployment Days Wet g g/cm2/year cm/year Period Solids Solids Dry g g/cm3 Station Deployed cm2 62 151 1.01 7.7 7.6 6-9/91 78.5 270.4 56 92 S-3A 8.1 50 62 1.01 8.2 163 11 92 78.5 326.3 --3B 140 1.01 7.27.190 78.5 215.4 65 62 -3A 11/91-2/92 7.3 7.2 ~3B н 90 78.5 218.6 65 62 142 1.01 83 78.5 146.4 61 62 89 1.01 5.0 4.9 2-5/92 --3A 62 90 1.01 5.0 5.0 H 83 78.5 147.6 61 -3B 6.4 6.4 59 62 126 1.01 5-8/92 91 78.5 213.6 -3A 6.2 6.3 123 1.01 \$1 91 78.5 211.5 58 62 -3B 5.0 62 106 1.01 5.1 97 78.5 173.7 61 -3A 11/92-2/93 5.2 97 78.5 175.4 63 62 111 1.01 5.3 -3B \*\* 78.5 102.5 58 62 59 1.01 3.3 3.2 84 -3Å 2-5/93 3.4 3.3 \*\* 78.5 103.4 59 62 61 1.01 84 -3B 7.4 7.3 62 118 1.01 78.5 197.4 60 74 -3A 7-9/93 7.3 7.4 45 195.9 60 62 118 1.01 74 78.5 -3B 7.6 1.01 7.739 78.5 107.1 60 62 64 -3A 9-12/93 7.0 62 60 1.01 7.1 --3B Ħ 39 78.5 110.3 54 8.3 12.8 78.5 289.0 59 46 171 0.64 11/94-3/95 96 -3A 6.6 6.4 mean= 5.4 5.6 73 0.96 78.5 133.2 55 60 S-4A 9-11/91 63 0.96 5.9 6.263 78.5 138.7 58 60 80 ... -4B 1.5 1.5 29 0.96 92 78.5 55.9 52 60 -4A 11/91-2/92 1.5 1.5 78.5 55.7 52 60 29 0.96 92 -4B 45 0.96 2.5 2.683 78.5 80.9 56 60 2-5/92 -4A 2.7 58 60 46 0.96 2.6 83 78.5 79.7 -4B 5.5 5.7 5-8/92 91 78.5 186.1 58 60 108 0.96 -4B 5.9 62 60 125 0.96 6.2 98 78.5 201.6 -4A 8-11/92 5.9 6.1 78.5 200.9 62 60 125 0.96 -4B £Ş 98 0.96 3.0 3.1 98 78.5 105.1 60 60 63 11/92-2/93 -4A 3.1 60 64 0.96 3.0 a 98 78.5 106.3 60 -4B 47 0.96 2.6 2.7 2-5/93 84 78.5 87.5 54 60 -4A

Table C8(cont): Sediment Accumulation Rates for Sitcum Waterway from Sediment Trap Data, June 1991 to June 1995.

SPM= Settling Particulate Matter

BS= Bottom Sediment

R.

7-9/93

41

9-12/93

14

ш

3/95-6/95

-4B

--4A

-4B

-4A

-4B

-4'A

-4B

\*=Estimated from Puget Sound Sediment Density Model (Crecelius, 1989)

78.5

78.5

78.5

78.5

78.5

78.5

78.5

84

70

70

43

43

98

98

using % solids data from in-situ bottom sediments. (dry density=wet density\*(BS% solids/100)

85.0

156.0

159.0

82.9

84.7

183.0

194.0

54

57

54

55

56

52

51

60

60

60

60

60

44

44

46

89

86

46

47

95

99

0.96

0.96

0.96

0.96

0.96

0.60

0.60

mean=

2.5

5.9

5.7

4.9

5.1

4.5

4.7

4.1

2.6

6.1

5.9

5.1

5.3

7.5

7.8

4.6

# Thea Foss Waterway

Table C9: Results of Conventionals and Metals Analysis of Settling Particulate Matter from Thea Foss Waterway,

Location				Hea	d near twin	Head near twin 96" Drains	IS				MICC	nanuci lica.	Midchannel near Uity Marina	IIIa
Station No.			·		TF-1	اسم						TF-1A	A	
Collection	11/92-	2/93-	7/93-	9/93	12/93-	3/94-	5/94-	8/94-		3/95	12/93-	3/94-	5/94-	8/94-
Period	2/93	5/93	9/93	12/93	3/94	5/94	8/94	11/94	3(7)	C6/0	3/34	5/94	0/74	11/54
Total Solids	43	42	38	43	43	33	37	35	42	27	41	41	40	38
TOC % Metals	6.8	6.7	7.6	11	12	10	7.6	11	9.6	9.6	7.6	5.8	5.5	5.6
Arsenic	Ла	па	ពង	па	22	22	па	na	'na	na	30	27	21	па
Cadmium	2.4	2.5	3.5	3.5	na	na	na	na	na	na	na	na	na	na
Copper	160	160	180	180	160	190	160	160	140	150	210	220	180	180
Mercury	0.80	0.65	0.70	0.60	0.63	0.58	0.64	0.56	0.47	0.59	0.86	1.0	1.0	0.94
Nickel	62	60	53	(2)	60	48	na	па	na	na	41	41	na	na
Lead	370	380	380	420	360	360	310	310	280	260	340	390	310	300
Zinc	470	490	350	510	490	460	340	430	310	260	360	350	260	270
04-44-4 M					TE-3									
Station No.	00111	0010		0,03	10,03	7-7	< 101 -	0/07	11/0/-	3/05-				
Collection	-7.6/11	-66/2	-66/1	-66/61	-66/21	-46/5	- 46/S	11/94	3/95	- CC/C				
rerioa	6617	061C	CC1C	rc 171										
Total Solids	43	38	4	40	38	35	ł	36	40	1				
TOC % Metals	5.5	6.8	5.4	5.9	6.2	5.8	1	5.4	5.9	I				
Arsenic	na	na	na	na	27	24	na	na	na	па				
Cadmium	7	1.9	2.9	£	na	na	na	na	na	na				
Copper	190	210	180	190	180	200	I	170	140	1				
Mercury	0.94	0.72	0.92	0.85	0.67	0.90	ł	1.0	0.72	1				
Nickel	45	41	38	39	39	37	na	na	na	na				
Lead	290	270	320	300	250	310	I	280	220	1				
7 inc	330	340	290	290	300	320	I	260	210	ł				

Table C9(cont.): Results of Conventionals and Metals Analysis of Settling Particulate Matter from Thea Foss Waterway, November 1992 to June 1995 (mg/kg, dry weight).

IIUIII IIICa LOSS Maici Way, AUVIIICU AVE War AVE And	11 000.T 1	מוכז אמץ ,			3					
Location				Mouth of	Mouth of Wheeler-Osgood Waterway	sgood Wa	terway			
Station No.					TF3	3				
Collection	11/92-	2/93-	7/93-	9/93-	12/93-	3/94-	5/94-	8/94-	11/94-	3/95-
Period	2/93	5/93	9/93	12/93	3/94	5/94	8/94	11/94	3/95	6/95
Total Solids	44	39	39	40	37	30	37	I	40	30
TOC %	5.1	6.5	5.4	6.3	6.4	5.5	5.0	I	5.3	6.1
Metals										
Arsenic	na	ពន	па	па	24	23	na	na	ла	na
Cadmium	1.9	1.9	2.8	2.9	па	na	na	na	na	na
Copper	270	310	200	220	210	240	180	1	160	210
Mercury	1.5	0.65	0.7	0.71	0.69	0.83	0.74	•	0.65	0.75
Nickel	43	45	36	40	41	38	na	na	na	na
Lead	250	210	240	260	220	240	190	I	170	180
Zinc	370	490	270	310	360	310	220	I	210	230
Location					Mouth	th				
Station No.					TF-4	4				*****
Collection	11/92-	2/93-	7/93-	9/93-	12/93-	3/94-	5/94-	8/94-	11/94-	3/95-
Period	2/93	5/93	9/93	12/93	3/94	5/94	8/94	11/94	3/95	6/95
Total Solids	43	39	39	42	37	30	31	33	38	19
TOC %	I	5.8	4.2	4.8	5.3	4.6	4.0	3.8	4.2	5.1
Metals										
Arsenic	na	na	na	па	22	22	na	na	Шâ	na
Cadmium	6.0	0.9	1.4	1.3	na	na	na	na	ла	na
Copper	140	120	140	150	150	150	120	120	67	110
Mercury	0.51	0.44	0.46	0.45	0.42	0.60	0.44	0.44	0.40	0.53
Nickel	39	40	27	31	35	29	นล	па	па	па
Lead	140	130	140	160	140	140	110	100	6	110
Zinc	210	200	180	210	210	210	140	130	120	130
-=No sample na=Not analyzed	zed				,					

Table C10: Summary of Semivolatile Organics and Butyltin\* Analysis of Settling Particulate Matter from Thea Foss Waterway, November 1992 to June 1995 (ug/kg, dry weight).

Location	Head near Twin 96" Drains								Mid-Channel						North o	f 15	5th Stre	et	Drain					
Station No.					TF-1							-17						TF-2						
-	11/92-		7/93-		2/93	4	5/94-	1	1/94–	-+	12/93-	~~~~	5/94-		11/92-		7/93-	12	2/93-	4	5/94	1	1/94	
Period	5/93		12/93		5/94		11/94	~	6/95		5/94		11/94		5/93		12/93		5/94		11/94		6/95	
TOC (%)	6.8		9.3		11		11		9.6	-	5.8		5.6	=	6.2		5.7		6.0		5.4		5.9	=
Semivolatiles																								
Acenaphthene	2000		400		390	j	270		380		220	j	220		260		270		200		220		190	
Acenaphthylene	440	i	310	i	210	j	210		240		230	j	290	ĺ	190		250		320		290		230	
Naphthalene	1600	-	800	•	510		580		690	Ì	560	j	820		380		710		560		830		530	u
Fluorene	1300	•	600		610	u	370		430		280	j	290		290		330		340		370		310	
Anthracene	2000		1300		1100		830		1000		640		700		320		700		870		860		780	
Phenanthrene	9300		6600		7800		3700		4000		1900		1600		2300	j	1900		1700		1700		2300	
Sum LPAH	17000	j	10000	j	1000	j	6000		6700		3800	j	3900		3700	j	4200		4000		4300		3800	
Fluoranthene	15000		12000		13000		7500		7200		3100		2900		4400		3100	•	3400		3200		4200	
Benzo(a)anthracene	6600		3700		5500		2500		2400		1500		1200		2500	j	1300		1500		1300		1500	
Chrysene	6600		6400		7000		4400		4000		2300		2000		2400	j	2000		1900		2300		2500	
Pyrene	9000		11000		9300		6900		6100		2300		4100		3000	j	4100		2300		4000		3900	
Benzofluoranthenes	12000		11000		13000		7700		7100		4000		2000		3700	j	3800		3200		1900		4000	
Benzo(a)pyrene	3400		4600		4100		3200		3000		2000		1800		850		1700		950		1800		1500	
Dibenzo(a,h)anthracene	940	j	930		960		540		510		350		300		240		320		260		340		120	u
Indeno(1,2,3-cd)pyrene	29000		4100		13000		2800		2600		4200		1500		770		1300		3700		1500		1400	
Benzo(g,h,i)perylene	1600	j	4000		1400		2800		2600		1700		1400		240		1400		180		1500		1300	
Sum HPAH	84000	j	58000		67000		38000		36000		21000		17000		18000	j	19000		17000		18000		20000	
Total PAH	100000	j	68000	j	77000	j	44000		42000		25000	j	21000		22000	j	23000		21000		22000		24000	
Phenol	370	j	350	u	190	j	100	j	210	u	180		350		160	u	150	•	130		130		140	
Pentachlorophenol	2600	u	2600	uj	230	j	720	uj	630	u	3500	u	1100	u	800	u	1100	u	1200		1300	u	610	
4-Methylphenol	510	u	270	j	170		170		310		93		2700		160	u	670		85	j	270		120	u
1-Methylnaphthalene	1200	j	-				-		-		-		-		200						-		-	
2-Methylnaphthalene	700	j	530		460	j	360		410	u	250	-	360		190		340		310		420		280	u
Carbazole	1700		1200		1300		640		670		280	-	190		320		170	-	210		180	_	350	
Benzoic Acid	9600		4000	uj	1100	j	4800	uj			1300	j	1800	uj		•		•		j	2200	-	420	
Benzyl Alcohol	170		350	u	450	u	130	u	130	u	350	u	110	u	160	j	350	u	120	u	130	u	120	u
Retene	1000	j	na		730				520		860		-		400				530		-		440	
Dibenzofuran	430	j	390		370	j	290		260		190	j	260		250		290		220		270		210	
Coprostanol	na		na		na		na		12000		na		na		na		na		na		na		120	
Diethyl phthalate	510	u	260	uj	500	u	100		130		380	•			-		220	uj		-		-	53	
Dimethyl phthalate	510	u	350		450	u	90	•	130	u	350		110			u	350		120		57	-	500	
Butyl benzyl phthalate	2500		2100	uj	2000		1300	j	1100		650	~	560		790		590		490	j	650	j	1300	
Di-n-butyl phthalate	490		630	uj		-	1600		710		230				-		350	~			2900		360	
Di-n-octyl phthalate	8200		1900				1100		380		350	•		u			220			u	660	u	7000	
Bis(2EH)phthalate	26000		21000	uj	2900	uj	16000		14000	u	94000		7400		96000		13000	uj	13000		13000		4800	
Butyltins*																							**	
Monobutyltin	470	~		u		uj			29	u					na		70		19	uj				u
Dibutyltin	240				310		360		150		370		420		na	•	140	u	86		280			u
Tributyltin	710	j	600		480		700		310		650		790		na		580		490		550		210	
Tetrabutyltin	240	u	i 68	u	13	uj	13	u	30	u	92	u	· · · · · · · · · · · · · · · · · · ·				73	u	140	u	14	u	23	u
nor Not analyzed		<b>`</b>				°							i- Fet		ated con	Cet	tration							

na= Not analyzed

-= No sample

u= Not detected at detection limit shown

j= Estimated concentration

uj= Estimated detection limit

\*= Reported as chloride
Mouth near D-Street Tank Farms Mouth of Wheeler-Osgood Waterway Location TF-4 TF-3 Station No. 11/94 -7/93-12/93 -5/94-11/94-11/92 -7/93-12/93-5/94-**Collection Period** 11/92-11/94 6/95 6/95 5/93 12/93 5/94 11/94 12/93 5/94 5/93 Period 5.7 4.6 4.5 5.0 3.8 4.7 5.0 5.8 5.9 6.0 TOC (%) Semivolatiles 150 j 220 ј 270 j 150 j 270160 j 310 210 280 460 Acenaphthene 220 ј 190 240 430 190 j 170 j 240 i 320 290 230 Acenaphthylene 440 450 u 730 200 j 460 550 570 710 390 u 410 Naphthalene 450 210 340 190 j 300 380 230320 320 820 Fluorene 950 470 620 570 390 750 670 2100 750 550 u Anthracene 1100 1700 1200 1500 2100 1700 1500 1700 2900 1800 Phenanthrene 3200 4900 2600 3700 3800 7200 4100 2100 i 2300 3300 Sum LPAH 2500 3600 1900 2900 2700 2000 3600 2800 3000 Fluoranthene 3100 1100 u 790 1200 950 u 720 1000 1400 uj 1200 1300 1200 Benzo(a)anthracene 1300 1800 1800 1700 1000 1700 2200 1900 1800 1900 Chrysene 3100 200 u 2900 3100 2200 3500 2600 3800 2500 1300 Pyrene 3100 2900 3800 2300 1700 2700 1500 2900 2500 3100 Benzofluoranthenes 210 u 1300 1300 1100 1100 u 1600 950 u 1600 900 Benzo(a)pyrene 130 u 230 290 ј 170 250 j 230 310 210 u 210 u 240 j 200 Dibenzo(a,h)anthracene 970 1100 800 1000 210 u 990 1100 3800 1100 Indeno(1,2,3-cd)pyrene 300 990 540 770 210 u 950 1200 940 130 u 1200 370 Benzo(g,h,i)perylene 14000 j 17000 j 11000 13000 5200 18000 15000 12000 11000 17000 j Sum HPAH 16000 17000 22000 14000 j 19000 14000 i 7600 j 15000 21000 25000 1 i ì Total PAH 160 u 270 u 210 u 210 270u 340 j 130 u. 260 u 210 120 u u Phenol 3400 u 1600 u 600 u 1100 u 1000 u 2700 uj 1800 u 1200 u 2600 u Pentachlorophenol 130 u 87 i 120 u 110 j 85 j 210 u 210 u 77 260 j 650 4-Methylphenol 130 u 96 j ..... ..... 1-Methylnaphthalene 200 280 u 100 j 270 i 410 250 490 380 230 u 200 300 2-Methylnaphthalene 340 u 91 j 120 u 150 j 270 u 150 210 u 130 260 u 590 Carbazole ù 4300 j 5400 7000 ui 2700 uj 3000 j 420 uj 5200 j 6700 5300 uj 2000 j Benzoic Acid 340 u 210 210 u 270 u 160 u 120 j 100 j 180 u 120 u u 130 uj Benzyl Alcohol 1100 400 350 610 420 Retene 400 \_ 220 u 160 250 430 300 210 u 150 j 220 ј 310 j 190 Dibenzofuran 6500 8500 u na na na na na na Coprostanol na na 270 uj 560 uj 160 uj 120 u 260 180 u 120 u 210 u 210 u 130 u uj Diethyl phthalate 160 u 85 j 340 u 210 u 80 j 130 u 260 u 180 u 120 u 210 u Dimethyl phthalate 1700 160 j 120 u 250 340 uj 520uj 890 u 590 380 450 Butyl benzyl phthalate 450 730 u 270 uj 270000 j 180 u 360 780 u 210 н 580 260 uj Di-n-butyl phthalate u 1400 340 u 790 u 120 u 210 u 210 u 180 u 590 u Di-n-octyl phthalate 130 u 260 u 4800 u 210 u 24000 ui 9100 uj 2300 uj 4600 u 7300 uj 590 9400 5200 uj Bis(2EH)phthalate Butyltins\* 150 j 70 u 44 uj 65 13 33 u 550 280 26 uj u na uj Monobutyltin 190 j 100 j 140 u 54 89 130 u 52 u 200 na 270 ui Dibutyltin 420 j 210 240 j 330 680 520 j 540 j 260 na 30 j Tributyltin 49 u 16 u 62 j 270 uj 27 u 14 u 17 u 70 u 67 u na Tetrabutyltin

Table C10(cont.): Summary of Semivolatile Organics and Butyltin\* Analysis of Settling Particulate Matter from Thea Foss Waterway collected November 1992 to June 1995 (ug/kg, dry weight).

na= Not analyzed

-= No sample

u= Not detected at detection limit shown

j= Estimated concentration

uj= Estimated detection limit

\*= Reported as chloride

Station Collection Period	TF-1 11/94-6/95	TF-2 11/94-6/95	TF-3 11/94-3/95	TF-3 3-6/95	TF-4 11/94-6/95
1242	130u	120u	210u	120u	180u
1254	320	310	220	310	120
1260	260	300	230	260	160
Total	580	610	450	570	280

Table C11: Summary of PCB analysis of settling particulate matter from Thea Foss Waterway, November 1994 to June 1995 (ug/kg, dry weight).

γ.

 $\overline{u}$  = Not detected at detection limit shown

	· · · ·	Number	Collection		SPM	BS		Dry	Gross	
Station	Deployment Period	Days Deployed	Area cm2	SPM Wet grms	Percent Solids	Percent Solids	SPM Dry g	Density* g/cm3	Accumulation g/cm2/year	Rate cm/year
		99	78.5	40.6	43	39	17	0.51	0.8	1.6
-1B	*	99	.78.5	41.2	43	39	18	0.51	0.8	1.6
-1A	2-5/93	82	78.5	80.2	42	39	34	0.51	1.9	3.7
-1B	ч	82	78.5	73.7	42	39	31	0.51	1.8	3.5
-1A	7-9/93	70	78.5	45.5	38	39	17	0.51	1.1	2.3
-1B	**	70	78.5	47.6	38	39	18	0.51	1.2	2.4
-1A	9-12/93	. 77	78.5	46.7	42	39	20	0.51	1.2	2.3
-1B	υ	77	78.5	44.2	43	39	19	0.51	1.1	2.2
-1A	12/93-3/94	106	78.5	83.7	43	39	36	0.51	1.6	3.1
-1B	n	106	78.5	85.3	43	39	37	0.51	1.6	3.1
-1A	35/94	55	78.5	50.4	32	- 39	16	0.51	1.3	2.6
-1B	*)	55	78.5	50.4	34	39	17	0.51	1.4	2.8
-1A	5-8/94	92	78.5	100.0	37	39	37	0.51	1.8	3.6
-1B		92	78.5	99.0	37	. 39	37	0.51	1.8	3.6
-1A	8-11/94	98	78.5	75.0	34	39	26	0.51	1.2	2.4
-1B	*)	98	78.5	76.0	35	39	26	0.51	. 1.2	2.4
-1A	11/94-3/95	97	78.5	95.0	42	39	40	0.51	. 1.9	3.7
-1B	v	97	78.5	96.0	42	39	41	0.51	1.9	3.8
-1A	3/95-6/95	98	78.5		28	39	81	0.51	3.9	7.6
-1B		. 98	78.5		27	39	93	0.51	4.4	8.7
								mean=	1.7	3.4
TF1A-	1 12/93-3/94	106	78.5	55.6	41	41	23	0.55	1.0	1.8
-1B	EF.	106	78.5		41	41	22	0.55	1.0	1.8
	3-5/94	55	78.5		41	41	37	0.55	3.1	5.7
-2B	41	55	78.5		40	41	36	0.55	3.1	5.6
	58/94	92	78.5		41	41	86	0.55	4.4	8.0
2B		92	78.5		<b>4</b> 1	41	85	0.55	4.3	7.8
	8-11/94	- 98	78.5		39	41	51	0.55	2.4	4.4
-2B	\$3	98	78.5		38	41	51	0.55	. 2.4	4.4
								mean=	2.7	4.9
TF-2A	11/92-2/93	99	78.5	33.8	43	46	15	0.64	0.7	1.1
-2B	18	99	78.5	33.8	43	46	14	0.64	0.7	1.1
-2A	2-5/93	82	78.5		37	46	14	0.64	0.8	1.3
-2B		82	78.5	39.8	38	46	15	0.64	0.9	. 1.3
-2A	7-9/93	70			45	46	32	0.64	2.1	3.3
-2B	a	70			42	46	30	0.64	2.0	3.1
-2A	9-12/93	77			41	46	19	0.64	1.2	1.8
-2B	#1	77			40	46	18	0.64	1.1	1.7
-2A	12/93-3/94	106			39	46	15		0.7	
-2B		106			38	46			0.7	
-2A	35/94	55			36	46			1.7	
-2B	61 E F F	55			34	46			1.6	
-2A	8-11/94	98			36	46			2.0	
-2B	"	98			37	46			2.1	
-2A	11/94-3/95	97			39	46			1.4	
-2B	4	97			40	46			1.4	
	Settling Partic							mean=	1.3	

Table C12: Sediment Accumulation Rates for Thea Foss Waterway from Sediment Trap Data, November 1992 to June 1995.

BS= Bottom Sediment

\*=Estimated from Puget Sound Sediment Density Model (Crecelius, 1989)

using % solids data from in-situ bottom sediments. (dry density=wet density\*(BS% solids/100)

Table C12(cont.): Sediment Accumulation Rates for Thea Foss Waterway from Sediment trap data, November 1992 – June 1995.

		Number	Collection		SPM	BS		Dry	Gross	Accumulation
	Deployment	Days	Area	SPM	Percent	Percent		Density*	Accumulation	Rate
Station		Deployed		Wet grms	Solids	Solids	Dry g	g/cm3	g/cm2/year	(cm/year)
TF-3A	11/92-2/93	99	78.5	41.5	43	40	18	0.53	0.8	1.6
-3B		. 99	78.5	41.4	45	40	19	0.53	0.9	1.6
-3A	2-5/93	82	78.5	38.5	40	40	15	0.53	0.9	1.6
-3B	6	82	78.5	38.3	38	40	15	0.53	0.8	1.6
-3A	7-9/93	70	78.5	76.2	41	40	31	0.53	2.1	3.9
-3B	H	70	78.5	73.6	39	40	29	0.53	1.9	3.6
-3A	9-12/93	77	78.5	51.4	39	40	20	0.53	1.2	2.3
3B	*1	77	78.5	51.9	40	40	21	0.53	1.3	2.4
-3A	12/93-3/94	106	78.5	33.4	.38	40	13	0.53	0.6	1.0
-3B		106	78.5	34.1	36	40	12	0.53	0.5	1.0
-3A	3-5/94	55	78.5	49.2	29	40	14	0.53	1.2	2.3
-3B	41	55	78.5	44.7	31	40	14	0.53	1.2	2.2
-3A	5-8/94	92	78.5	141.0	37	40	52	0.53	2.6	5.0
-3B		92	78.5	- 133.0	37	40	49	0.53	2.5	4,6
-3A	11/94-3/95	97	78.5	53.0	40	. 40	21	0.53	1.0	1.9
3B	12	97	78.5	54.0	40	40	22	0.53	1.0	2.0
-3A	3/95-6/95	98	78.5	89.0	31	40	27	0.53	1.3	2.5
-3B	65	. 98	78.5	91.0	30	40	27	0.53	1.3	2.5
							•	mean=	1.3	
TF-4A	11/92-2/93	99	78.5	23.7	43	-64	10	1.07	0.5	
-4B	16	99	78.5	26.5	42	64	11	1.07	0.5	0.5
-4A	2-5/93	82	78.5	30.5	39	64	· 12	1.07	0.7	0.6
-4B	. "	82	78.5	29.7	38	64	11	1.07	0.6	0.6
-4A	79/93	69	78.5	57.9	- 38	64	22	1.07	1.5	1.4
-4B	H	69	78.5	56.9	39	64	22	1.07	1.5	1.4
-4A	9-12/93	77	78.5	39.5	41	64	16	1.07	-1.0	0.9
-4B		77	78.5	39.3	42	64	17	1.07	1.0	0.9
-4A	12/93-3/94	106	78.5	28.8	38	64	11	1.07	0.5	0.5
-4B	н,	106	78.5	31.3	37	64	11	1.07	0.5	0.5
-4A	35/94	55	78.5	37.9	30	64	11	1.07	1.0	0.9
-4B	41	55	78.5	39.9	30	64	12	1.07	1.0	0.9
-4A	5-8/94	92	78.5	108.0	30	64	32	1.07	1.6	1.5
-4B		92			32	64	34	1.07	1.7	1.6
-4A	8-11/94	98			34	64	27	1.07	1.3	1.2
-4B	68	98			31	64	25	1.07	1.2	. 1.1
-4A	11/94-3/95	97			38	64	. 15		0.7	0.7
-4A	3/95-6/95	98			16	64	12		0.6	
4B	11	98			23	64	15		0.7	
	Settling Partic							mean≔	0.95	

SPM= Settling Particulate Matter

BS= Bottom Sediment

\*=Estimated from Puget Sound Sediment Density Model (Crecelius, 1989)

using % solids data from in-situ bottom sediments. (dry density=wet density\*(BS% solids/100)

### Appendix D

#### Physical/Chemical Analysis of Bottom Sediments (Dry Weight) December 1989, January 1991, and November 1994

Hylebos Waterway Table D1: Conventionals and Metals Analysis Table D2: Volatiles, Semivolatiles, and PCB Analysis Table D3: Butyltins Analysis

Sitcum Waterway Table D4: Conventionals and Metals Analysis Table D5: Semivolatiles Analysis

Thea Foss Waterway Table D6: Conventionals and Metals Analysis Table D7: Semivolatiles Analysis Table D8: Butyltins Analysis

# Hylebos Waterway

Table D1: Results of Conventionals and Metals Analysis of Bottom Sediments from Hylebos Waterway, January 1991 and November 1994.

Location	Uppe	Upper Turning	g	@ General	neral	@ Elf	f	Near Lincoln	lcoln			@ Occidental	ental	r	
		Basin	,	Metals	als	Atochem	an a	Drain		Near Cenex	nex	Chemical	al	Mouth	T
Station No.	÷	Ш-1		H-2	5	H-3		H4		H-5		H6		H-7	
Collection Date	1/01	(Ren)	11/94	1/91	11/94	1/91	11/94	1/91	11/94	1/91	11/94	1/91	11/94	1/91	11/94
Depth (ft) @ MLLW		(Jan )		32				22		25		30		31	
Total Solids (%)	47	45	37	41	44	41	41	46	56	49	53	52	50	64	59
TOC (%)	4,4	4.6	4.9	5.2	3.4	4.3	3.6	4.0	3.1	5.3	12	2.7	2.1	1.8	1.7
Grain Size (%)															
Gravel (>2mm)	7	0	6	0	4	0		5		16	31	0	0	0	*1
Sand (2mm-62um)	26	61	10	28	27	22	23	45	61	38	52	32	21	50	42
Silt (62um-4um)	46	60	62	39	48	45	\$	31	28	26	35	42	58	44	42
Clay (<4um)	27	21	27	33	21	33	22	22	10	20	6	26	21	16.	15
Metals (mg/kg, dry)															
Antimony	9.0	10	na	42	na	14	na	17	na	9.0	na	3.9	na	2.2 u	na
Arsenic	62	56	38	86	30	49	35	52	20	37	16	18	14	15	11
Cadmium	0.7	0.9	0.89 j	1.5	0.7 j	0.8	1.0 j	0.7	0.4 j	0.6	0.3 u	0.4	na	0.3	0.3 u
Copper	93	120	140	220	120	150	130	150	98	120	63	120	100	98	72
Mercury	0.22	0.26	0.35	0.46	0.43	0.45	0.45	0.30	0.19	0.28	0.22	0.23	0.25	0.16	0.14
Lead	55	69	63	160	67	76	88	92	48	84	53	100	100	43	39
Nickel	44	47	160	53	69	49	96	42	110	36	130	40	160	26	06
Zinc	190	230	230	540	200	210	180	200	110	160	95	140	110	120	67
u=Not detected at detection limit shown	ion limit	shown													

j=Estimated concentration na=Not analyzed (Rep)=Replicate sample Table D2: Summary of Volatiles, Semivolatiles and PCBs Detected in Bottom Sediments from Hylebos Waterway, January 1991 and November 1994 (ug/kg, dry weight).

Location	l	Jppe	r Turning Basin			@ Gen Met				@ Elf .tochem		Linc Dra	
Station No.			H-1			H-		ļ	2	H-3		H-	
Collection Date Depth @ MLLW (ft)	1/91 23		Rep	11/94	<b>†</b>	1/91 28	11/94	Ī	1/91 31	Rep	11/94	1/91 23	11/94
TOC (%)	4.4		4.6	4.9		5.2	3.4		4.2	4.3	3.6	4.0	3.1
Volatiles													÷
Acetone	na		na	na	ł	na	na	Į	na	na	na	na	na
Carbon Disulfide	na		na	na		na	na		na	na	na	na	na
Cis 1,2-Dichloroethene	na		na	na		na	na		na	na	na	na	na
Trichloroethene	na		na	na	- I-	na	na		na	na	na	na na	na na
Tetrachloroethene	na		na	na		na	na na		na na	na na	na na	na	na
Chloroform	na		na na	na na	ļ	na na	na		na	na	na	na	na
Isopropyltoluene Semivolatiles	na		na	na		na	1162		1144	int	Inte	ma	
Acenaphthene	32	;	80 u	35 j	.	94 u	70. j		29 ј	na	140 j	31 j	32 j
Acenaphthylene		i	33 j	64 j		36 j	59 j		23 j	na	86 j	23 j	36 j
Naphthalene	80	*	80 u	63 j	· I	94 u	140 j		50 j	na	140 j	42 i	110 j
Fluorene		i	27 j	250 i		40 j	96 j		54	na	160 j	48	43 j
Anthracene	170	J	170	290 j		240	300 <sup>°</sup>	l	170	na	570 <sup>~</sup>	130	130 j
Phenanthrene	230		180	310	1	350	430		320	na	1200	330	210
Sum LPAH	510	j	410 j	1000 j	i	670 j	1100 j		650 j	na	2300 ј	600 j	560 j
Fluoranthene	790		930	1500		1300	1600	-	1200	na	2300	750	600
Benzo(a)anthracene	340		400	660	ļ	660	770		580	na	1200	390	280
Chrysene	730		820	1700		1800	2000		1500 .	na	2700	810	690
Pyrene	550		740	1300		1400	1800		1400	na	3200	680	900
Benzofluoranthenes	870		1100	2800		2300	3000		2100	na	4000	1100	1100
Benzo(a)pyrene	170		210	780		400	900		390	na	1400	210	330
Dibenzo(a,h)anthracene	80	u	58	260 1	uj	130 j	320		140	na	370	79	150 uj
Indeno(1,2,3-cd)pyrene	120		190	810		300	900		310	na	1300	210	350
Benzo(g,h,i)perylene	61	u	<u>80 u</u>	730	_	47 j	880		<u>59 j</u>	na	1200	33	350 4800
Sum HPAH	3600		4400	11000	.	8300 j	12000	1	7700 j	na	18000 20000 j	4300 4900 j	
Total PAH	4100	•	4800 j 120	12000 j 68		9000 j 190 u	<u>13000 j</u> 300		8400 j 44 j	na na	20000 j 200 u	4900 j 33 j	<u> </u>
Phenol		j u	120 400 u	68 j 2500 i		470 u	2000 u		230 u	na	200 u	210 u	
Pentachlorophenol 2-Methylnaphthalene			400 u 80 u	2300 51		470 u 94 u			230 u 33 j	na	2000 u 86 j	210 u 23 j	41 j
Carbazole	na	u	na	120	)	na na	140 j		na	na	150 j	na	150 u
Hexachloroethane	120	21	160 u	250	,,	190 u	200 u	1	48	na	200 u	80 u	
Hexachlorobenzene	61		80 u		u	94 u		- i	34 j	na	200 u	41 u	
Hexachlorobutadiene	120	ŭ	160 u		u	190 u	200 u		93 u	na	31 j	82 u	150 u
Benzoic Acid	610		800 u	1700	_	940 u			210 ј	na	2000 uj	1	
Retene	na		na	200	uj	па	່280 <sup>°</sup>		na	na	280	na	220
Dibenzofuran	80	u	80 u	46	j	94 u	89 <del>j</del>		37 ј	na	89 j	35 j	- 39 j
Diethylphthalate	41	u		0.00	j	94 u	200 u	ij	46 u	na	200 uj	41 u	150 u
Dimethylphthalate	27	j	80 u	140	uj	94 u			39 j	na	200 u	34 j	150 u
Butylbenzylphthalate	8Ò	u	80 u	710	j	100	200 u	ij	46 u	na	200 uj	1 V	770 u
Di-n-Butylphthalate	61		80 u		uj	94 u		~ }	27 ј	na	2200	34 j	290 u
Di-n-Octylphthalate	41	u	80 u		uj	94 u		ij	46 u	na	980 uj		
Bis(2EH)phthalate	650		790		uj	1200	1500		820	na	760 uj	1	640
Isophorone	41	u	80 u		uj	94 u			46 u	na	17 j	41 u	
Coprostanol	na		na		j	na	2300 j		na	na	2300 j	na	690 j
N-Nitrodiphenylamine	41		80 u	250	÷	94 u			46 u	na	200 u	41 u	
1,4–Dichlorobenzene 1,2,4–Trichlorobenzene	41 41	u u	80 и 80 и	250 250		94 u 94 u		*	46 u 46 u	na na	200 u 200 u	41 u 41 u	
PCB													
1242	85	u	na	120	u	96 v	120 j	.	100 u	100 u	ı 150 j	85 u	ı 77 j
1254	220		na	300		790	680 j		1100	1700	2000 j	660	440 j
1260	83	u	na	220		96 i			460	1300	860 j	420	440 j
													960 j

-=No sample

(Rep)=Replicate sample

na=Not analyzed

j=Estimated concentration

Table D2(cont): Summary of Volatiles, Semivolatiles and PCBs Detected in Bottom Sediments from Hylebos Waterway, January 1991 and November 1994 (ug/kg, dry weight).

Near Cenex         Chemical         Mouth           Station No.         H=5         H=6         H=7           Collection Date         1/91         11/94         1/91         11/94         1/91         11/94           Depth @ MLLW (ħ)         19         1         1/91         11/94         1/91         11/94         1/91         11/94           TOC (\$\$)         5.3         12         2.6         2.8         2.1         1.8         1.7         1/9         2.0 uj           Carbon Disulfide         90 j         3.9 uj         1.5 uj         2 j         2.1 1.5 j         1.1 u         0.6 j         1.8 u           Trichioroethene         0.8 j         3.5 uj         2 j         1.5 j.1 uj         1.4 u         1.8 u           Isomivolatiles         2         2.0         2.8         2.1         2.5         1.5 j.1 uj         1.8 u           Acenaphtylene         47 j         1.1 j         2.6 j         5.1 uj         1.8 u         1.8 u           Semivolatiles         320         150 j         141 j         na         65 j         171 j         54 j           Acenaphtylene         120 j         2.00 j         700 na         120 j         26 j	Location		Ī	@(	Occidental	T		·
Station No. $H-5$ $H-6$ $H-7$ Collection Date         1/91         1/91         Rep         11/94         1/91         31          31           TOC (%)         5.3         12         2.6         2.8         2.1         1.8         1.7           Volatiles         90         j         39         uj         1         j         1         j         6.6         j         1.8         1.7           Volatiles         90         j         39         uj         15         uj         2         uj         49         uj         1.7         uj         20         0.6         j         1.8         u         1         u         1.8         u         1         u         1.8         u         1.8         u         1.0         1.0         1.0         1.0         1.0         1.0         1.0         1.0		Near Ce	nex				Mouti	ı
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Station No.				H6		H-7	
$\begin{array}{ c c c c c c c c c c c c c c c c c c c$	Collection Date	1/91	11/94	1/91	Rep	11/94		11/94
VolatilesPointPointPointPointPointPointAcetone90j39uj15uj2uj49uj17uj20ujCarbon Disulfide1j3.5uj223.11u0.57j11u1.8ujTrichloroethene0.8j3.5u2j1.5j1.4u1.8ujChloroform2u3.5uj2j0.8j1.1u1.8ujSentivolatiles2u582u2u4.4uj1u1.8ujSentivolatiles320150j41jna65j1.7j54jAceaaphtaylene470j100j26jna177j20j50jHuanathrene19001000410na64029070360j1200jSum LPAH3600j23002400900na1100430810jBenzofuyanthracene1300900410na530170360jPyrene1700200640na950220j70jBenzofuyanthracene160190ujna1600100100430830j <t< td=""><td>Depth @ MLLW (ft)</td><td>19</td><td></td><td>31</td><td></td><td></td><td>31</td><td></td></t<>	Depth @ MLLW (ft)	19		31			31	
	TOC (%)	5.3	12	2.6	2.8	2.1	1.8	1.7
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$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Acetone	90 j	<b>u</b>		•	¥	•	•
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$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Cis 1,2-Dichloroethene	~						
$\begin{array}{c} \begin{tabular}{ c c c c c c c c c c c c c c c c c c c$								
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Semivolatiles Acenaphthene 320 150 j 41 j na 65 j 17 j 54 j Acenaphthene 360 230 70 na 120 j 26 j 150 j Fluorene 360 230 70 na 120 j 26 j 150 j Phenanthrene 120 340 61 na 170 26 j 120 j Anthracene 350 420 200 na 360 97 360 j Phenanthrene 1900 1000 410 na 640 J 480 j 1200 j Fluoranthene 1900 2300 j 810 j na 1400 J 480 j 1200 j Fluoranthene 1300 900 410 na 530 170 360 j Chrysene 1800 1600 730 na 1100 300 540 j Pyrene 1700 2000 640 na 950 220 j 770 j Benzo(a)nthracene 160 190 uj Phenzo(a),anthracene 160 190 uj Phenzo(a),anthracene 160 190 uj Toda PAH 9800 j 10000 4200 na 1200 380 830 j Dibenzo(a,h)arthracene 160 190 uj Toda PAH 9800 j 10000 4200 na 100 300 240 j Benzo(g,h,h)arthracene 160 190 uj Toda PAH 9800 j 10000 4200 j na 6200 180 j 4100 j Benzo(g,h,h)perylene 68 j 400 41 j na 300 30 u 210 j Enzo(g,h,h)perylene 68 j 400 41 j na 6200 j 53 0 j 5300 j Toda PAH 9800 j 10000 4200 j na 6200 1800 j 4100 j Sum HPAH 9800 j 10000 4200 j na 6200 1800 j 4100 j Portachlorophenol 180 u 1900 u 89 j na 1600 u 150 u 540 uj Pentachlorophenol 180 u 1900 u 87 j na 160 u 17 j 54 uj Pentachlorophenol 73 u 32 j 46 j na 83 j 61 u 32 j Benzo(a) j 190 u 32 j 46 j na 83 j 61 u 32 j Benzo(a) j 190 u 32 u 39 j Hexachlorobtarane 37 u 190 u 62 na 160 u 61 u 54 uj Hexachlorobtarane 37 u 190 u 38 u na 160 u 30 u 270 uj Benzo(brohene 37 u 190 u 38 u na 160 u 30 u 540 uj Dibenzofuran 180 300 159 na 760 j 2300 j 300 u 39 j Hexachlorobtarane 37 u 190 u 38 u na 160 u 30 u 54 uj Dibenzofuran 180 300 u 190 u 38 u na 160 u 30 u 54 uj Dibenzofuran 180 300 u 190 u 38 u na 160 u 30 u 54 uj Dibenzofuran 180 300 u 190 u 38 u na 54 uj Dibenzofuran 180 300 u 190 u 38 u na 54 uj Dibenzofuran 180 u 300 u 54 uj Dihenzofuran 180 300 u 59 na 73 j 23 j 87 j Dibenzofuran 180 300 u 270 uj Baz 160 u 30 u 54 uj Di-n-Octylphthalate 37 u 190 u 38 u na 610 u 30 u 54 uj Di-n-Octylphthalate 37 u 190 u 38 u na 56 j 30 u 54 uj Di-n-Octylphthalate 37 u 190 u 38 u na 56 j 30 u 54 uj Di-n-Octylphthalate 37 u 190 u 38 u na 56 j 30				~				
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Anthracene $850$ $420$ $200$ na $360$ $97$ $360$ jPhenanthrene $1900$ $1000$ $410$ na $640$ $290$ $470$ jSum LPAH $3600$ $j$ $2300$ $j$ $810$ jna $1400$ j $480$ j $1200$ jFluoranthene $2300$ $2400$ $900$ na $1100$ $430$ $810$ jBenzo(a)anthracene $1300$ $900$ $410$ na $530$ $170$ $360$ jChrysene $1700$ $2000$ $640$ na $950$ $220$ $j$ $770$ jBenzo(a)pyrene $1700$ $2200$ na $1200$ $380$ $830$ jBenzo(a,h)anthracene $160$ $190$ $uj$ $75$ na $160$ $100$ $330$ $j$ Dienzo(a,h,h)erpriene $68$ $j$ $400$ $110$ $200$ $j$ $sac$ $j$ $j$ Indeno(1,2,3-cd)pyrene $66$ $j$ $4200$ $j$ $na$ $600$ $1100$ $j$ $j$ Sum HPAH $9800$ $j$ $10000$ $4200$ $j$ $na$ $600$ $i$ $170$ $j$ $54$ $uj$ PhenolPhenol $10000$ $i$ $4200$ $j$ $na$ $66$ $i$ $na$ $i$ $i00$ $j$ Phenol $1000$ $i$ $1900$ $i$ $72$ $na$ $n60$ $i$ $17$ $j$ $54$ $uj$ <t< td=""><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td></t<>								
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Phenol100j100j72jna160i17j54ujPentachlorophenol180u1900u89jna1600u150u540uj2-Methylnaphthalene19029045na98j12j81jCarbazolena150jnana66jna100jHexachlorobtane73u190u62na160u61u54ujHexachlorobutadiene73u190u87na94j30u39jHexachlorobutadiene73u32j46jna83j61u32jBenzoic Acid230j1900uj200jna1600uj300u190ujRetenena8900nana1600u30u54ujDibenzofuran18030059na93.j23j87jDiethylphthalate37u1900uj38una160u30u54ujDimethylphthalate37u190uj38una160u30u54ujDi-n-Octylphthalate37u190uj38un	Sum HPAH	•		; v	na			
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Hexachloroethane73 u190 u62na160 u61 u54 ujHexachlorobenzene37 u190 u87na94 j30 u39 jHexachlorobutadiene73 u32 j46 jna83 j61 u32 jBenzoic Acid230 j1900 uj200 jna1600 uj300 u190 ujRetenena8900nana34030 u54 uDibenzofuran18030059na93 j23 j87 jDiethylphthalate37 u1900 uj38 una160 u30 u54 ujButylbenzylphthalate37 u190 uj38 una160 u30 u54 ujDi-m-Butylphthalate37 u190 uj38 una160 uj30 u54 ujDi-m-Octylphthalate37 u190 uj38 una160 uj30 u54 ujBis(2EH)phthalate37 u940 uj38 una160 uj30 u54 ujIsophorone37 u190 u38 una160 uj30 u54 ujCoprostanolna1500 jnana230 jna54 ujN-Nitrodiphenylamine37 u190 u38 una160 u30 u54 uj1,4-Dichlorobenzene37 u190 u38 una160 u30 u54 uj1,242100 u94 u10 u38 una160 u30 u54 uj1,242100 u3						•		
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$\begin{array}{c ccccccccccccccccccccccccccccccccccc$		37 u	190 u	38 u	na	160 u	30 u	54 uj
PCB         1242         100 u         94 u         71 u         na         250 j         63 u         120 j           1254         270         180 j         310         na         610 j         63 u         150 j           1260         1100         550 j         280         na         720 j         150         240 j		37 u	190 u	38 u	na	26 ј	30 u	
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$		37 u	190 u	38 u	na	36 j	30 u	54 uj
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1254270180 j310na610 j63 u150 j12601100550 j280na720 j150240 j		100	Q4	71 n	na	250 +	63 11	120 i
1260 1100 550 j 280 na 720 j 150 240 j								
				1				

u=Not detected at detection limit shown

-=No sample (Rep)=Replicate sample

uj=Estimated detection limit na=Not analyzed

j=Estimated concentration

Table D3: Results of Butyltin\* Analysis of Bottom Sediments from Hylebos Waterway, November 1994 (ug/kg, dry weight).

	Upper Turning	@ General	@ Elf	Near Lincoln		@ Occidental	
	Basin	Metals	Atochem	Drain	Near Cenex	Chemical	Mouth
Station	H-1	H-2	H-3	H-4	H-5	· H-6	H-7
Monobutyltin	100	34 j	120	. 35	47	110	23
Dibutvltin	160 i	210 j	44	380 j	39 uj	26 uj	92 j
Tributvltin	370	280 j	360	820	180	290	150
Tetrabutyltin		4.3 j	2.6 j	30	0.6 j	14 u	2 j

\*= Reported as chloride

u=Not detected at detection limit shown

j-Estimated concentration

## Sitcum Waterway

Table D4: Results of Conventionals and Metals Analysis of Bottom Sediments from Sitcum Waterway, January 1991 and November 1994 (mg/kg, dry weight)

;

Location	Head @	0	Sealand	q	Te	Terminal 7 @		Mouth @	0
	North Corner Drain	er Drain	Terminal	nal		Berth B		Terminal 7	al 7
Station No.	S-1		S-2			S-3		S-4	
Collection Date	1/91	11/94	1/91	11/94	1/91	(Rep)	11/94	1/91	11/94
Depth @ MLLW (ft)	22		44	na	47			47	
Total Solids (%)	57	52	62	na	62	61	46	09	44
TOC (%)	1.5	1.7	1.5	na		2.0	 	1.9	1,1
Grain Size (%)									
Gravel (>2mm)	80	+1	<b>*</b> (	na	-	0	0	0	0
Sand (2mm-62um)	22	27	18	na	39	41	<b>7-1</b>	14	0
Silt (62um-4um)	50	54	61	na	43	42	LL	67	LL
Clay (<4um)	20	18	20	na	18	17	22	19	23
Metals									
Arsenic	25	11	28	na	20	24	12	Ĺ	16
Copper	160	130	120	na	150	160	55	64	59
Lead	290	230	160	na	370	370	19.	73	21
Zinc	340	250	190	IJA	450	470	69	120	73
na=Not analyzed									

(Rep)=Replicate sample

Location	Head	1@			Т	erminal 7 @		Mouth	@
,	North Cor	mer Drain	Sealand Terr	minal		Berth B		Termin	al 7
Station No.	S-	-1	S-2			S-3		S-4	
Collection Date	1/91	. 11/94	1/91		1/91	(Rep)	11/94	1/91	11/94
Depth @ MLLW (ft)	22	****	44	na	47			47	
Naphthalene	54	190 j	48 j	na	64 j	280	190 u	36 j	32 j
Acenaphthene	56	110 j	21 ј	na	250	540	190 u	17 j	200 u
Acenaphthylene	24 j	72 ј	11 ј	na	20 ј	28 ј	15 ј	31 u	200 u
Fluorene	170	150 j	27 ј	na	290	580	190 u	22 ј	200 uj
Anthracene	280	390	66	na	440	620	29 ј	50	200 uj
Phenanthrene	820	860	120	na	1300	3500	73 j	130	74 j
Sum LPAH	1400 j	1800	300 j	na	2400 j	5500 j	120	260 j	500 j
Fluoranthene	1400	2400	290	na	2000	3600	120 j	300	110 j
Benzo(a)anthracene	660	800	150	na	770	1100	190 u	120	200 u
Chrysene	950	1300	300	na	950	1300	100 j	190	120 j
Pyrene	1100	2000	290	na	1300	2300	140 j	280	140 j
Benzofluoranthenes	1400	1500	330	na	1000	1200	330 uj	230	190 j
Benzo(a)pyrene	360	680	130	na	300	350	190 uj	89	63 j
Dibenzo(a,h)anthracene	120	190 <sup>°</sup> uj	82	na	110	78	190 uj	56 j	200 uj
Indeno(1,2,3-cd)pyrene	360	420	150	na	260	250	- 190 uj	93	71 ј
Benzo(g,h,i)perylene	32 ј	360	67 ј	na	47 j	30 j	190 uj	58 j	55 j
Sum HPAH	6400 ј	9700	1800 j	na	6700 j	10000 ј	1500	1400 j	950 j
Total PAH	7800 j	12000	2100 ј	na	9100 j	16000 j	1600	1700 j	1500 j
Phenol	, 34 j	190 u	94 j	na	100 u	71 j	190 u	240	200 u
2-Methylnaphthalene	48	91 j	25 ј	na	87	. 310	17 ј	37	24 ј
Carbazole	na	150 ј	na	na	na	na	190 u	na	200 u
Coprostanol	na	810 j	na	na	na	na	na	na	
Benzoic Acid	160 j	1900 uj	320 u	na	300 u	86 j	1900 uj	81_j	2000 uj
Retene	na	310	na	na	na	na	120 j	na	110 j
Dibenzofuran	88	140 j	28 j	na	230	690	190 uj	28 j	200 uj
Dimethylphthalate	320	290	21 ј	na	33	38	190 u	12	200 u
Butylbenzylphthalate	32 u	950 uj	32 u	na	30 j	30 u	190 uj	30 u	200 uj
Di-n-butylphthalate	32 u	11000 uj	32 u	na	31 u	30 u	3700 uj	30 j	7100 uj
Bis(2EH)phthalate	730	1100 uj	180	na	290	300	200 uj	160	270 uj

Table D5: Summary of Semivolatile Organics Detected in Bottom Sediments from Sitcum Waterway, January 1991 and November 1994 (ug/kg, dry weight).

-=No sample

u=Not detected at detection limit shown

j=Estimated concentration

uj=Estimated detection limit

(Rep)=Replicate sample na=Not analyzed

## Thea Foss Waterway

Location		Hea	1 near Twin		·	No	rth of 15th	
		9	5" Drains		1	Sti	reet Drain	
Station No.			TF-1				TF-2	
Collection Date	12/89	1/91	(Rep)	11/94	(Rep)	12/89	1/91	11/94
Total Solids (%)	37	40	41	38	38	48	44	46
TOC (%)	7.2	8.9	6.7	8.5	8.8	6.3	5.4	6.8
Grain Size (%)								
Gravel (>2mm)	0	0	1	1	0	6	0	1
Sand (2mm-62um)	25	30	43	21	21	53	13	55
Silt (62um-4um)	59	49	37	55	57	31	66	37
Clay (<4um)	16	22	19	23	22	10	21	17
Metals (mg/kg, dry)								
Cadmium	3.1	na	na	3.0	3.1	2.0	na	2.2
Copper	170	na	na	190	190	140	na	130
Mercury	0.71	na	na	1.8	0.6	0.84	na	0.66
Nickel	86	na	na	190	44	44	na	28
Lead	470	na	, na	340	310	500	na	370
Zinc	520	na	na	400	430	400	na	310
Location	Mouth	of Wheeler	•					
	Osgo	od Waterwa	у			Mout	h	
Station No.		TF-3				TF-4	Ļ	
Collection Date	12/89	1/91	11/94		12/89	(Rep)	1/91	11/94
Total Solids (%)	38	42	40		49	75	73	59

Table D6: Results of Conventionals and Metals Analysis of Bottom Sediments from Thea Foss Waterway, December 1989, January 1991 and November 1994 (mg/kg, dry weight).

na=Not analyzed

TOC (%)

Grain Size (%) Gravel (>2mm)

Sand (2mm-62um)

Metals (mg/kg, dry)

Silt (62um-4um)

Clay (<4um)

Cadmium

Copper

Mercury

Nickel

Lead

Zinc

5.7

0

10

64

26

3.6

250

0.92

46

300

360

4.5

0

22

53

25

na

na

na

na

na

na

5.5

0

12

58

30

2.2 j

270

0.8

160

220

300

3.9

0

28

52

20

1.4

140

0.58

42

170

190

1.4

5

86

6

3

0.4

89

34

140

200

0.09

2.3

5

79

11

5

na

na

na <sup>.</sup>

na

na

na

2.5

3

52

35

10

0.4 j

98

0.35

110

67

100

j=Estimated concentration

(Rep)=Replicate sample

Table D7: Summary of Semivolatile Organics Detected in Bottom Sediments from Thea Foss Waterway, December 1989 and November 1994 (ug/kg, dry weight).

Location	Hea	ad near		Nort	h of	15th		Mouth	of		Mouth ne	ar E	)-Street	
	Twin 9	6" Drains		Stre	et D	rain		Wheeler-C	sgood		Tank	Fai	ms	
Station No.	Т	F-1		ុរា	[F-2	2		TF-:	3		T	F-4		
Collection Date	12/89	11/94		12/89		11/94		12/89	11/94		12/89		11/94	
Depth @ MLLW (ft)	14			15			1	18			30			
Acenaphthene	500	250	j	300	j	550		200 j	220		56	j	300	
Acenaphthylene	310	j 260	j	190	j	290		280 j	330		170		340	
Naphthalene	600	590		260	j	960		260 ј	870		160	j	640	
Fluorene	910	440		540		670		270 ј	340		140	j	430	
Anthracene	1900	1400		1100		1800		880	850		560		990	
Phenanthrene	8000	4800		4200		6900		1400	1500		950		2500	
Sum LPAH		j 7700	j	6600	j	11000		3300 j	4100		2000	j	5200	
Fluoranthene	19000	11000		7400		8900		3300	2700		1800		3400	
Benzo(a)anthracene	7200	4000		2900		3800		2000	1200		930		1600	
Chrysene	8600	6700		3600		5000		2600	2000		1100 '		2300	
Pyrene	16000	9700		6800		9500		4500	4100		2200		4200	
Benzofluoranthenes	15000	11000		5800		7300		4600	3600		2000		3300	
Benzo(a)pyrene	5200	4600		2600		38000		2200	1700		1000		2100	
Dibenzo(a,h)anthracene	1900	950		730		560		580	400	uj	360		400	
Indeno(1,2,3-cd)pyrene	3100	4700		1800		2700		980	1500		640		1300	
Benzo(g,h,i)perylene	3300	4000		1600		2400		1200	1400		810		1200	
Sum HPAH	79000	56000		33000		78000		22000	19000		11000		20000	
Total PAH	91000	j 64000	j	40000	j	89000		25000 ј	23000		13000	j	25000	•
Phenol	na	220	j	na		160	-	na	270		na		150	
Pentachlorophenol	na	250	j	na		100	j	na	2100	u	na		1500	u
4-Methylphenol	na		j	na		300		na	160	j	na		120	j
2-Methylnaphthalene	210	300	j	110	j	480		100 j	400		58	j	300	
Carbazole	2200	960		1200		830		240 j	210	-	110	j	270	
Benzoic Acid	na	2200	uj	na		590		na	2100	uj	na		1500	
Benzyl Alcohol	na	170 .	j	na		90	j	na	210	u	na		150	u
Retene	na	480		na		480	· ·	na	460		na		370	
Dibenzofuran	300	300		230	j	370		120 j	240		57	j	250	•
Diethyl phthalate	160	u 280	u	260		210	u	120 j	210	u	110	u,	150	u
Dimethyl phthalate	220	j 110	j	130	j	210	u	130 j	210	u	110	u	150	u
Butyl benzyl phthalate	2100	1400 .	j	11000		1800	j	340	330	j	130	j	740	u
Di-n-Butyl phthalate	450	460	uj	250000		310	uj	150 u	210	u	110	u	150	u
Di-n-Octyl phthalate	1800	1400	uj	540		1000		150 u	1000	uj	110	<b>u</b> .	740	u
Bis(2-ethylhexyl)phthalate	21000	17000	uj	31000		12000	uj	6100	43000	uj	2600		1300	u
1,4-Dichlorobenzene	na	89	j	na		270		na	210	u	na		150	
Coprostanol	na	6700		na		4200	u	na	3000	j	na na		1000	j
u-Not detected at detection	1				~~~~~									******

u=Not detected at detection limit shown

j=Estimated concentration

na= not analyzed

Table D8: Results of Butyltin\* Analysis of Bottom Sediments from Thea Foss Waterway, December 1989, January 1991, and November 1994 (ug/kg, dry weight).

Location		Head near Twin	r Twin		No:	North of 15th	
		96" Drains	tains		Sti	Street Drain	
Station		TF-1	+(			TF-2	
Collection Date	12/89	1/91	(Rep)	11/94	12/89	1/91	11/94
Monobutyltin	16 u	21	15	66	14 u	25	82
Dibutyltin	160	33	25	160 j	94	46	120 j
Tributyltin	160	65	59	320	110	06	210
Tetrabutyltin	19 u	5.9 j	5.5 u	12 u	17 u	5.3 u	12 u
Location	Mouth	Mouth of Wheeler-	-Tr		Mouth near D-Street	D-Street	
	Osgoo	Osgood Waterway	ay		Tank Farms	rms	
Station		TF-3.	,		TF-4		
Collection Date	12/89	<u>1/91</u>	11/94	12/89	(Rep)	1/91	11/94
Monobutyltin	400	65	78 j	13 u	0°6	9.3	20
Dibutyltin	220	4	260 j	82	12	I.4 u	100 j
Tributyltin	410	110	260 j	120	15	10	140
Tetrabutyltin	20 u	5.4 u	23 uj	16 u	11 u	3.5 u	17 u
*= Reported as chloride		****					
u=Not detected at detection limit shown	ion limit shov	UN					
j-Estimated concentration	u						
(Rep)=Replicate sample							
mi-Estimated detection limit	imit						

uj=Estimated detection limit